

Prevalence of human leukocyte antigen HLA-B*57:01 in individuals with HIV in West and Central Africa

Malewe Kolou, Armel Poda, Zelica Diallo, Esther Konou, Tatiana Dokpomiwa, Jacques Zoungrana, Mounerou Salou, Lionèle Mba-Tchounga, André Bigot, Abdoul-Salam Ouedraogo, et al.

► To cite this version:

Malewe Kolou, Armel Poda, Zelica Diallo, Esther Konou, Tatiana Dokpomiwa, et al.. Prevalence of human leukocyte antigen HLA-B*57:01 in individuals with HIV in West and Central Africa. *BMC Immunology, BioMed Central*, 2021, 22 (1), pp.48. 10.1186/s12865-021-00427-7 . inserm-03310669

HAL Id: inserm-03310669

<https://www.hal.inserm.fr/inserm-03310669>

Submitted on 30 Jul 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH

Open Access



Prevalence of human leukocyte antigen HLA-B*57:01 in individuals with HIV in West and Central Africa

Malewe Kolou¹, Armel Poda², Zelica Diallo³, Esther Konou¹, Tatiana Dokpomiwa⁴, Jacques Zoungrana², Mounerou Salou¹, Lionèle Mba-Tchounga⁵, André Bigot⁴, Abdoul-Salam Ouedraogo⁶, Marielle Bouyout-Akoutet⁷, Didier K. Ekouevi^{8,9*} and Serge P. Eholie^{3†}

Abstract

Background: The presence of the human leukocyte antigen HLA-B*57:01 is associated with the development of a hypersensitivity reaction to abacavir (ABC). Limited data exist on HLA-B*57:01 prevalence in individuals with HIV-1 in Africa. This study aimed to estimate HLA-B*57:01 prevalence in individuals with HIV-1 in West and Central Africa.

Methods: A cross-sectional study was conducted in four countries in West and central Africa (Burkina-Faso, Côte d'Ivoire, Gabon, and Togo) from January 2016 to February 2020 to determine the status of HLA-B*57:01 in adults with HIV-1. The presence of HLA-B*57:01 was determined by using Single Specific Primer-Polymerase Chain Reaction (SSP-PCR) in blood samples. Prevalence rates were stratified based on country.

Results: A total of 4016 (69.8% women) individuals with HIV were enrolled. Their median age was 45, and the interquartile range was 38–52. We included 500 (12.4%) patients in Burkina-Faso, 1453 (36.2%) in Côte d'Ivoire, 951 (23.7%) in Gabon, and 1112 (27.7%) in Togo. The overall HLA-B*57:01 prevalence was 0.1% [95% CI: 0.0–0.2%]. The prevalence of HLA-B*57:01 was similar according to the four countries. Only one case was reported in each country except Togo, with no cases.

Conclusions: HLA-B*57:01 prevalence is low in individuals with HIV in West and central Africa, and there is no difference among countries. This study does not confirm the utility of HLA-B*57:01 allele testing for abacavir use in this region.

Keywords: Abacavir, Antiretroviral, HLA-B*57:01, HIV infection, Prevalence, West, And Central Africa

* Correspondence: didier.ekouevi@gmail.com

†Didier K. Ekouevi and Serge P. Eholie are equal authorship

⁸Département de santé Publique, Université de Lomé, Faculté des Sciences de la santé, Lomé, Togo

⁹Centre Inserm 1219 & Institut de Santé Publique d'épidémiologie et de développement, Université de Bordeaux, Bordeaux, France

Full list of author information is available at the end of the article



© The Author(s). 2021 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Background

According to UNAIDS, new HIV infections have been reduced by 23% since 2010, largely due to a substantial 38% decrease in Eastern and Southern Africa [1]. Despite this reduction in new HIV infections, there were 690,000 AIDS-related deaths in 2019 [1]. The rollout of combination antiretroviral therapy (ART) has significantly changed the natural history of HIV infection [2]. Advances in treatment strategies have led to a spectacular drop in the mortality rate of people living with HIV (PLHIV) worldwide [3]. However, of the 38 million people living with HIV worldwide, only 25.4 million are on treatment [1]. Abacavir, a nucleoside HIV reverse transcriptase inhibitor, is one of the drugs used in the management of HIV infection. It is a key component of the first-line antiretroviral (ART) regimens recommended by the US Department of Health and Human Services (DHHS) [4]. A randomized, double-blind study from 2015 called for positioning the dolutegravir/abacavir/lamivudine regimen as a first-line option for treatment-naïve HIV-positive patients [5]. Indeed, of the 833 randomized participants, 71% in the dolutegravir/abacavir/lamivudine group and 63% in the efavirenz/tenofovir/emtricitabine group maintained viral loads of < 50 copies per milliliter up to 144 weeks after initiation of treatment ($P = 0.01$) [5]. The higher efficacy was mainly due to fewer dropouts due to adverse events in the dolutegravir/abacavir/lamivudine group [dolutegravir/abacavir/lamivudine group, 13 (3%); efavirenz/tenofovir/emtricitabine group, 48 (11%)] [5]. Several studies have provided clear benefits and recommended abacavir-based treatment regimens [6–8]. According to the recent World Health Organization (WHO) guidelines in low-income countries, especially in West and Central Africa, abacavir is recommended as an alternative to first-line regimens in children and, in special circumstances in adults [9].

The main limitation to the use of abacavir is a frequent, early and potentially fatal Type IV hypersensitivity reaction [10]. Data from several clinical trials suggest that an abacavir-induced hypersensitivity reaction occurs in 2.3–9% of patients exposed to abacavir [11–13]. In a cohort of 14,310 individuals with HIV-1 examined in 93 centers across Europe, Israel and Argentina, 2101 (64.1%) were forced to abandon abacavir treatment [14]. For 167 (5.1%) patients, the reason for discontinuing treatment was the occurrence of a hypersensitivity reaction, and the overall incidence of hypersensitivity reactions in the study was estimated at 22.1 (95% confidence interval [CI] 18.7–25.4) per 100 person-years [14].

Human leukocyte antigen (HLA)-B*57:01, an allele of the major histocompatibility complex, has been strongly associated with the risk of an abacavir hypersensitivity reaction [15]. A prospective, randomized, multicenter, double blind, prospective study has shown that screening

for HLA-B*57:01 can reduce the incidence of hypersensitivity reactions to abacavir [16]. Several studies have been conducted to estimate the prevalence of HLA-B*57:01 in many populations around the world. A meta-analysis by the Clinical Pharmacogenetics Implementation Consortium (CPIC) of data published between 1950 and 2011 allowed the mapping of the distribution of HLA-B*57:01 in different ethnic populations [17]. The combined analyses covered 35,630 Europeans, 1321 South Americans, 8570 Africans, 1029 Middle Easterners, 3391 Mexicans and 12,501 Asians. Europeans recorded the highest frequency of HLA-B*57:01, with proportions up to 14.1% [17]. Among South Americans and Mexicans, 2.6 and 2.2%, respectively, were carriers of HLA-B*57:01 [17]. This proportion was 2.5% for Middle East residents and 12.6% for Asian residents [17]. African populations recorded the lowest prevalence with an estimated overall frequency of 1.0% [17].

The majority of HLA-B*57:01 prevalence surveys among people from Africa are conducted among African Americans [18–21]. However, some studies have been carried out in sub-Saharan Africa, notably in Nigeria, Kenya, South Africa and Uganda [22–26], but data on the distribution of HLA-B*57:01 in the population of Sub-Saharan Africa remain scarce.

In Burkina Faso, according to a study carried out in 2014 among 51 HIV sero-discordant couples, 78% of individuals with HIV were carriers of the HLA-B*57:01 gene [27]. This high frequency, in contrast to what is generally reported in African populations, has led the authors of this Burkinabe study to recommend large cohort studies to confirm their results [27]. In the Ivory Coast, Gabon and Togo, to our knowledge, there are no data on the prevalence of the HLA-B*57:01 gene. The objective of this study was to estimate the prevalence of the HLA-B*57:01 gene in individuals with HIV in four West and Central African countries (Burkina Faso, Ivory Coast, Gabon and Togo) and to measure the association between HLA-B*57:01 gene carrying and ethnic subgroups in West and Central Africa.

Methods

A multicounty, multicentric cross-sectional study was conducted from January 2016 to February 2020 in four countries in Central and West Africa: Togo, Burkina Faso, Ivory Coast and Gabon (Fig. 1).

Study population

Participants were recruited from clinical centers chosen based on existing partnerships and centers' involvement in clinical research projects on HIV. Those clinical centers were in four countries. In Burkina Faso, the clinical center was the Day Hospital, University Hospital Sourou Sanon (Bobo-Dioulasso); The Support, Research and Training



Fig. 1 Participating countries located in West and Central Africa. Figure 1 shows location of participating countries across West and Central Africa. These countries appear with red mark on the map. The figure was created by us, using Openstreetmap and Adobe Photoshop CC 2019 software version 20.0.4 by Adobe Systems Incorporated© (<https://www.adobe.com/fr/products/photoshop.html#>)

Center (CePReF) (Abidjan); the Infectious and Tropical Diseases Service (SMIT); Treichville University Hospital (Abidjan); and the medical center for monitoring blood donors (CMSDS), CNTS, Treichville (Abidjan), which was included in Côte d'Ivoire. Participants from Gabon were enrolled at the Méléne Hospital in Libreville, the Parasitology Mycology Laboratory at the Faculty of Medicine of the University of Health Sciences and at the outpatient Treatment Center (CTA) of Oyem and Koulamoutou. Finally, in Togo, two centers were involved: the day hospital of the infectious diseases department of the Sylvanus Olympio University Hospital and the Laboratory of Molecular Biology and Immunology of the Faculty of Health Sciences of the University of Lomé (BIOLIM-FSS/UL).

Participants were eligible to participate in the study if the following four criteria were met: (i) infected with HIV, regardless of subtype (HIV-1, HIV-2, HIV-1 + 2); (ii) being 18 years of age or older; (iii) being from West or Central Africa; (iv) having consulted in one of the study centers and residing in the study city; and (v) having given free, informed and written consent, signed by the participant and the investigating physician on the day of inclusion and before any sampling required by the study.

Sample size estimation

The sample size of participants was calculated using a single proportion population formula with a 95% confidence

level. We hypothesized that the prevalence of HLA-B*57:01 allele among black Africans is less than 1% with a 1% margin error. Considering a 10% nonresponse rate, the minimum number of participants per country was estimated at 418 and a minimum sample size was estimated at 1672 for the four countries participating to the study.

Data collection

After eligibility screening and written informed consent, sociodemographic characteristics and HIV epidemiological data were collected using a standardized questionnaire. The questionnaire was administered by a trained study team during a face-to-face interview. After the interview, information on biological variables and ART treatment was obtained from another data source (biological database and pharmacy). Sociodemographic and clinical characteristics (age, sex, nationality, ethnic group, WHO clinical stage, history of antiretroviral therapy) were collected. Biological variables collected were absolute CD4 count, HIV type, HLA type and presence of HLA-B*57:01. From each patient, whole blood samples were collected at the elbow fold in two EDTA tubes and were used for the detection of the HLA-B*57:01 allele.

Laboratory procedures

Sample preparation and nucleic acid extraction

Blood samples were centrifuged at 3500 tours/min. Buffy coats were collected, aliquoted and stored at -80°C .

DNA was extracted from buffy coat samples using the QIAamp® DNA Mini kit according to the manufacturer's instructions. The DNA eluate was stored at -80°C and sent to *Laboratoire d'Histocompatibilité des Cliniques Universitaire Saint Luc* in Belgium, where HLA-B*57:01 typing was performed.

HLA-B*57:01 typing

A qPCR-SSP DNA generic typing test (LinkSêq™ HLA-B*57:01-ONE LAMBDA) was used. Specifically, generic real-time PCR was performed in 96-well plates. The test separately detected HLA-B*57:01 and closely related HLA-B57 alleles (HLA-B*57:02/HLA-B*57:03) for each sample. As recommended by the manufactured protocol, DNA samples were added immediately after isolation to each well of the tray to the Taq polymerase reconstituted with a dNTP-buffer mix (Micro SSP D-mix). Each typing tray included a negative control reaction tube that detects the presence of the internal control PCR product. PCR amplification was carried out according to standard procedures (One Lambda, Inc.) in LightCycler® 480 Real-Time PCR System-Roche. SureTyper software was then used to analyze and interpret the melt curves generated from each well, and the results were thus obtained.

Statistical analysis

Data describing clinical and demographic patient characteristics were summarized using medians with interquartile ranges (IQRs) for continuous variables and frequencies and proportions for categorical variables. Prevalence was estimated with their 95% confidence interval (95% CI). Fisher's exact tests was used to compare categorical variables. All analyses were performed using R© version 3.4.3 software, and the level of significance was set at 5%.

Ethical considerations

Ethical approval was obtained from the ethics committees in Burkina Faso, Ivory Coast, Gabon and Togo before the start of the inclusion. Potential participants were informed about the study purpose and procedures, potential risks and protections. Those willing to participate were invited to sign a consent prior to participation.

Results

Population characteristics

A total of 4016 HIV-positive patients from four countries were included in this study. Table 1 presents the number of patients enrolled in each country in West and Central Africa.

The median age of the participants was 45 years (IQR = [38–52]), and more than two-thirds (69.8%) were female. Most participants were 40–49 years old (38.0%),

Table 1 Number of participant per countries

	Frequency	Proportion (%)
Côte d'Ivoire	1453	36.2
Gabon	951	23.7
Togo	1112	27.7
Burkina Faso	500	12.4
Total	4016	100.0

and 46.9% had at least a secondary school level of education. The majority of patients were on ART ($n = 3510$; 87.4%), and among them, 117 (3.3%) were on ART containing abacavir.

More than two-thirds (62.4%) had been diagnosed with HIV for more than 5 years (at the time of the survey). A higher proportion of them (90.9%) had HIV-1 status, and approximately one-quarter (26.3%) were at WHO stage I. Other demographic, clinical, and laboratory characteristics of all participants by country are presented in Table 2.

Prevalence of HLA-B57 and HLA- B*57:01 per country

Of the 4016 patients who underwent HLA typing, 259 were positive for HLA-B57, allowing the overall prevalence of the HLA-B57 allele to be estimated at 6.4% (95% CI: 5.7–7.3). There was no significant difference in the prevalence of the HLA-B057 allele according to country ($p = 0.238$), sex ($p = 0.078$), or age ($p = 0.276$) (Table 3).

Prevalence HLA- B*57:01 per country

Among 259 patients with the HLA-B57 allele, three were positive for the HLA-B*57:01 allele. The overall prevalence in the population was 0.1% (95% CI: 0.0–0.2). One case was reported in Côte d'Ivoire, Gabon and Burkina Faso, and no case was reported in Togo (Table 3).

Discussion

To our knowledge, this study is the first large multi-country study conducted in four countries in the West ($n = 3$) and Central ($n = 1$) African regions of the prevalence of the HLA-B*57:01 allele among individuals with HIV. According to the latest epidemiological data, West and Central Africa is home to 4.9 million people living with HIV. HIV prevalence among adults is 1.4%, which is relatively low compared to East and Southern Africa. In this region, the number of people accessing treatment rose significantly from 860,000 in 2010 to 2.9 million in 2019. HLA-B*57:01 is a generic marker of clinical importance used in several countries, specifically in developed countries, to decrease abacavir-related hypersensitivity reactions. Guidelines in North America and Europe recommend routine screening for HLA-B*57:01 prior to initiation of abacavir therapy [28]. Ideally, since

Table 2 Demographic, clinical and laboratory characteristics of individuals with HIV in West and Central Africa (2016–2020)

	Côte d'Ivoire (n = 1453)	Gabon (n = 951)	Togo (n = 1112)	Burkina Faso (n = 500)	Total (n = 4016)
Sex					
Male	483 (33.2)	303 (31.9)	329 (29.6)	96 (19.2)	1211 (30.2)
Female	970 (66.8)	648 (68.1)	783 (70.4)	404 (80.8)	2805 (69.8)
Age (years), Median [IQR]	46 [40–53]	44 [36–52]	44 [37–51]	44 [38–51]	45 [38–52]
Age (years)					
< 20	3 (0.2)	21 (2.2)	12 (1.1)	1 (0.2)	37 (0.9)
20–29	55 (3.8)	88 (9.3)	76 (6.8)	19 (3.8)	238 (5.9)
30–39	283 (19.5)	238 (25.0)	267 (24.0)	142 (28.4)	930 (23.2)
40–49	584 (40.2)	322 (33.9)	426 (38.3)	196 (39.2)	1528 (38.0)
50–59	399 (27.5)	198 (20.8)	256 (23.0)	118 (23.6)	971 (24.2)
≥ 60	129 (8.9)	84 (8.8)	75 (6.7)	24 (4.8)	312 (7.8)
Education level					
None	499 (34.3)	186 (19.6)	199 (17.9)	202 (40.4)	1086 (27.0)
Primary	301 (20.7)	148 (15.6)	390 (35.1)	209 (41.8)	1048 (26.1)
Secondary	435 (29.9)	448 (47.1)	454 (40.8)	84 (16.8)	1421 (35.4)
University	218 (15.0)	169 (17.8)	69 (6.2)	5 (1.0)	461 (11.5)
Time since HIV diagnosis (years)					
< 1	78 (5.4)	135 (14.2)	126 (11.3)	20 (4.0)	359 (8.9)
[1–3]	132 (9.1)	170 (17.9)	160 (14.4)	43 (8.6)	505 (12.6)
[3–5]	155 (10.7)	147 (15.5)	144 (12.9)	52 (10.4)	498 (12.4)
≥ 5	1065 (73.3)	389 (40.9)	666 (59.9)	384 (76.8)	2504 (62.4)
Missing	23 (1.6)	110 (11.6)	16 (1.4)	1 (0.2)	150 (3.7)
CD4 cell count, Median [IQR]	230 [127–367]	237 [138–440]	169 [84–297]	227 [110–333]	210 [106–340]
Baseline CD4 cell count (cells/mm3)					
< 200	563 (38.7)	78 (8.2)	549 (49.4)	201 (40.2)	1391 (34.6)
[200–350]	407 (28.0)	56 (5.9)	243 (21.9)	148 (29.6)	854 (21.3)
[350–500]	177 (12.2)	25 (2.6)	111 (10.0)	58 (11.6)	371 (9.2)
≥ 500	188 (12.9)	40 (4.2)	52 (4.7)	47 (9.4)	327 (8.1)
Missing	118 (8.1)	752 (79.1)	157 (14.1)	46 (9.2)	1073 (26.7)
HIV status					
HIV-1	1370 (94.3)	734 (77.2)	1078 (96.9)	469 (93.8)	3651 (90.9)
HIV-2	37 (2.5)	4 (0.4)	7 (0.6)	17 (3.4)	65 (1.6)
HIV-1 + 2	34 (2.3)	54 (5.7)	1 (0.1)	14 (2.8)	103 (2.6)
Missing	12 (0.8)	159 (16.7)	26 (2.3)	0 (0.0)	197 (4.9)
WHO stage					
Stage I	586 (40.3)	15 (1.6)	323 (29.0)	134 (26.8)	1058 (26.3)
Stage II	194 (13.4)	24 (2.5)	298 (26.8)	106 (21.2)	622 (15.5)
Stage III	236 (16.2)	31 (3.3)	180 (16.2)	179 (35.8)	626 (15.6)
Stage IV	30 (2.1)	1 (0.1)	87 (7.8)	25 (5.0)	143 (3.6)
Missing	407 (28.0)	880 (92.5)	224 (20.1)	56 (11.2)	1567 (39.0)
Abacavir based regimen^a					
No	1277 (98.6)	565 (87.9)	1053 (98.2)	498 (99.6)	3393 (96.7)
Yes	18 (1.4)	78 (12.1)	19 (1.8)	2 (0.4)	117 (3.3)

^aAmong 3510 on ARV treatment

Table 3 Prevalence of HLA-B57 and HLA-B*57:01 per country in West and Central Africa (2016–2020)

	N	HLA-B57 ⁺			HLA-B*57:01 ⁺⁺		
		n	%	95 CI%	n	%	95CI%
Côte d'Ivoire	1453	107	7.4	[6.1–8.8]	1	0.1	[0.0–0.4]
Gabon	951	51	5.4	[4.0–7.0]	1	0.1	[0.0–0.6]
Togo	1112	72	6.5	[5.1–8.1]	0	0.0	[0.0–0.3]
Burkina Faso	500	29	5.8	[3.9–8.2]	1	0.2	[0.0–0.1]
Total	4016	259	6.4	[5.7–7.3]	3	0.1	[0.0–0.2]

⁺*p*-value = 0.238; ⁺⁺*p*-value = 0.571
95 CI% 95% Confidence Interval

HLA-B*57:01 varies among different populations, it is ascertaining HLA-B*57:01 prevalence before decoding or not performing systematic screening before initiating abacavir-based ART regimens. In our study, we screened HIV-positive patients on ART or not for HLA-B*57:01 carriage.

We estimated an overall HLA-B*57:01 prevalence rate of 0.07%, ranging from 0% in Togo to 0.2% in Burkina-Faso. A similar survey was conducted in Nigeria, located in the West African region between April 2016 and April 2017 in five HIV treatment facilities. In this study, 1504 adults were enrolled. Of these, 132 (9.1%) were HLA-B57 positive using a nonspecific low-resolution HLA-B*57:01 primer mix. On further analysis, none of the 132 samples (0%) had the HLA-B*57:01 allele [22]. Another survey was conducted in a different population and reported a high prevalence of HLA-B*57:01. This is the case in Colombia, with a prevalence rate of 2.7% in Colombian HIV-infected individuals. The prevalence was 4% for whites, 2.6% for other races, and 1.9% for Afro-Colombians [29]. Other studies have confirmed a low prevalence of HLA-B*57:01 in the African population. In the United States, there is a lower frequency of the HLA-B*57:01 allele in African Americans, with a reported frequency between 2.3 and 4% [20, 30, 31]. Among White subjects, prevalence was 7.93%. Among black subjects, only two (both Ugandan) were HLA-B*57:01 positive, giving a rate of 0.26% [32].

Our study had a few limitations. We did not conduct an analysis based on race in our survey. Other races are indeed rare in individuals with HIV in West and central Africa, and this variable is not collected routinely in databases. Further studies should explore including race (i.e., mestizo), as in our sample, one person in Gabon with the mestizo race was screened positive. Some studies have reported that the heterogeneity of HLA-B*57:01 prevalence is mostly dependent upon race and ethnicity heritage [16, 33]. Another limitation is that we did not perform logistic regression with the low prevalence observed to identify factors associated with the presence of HLA-B*57:01. Another limitation to the study is that

HLA typing were performed on patients mostly already on ART. This design could induce some bias towards excluding patients already having had hypersensitivity reaction to abacavir. Finally, HLA-B*57:01-negative patients starting abacavir-containing regimens (3%) were not followed up; therefore, the incidence of HSR was not assessed.

Since abacavir is included in the first-line regimen, the question of hypersensitivity to abacavir should be important to document. However, to our knowledge, there are no guidelines for screening patients before ART initiation in Africa. Before abacavir should be wisely used in Africa, it is necessary to explore the prevalence of HLA-B*57:01. The application of routine HLA-B*57:01 testing to other racial populations deserves further investigation. In these four countries, antiretroviral therapy is available and free of charge for patients. Currently, the first-line regimen includes 2 NRTIs and dolutegravir in accordance with the WHO guidelines or 2 NRTIs and efavirenz. An optimized NRTI backbone should be used, such as zidovudine (AZT), tenofovir or abacavir (ABC), and vice versa.

In addition, HLA-B*57:01 screening was not performed routinely because of the lack of equipment and reference laboratories specializing in immunologic tests in the four countries. This is the main reason all-immunological analyses were performed in Belgium, which is specialized in this analysis to avoid heterogeneity between the tests.

Recently, the value of HLA-B*57:01 screening prior to prescribing abacavir is a concern in areas where its prevalence is low [34, 35]. In the context of low-income countries where pharmacogenetic screening is not available and based on this result with a low prevalence of HLA-B*57:01, we could not recommend the testing of HLA-B*57:01. Despite the low prevalence of HLA-B*57:01, clinical follow-up of patients starting an abacavir-based regimen is mandatory. It is therefore important to educate the patient about the possible symptoms of a hypersensitivity reaction, especially at the beginning of treatment, so that the patient can easily identify them and return for further consultation. Any suspicion of hypersensitivity to abacavir should lead to discontinuation of treatment and the initiation of rigorous clinical follow-up [36, 37]. Screening for HLA-B*57:01 prior to initiation of any therapy with abacavir has been shown to be beneficial [38, 39].

Conclusion

HLA-B*57:01 is rare in people living with HIV, in the four countries participating to this survey, and there are no significant differences among countries. HLA-B*57:01 screening may not be a cost-effective strategy to reduce the risk associated with abacavir hypersensitivity in these

countries. Indeed, the prevalence of HLA-B*57:01 allele positivity was low in the study population. These results cannot be generalized to the West and Central African region as prevalence may be much higher in other countries and among other ethnicities in the same regions of Africa.

However, despite the low prevalence of the HLA-B*57:01 allele, rigorous monitoring should be maintained in any patient on ABC since the possibility of occurrence of HSR to this drug should not be definitively ruled out.

Acknowledgments

We are thankful to the individuals who agreed to participate in this study and the staff working on the project who performed data collection for the study. We also acknowledge the technical support of the work of the teams of the *Laboratoire d'Histocompatibilité des Cliniques Universitaire Saint Luc* in Belgium involved in this study. Finally, we thank Alexandra Bitty-Anderson, Abidjan, Côte d'Ivoire, who edited the English version of this paper.

Authors' contributions

DKE and SPE conceived the study protocol. DKE, SPE, MK and MBA drafted the analysis plan and wrote the first draft of the manuscript. DKE performed the statistical analysis. ZD, AP, JZ and LMT collected and supervised the data collection. EK, MK, MS, ASO, AB and TD performed biological analysis. All authors contributed to data analysis through review and interpretation of the results. All authors read, revised and approved the final manuscript.

Funding

This project was funded by Viiv Healthcare.

Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate

This study was approved by: i) the Bioethics Committee for Health Research from the Togo Ministry of Health (CBRS N°005/2016/MSPS/CAB/SG/DPML/CBRS); ii) the Health Research Ethics Committee from the Burkina Faso ministry of higher education, scientific research and innovation (N°2017–9–150); iii) the National Research Ethics Committee from Ministry of Health and Public Hygiene of Ivory Coast (115/MSPH/CNER-km); iv) the national ethics committee of Gabon (N°0044/2016/SG/P). All methods were carried out in accordance with relevant guidelines and regulations in the Ethical Approval and Consent to Participate. Potential participants were told about the study purpose and procedures, potential risks and protections, and compensation. Written informed consent was obtained prior to participation.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Laboratoire de Biologie Moléculaire et d'Immunologie (BIOLIM), Université de Lomé, Faculté des Sciences de la santé, Lomé, Togo. ²Department of Infectious Diseases, Université Nazi Boni, Bobo-Dioulasso, Burkina Faso. ³Département de Dermatologie et Maladies Infectieuses, Université Félix Houphouët-Boigny, UFR des Sciences Médicales, Abidjan, Côte d'Ivoire. ⁴Department of Pharmacy, Faculty of Health Science, University of Abomey-Calavi, Cotonou, Benin. ⁵Programme PACCI, Site de recherche ANRS de Côte d'Ivoire, Abidjan, Côte d'Ivoire. ⁶Department of Medical Bacteriology and Virology, Université Nazi BONI, CHU Sourou Sanou, Bobo-Dioulasso, Burkina Faso. ⁷Department of Parasitology, Mycology and Tropical Medicine, Université des Sciences de la Santé, Libreville, Gabon. ⁸Département de santé Publique, Université de Lomé, Faculté des Sciences de la santé, Lomé, Togo. ⁹Centre Inserm 1219 & Institut de Santé Publique d'épidémiologie et de développement, Université de Bordeaux, Bordeaux, France.

Received: 24 January 2021 Accepted: 11 May 2021

Published online: 22 July 2021

References

1. Joint United Nations Programme on HIV/AIDS (UNAIDS). Global Aids Update 2020 – Seizing the moment: tackling entrenched inequalities to end epidemics. Geneva: UNAIDS; 2020. p. 40.
2. Parashar S, Collins AB, Montaner JS, Hogg RS, Milloy MJ. Reducing rates of preventable HIV/AIDS-associated mortality among people living with HIV who inject drugs. *Curr Opin HIV AIDS*. 2016;11(5):507–13. <https://doi.org/10.1097/COH.0000000000000297>.
3. Joint United Nations Programme on HIV/AIDS (UNAIDS). The gap report. Geneva: UNAIDS; 2014. p. 422. Available from: <https://tinyurl.com/y5mr8ahf>
4. Lee GQ, McCluskey S, Boum Y 2nd, Hunt PW, Martin JN, Bangsberg DR, et al. Brief report: should Abacavir be a first-line alternative for adults with HIV in sub-Saharan Africa? *J Acquir Immune Defic Syndr*. 2017;76(2):188–92. <https://doi.org/10.1097/QAI.0000000000001487>.
5. Walmsley S, Baumgarten A, Berenguer J, Felizarta F, Florence E, Khuong-Josse MA, et al. Brief Report: Dolutegravir plus Abacavir/Lamivudine for the treatment of HIV-1 infection in antiretroviral therapy-naïve patients: week 96 and week 144 results from the single randomized clinical trial. *J Acquir Immune Defic Syndr*. 2015; 70(5):515–519. doi: <https://doi.org/10.1097/QAI.0000000000000790>. Erratum in: *J Acquir Immune Defic Syndr*. 2016; 71(1):e33.
6. Cruciani M, Malena M. Combination dolutegravir-abacavir-lamivudine in the management of HIV/AIDS: clinical utility and patient considerations. *Patient Prefer Adherence*. 2015;9:299–310. <https://doi.org/10.2147/PPA.S65199>.
7. Fernandez JV, Munir A. Abacavir. In: StatPearls; 2020. [Cited November 14, 2020]. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK537117/>.
8. Bossacoma Busquets F, Noguera-Julian A, Sanchez E, Fortuny C. Dolutegravir plus abacavir/lamivudine works in adolescents, but size matters. *J Antimicrob Chemother*. 2017;72(10):2958–60. <https://doi.org/10.1093/jac/dkx235>.
9. World Health Organization (WHO). Update of recommendations on first and second line antiretroviral regimens. Geneva: WHO; 2019. p. 16.
10. Haute Autorité de Santé (HAS). Détection de l'allèle HLA-B*57:01 préalable au traitement par abacavir. Saint-Denis La Plaine: HAS; 2009. p. 42.
11. Clay PG. The abacavir hypersensitivity reaction: a review. *Clin Ther*. 2002; 24(10):1502–14. [https://doi.org/10.1016/S0149-2918\(02\)80057-1](https://doi.org/10.1016/S0149-2918(02)80057-1).
12. Hetherington S, McGuirk S, Powell G, Cutrell A, Naderer O, Spreen B, et al. Hypersensitivity reactions during therapy with the nucleoside reverse transcriptase inhibitor abacavir. *Clin Ther*. 2001;23(10):1603–14. [https://doi.org/10.1016/S0149-2918\(01\)80132-6](https://doi.org/10.1016/S0149-2918(01)80132-6).
13. Rodriguez-Novoa S, Soriano V. Current trends in screening across ethnicities for hypersensitivity to abacavir. *Pharmacogenomics*. 2008;9(10):1531–41. <https://doi.org/10.2217/14622416.9.10.1531>.
14. Bannister WP, Friis-Møller N, Mocroft A, Viard JP, van Lunzen J, Kirk O, et al. Incidence of abacavir hypersensitivity reactions in euroSIDA. *Antivir Ther*. 2008;13(5):687–96.
15. Zhang H, Zhang T, Zhao H, Han N, Zhou H, He Y, et al. Low prevalence of human leukocyte antigen-B*5701 in HIV-1-infected Chinese subjects: a prospective epidemiological investigation. *AIDS Res Ther*. 2015;12(1):28. <https://doi.org/10.1186/s12981-015-0064-9>.
16. Mallal S, Phillips E, Carosi G, Molina JM, Workman C, Tomazic J, et al. HLA-B*5701 screening for hypersensitivity to abacavir. *N Engl J Med*. 2008;358(6): 568–79. <https://doi.org/10.1056/NEJMoa0706135>.
17. Martin MA, Klein TE, Dong BJ, Pirmohamed M, Haas DW, Kroetz DL; Clinical pharmacogenetics implementation consortium. clinical pharmacogenetics implementation consortium guidelines for HLA-B genotype and abacavir dosing: Supplementary material. 2012 [Cited November 12, 2020]. Available from: <https://tinyurl.com/y42rso8r>.
18. Waters LJ, Mandalia S, Gazzard B, Nelson M. Prospective HLA-B*5701 screening and abacavir hypersensitivity: a single centre experience. *AIDS*. 2007;21(18):2533–4. <https://doi.org/10.1097/QAD.0b013e328273bc07>.
19. Watson ME, Patel LG, Ha B, Wannamaker P, Cuffe R, Shaefer M. A study of HIV provider attitudes toward HLA-B 5701 testing in the United States. *AIDS Patient Care STDs*. 2009;23(11):957–63. <https://doi.org/10.1089/apc.2008.0245>.
20. Cao K, Hollenbach J, Shi X, Shi W, Chopek M, Fernández-Viña MA. Analysis of the frequencies of HLA-A, B, and C alleles and haplotypes in the five major ethnic groups of the United States reveals high levels of diversity in these loci and

- contrasting distribution patterns in these populations. *Hum Immunol.* 2001;62(9):1009–30. [https://doi.org/10.1016/S0198-8859\(01\)00298-1](https://doi.org/10.1016/S0198-8859(01)00298-1).
21. Gao X, Single RM, Karacki P, Marti D, O'Brien SJ, Carrington M. Diversity of MICA and linkage disequilibrium with HLA-B in two north American populations. *Hum Immunol.* 2006;67(3):152–8. <https://doi.org/10.1016/j.humimm.2006.02.009>.
 22. Agbaji OO, Akanbi MO, Otoh I, Agaba PA, Akinsola R, Okolie V, et al. Absence of human leukocyte antigen-B*57:01 amongst patients on antiretroviral therapy in Nigeria: implications for use of abacavir. *Niger Postgrad Med J.* 2019;26(4):195–8. https://doi.org/10.4103/npmj.npmj_75_19.
 23. Puthanakit T, Bunupuradah T, Kosalaraksa P, Vibol U, Hansudewechakul R, Ubolyam S, et al. Prevalence of human leukocyte antigen-B*5701 among HIV-infected children in Thailand and Cambodia: implications for abacavir use. *Pediatr Infect Dis J.* 2013;32(3):252–3. <https://doi.org/10.1097/INF.0b013e3182745dba>.
 24. Shah R, Nabiswa H, Okinda N, Revathi G, Hawken M, Nelson M. Prevalence of HLA-B*5701 in a Kenyan population with HIV infection. *J Inf Secur.* 2018;76(2):212–4.
 25. Masebe T, Bessong PO, Ndip RN, Meyer D. Genetic variants of APOC3 promoter and HLA-B genes in an HIV infected cohort in northern South Africa: a pilot study. *Int J Mol Sci.* 2014;15(7):11403–15. <https://doi.org/10.3390/ijms150711403>.
 26. Munderi P, Snowden WB, Walker AS, Kityo C, Mosteller M, Kabuyeet G, et al. Distribution of HLA-B alleles in a Ugandan HIV-infected adult population: NORA pharmacogenetic substudy of DART. *Trop Med Int Health.* 2011;16(2):200–4. <https://doi.org/10.1111/j.1365-3156.2010.02688.x>.
 27. Kagoné TS, Bisseye C, Méda N, Testa J, Pietra V, Kania D, et al. A variant of DC-SIGN gene promoter associated with resistance to HIV-1 in serodiscordant couples in Burkina Faso. *Asian Pac J Trop Med.* 2014;7S1:S93–6.
 28. European AIDS. Clinical Society (EACS). European Guidelines for Treatment of HIV-Infected Adults in Europe, version 8.0. Brussels: EACS; 2015. p. 94.
 29. Martínez Buitrago E, Oñate JM, García-Goez JF, Álvarez J, Lenis W, Sañudo LM, et al. HLA-B*57:01 allele prevalence in treatment-Naïve HIV-infected patients from Colombia. *BMC Infect Dis.* 2019;19(1):793. <https://doi.org/10.1186/s12879-019-4415-3>.
 30. Hughes AR, Mosteller M, Bansal AT, Davies K, Haneline SA, Lai EH, et al. Association of genetic variations in HLA-B region with hypersensitivity to abacavir in some, but not all, populations. *Pharmacogenomics.* 2004;5(2):203–11. <https://doi.org/10.1517/phgs.5.2.203.27481>.
 31. Nolan D, Gaudieri S, Mallal S. Pharmacogenetics: a practical role in predicting antiretroviral drug toxicity? *J HIV Ther.* 2003;8(2):36–41.
 32. Orkin C, Sadiq ST, Rice L, Jackson F. UK EPI team. Prospective epidemiological study of the prevalence of human leukocyte antigen (HLA)-B*5701 in HIV-1-infected UK subjects. *HIV Med.* 2010;11(3):187–92. <https://doi.org/10.1111/j.1468-1293.2009.00762.x>.
 33. Hughes CA, Foisy MM, Dewhurst N, Higgins N, Robinson L, Kelly DV, et al. Abacavir hypersensitivity reaction: an update. *Ann Pharmacother.* 2008;42(3):387–96. <https://doi.org/10.1345/aph.1K522>.
 34. Kapoor R, Martínez-Vega R, Dong D, Tan SY, Leo YS, Lee CC, et al. Reducing hypersensitivity reactions with HLA-B*5701 genotyping before abacavir prescription: clinically useful but is it cost-effective in Singapore? *Pharmacogenet Genomics.* 2015;25(2):60–72. <https://doi.org/10.1097/FPC.000000000000107>.
 35. To SW, Chen JH, Wong KH, Chan KC, Tsang OT, Yam WC. HLA-B*5701 genetic screening among HIV-1 infected patients in Hong Kong: is this a practical approach in Han-Chinese? *Int J STD AIDS.* 2013;24(1):50–2.
 36. Nolan D. HLA-B*5701 screening prior to abacavir prescription: clinical and laboratory aspects. *Crit Rev Clin Lab Sci.* 2009;46(3):153–65. <https://doi.org/10.1080/10408360902937817>.
 37. Squires KE, Young B, DeJesus E, Bellos N, Murphy D, Sutherland-Phillips DH, et al. Safety and efficacy of a 36-week induction regimen of abacavir/lamivudine and ritonavir-boosted atazanavir in HIV-infected patients. *HIV Clin Trials.* 2010;11(2):69–79. <https://doi.org/10.1310/hct1102-69>.
 38. Hughes DA, Vilar FJ, Ward CC, Alfirevic A, Park BK, Pirmohamed M. Cost-effectiveness analysis of HLA B*5701 genotyping in preventing abacavir hypersensitivity. *Pharmacogenetics.* 2004;14(6):335–42. <https://doi.org/10.1097/00008571-200406000-00002>.
 39. Schackman BR, Scott CA, Walensky RP, Losina E, Freedberg KA, Sax PE. The cost-effectiveness of HLA B*5701 genetic screening to guide initial antiretroviral therapy for HIV. *AIDS.* 2008;22(15):2025–33. <https://doi.org/10.1097/QAD.0b013e3283103ce6>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

