

Polygenic risk scores and breast and epithelial ovarian cancer risks for carriers of BRCA1 and BRCA2 pathogenic variants

Daniel Barnes, Matti Rookus, Lesley McGuffog, Goska Leslie, Thea Mooij, Joe Dennis, Nasim Mavaddat, Julian Adlard, Munaza Ahmed, Kristiina Aittomäki, et al.

► **To cite this version:**

Daniel Barnes, Matti Rookus, Lesley McGuffog, Goska Leslie, Thea Mooij, et al.. Polygenic risk scores and breast and epithelial ovarian cancer risks for carriers of BRCA1 and BRCA2 pathogenic variants. *Genetics in Medicine*, Nature Publishing Group, 2020, 22 (10), pp.1653-1666. 10.1038/s41436-020-0862-x . inserm-03193473

HAL Id: inserm-03193473

<https://www.hal.inserm.fr/inserm-03193473>

Submitted on 8 Apr 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Open

Polygenic risk scores and breast and epithelial ovarian cancer risks for carriers of *BRCA1* and *BRCA2* pathogenic variants

A full list of authors and their affiliations appears at the end of the paper.

Purpose: We assessed the associations between population-based polygenic risk scores (PRS) for breast (BC) or epithelial ovarian cancer (EOC) with cancer risks for *BRCA1* and *BRCA2* pathogenic variant carriers.

Methods: Retrospective cohort data on 18,935 *BRCA1* and 12,339 *BRCA2* female pathogenic variant carriers of European ancestry were available. Three versions of a 313 single-nucleotide polymorphism (SNP) BC PRS were evaluated based on whether they predict overall, estrogen receptor (ER)-negative, or ER-positive BC, and two PRS for overall or high-grade serous EOC. Associations were validated in a prospective cohort.

Results: The ER-negative PRS showed the strongest association with BC risk for *BRCA1* carriers (hazard ratio [HR] per standard deviation = 1.29 [95% CI 1.25–1.33], $P = 3 \times 10^{-72}$). For *BRCA2*, the strongest association was with overall BC PRS (HR = 1.31 [95% CI 1.27–1.36], $P = 7 \times 10^{-50}$). HR estimates decreased significantly with

age and there was evidence for differences in associations by predicted variant effects on protein expression. The HR estimates were smaller than general population estimates. The high-grade serous PRS yielded the strongest associations with EOC risk for *BRCA1* (HR = 1.32 [95% CI 1.25–1.40], $P = 3 \times 10^{-22}$) and *BRCA2* (HR = 1.44 [95% CI 1.30–1.60], $P = 4 \times 10^{-12}$) carriers. The associations in the prospective cohort were similar.

Conclusion: Population-based PRS are strongly associated with BC and EOC risks for *BRCA1/2* carriers and predict substantial absolute risk differences for women at PRS distribution extremes.

Genetics in Medicine (2020) 22:1653–1666; <https://doi.org/10.1038/s41436-020-0862-x>

Key words: *BRCA1/2*; breast cancer; ovarian cancer; PRS; genetics

INTRODUCTION

Pathogenic variants in *BRCA1* and *BRCA2* are associated with high risk of developing breast and ovarian cancers.^{1,2} A recent study of *BRCA1/2* carriers estimated the average risk of developing breast cancer by age 80 years to be 72% for *BRCA1* and 69% for *BRCA2* carriers.² Corresponding ovarian cancer risks were 44% for *BRCA1* and 17% for *BRCA2* carriers. This and previous studies have demonstrated that cancer risks for *BRCA1/2* carriers increase with an increasing number of affected first- or second-degree relatives,² suggesting genetic or other familial factors modify cancer risks for *BRCA1/2* carriers. Consistent with this observation, common breast and ovarian cancer susceptibility single-nucleotide polymorphisms (SNPs), identified through genome-wide association studies (GWAS) in the general population, have been shown to modify breast and ovarian cancer risks for *BRCA1/2* carriers.^{3–7}

Polygenic risk scores (PRS) based on the combined effects of disease-associated SNPs, can lead to significant levels of breast and ovarian cancer risk stratification in the general population.^{8,9} It has also been demonstrated that PRS can result in large absolute risk differences of developing these cancers for *BRCA1/2* carriers.¹⁰ The largest study to date was a retrospective cohort study of 23,463 carriers using a

PRS based on up to 88 breast cancer susceptibility SNPs and a PRS based on up to 17 ovarian cancer susceptibility SNPs.¹⁰

Recent population-based GWAS identified an additional 72 breast and 12 ovarian cancer susceptibility SNPs.^{6,7,11} Based on these data, PRS have been constructed that include SNPs associated at both genome-wide and sub-genome-wide significance levels. The best performing PRS for breast cancer includes 313 SNPs.¹²

It is therefore important to understand how the most recently developed breast and ovarian cancer PRS modify cancer risks for *BRCA1/2* carriers, as this information will be necessary for implementation studies to evaluate how their application influences cancer risk management for women with pathogenic variants in these genes. In this study, we used the largest sample of women with pathogenic *BRCA1/2* variants currently available to assess the associations between the most recently developed PRS with cancer risks for *BRCA1/2* carriers. We evaluated how these PRS associations vary with age, cancer family history, and *BRCA1/2* gene variant characteristics. We further validated the associations for the first time in a prospective cohort of carriers and investigated implications for cancer risk prediction.

Correspondence: Daniel R. Barnes (drb54@medschl.cam.ac.uk)

Submitted 27 November 2019; revised 28 May 2020; accepted: 29 May 2020

Published online: 15 July 2020

MATERIALS AND METHODS

Retrospective cohort study participants

Study participants were enrolled through 63 studies from 29 countries contributing to the Consortium of Investigators of Modifiers of *BRCA1/2* (CIMBA).¹³ Eligibility was restricted to women who were ≥ 18 years old at recruitment and carried a pathogenic *BRCA1/2* variant. CIMBA collected information on year of birth, variant description, age at study recruitment and last follow-up, age at breast and ovarian cancer (including invasive ovarian, fallopian tube, or peritoneal) diagnosis, age/date at bilateral prophylactic mastectomy, and number of first- and second-degree relatives with breast or ovarian cancer. Related individuals were tracked through a unique family identifier. The majority of study participants were recruited through cancer genetics clinics and enrolled in regional or national research studies. Variants were categorized according to their predicted or known effect on cellular protein expression: class I included loss-of-function pathogenic variants expected to result in unstable or no protein; class II included variants likely to yield stable mutant proteins.¹⁴ Breast cancer pathology data were available from pathology reviews, tumor registry records, medical records or pathology records, and from tissue microarray immunohistochemical staining.¹⁵

The genotyping, quality control and imputation processes have been described in detail previously^{6,7} (brief description provided in supplement). The present study was restricted to carriers of *BRCA1/2* pathogenic variants of European ancestry, determined using genetic data and multidimensional scaling.^{6,7}

Breast cancer PRS

The methods for calculating the PRS are described in the Supplementary material. We evaluated three versions of the published breast cancer PRS based on the same 313 SNPs, with different weights optimized to predict the risk of overall (PRS_{BC}), ER-negative (PRS_{ER-}), or ER-positive (PRS_{ER+}) breast cancer¹² (Table S1).

The breast cancer PRS were standardized using the standard deviations (SDs) of the corresponding PRS in population-based controls. Therefore, the estimated hazard ratios (HRs) from this study are directly comparable with odds ratios (ORs) estimated from population-based data.¹²

Epithelial ovarian cancer PRS

We constructed ovarian cancer PRS based on ovarian cancer susceptibility SNPs identified through GWAS.⁷ Two ovarian cancer PRS were constructed: one for all invasive epithelial ovarian cancer (EOC) using 30 SNPs (PRS_{EOC}); and one for predicting high-grade serous (HGS) EOC using 22 SNPs (PRS_{HGS}) (Supplementary material, Table S2). HGS is the predominant EOC histotype in *BRCA1/2* tumors.¹⁶

The PRS SDs in unaffected women in our sample were used to standardize PRS_{EOC} and PRS_{HGS}.

Associations between PRS and breast cancer risk

Associations between PRS and breast cancer risk for *BRCA1/2* carriers were assessed using the CIMBA retrospective cohort.

Study participants were censored at the first of (1) breast cancer diagnosis, (2) ovarian cancer diagnosis, (3) risk-reducing bilateral mastectomy, (4) last follow-up, or (5) age 80 years. Participants with a first breast cancer diagnosis were considered affected. To account for nonrandom sampling with respect to disease status, associations were evaluated using weighted Cox regression.^{17,18} This involved assigning age- and disease-specific sampling weights, such that observed weighted age-specific incidences agreed with established incidences for *BRCA1/2* pathogenic variant carriers (Supplementary material).¹⁹

We assessed the associations between three breast cancer PRS with the risk of overall breast cancer, and separately with ER-positive or ER-negative breast cancer risk. Models were stratified by country and Ashkenazi Jewish ancestry and were adjusted for birth cohort and the first four ancestry informative principal components calculated separately by genotyping array (Supplementary material). We fitted models adjusting for family history of breast cancer in first- and second-degree relatives to determine whether cancer family history was a confounder of PRS associations. Family history was coded as no family history, or one relative, or two or more relatives diagnosed with breast cancer. Robust variances were calculated to account for the inclusion of related individuals by clustering on family membership. All models were fitted separately in *BRCA1* and *BRCA2* carriers.

We fitted separate models in which the PRS was assumed to be (1) continuous and (2) categorical based on PRS percentiles determined by the PRS distribution in unaffected carriers. We tested for variation in the association of the PRS by age by fitting Cox regression models in which the PRS was a time-varying covariate, with age as the time scale, that included a PRS main effect and a PRS-by-age interaction term. Heterogeneity in the associations across countries was assessed by fitting models with a PRS-country interaction term. A likelihood ratio test (LRT) was used to assess statistical significance of interaction terms by comparing the models with the interaction against a model without the interaction term (Supplementary material). Similarly, LRTs were used to compare the fit of nested models.

Previous studies have demonstrated that cancer risks for *BRCA1/2* carriers vary by pathogenic variant location or functional effect.^{2,20} To investigate whether the PRS associations varied by *BRCA1/2*-variant location, we fitted models that included a PRS by location interaction. Variants were grouped into regions by nucleotide position on the basis of previously reported differences in breast or ovarian cancer risks. *BRCA1* variants were grouped in three regions (5' to c.2281, c.2282 to c.4071, and c.4072 to 3').^{20,21} The *BRCA2* ovarian cancer cluster region (OCCR) was used to define the variant location groups.^{20,22} Two *BRCA2* OCCR definitions were used: "narrow" (5' to c.3846, c.3847 to c.6275, c.6276 to 3') and "wide" (5' to c.2831, c.2832 to c.6401, c.6402 to 3'). We also investigated variation in PRS associations by the predicted variant effect on protein stability/expression (class I versus class II, defined above).

To assess the associations with ER-specific breast cancer risk, a similar censoring process was used except the event of interest was diagnosis of either ER-positive or ER-negative breast cancer. Affected carriers with the alternative ER status to the outcome of interest were censored at that diagnosis. Carriers with missing ER status were excluded from the analysis.

Associations with epithelial ovarian cancer risk

The associations with EOC risk were evaluated following a similar process. However, women were censored at bilateral risk-reducing salpingo-oophorectomy (RRSO) rather than bilateral mastectomy. Carriers with a first ovarian cancer diagnosis were assumed to be affected in this analysis. We also fitted models that adjusted for family history of ovarian cancer in first- and second-degree relatives, coded as no family history, or one relative, or two or more relatives diagnosed with the disease.

The discriminatory ability of each PRS was assessed by Harrell's C-statistic²³ stratified by country and Ashkenazi Jewish ancestry and adjusted for birth cohort and principal components.²⁴ Standard errors were estimated using 1000 bootstrap replications.

Validation in prospective cohorts

The PRS associations were further evaluated using prospective cohort data. The prospective cohort included pathogenic variant carriers from the *BRCA1* and *BRCA2* Cohort Consortium (BBCC)² and CIMBA¹³ who were unaffected at recruitment (informed consent and baseline questionnaire). The BBCC included data from the International *BRCA1/2* Carrier Cohort Study (IBCCS), Breast Cancer Family Registry (BCFR), and the Kathleen Cunningham Foundation Consortium for Research into Familial Breast Cancer (kConFab) (details in Supplementary material).² Only women of European ancestry were included in the analysis. All prospective cohort participants were genotyped as part of the CIMBA effort described above. However, prospective analyses considered only the prospective follow-up period from the time at recruitment of each participant into the study. Thus, the analysis time considered in the prospective and retrospective analyses were completely distinct. Associations were evaluated using Cox regression, separately for *BRCA1* and *BRCA2* carriers. The censoring process and analysis are described in detail in the Supplementary material.

Predicted age-specific cancer risks by PRS

Retrospective analysis HR estimates were used to predict age-specific absolute risks of developing breast and ovarian cancer by PRS percentiles following a previously published method.²⁵ To ensure consistency with known cancer risks for *BRCA1/2* carriers, average age-specific cancer incidences were constrained over PRS percentile categories to agree with external estimates of cancer incidences for carriers² (Supplementary material). We also calculated absolute breast cancer risks for carriers in the absence or presence of cancer family history and by *BRCA2*

variant location, assuming external average cancer incidences by family history and variant location.² The absolute risks were used to calculate 10-year cancer risks at each age by different PRS percentiles (Supplementary material).

Ethics statement

All study participants provided written informed consent and participated in research or clinical studies at the host institute under ethically approved protocols. The studies and their approving institutes are listed as a separate online Supplement (Ethics Statement).

All statistical tests were two-sided. Retrospective and prospective cohort analyses were performed using R 3.5.1. Age-varying PRS and discrimination analyses were conducted using Stata 13.1 (Supplementary material).

RESULTS

The CIMBA retrospective cohort consisted of 18,935 *BRCA1* carriers (9473 diagnosed with breast and 2068 with ovarian cancer) and 12,339 *BRCA2* carriers (6332 with breast and 718 with ovarian cancer, Table S3).

The SNPs included in the PRS were well imputed on both genotyping platforms (Supplementary material, Figs. S1, S2, Tables S1, S2). The average PRS were larger for women diagnosed with cancer, compared with unaffected carriers (Table S3), but the PRS SDs were similar in unaffected and affected carriers (Table S3).

Associations with breast cancer risk

Table 1 shows the associations between PRS_{BC}, PRS_{ER-}, and PRS_{ER+} and overall breast cancer risk for carriers using the CIMBA retrospective cohort data. PRS_{ER-} yielded the strongest association for *BRCA1* carriers (per SD HR = 1.29, 95% CI = 1.25–1.33, $P = 3 \times 10^{-72}$). For *BRCA2* carriers, the strongest associations were found for PRS_{BC} (per SD HR = 1.31, 95% CI = 1.27–1.36, $P = 7 \times 10^{-50}$) and PRS_{ER+} (per SD HR = 1.31, 95% CI = 1.26–1.36, $P = 6 \times 10^{-49}$). Adjusting for breast cancer family history yielded similar associations between the PRS and breast cancer risk to those observed in the unadjusted models (Table 1). Family history was significantly associated with risk in all models.

The PRS_{ER-} and PRS_{BC} were used for subsequent *BRCA1* and *BRCA2* carrier analyses, respectively. There was no statistically significant evidence of heterogeneity in the country-specific HR estimates (*BRCA1* $P_{LRT} = 0.26$, *BRCA2* $P_{LRT} = 0.64$; Fig. S3). The estimated HRs for each PRS percentile category (Table 2) were consistent with the HRs predicted under models with the continuous PRS (estimated above), but were attenuated compared to the HRs expected under the population-based PRS distributions (Fig. 1a, b). Models estimating PRS percentile-specific associations did not fit significantly better than models in which PRS were continuous (*BRCA1* carriers $P_{LRT} = 0.18$; *BRCA2* carriers $P_{LRT} = 0.99$). The HRs for the breast cancer association decreased with age (Table 2; PRS-by-age interaction HRs: *BRCA1* HR = 0.996, $P = 0.003$; *BRCA2* HR = 0.994,

ARTICLE

Table 1 PRS associations with breast and ovarian cancer risks for *BRCA1* and *BRCA2* pathogenic variant carriers using the CIMBA retrospective cohort data.

Outcome	PRS	<i>BRCA1</i> carriers					<i>BRCA2</i> carriers				
		Unaffected/ affected	No FH ^a adjustment HR (95% CI) P	FH adjusted HR (95% CI) P			Unaffected/ affected	No FH adjustment HR (95% CI) P	FH adjusted HR (95% CI) P		
Breast cancer	BC	9462/ 9473	1.20 (1.17–1.23) 1.29 (1.25–1.33)	1.15×10 ⁻³⁹ 3.03×10⁻⁷² 6.93×10⁻²⁹	1.20 (1.17–1.23) 1.29 (1.25–1.33)	9.54×10 ⁻⁴⁰ 1.02×10⁻⁷¹ 5.50×10⁻²⁹	6007/ 6332	1.31 (1.27–1.36) 1.23 (1.19–1.28) 1.31 (1.26–1.36)	7.11×10⁻⁵⁰ 4.06×10 ⁻²⁹ 6.12×10 ⁻⁴⁹	1.31 (1.26–1.36) 1.23 (1.18–1.27) 1.30 (1.26–1.35)	6.54×10⁻⁴⁸ 6.72×10 ⁻²⁸ 5.10×10 ⁻⁴⁷
	ER-										
	ER+										
ER-negative breast cancer	BC	10,138/ 3263	1.09 (1.05–1.13) 1.23 (1.18–1.28)	3.69×10 ⁻⁶ 2.39×10⁻²⁷ 4.58×10⁻³	1.09 (1.05–1.13) 1.23 (1.18–1.27)	4.44×10 ⁻⁶ 1.08×10⁻²⁶ 4.93×10⁻³	8049/ 703	1.20 (1.11–1.30) 1.31 (1.21–1.43) 1.17 (1.08–1.26)	4.90×10 ⁻⁶ 1.15×10⁻¹⁰ 1.36×10 ⁻⁴	1.19 (1.10–1.29) 1.29 (1.19–1.41) 1.15 (1.07–1.25)	1.91×10 ⁻⁵ 9.98×10⁻¹⁰ 3.91×10 ⁻⁴
	ER-										
	ER+										
ER-positive breast cancer	BC	12,376/ 1025	1.44 (1.35–1.53) 1.29 (1.21–1.38) 1.44 (1.35–1.54)	3.88×10⁻²⁸ 2.94×10⁻¹⁵ 3.94×10⁻²⁸	1.44 (1.35–1.54) 1.29 (1.21–1.37) 1.45 (1.35–1.54)	1.25×10⁻²⁷ 9.25×10⁻¹⁵ 1.12×10⁻²⁷	6440/ 2312	1.37 (1.31–1.44) 1.22 (1.16–1.28) 1.38 (1.32–1.45)	2.95×10 ⁻⁴⁰ 1.88×10⁻⁴² 1.93×10 ⁻¹⁵	1.36 (1.30–1.43) 1.21 (1.15–1.27) 1.37 (1.31–1.44)	6.28×10 ⁻³⁸ 5.99×10⁻⁴⁰ 1.54×10 ⁻¹⁴
	ER-										
	ER+										
Ovarian cancer	EOC	16,867/ 2068	1.31 (1.24–1.39) 1.32 (1.25–1.40)	1.49×10 ⁻²¹ 3.01×10⁻²²	1.31 (1.24–1.39) 1.32 (1.25–1.40)	2.36×10 ⁻²¹ 5.18×10⁻²²	11,621/ 718	1.43 (1.29–1.59) 1.44 (1.30–1.60)	1.81×10 ⁻¹¹ 4.34×10⁻¹²	1.42 (1.28–1.58) 1.43 (1.29–1.59)	3.40×10 ⁻¹¹ 7.54×10⁻¹²
	HGS										

BC breast cancer, CI confidence interval, CIMBA Consortium of Investigators of Modifiers of *BRCA1/2*, ER- estrogen receptor negative, ER+ estrogen receptor positive, EOC epithelial ovarian cancer, FH family history, HGS high-grade serous, HR hazard ratio, PRSpolygenic risk score.

Rows in bold represent the best performing PRS for each particular outcome.

^aFamily history in first- and second-degree relatives: coded as no family history, or one relative, or two or more relatives diagnosed with the disease.

$P = 9.40 \times 10^{-5}$). The HRs for the PRS associations with breast cancer risk did not differ by variant location (Table 2: *BRCA1* $P_{LRT} = 0.17$; *BRCA2* $P_{LRT} \geq 0.27$). However, the associations differed by the predicted effect of the gene variant on protein stability/expression: the HRs for the PRS associations with breast cancer risk were larger for carriers with class II (stable mutant proteins) versus class I (unstable/no protein) variants (Table 2, *BRCA1*: class I HR = 1.26 [95% CI = 1.22–1.30], class II HR = 1.38 [1.30–1.46], $P_{\text{difference}} = 0.011$; *BRCA2*: class I HR = 1.30 [95% CI = 1.25–1.35], class II HR = 1.72 [95% CI = 1.44–2.06], $P_{\text{difference}} = 0.003$).

Under the age-varying PRS models, the C-statistic for PRS_{ER-} was 0.60 (95% CI = 0.59–0.61) for *BRCA1* carriers, and for the PRS_{BC} for *BRCA2* carriers 0.65 (95% CI = 0.63–0.67). Under models that did not include the age-varying PRS, the estimated C-statistics were 0.58 (95% CI = 0.57–0.59) and 0.60 (95% CI = 0.59–0.62) for *BRCA1* and *BRCA2* carriers, respectively.

Associations with ER-specific breast cancer risk

The strongest PRS associations with ER-negative disease were observed for PRS_{ER-} for both *BRCA1* (per SD HR = 1.23, 95% CI = 1.18–1.28, $P = 2 \times 10^{-27}$) and *BRCA2* (HR = 1.31, 95% CI = 1.21–1.43, $P = 1 \times 10^{-10}$) carriers (Table 1). The PRS_{BC} and PRS_{ER+} showed the strongest associations with ER-positive disease for *BRCA1* and *BRCA2* carriers with similar HR estimates for PRS_{BC} and PRS_{ER+} (Table 1). The associations remained similar after adjusting for family history of breast cancer (Table 1).

Associations with epithelial ovarian cancer risk

The 30-SNP PRS_{EOC} was strongly associated with EOC risk for *BRCA1* (per SD HR = 1.31, 95% CI = 1.24–1.39,

$P = 1 \times 10^{-21}$) and *BRCA2* (per SD HR = 1.43, 95% CI = 1.29–1.59, $P = 2 \times 10^{-11}$) carriers (Table 1). The 22-SNP PRS_{HGS}, based only on SNPs showing associations with high-grade serous EOC, showed similar associations (Table 1, *BRCA1* HR = 1.32, 95% CI = 1.25–1.40, $P = 3 \times 10^{-22}$; *BRCA2* HR = 1.44, 95% CI = 1.30–1.60, $P = 4 \times 10^{-12}$). Adjusting for family history of ovarian cancer yielded similar associations to unadjusted models (Table 1).

PRS_{HGS} was used for downstream analyses for *BRCA1* and *BRCA2* carriers. There was no evidence of heterogeneity in the PRS_{HGS} associations across countries (Fig. S3: *BRCA1* $P_{LRT} = 0.08$; *BRCA2* $P_{LRT} = 0.97$). For both *BRCA1* and *BRCA2* carriers the estimated HRs by PRS percentile categories (Table 2) were consistent with those expected under the theoretical population-based PRS distributions (Fig. 1c, d). There was no evidence that the PRS_{HGS} association with EOC risk varied by age (*BRCA1* $P = 0.35$; *BRCA2* $P = 0.14$). The associations between PRS_{HGS} and EOC risk varied by *BRCA1* variant location ($P_{LRT} = 8.7 \times 10^{-3}$), with a larger HR for variants in the central region of *BRCA1* (central region HR = 1.50, 95% CI = 1.35–1.66; 5' to c.2281 region HR = 1.30, 95% CI = 1.18–1.42; c.4072 to 3' region HR = 1.21, 95% CI = 1.10–1.33). There was little evidence to support differences in the associations by *BRCA2* variant location (Table 2). There was no evidence of differences in the associations by the *BRCA1* variant predicted effect on protein expression ($P_{\text{difference}} = 0.85$).

The C-statistics for PRS_{HGS} were estimated to be 0.604 (95% CI = 0.582–0.626) for *BRCA1* and 0.667 (95% CI = 0.636–0.699) for *BRCA2* carriers.

Prospective cohort associations

The breast cancer prospective cohort included 2088 *BRCA1* carriers with 297 incident cases and 1757 *BRCA2* carriers

Table 2 Categorical PRS, age-varying and pathogenic variant characteristic specific PRS associations with cancer risks for BRCA1 and BRCA2 carriers, using data from the CIMBA retrospective cohort.

Model	Category	Breast cancer			Ovarian cancer							
		BRCA1 carriers: PRS _{ER} HR (95% CI)	P	P _{LRT}	BRCA2 carriers: PRS _{BC} HR (95% CI)	P	P _{LRT}	BRCA2 carriers: PRS _{HGS} HR (95% CI)	P	P _{LRT}		
Categorical PRS percentiles (%)	0-5	0.59 (0.50-0.70)			0.52 (0.42-0.64)			0.68 (0.50-0.92)			0.40 (0.20-0.79)	
	5-10	0.69 (0.59-0.80)			0.60 (0.49-0.73)			0.80 (0.59-1.09)			0.47 (0.24-0.91)	
	10-20	0.77 (0.69-0.86)			0.69 (0.59-0.80)			1.01 (0.81-1.26)			0.53 (0.33-0.85)	
	20-40	0.91 (0.84-1.00)			0.82 (0.73-0.92)			0.96 (0.80-1.15)			0.83 (0.60-1.14)	
	40-60	1.00 [reference]			1.00 [reference]			1.00 [reference]			1.00 [reference]	
	60-80	1.12 (1.03-1.21)			1.05 (0.94-1.18)			1.16 (0.97-1.39)			0.97 (0.71-1.33)	
	80-90	1.38 (1.25-1.53)			1.21 (1.06-1.38)			1.57 (1.28-1.91)			1.38 (0.95-2.00)	
	90-95	1.55 (1.37-1.75)			1.44 (1.21-1.71)			1.86 (1.44-2.41)			1.36 (0.86-2.15)	
	95-100	1.61 (1.43-1.82)			1.69 (1.45-1.98)			2.24 (1.76-2.84)			2.03 (1.31-3.15)	
	PRS	1.517 (1.359-1.694)	1.04×10 ⁻¹³	0.017	1.721 (1.498-1.977)	1.75×10 ⁻¹⁴	2.27×10 ⁻³	1.507 (1.125-2.020)	6.02×10 ⁻³	0.41	2.183 (1.263-3.774)	5.17×10 ⁻³
PRS × age	0.996 (0.993-0.999)	3.27×10 ⁻³		0.994 (0.991-0.997)	9.40×10 ⁻⁵		0.997 (0.991-1.003)	0.35		0.992 (0.982-1.003)	0.14	
Gene	Class I	1.26 (1.22-1.30)	0.011 ^b	5.29×10 ⁻³	1.30 (1.25-1.35)	3.20×10 ⁻³ ^b	0.046	1.33 (1.24-1.43)	0.85 ^b	N/A ^c	N/A ^c	
pathogenic variant class	Class II	1.38 (1.30-1.46)		0.17	1.72 (1.44-2.06)			1.32 (1.18-1.47)		8.73×10 ⁻³	N/A	
BRCA1 pathogenic variant location	c.2282-c.4071	1.25 (1.19-1.31)			N/A			1.50 (1.35-1.66)				
	5' to c.2281	1.28 (1.22-1.34)						1.30 (1.18-1.42)				
	c.4072 to 3'	1.34 (1.28-1.41)						1.21 (1.10-1.33)				
BRCA2 pathogenic variant location (narrow)	c.3847-c.6275	N/A					0.27	N/A		1.48 (1.24-1.76)	0.96	
	5' to c.3846	1.26 (1.17-1.34)						1.41 (1.17-1.69)				
	c.6276 to 3'	1.37 (1.29-1.46)						1.43 (1.20-1.70)				
BRCA2 pathogenic variant location (wide)	c.2831-c.6401	N/A					0.33	N/A		1.48 (1.26-1.75)	0.90	
	5' to c.2830	1.26 (1.17-1.37)						1.37 (1.13-1.68)				
	c.6402 to 3'	1.37 (1.29-1.46)						1.43 (1.20-1.71)				

Class I pathogenic variant refers to loss-of-function pathogenic variants expected to result in unstable or no protein; class II pathogenic variant refers to pathogenic variants likely to yield stable mutant proteins. P value for the Wald test statistic unless otherwise stated. LRT compares the models with an interaction term against the model without the interaction term.

BC breast cancer, CI confidence interval, CIMBA Consortium of Investigators of Modifiers of BRCA1/2, ER-estrogen receptor negative, HGS high-grade serous, HR hazard ratio, LRT likelihood ratio test, N/A not applicable.

^aAge in years.

^bP value for the difference in HR for class I carriers vs. the HR for class II carriers.

^cNumber of affected class II carriers was too small to make meaningful inference.

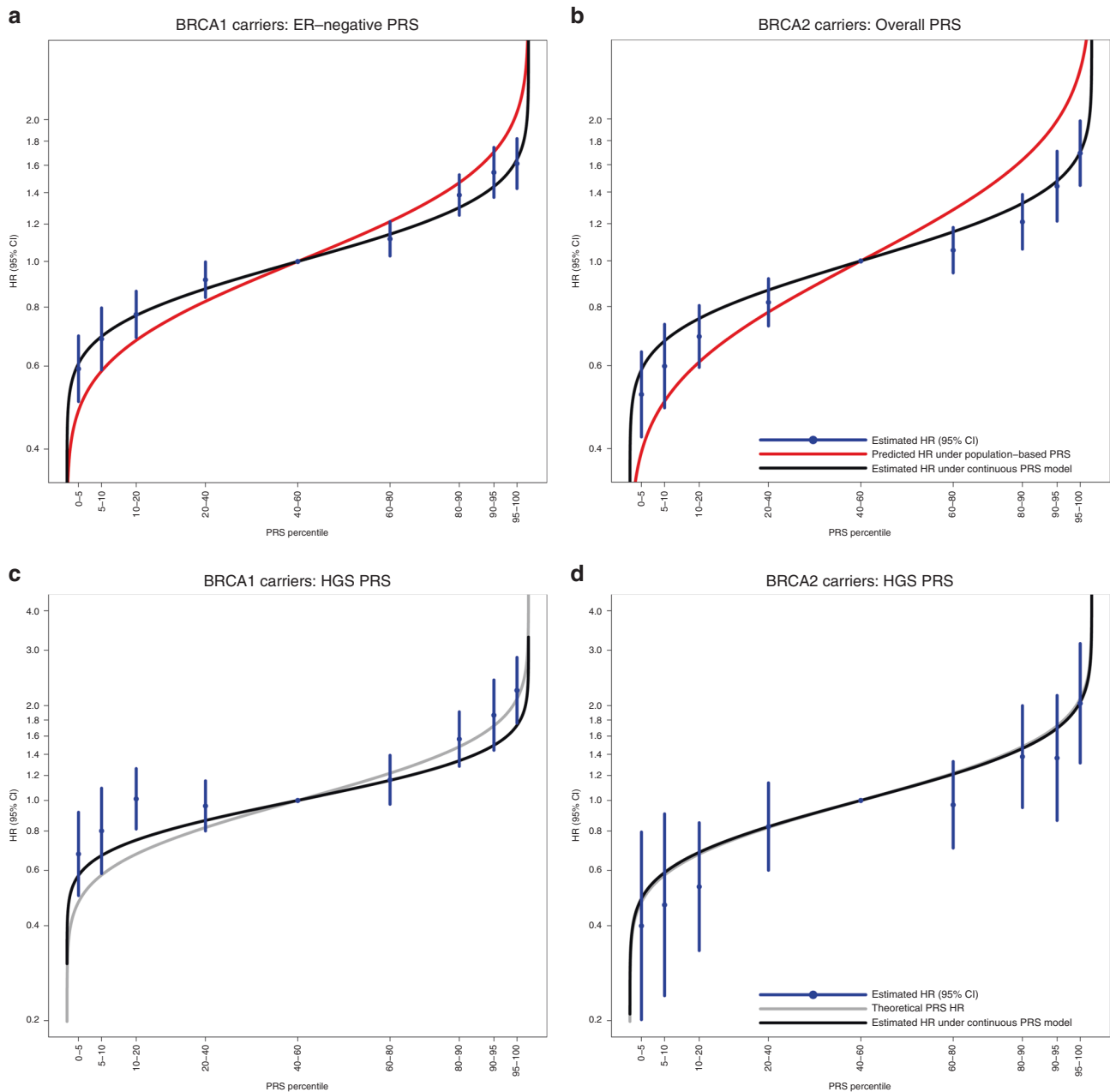


Fig. 1 Associations with specific polygenic risk score (PRS) percentiles. The PRS percentile thresholds were determined in the sets of unaffected carriers for the disease under assessment. Table 2 shows the estimated hazard ratios (HRs). The black curve represents the expected HRs assuming the per standard deviation HR estimates in *BRCA1* and *BRCA2* carriers based on the continuous PRS models (Table 1). (a) PRS_{ER-} percentile-specific associations with breast cancer risk for *BRCA1* carriers. The red curve represents the expected HRs over the PRS percentile distribution, assuming the per SD odds ratio (OR) estimate from the population-based validation studies from Mavaddat et al.¹² (OR = 1.45 per PRS_{ER-} standard deviation). (b) PRS_{BC} percentile-specific associations with breast cancer risk for *BRCA2* carriers. The red curve represents the expected HRs over the PRS percentile distribution, assuming the per SD OR estimate from the population-based validation studies from Mavaddat et al.¹² (OR = 1.61 per PRS_{BC} standard deviation). (c) PRS_{HGS} percentile-specific associations with ovarian cancer risk for *BRCA1* carriers. (d) PRS_{HGS} percentile-specific associations with ovarian cancer risk for *BRCA2* carriers. The gray curve (c and d only) represents the theoretical HRs across the PRS distribution, calculated by assuming external single-nucleotide polymorphism (SNP) effect sizes and allele frequencies for SNPs contributing to the PRS. CI confidence interval, ER estrogen receptor, HGS high-grade serous.

with 215 incident cases (Table S4). The PRS_{ER-} was associated with breast cancer risk for *BRCA1* carriers (per SD HR = 1.28, 95% CI = 1.14–1.44, $P = 4.4 \times 10^{-5}$). For *BRCA2* carriers, PRS_{BC} was associated with breast cancer risk with a per SD HR = 1.36 (95% CI = 1.17–1.57, $P = 4.3 \times 10^{-5}$) (Table 3).

The ovarian cancer prospective cohort comprised 3152 *BRCA1* carriers with 108 incident cases and 2495 *BRCA2* carriers with 56 incident cases (Table S4). The PRS_{HGS} was associated with EOC risk for both *BRCA1* (HR = 1.28, 95% CI = 1.06–1.55, $P = 0.011$) and *BRCA2* (HR = 1.45, 95% CI = 1.13–1.86, $P = 0.003$) carriers (Table 3).

Table 3 Associations of the best performing PRS in the prospective cohort of *BRCA1* and *BRCA2* carriers.

Outcome	PRS	Number of women at risk	Incident cancers	HR (95% CI)	<i>P</i>	
Breast cancer	<i>BRCA1</i> carriers	ER-	2088	297	1.28 (1.14–1.44)	4.44×10^{-5}
	<i>BRCA2</i> carriers	BC	1757	215	1.36 (1.17–1.57)	4.26×10^{-5}
Ovarian cancer	<i>BRCA1</i> carriers	HGS	3152	108	1.28 (1.06–1.55)	1.08×10^{-2}
	<i>BRCA2</i> carriers	HGS	2495	56	1.45 (1.13–1.86)	3.29×10^{-3}

Number of women at risk is the number of pathogenic variant carriers unaffected at baseline. Incident cancers is the number of women who developed breast/ovarian cancer during the follow-up period.

BC breast cancer, CI confidence interval, ER- estrogen receptor negative, HGS high-grade serous, HR hazard ratio, PRS polygenic risk score.

Absolute risks of cancer by PRS percentiles

We estimated age-specific and 10-year absolute risks of developing breast and ovarian cancer across different PRS percentiles (Figs. 2 and S4). *BRCA1* carriers at the 5th and 95th percentiles of the PRS_{ER-} distribution were predicted to have breast cancer risks to age 80 years of 59% and 83%, respectively. The corresponding risks for *BRCA2* carriers based on PRS_{BC} were 57% and 81%. Although PRS associations were not altered by family history adjustment in the models, and there was no significant evidence of interaction between PRS and variant location, both of these factors remain significant predictors of breast cancer risk (in addition to PRS). Therefore, family history and variant location can be considered jointly with the PRS to predict cancer risks for *BRCA1/2* carriers (Figs. S5–S9). For example, breast cancer risk to age 80 years for *BRCA2* carriers with no family history at the 5th and 95th percentiles of the PRS were predicted to be 43% and 67%, respectively, compared with 62% and 85% for those with a family history. The risks of developing ovarian cancer by age 80 years were 30% and 59% for *BRCA1* carriers at the 5th and 95th percentiles of the PRS_{HGS} distribution. The corresponding risks for *BRCA2* carriers were 10% and 28%, respectively.

DISCUSSION

We investigated the associations between a recently reported PRS for breast cancer, based on 313 SNPs, and a PRS for EOC, based on 30 SNPs, with cancer risks for *BRCA1* and *BRCA2* carriers. The associations were evaluated in a large retrospective cohort and separately in a prospective cohort of *BRCA1/2* carriers.

The results demonstrate that the PRS developed using population-based data are also associated with breast and ovarian cancer risk for women with *BRCA1/2* pathogenic variants. The PRS developed for predicting ER-negative breast cancer showed the strongest association with breast cancer risk for *BRCA1* carriers, while for *BRCA2* carriers the PRS developed for predicting overall breast cancer risk performed best. The associations were unchanged after adjusting for cancer family history and were similar between the retrospective and prospective studies. There was evidence that the magnitude of the PRS associations decreased with increasing age for *BRCA1* and *BRCA2* carriers. There was evidence for differences in associations by the predicted effects of variants on protein stability/expression, with the breast cancer PRS

having a larger effect for carriers of variants predicted to yield a stable protein. For ovarian cancer, the PRS developed for predicting overall or HGS EOC demonstrated similar evidence of association with EOC risk, for both *BRCA1* and *BRCA2* carriers. The results are consistent with findings from a previous CIMBA study, based on fewer samples and fewer SNPs, which demonstrated that PRS can lead to large differences in absolute risks of developing breast and ovarian cancers for female *BRCA1/2* carriers.¹⁰

The estimated HR associations for the PRS with breast cancer risk from this study were smaller than the estimated ORs from the population-based study in which they were derived.¹² This difference is unlikely to be an overestimation of the ORs in the general population (“winner’s curse”²⁶), because the effect sizes were estimated in prospective studies that were independent of the data used in their development.^{12,27} Adjustment for family history, a potential confounder in this study, did not influence the associations. Therefore, these most likely represent real differences, in which PRS modify breast cancer risk for *BRCA1/2* carriers to a smaller relative extent than the general population. This meaningful attenuation must be considered when using population-based PRS to predict breast cancer risk for *BRCA1/2* carriers and should be incorporated into breast cancer risk prediction models.²⁸

The departure from the multiplicative model for the joint effects of PRS (or some subset of SNPs) and *BRCA1/2* pathogenic variants might simply reflect the high absolute risks for *BRCA1/2* carriers. That is, women with the highest polygenic risk are likely to develop breast cancer at a young age, so that the relative risk associated with the PRS will diminish with age. It is interesting that the decreasing age effect appeared stronger for carriers than the general population, while the relative risk below age 50 years was more comparable with that seen in the general population.¹² We found that the breast cancer HRs were significantly elevated for carriers of variants that are predicted to generate a stable mutant protein (class II variants). These elevated HRs were similar to the corresponding ORs for association between the PRS and ER-negative (OR = 1.47) and ER-positive (OR = 1.74) breast cancer reported in the general population.¹² The vast majority of individuals in the general population would be expected to be noncarriers with intact *BRCA1/2* protein expression in at-risk tissues, so this observation suggests that some SNPs in the PRS may exert

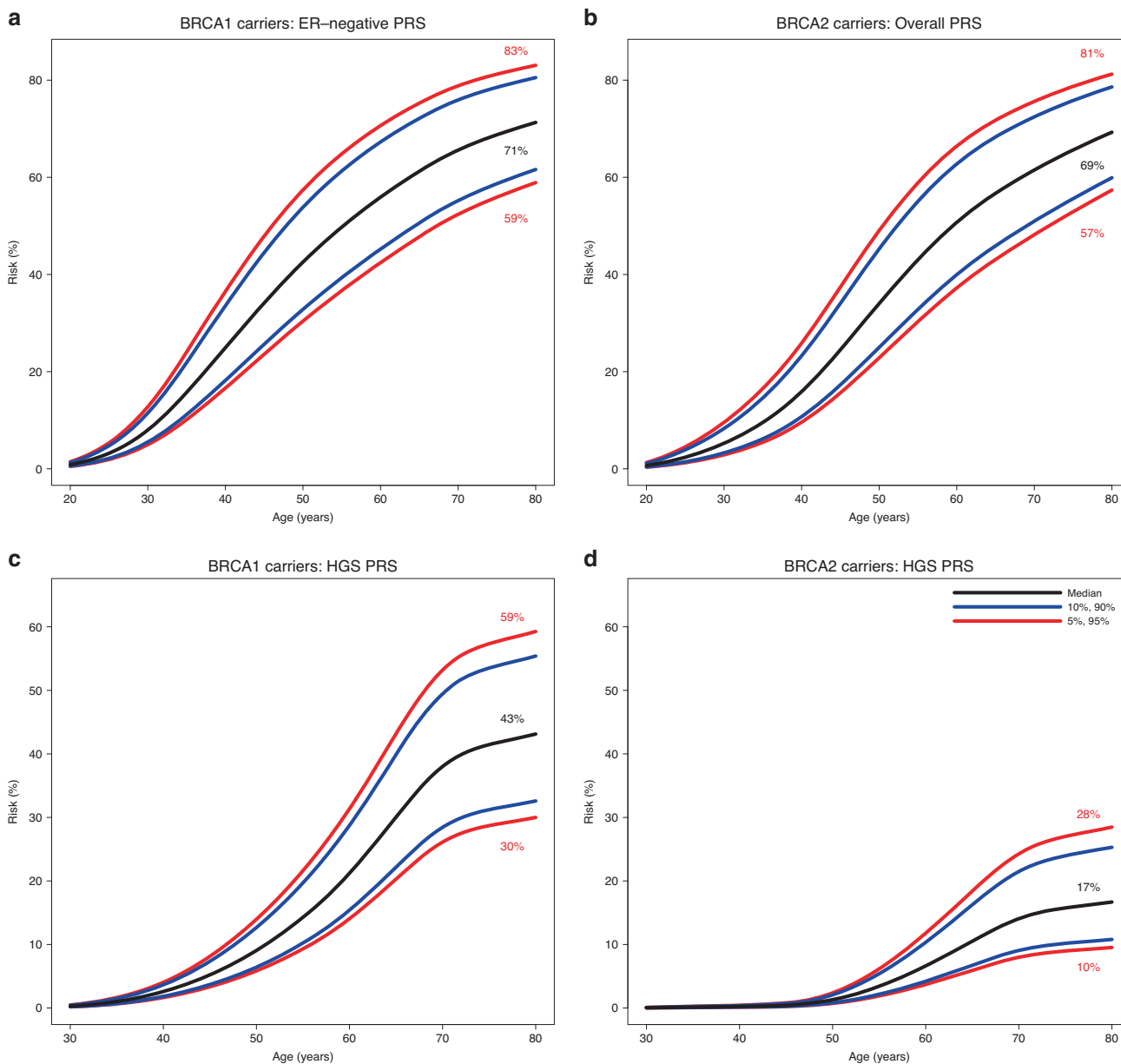


Fig. 2 Predicted absolute risks of developing breast and ovarian cancer by polygenic risk score (PRS) percentile. Risks were calculated assuming the retrospective cohort hazard ratio (HR) estimates (Tables 1, 2). (a) Predicted absolute risks of developing breast cancer for *BRCA1* carriers by percentiles of the PRS_{ER-}. (b) Predicted absolute risks of developing breast cancer for *BRCA2* carriers by percentiles of the PRS_{BC}. (c) Predicted absolute risks of developing ovarian cancer for *BRCA1* carriers by the percentiles of the PRS_{HGS}. (d) Predicted absolute risks of developing ovarian cancer for *BRCA2* carriers by percentiles of the PRS_{HGS}. ER estrogen receptor, HGS high-grade serous.

their effect on proteins that interact with stable wildtype or mutant *BRCA1* or *BRCA2* protein.

We used the ER-specific PRS to assess associations with ER-positive and ER-negative breast cancer for *BRCA1/2* carriers. As expected, the PRS developed for ER-positive breast cancer in the general population was the most predictive of ER-positive breast cancer risk for both *BRCA1* and *BRCA2* carriers, and the PRS developed for ER-negative breast cancer was the most predictive of ER-negative breast cancer for both *BRCA1* and *BRCA2* carriers, in line with known differences in ER expression between *BRCA1*- and *BRCA2*-related

tumors.^{29,30} These results suggest that further risk prediction improvements can be achieved by estimating the risk of developing ER-specific breast cancer for *BRCA1/2* carriers.

Unlike the breast cancer PRS, no systematic evaluation of EOC PRS has been reported in the general population. We therefore included only SNPs identified through GWAS for EOC and its histotypes, using the reported effect sizes as PRS weights. We found that a PRS constructed on the basis of the associations between SNPs and HGS EOC was the most predictive for both *BRCA1* and *BRCA2* carriers, in line with the fact that the majority of tumors in both *BRCA1* and

BRCA2 carriers are HGS.¹⁵ The estimated HR for PRS_{HGS} was larger for *BRCA2* carriers compared with the *BRCA1* carrier HR estimate. This pattern had been observed previously, based on a smaller sample size and fewer SNPs, but the difference between the HRs observed here is smaller than that reported previously.¹⁰

Predicted absolute risks for *BRCA1* carriers at the 5th and 95th PRS percentiles at age 50 years varied from 31% to 58% for breast, and from 5% to 13% for ovarian cancer. By age 80 years, they varied from 59% to 83% for breast and from 30% to 59% for ovarian cancer. The corresponding absolute risks for *BRCA2* carriers by age 50 years ranged from 23% to 49% and by age 80 years from 57% to 81% for breast cancer. The ovarian cancer risks by age 80 years varied from 10% to 28%. We also observed differences in the 10-year age-specific risks of cancer for different PRS distribution percentiles (Fig. S4). For example, the estimated 10-year risk of developing breast cancer at age 40 years was 17% and 34% for *BRCA1* carriers at the 5th and 95th percentiles of the PRS for ER-negative breast cancer, respectively. We found no significant attenuation of the PRS associations when adjusting for family history, and no evidence of interaction between PRS and pathogenic variant location. However, family history and variant location are both associated with cancer risk for *BRCA1/2* carriers.^{2,20–}

²² Taken together, the results suggest that when family history and PRS are considered jointly, or when variant location and PRS are considered jointly, both factors influence the risk of developing breast cancer for *BRCA1/2* carriers. As a consequence, the differences in absolute risk become larger when the PRS is considered together with family history or variant location (Figs. S5–S9) and demonstrate that the PRS should be considered in combination with other risk factors to provide comprehensive cancer risks for *BRCA1/2* carriers.

Strengths of this study include the large cohort sample sizes of *BRCA1/2* carriers and use of independent prospective cohort data to validate PRS associations with cancer risks. The similarity in association estimates between the retrospective and prospective analyses suggests that retrospective estimates have not been strongly influenced by potential biases (e.g., survival bias). As the PRS analyzed in this study were originally developed and validated in population-based studies, the associations reported here represent independent evaluations of the PRS in *BRCA1/2* carriers. The analyses were also adjusted for cancer family history, hence associations are unlikely to be biased due to confounding.

Limitations of this study include the fact that tumor ER status information was missing on a substantial proportion of the study population. Therefore, we were unable to assess associations with ER-specific breast cancer in the entire sample of *BRCA1/2* carriers. The use of PRS developed in the general population means that if there are *BRCA1*- or *BRCA2*-specific modifier SNPs,^{4,5} these may not have been included in the PRS. Therefore, alternative approaches should also investigate developing PRS using data directly from *BRCA1* and *BRCA2* carriers, although much larger sample sizes will be required. We did not present confidence intervals for the

predicted PRS-specific absolute risks of breast or ovarian cancer, and the absolute PRS-specific risks by variant location and family history. These predictions critically depend on external cancer incidence estimates for *BRCA1/2* pathogenic variant carriers,² which themselves are uncertain and therefore should only be used as a general guide. Future studies should aim to factor in uncertainty in the predicted risks based on all parameters. In addition, the PRS-specific absolute cancer risks overall and by family history or pathogenic variant location should be validated in much larger prospective studies of unaffected carriers. Finally, the present analyses were limited to carriers of European ancestry. Hence the results presented may not be applicable to *BRCA1/2* carriers of Asian, African, and other non-European ancestries.

PRS are now being used in risk-stratified screening trials and other implementation studies in the general population.³¹ They are commercially available and have been incorporated in comprehensive cancer risk prediction models.^{28,32} The findings of this study indicate that these PRS, in combination with established risk modifiers (e.g. family history and pathogenic variant characteristics) can be used to provide more personalized cancer risk predictions for carriers, which may assist clinical management decisions. It is therefore important to undertake relevant implementation studies to determine the optimal way of incorporating these PRS into genetic counseling and risk management, and to assess whether PRS on their own or in combination with other risk factors influence the short- or long-term clinical management decisions that female *BRCA1/2* carriers make. Furthermore, the available risk models incorporating the effects of *BRCA1/2* pathogenic variants^{28,32} and PRS should be validated in large prospective studies of carriers.

SUPPLEMENTARY INFORMATION

The online version of this article (<https://doi.org/10.1038/s41436-020-0862-x>) contains supplementary material, which is available to authorized users.

ACKNOWLEDGEMENTS

Full acknowledgements and funding details can be found in the Supplementary material. The following consortia and studies contributed to this research and are listed as authors: **The Genetic Modifiers of *BRCA1* and *BRCA2* (GEMO) Study Collaborators:** Pascaline Berthet, Chrystelle Colas, Marie-Agnès Collonge-Rame, Capucine Delnatte, Laurence Faivre, Paul Gesta, Sophie Giraud, Christine Lasset, Fabienne Lesueur, Véronique Mari, Noura Mebirouk, Emmanuelle Mouret-Fourme, Hélène Schuster, Dominique Stoppa-Lyonnet. **Epidemiological Study of Familial Breast Cancer (EMBRACE) Collaborators:** Julian Adlard, Munaza Ahmed, Antonis Antoniou, Daniel Barrowdale, Paul Brennan, Carole Brewer, Jackie Cook, Rosemarie Davidson, Douglas Easton, Ros Eeles, D. Gareth Evans, Debra Frost, Helen Hanson, Louise Izatt, Kai-ren Ong, Lucy Side, Aoife O'Shaughnessy-Kirwan, Marc Tischkowitz, Lisa Walker. **Kathleen Cuningham Foundation Consortium for research into Familial Breast cancer (kConFab) Investigators:** Georgia Chenevix-

ARTICLE

Trench, Kelly-Anne Phillips, Amanda Spurdle. **Hereditary Breast and Ovarian Cancer Research Group Netherlands (HEBON) Investigators:** Marinus Blok, Peter Devilee, Frans Hogervorst, Maartje Hooning, Marco Koudijs, Arjen Mensenkamp, Hanne Meijers-Heijboer, Matti Rookus, Klaartje van Engelen. **French National BRCA1 and BRCA2 mutations carrier cohort (GENEPSO) Investigators:** Nadine Andrieu, Catherine Noguès. **The Consortium of Investigators of Modifiers of BRCA1 and BRCA2 (CIMBA):** All authors are members of CIMBA.

DISCLOSURE

G.P. has received honoraria from Novartis, Amgen, Roche, Pfizer, and AstraZeneca. The other authors declare no conflicts of interest.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

REFERENCES

1. Antoniou A, Pharoah PDP, Narod S, et al. Average risks of breast and ovarian cancer associated with BRCA1 or BRCA2 mutations detected in case Series unselected for family history: a combined analysis of 22 studies. *Am J Hum Genet.* 2003;72:1117–1130.
2. Kuchenbaecker KB, Hopper JL, Barnes DR, et al. Risks of breast, ovarian, and contralateral breast cancer for BRCA1 and BRCA2 mutation carriers. *JAMA.* 2017;317:2402–2416.
3. Antoniou AC, Spurdle AB, Sinilnikova OM, et al. Common breast cancer-predisposition alleles are associated with breast cancer risk in BRCA1 and BRCA2 mutation carriers. *Am J Hum Genet.* 2008;82:937–948.
4. Couch FJ, Wang X, McGuffog L, et al. Genome-wide association study in BRCA1 mutation carriers identifies novel loci associated with breast and ovarian cancer risk. *PLoS Genet.* 2013;9:e1003212.
5. Gaudet MM, Kuchenbaecker KB, Vijai J, et al. Identification of a BRCA2-specific modifier locus at 6p24 related to breast cancer risk. *PLoS Genet.* 2013;9:e1003173.
6. Milne RL, Kuchenbaecker KB, Michailidou K, et al. Identification of ten variants associated with risk of estrogen-receptor-negative breast cancer. *Nat Genet.* 2017;49:1767–1778.
7. Phelan CM, Kuchenbaecker KB, Tyrer JP, et al. Identification of 12 new susceptibility loci for different histotypes of epithelial ovarian cancer. *Nat Genet.* 2017;49:680–691.
8. Mavaddat N, Pharoah PDP, Michailidou K, et al. Prediction of breast cancer risk based on profiling with common genetic variants. *J Natl Cancer Inst.* 2015;107:djv036.
9. Yang X, Leslie G, Gentry-Maharaj A, et al. Evaluation of polygenic risk scores for ovarian cancer risk prediction in a prospective cohort study. *J Med Genet.* 2018;55:546–554.
10. Kuchenbaecker KB, McGuffog L, Barrowdale D, et al. Evaluation of polygenic risk scores for breast and ovarian cancer risk prediction in BRCA1 and BRCA2 mutation carriers. *J Natl Cancer Inst.* 2017;109:djw302.
11. Michailidou K, Lindström S, Dennis J, et al. Association analysis identifies 65 new breast cancer risk loci. *Nature.* 2017;551:92–94.
12. Mavaddat N, Michailidou K, Dennis J, et al. Polygenic risk scores for prediction of breast cancer and breast cancer subtypes. *Am J Hum Genet.* 2019;104:21–34.
13. Chenevix-Trench G, Milne RL, Antoniou AC, et al. An international initiative to identify genetic modifiers of cancer risk in BRCA1 and BRCA2 mutation carriers: the Consortium of Investigators of Modifiers of BRCA1 and BRCA2 (CIMBA). *Breast Cancer Res.* 2007;9:104.
14. Antoniou AC, Sinilnikova OM, Simard J, et al. RAD51 135G→C modifies breast cancer risk among BRCA2 mutation carriers: results from a combined analysis of 19 studies. *Am J Hum Genet.* 2007;81:1186–1200.
15. Mavaddat N, Barrowdale D, Andrulis IL, et al. Pathology of breast and ovarian cancers among BRCA1 and BRCA2 mutation carriers: results from the Consortium of Investigators of Modifiers of BRCA1/2 (CIMBA). *Cancer Epidemiol Biomarkers Prev.* 2012;21:134–147.
16. Lakhani SR, Manek S, Penault-Llorca F, et al. Pathology of ovarian cancers in BRCA1 and BRCA2 carriers. *Clin Cancer Res.* 2004;10:2473–2481.
17. Antoniou AC, Goldgar DE, Andrieu N, et al. A weighted cohort approach for analysing factors modifying disease risks in carriers of high-risk susceptibility genes. *Genet Epidemiol.* 2005;29:1–11.
18. Barnes DR, Lee A, Investigators E, kConFab I, Easton DF, Antoniou AC. Evaluation of association methods for analysing modifiers of disease risk in carriers of high-risk mutations. *Genet Epidemiol.* 2012;36:274–291.
19. Antoniou AC, Cunningham AP, Peto J, et al. The BOADICEA model of genetic susceptibility to breast and ovarian cancers: updates and extensions. *Br J Cancer.* 2008;98:1457–1466.
20. Rebbeck TR, Mitra N, Wan F, et al. Association of type and location of BRCA1 and BRCA2 mutations with risk of breast and ovarian cancer. *JAMA.* 2015;313:1347–1361.
21. Thompson D, Easton D, Breast Cancer Linkage Consortium. Variation in BRCA1 cancer risks by mutation position. *Cancer Epidemiol Biomarkers Prev.* 2002;11:329–336.
22. Thompson D, Easton D, Breast Cancer Linkage Consortium. Variation in cancer risks, by mutation position, in BRCA2 mutation carriers. *Am J Hum Genet.* 2001;68:410–419.
23. Harrell FE. Evaluating the yield of medical tests. *JAMA.* 1982;247:2543–2546.
24. White IR, Rapsomaniki E, Emerging Risk Factors Collaboration. Covariate-adjusted measures of discrimination for survival data. *Biom J.* 2015;57:592–613.
25. Antoniou AC, Beesley J, McGuffog L, et al. Common breast cancer susceptibility alleles and the risk of breast cancer for BRCA1 and BRCA2 mutation carriers: implications for risk prediction. *Cancer Res.* 2010;70:9742–9754.
26. Xiao R, Boehnke M. Quantifying and correcting for the winner's curse in genetic association studies. *Genet Epidemiol.* 2009;33:453–462.
27. Läll K, Lepamets M, Palover M, et al. Polygenic prediction of breast cancer: comparison of genetic predictors and implications for risk stratification. *BMC Cancer.* 2019;19:557.
28. Lee A, Mavaddat N, Wilcox AN, et al. BOADICEA: a comprehensive breast cancer risk prediction model incorporating genetic and nongenetic risk factors. *Genet Med.* 2019;1708–1718.
29. Mavaddat N, Barrowdale D, Andrulis IL, et al. Pathology of breast and ovarian cancers among BRCA1 and BRCA2 mutation carriers: results from the Consortium of Investigators of Modifiers of BRCA1/2 (CIMBA). *Cancer Epidemiol Biomarkers Prev.* 2012;21:134–147.
30. Lee AJ, Cunningham AP, Kuchenbaecker KB, et al. BOADICEA breast cancer risk prediction model: updates to cancer incidences, tumour pathology and web interface. *Br J Cancer.* 2014;110:535–545.
31. Antoniou A, Anton-Culver H, Borowsky A, et al. A response to "Personalised medicine and population health: breast and ovarian cancer". *Hum Genet.* 2019;138:287–289.
32. IBIS. IBIS breast cancer risk evaluation tool. 2017. <http://www.ems-trials.org/riskevaluator/>.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2020

Daniel R. Barnes, PhD¹, Matti A. Rookus, PhD², Lesley McGuffog¹, Goska Leslie, MEng¹,
 Thea M. Mooij, MSc², Joe Dennis, MSc¹, Nasim Mavaddat, PhD¹, Julian Adlard, MD³,
 Munaza Ahmed, MD(Res), FRCP⁴, Kristiina Aittomäki, MD, PhD⁵, Nadine Andrieu, PhD^{6,7,8,9},
 Irene L. Andrulis, PhD^{10,11}, Norbert Arnold, PhD^{12,13}, Banu K. Arun, MD¹⁴, Jacopo Azzollini, MD¹⁵,
 Judith Balmaña, MD, PhD^{16,17}, Rosa B. Barkardottir, CandSci^{18,19}, Daniel Barrowdale, BSc¹,
 Javier Benitez, PhD^{20,21}, Pascaline Berthet, MD²², Katarzyna Białkowska, MSc²³,
 Amie M. Blanco, MS²⁴, Marinus J. Blok, PhD²⁵, Bernardo Bonanni, MD²⁶,
 Susanne E. Boonen, MD, PhD²⁷, Åke Borg, PhD²⁸, Aniko Bozsik, PhD²⁹, Angela R. Bradbury, MD³⁰,
 Paul Brennan, MBBS, FRCP³¹, Carole Brewer, MD³², Joan Brunet, MD, PhD³³, Sandra S. Buys, MD³⁴,
 Trinidad Caldes, MD³⁵, Maria A. Caligo, PhD³⁶, Ian Campbell, PhD^{37,38},
 Lise Lotte Christensen, MSc, PhD³⁹, Wendy K. Chung, MD, PhD⁴⁰, Kathleen B. M. Claes, PhD⁴¹,
 Chrystelle Colas, MD, PhD⁴², GEMO Study Collaborators, EMBRACE Collaborators,
 Marie-Agnès Collonge-Rame, MD⁴³, Jackie Cook, MD⁵⁰, Mary B. Daly, MD, PhD⁶²,
 Rosemarie Davidson, MD⁵¹, Miguel de la Hoya, PhD³⁵, Robin de Putter, MD⁴¹,
 Capucine Delnatte, MD⁴⁴, Peter Devilee, PhD^{63,64}, Orland Diez, PhD^{65,66}, Yuan Chun Ding, PhD⁶⁷,
 Susan M. Domchek, MD⁶⁸, Cecilia M. Dorfling, MSc⁶⁹, Martine Dumont, PhD⁷⁰, Ros Eeles, MD, PhD⁵²,
 Bent Ejlersen, MD⁷¹, Christoph Engel, MD⁷², D. Gareth Evans, MD, PhD^{53,54},
 Laurence Faivre, MD, PhD^{45,73}, Lenka Foretova, MD, PhD⁷⁴, Florentia Fostira, PhD⁷⁵,
 Michael Friedlander, MD, PhD⁷⁶, Eitan Friedman, MD, PhD^{77,78}, Debra Frost, ONC¹,
 Patricia A. Ganz, MD⁷⁹, Judy Garber, MD, MPH⁸⁰, Andrea Gehrig, MD⁸¹, Anne-Marie Gerdes, MD⁸²,
 Paul Gesta, MD⁸³, Sophie Giraud, MD, PhD⁴⁶, Gord Glendon, MSc¹⁰, Andrew K. Godwin, PhD⁸⁴,
 David E. Goldgar, PhD⁸⁵, Anna González-Neira, PhD²¹, Mark H. Greene, MD⁸⁶,
 Daphne Gschwantler-Kaulich, MD⁸⁷, Eric Hahnen, PhD^{88,89}, Ute Hamann, PhD⁹⁰,
 Helen Hanson, MD, FRCP⁹¹, Julia Hentschel, PhD⁹², Frans B. L. Hogervorst, PhD⁹³,
 Maartje J. Hooning, PhD⁹⁴, Judit Horvath, MD, PhD⁹⁵, Chunling Hu, MD, PhD⁹⁶,
 Peter J. Hulick, MD^{97,98}, Evgeny N. Imyanitov, MD⁹⁹, kConFab Investigators, HEBON Investigators,
 GENEPSO Investigators, Claudine Isaacs, MD¹⁰⁸, Louise Izatt, PhD⁵⁵, Angel Izquierdo, MD, MPH³³,
 Anna Jakubowska, PhD^{23,109}, Paul A. James, MBBS, PhD^{38,110}, Ramunas Janavicius, MD, PhD^{111,112},
 Esther M. John, PhD¹¹³, Vijai Joseph, PhD¹¹⁴, Beth Y. Karlan, MD^{115,116}, Karin Kast, MD¹¹⁷,
 Marco Koudijs, PhD¹⁰³, Torben A. Kruse, PhD¹¹⁸, Ava Kwong, MD^{119,120,121}, Yael Laitman, MD⁷⁷,
 Christine Lasset, MD, PhD^{47,122}, Conxi Lazaro, PhD³³, Jenny Lester, MPH^{115,116},
 Fabienne Lesueur, PhD^{6,7,8,9}, Annelie Liljegren, MD, PhD¹²³, Jennifer T. Loud, DNP, CRNP⁸⁶,
 Jan Lubiński, MD, PhD²³, Phuong L. Mai, MD, MS¹²⁴, Siranoush Manoukian, MD¹⁵,
 Véronique Mari, MD⁴⁸, Noura Mebirouk, PhD^{6,7,8,9}, Hanne E. J. Meijers-Heijboer, PhD¹⁰⁵,
 Alfons Meindl, PhD¹²⁵, Arjen R. Mensenkamp, PhD¹⁰⁴, Austin Miller, PhD¹²⁶,
 Marco Montagna, PhD¹²⁷, Emmanuelle Mouret-Fourme, MD⁴², Semanti Mukherjee, PhD¹²⁸,
 Anna Marie Mulligan, MBCh^{129,130}, Katherine L. Nathanson, PhD⁶⁸, Susan L. Neuhausen, PhD⁶⁷,
 Heli Nevanlinna, PhD¹³¹, Dieter Niederacher, PhD¹³², Finn Cilius Nielsen, MD¹³³,
 Liene Nikitina-Zake, MD, PhD¹³⁴, Catherine Noguès, MD¹⁰⁷, Edith Olah, PhD, DSc²⁹,
 Olufunmilayo I. Olopade, MD¹³⁵, Kai-ren Ong, MD⁵⁶, Aoife O'Shaughnessy-Kirwan, PhD⁵⁸,
 Ana Osorio, PhD^{20,21}, Claus-Eric Ott, MD¹³⁶, Laura Papi, MD, PhD¹³⁷, Sue K. Park, MD, PhD^{138,139,140},
 Michael T. Parsons, PhD¹⁰⁰, Inge Sokilde Pedersen, PhD^{141,142,143}, Bernard Peissel, MD¹⁵,
 Ana Peixoto, MSc¹⁴⁴, Paolo Peterlongo, PhD¹⁴⁵, Georg Pfeiler, MD¹⁴⁶,
 Kelly-Anne Phillips, MD^{37,38,101,102}, Karolina Prajzencanc, MSc²³, Miquel Angel Pujana, PhD¹⁴⁷,
 Paolo Radice, PhD¹⁴⁸, Juliane Ramser, PhD¹⁴⁹, Susan J. Ramus, PhD^{150,151,152}, Johanna Rantala, PhD¹⁵³,
 Gad Rennert, MD, PhD¹⁵⁴, Harvey A. Risch, MD, PhD¹⁵⁵, Mark Robson, MD¹²⁸,
 Karina Rønlund, MD, PhD¹⁵⁶, Ritu Salani, MD, MBA¹⁵⁷, Hélène Schuster, MD^{49,158,159},
 Leigha Senter, MS¹⁶⁰, Payal D. Shah, MD³⁰, Priyanka Sharma, MD¹⁶¹, Lucy E. Side, MD⁵⁷,
 Christian F. Singer, MD, MPH¹⁴⁶, Thomas P. Slavin, MD¹⁶², Penny Soucy, PhD⁷⁰,
 Melissa C. Southey, PhD^{163,164,165}, Amanda B. Spurdle, PhD¹⁰⁰, Doris Steinemann, PhD¹⁶⁶,
 Zoe Steinsnyder, BS¹²⁸, Dominique Stoppa-Lyonnet, MD, PhD^{42,167,168}, Christian Sutter, PhD¹⁶⁹,

ARTICLE

Yen Yen Tan, PhD⁸⁷, Manuel R. Teixeira, MD, PhD^{144,170}, Soo Hwang Teo, PhD^{171,172}, Darcy L. Thull, MS¹⁷³, Marc Tischkowitz, MD, PhD^{59,60}, Silvia Tognazzo, MSc¹²⁷, Amanda E. Toland, PhD¹⁷⁴, Alison H. Trainer, MBBS, PhD^{110,175}, Nadine Tung, MD¹⁷⁶, Klaartje van Engelen, MD¹⁰⁶, Elizabeth J. van Rensburg, PhD⁶⁹, Ana Vega, PhD^{177,178,179}, Jeroen Vierstraete, MSc⁴¹, Gabriel Wagner, MSc¹⁴⁶, Lisa Walker, PhD⁶¹, Shan Wang-Gohrke, MD, PhD¹⁸⁰, Barbara Wappenschmidt, MD^{88,89}, Jeffrey N. Weitzel, MD¹⁶², Siddhartha Yadav, MBBS¹⁸¹, Xin Yang, PhD¹, Drakoulis Yannoukakos, PhD⁷⁵, Dario Zimbalatti, MD¹⁵, Kenneth Offit, MD, MPH^{114,128}, Mads Thomassen, PhD¹¹⁸, Fergus J. Couch, PhD⁹⁶, Rita K. Schmutzler, MD^{88,89,182}, Jacques Simard, PhD⁷⁰, Douglas F. Easton, PhD^{1,183}, Georgia Chenevix-Trench, PhD¹⁰⁰ and Antonis C. Antoniou, PhD¹, on behalf of the Consortium of Investigators of Modifiers of BRCA and BRCA2

¹Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK; ²The Netherlands Cancer Institute, Department of Epidemiology (PSOE), Amsterdam, The Netherlands; ³Chapel Allerton Hospital, Yorkshire Regional Genetics Service, Leeds, UK; ⁴Great Ormond Street Hospital for Children NHS Trust, North East Thames Regional Genetics Service, London, UK; ⁵University of Helsinki, Department of Clinical Genetics, Helsinki University Hospital, Helsinki, Finland; ⁶Inserm U900, Genetic Epidemiology of Cancer team, Paris, France; ⁷Institut Curie, Paris, France; ⁸Mines ParisTech, Fontainebleau, France; ⁹Department of Life & Health Sciences, PSL University, Paris, France; ¹⁰Lunenfeld-Tanenbaum Research Institute of Mount Sinai Hospital, Fred A. Litwin Center for Cancer Genetics, Toronto, ON, Canada; ¹¹University of Toronto, Department of Molecular Genetics, Toronto, ON, Canada; ¹²University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Department of Gynaecology and Obstetrics, Kiel, Germany; ¹³University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Institute of Clinical Molecular Biology, Kiel, Germany; ¹⁴University of Texas MD Anderson Cancer Center, Department of Breast Medical Oncology, Houston, TX, USA; ¹⁵Fondazione IRCCS Istituto Nazionale dei Tumori di Milano, Unit of Medical Genetics, Department of Medical Oncology and Hematology, Milan, Italy; ¹⁶Vall d'Hebron Institute of Oncology, High Risk and Cancer Prevention Group, Barcelona, Spain; ¹⁷University Hospital of Vall d'Hebron, Department of Medical Oncology, Barcelona, Spain; ¹⁸Landsþítali University Hospital, Department of Pathology, Reykjavik, Iceland; ¹⁹University of Iceland, BMC (Biomedical Centre), Faculty of Medicine, Reykjavik, Iceland; ²⁰Centro de Investigación en Red de Enfermedades Raras (CIBERER), Madrid, Spain; ²¹Spanish National Cancer Research Centre (CNIO), Human Cancer Genetics Programme, Madrid, Spain; ²²Centre François Baclesse, Département de Biopathologie, Caen, France; ²³Pomeranian Medical University, Department of Genetics and Pathology, Szczecin, Poland; ²⁴University of California San Francisco, Cancer Genetics and Prevention Program, San Francisco, CA, USA; ²⁵Maastricht University Medical Center, Department of Clinical Genetics, Maastricht, The Netherlands; ²⁶IEO, European Institute of Oncology IRCCS, Division of Cancer Prevention and Genetics, Milan, Italy; ²⁷Zealand University Hospital, Clinical Genetic Unit, Department of Paediatrics, Roskilde, Denmark; ²⁸Lund University, Division of Oncology and Pathology, Department of Clinical Sciences Lund, Lund, Sweden; ²⁹National Institute of Oncology, Department of Molecular Genetics, Budapest, Hungary; ³⁰Perelman School of Medicine at the University of Pennsylvania, Department of Medicine, Abramson Cancer Center, Philadelphia, PA, USA; ³¹Institute of Genetic Medicine, International Centre for Life, Northern Genetic Service, Newcastle upon Tyne, UK; ³²Royal Devon & Exeter Hospital, Department of Clinical Genetics, Exeter, UK; ³³ONCOBELL-IDIBELL-IDIBGI-IGTP, Catalan Institute of Oncology, CIBERONC, Hereditary Cancer Program, Barcelona, Spain; ³⁴Huntsman Cancer Institute, Department of Medicine, Salt Lake City, UT, USA; ³⁵CIBERONC, Hospital Clínico San Carlos, IdISSC (Instituto de Investigación Sanitaria del Hospital Clínico San Carlos), Molecular Oncology Laboratory, Madrid, Spain; ³⁶University Hospital, SOD Genetica Molecolare, Pisa, Italy; ³⁷Peter MacCallum Cancer Center, Melbourne, VIC, Australia; ³⁸The University of Melbourne, Sir Peter MacCallum Department of Oncology, Melbourne, VIC, Australia; ³⁹Aarhus University Hospital, Department of Clinical Medicine, Aarhus, Denmark; ⁴⁰Columbia University, Departments of Pediatrics and Medicine, New York, NY, USA; ⁴¹Ghent University, Centre for Medical Genetics, Ghent, Belgium; ⁴²Institut Curie, Service de Génétique, Paris, France; ⁴³CHU de Besançon, Service de Génétique, Besançon, France; ⁴⁴CHU Nantes, Laboratoire de génétique moléculaire, Nantes, France; ⁴⁵Centre Georges-François Leclerc, Unité d'oncogénétique, Centre de Lutte Contre le Cancer, Dijon, France; ⁴⁶Hospices Civils de Lyon, Department of Genetics, Bron, France; ⁴⁷Centre Léon Bérard, Unité de Prévention et d'Epidémiologie Génétique, Lyon, France; ⁴⁸Centre Antoine Lacassagne, Département d'Hématologie-Oncologie Médicale, Nice, France; ⁴⁹Unité d'Oncogénétique Centre de Lutte contre le Cancer Paul Strauss, Strasbourg, France; ⁵⁰Sheffield Children's Hospital, Sheffield Clinical Genetics Service, Sheffield, UK; ⁵¹Queen Elizabeth University Hospitals, Department of Clinical Genetics, Glasgow, UK; ⁵²The Institute of Cancer Research and Royal Marsden NHS Foundation Trust, Oncogenetics Team, London, UK; ⁵³The University of Manchester, Manchester Academic Health Science Centre, Manchester Universities Foundation Trust, St. Mary's Hospital, Genomic Medicine, Division of Evolution and Genomic Sciences, Manchester, UK; ⁵⁴Manchester Academic Health Science Centre, Manchester Universities Foundation Trust, St. Mary's Hospital, Genomic Medicine, North West Genomics hub, Manchester, UK; ⁵⁵Guy's and St Thomas' NHS Foundation Trust, Clinical Genetics, London, UK; ⁵⁶Birmingham Women's Hospital Healthcare NHS Trust, West Midlands Regional Genetics Service, Birmingham, UK; ⁵⁷Princess Anne Hospital, Southampton, UK; ⁵⁸Cambridge University Hospitals NHS Foundation Trust, East Anglian Medical Genetics Service, Cambridge, UK; ⁵⁹McGill University, Program in Cancer Genetics, Departments of Human Genetics and Oncology, Montréal, QC, Canada; ⁶⁰University of Cambridge, Department of Medical Genetics, National Institute for Health Research Cambridge Biomedical Research Centre, Cambridge, UK; ⁶¹Oxford University Hospitals, Oxford Centre for Genomic Medicine, Oxford, UK; ⁶²Fox Chase Cancer Center, Department of Clinical Genetics, Philadelphia, PA, USA; ⁶³Leiden University Medical Center, Department of Pathology, Leiden, The Netherlands; ⁶⁴Leiden University Medical Center, Department of Human Genetics, Leiden, The Netherlands; ⁶⁵Vall d'Hebron Institute of Oncology (VHIO), Oncogenetics Group, Barcelona, Spain; ⁶⁶University Hospital Vall d'Hebron, Clinical and Molecular Genetics Area, Barcelona, Spain; ⁶⁷Beckman Research Institute of City of Hope, Department of Population Sciences, Duarte, CA, USA; ⁶⁸University of Pennsylvania, Basser Center for BRCA, Abramson Cancer Center, Philadelphia, PA, USA; ⁶⁹University of Pretoria, Department of Genetics, Arcadia, South Africa; ⁷⁰Centre Hospitalier Universitaire de Québec – Université Laval Research Center, Genomics Center, Québec City, QC, Canada; ⁷¹Rigshospitalet, Copenhagen University Hospital, Department of Oncology, Copenhagen, Denmark; ⁷²University of Leipzig, Institute for Medical Informatics, Statistics and Epidemiology, Leipzig, Germany; ⁷³DHU Dijon, Centre de Génétique, Dijon, France; ⁷⁴Masaryk

Memorial Cancer Institute, Department of Cancer Epidemiology and Genetics, Brno, Czech Republic; ⁷⁵National Centre for Scientific Research 'Demokritos', Molecular Diagnostics Laboratory, INRASTES, Athens, Greece; ⁷⁶NHMRC Clinical Trials, ANZ GOTG Coordinating Centre, Camperdown, NSW, Australia; ⁷⁷Chaim Sheba Medical Center, The Susanne Levy Gertner Oncogenetics Unit, Ramat Gan, Israel; ⁷⁸Tel Aviv University, Sackler Faculty of Medicine, Ramat Aviv, Israel; ⁷⁹Jonsson Comprehensive Cancer Centre, UCLA, Schools of Medicine and Public Health, Division of Cancer Prevention & Control Research, Los Angeles, CA, USA; ⁸⁰Dana-Farber Cancer Institute, Cancer Risk and Prevention Clinic, Boston, MA, USA; ⁸¹University Würzburg, Department of Human Genetics, Würzburg, Germany; ⁸²Rigshospitalet, Copenhagen University Hospital, Department of Clinical Genetics, Copenhagen, Denmark; ⁸³CH Niort, Service Régional Oncogénétique Poitou-Charentes, Niort, France; ⁸⁴University of Kansas Medical Center, Department of Pathology and Laboratory Medicine, Kansas City, KS, USA; ⁸⁵Huntsman Cancer Institute, University of Utah School of Medicine, Department of Dermatology, Salt Lake City, UT, USA; ⁸⁶Clinical Genetics Branch, Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, MD, USA; ⁸⁷Medical University of Vienna, Dept of OB/GYN, Vienna, Austria; ⁸⁸Faculty of Medicine and University Hospital Cologne, University of Cologne, Center for Familial Breast and Ovarian Cancer, Cologne, Germany; ⁸⁹Faculty of Medicine and University Hospital Cologne, University of Cologne, Center for Integrated Oncology (CIO), Cologne, Germany; ⁹⁰German Cancer Research Center (DKFZ), Molecular Genetics of Breast Cancer, Heidelberg, Germany; ⁹¹St George's NHS Foundation Trust, Southwest Thames Regional Genetics Service, London, UK; ⁹²University Hospital Leipzig, Institute of Human Genetics, Leipzig, Germany; ⁹³The Netherlands Cancer Institute - Antoni van Leeuwenhoek hospital, Family Cancer Clinic, Amsterdam, The Netherlands; ⁹⁴Erasmus MC Cancer Institute, Department of Medical Oncology, Family Cancer Clinic, Rotterdam, The Netherlands; ⁹⁵University of Münster, Institute of Human Genetics, Münster, Germany; ⁹⁶Mayo Clinic, Department of Laboratory Medicine and Pathology, Rochester, MN, USA; ⁹⁷NorthShore University HealthSystem, Center for Medical Genetics, Evanston, IL, USA; ⁹⁸The University of Chicago Pritzker School of Medicine, Chicago, IL, USA; ⁹⁹N.N. Petrov Institute of Oncology, St. Petersburg, Russia; ¹⁰⁰QIMR Berghofer Medical Research Institute, Department of Genetics and Computational Biology, Brisbane, QLD, Australia; ¹⁰¹The University of Melbourne, Department of Medicine, St Vincent's Hospital, Fitzroy, VIC, Australia; ¹⁰²The University of Melbourne, Centre for Epidemiology and Biostatistics, Melbourne School of Population and Global Health, Melbourne, VIC, Australia; ¹⁰³University Medical Center Utrecht, Department of Medical Genetics, Utrecht, The Netherlands; ¹⁰⁴Radboud University Medical Center, Department of Human Genetics, Nijmegen, The Netherlands; ¹⁰⁵Amsterdam UMC, location AMC, Department of Clinical Genetics, Amsterdam, The Netherlands; ¹⁰⁶Amsterdam UMC, location VUmc, Department of Clinical Genetics, Amsterdam, The Netherlands; ¹⁰⁷Oncogénétique Clinique and Aix Marseille Univ, INSERM, IRD, SESSTIM, Institut Paoli-Calmettes, Département d'Anticipation et de Suivi des Cancers, Marseille, France; ¹⁰⁸Lombardi Comprehensive Cancer Center, Georgetown University, Washington, DC, USA; ¹⁰⁹Pomeranian Medical University, Independent Laboratory of Molecular Biology and Genetic Diagnostics, Szczecin, Poland; ¹¹⁰Peter MacCallum Cancer Center, Parkville Familial Cancer Centre, Melbourne, VIC, Australia; ¹¹¹Vilnius University Hospital Santariskiu Clinics, Hematology, Oncology and Transfusion Medicine Center, Department of Molecular and Regenerative Medicine, Vilnius, Lithuania; ¹¹²State Research Institute Centre for Innovative Medicine, Vilnius, Lithuania; ¹¹³Stanford Cancer Institute, Stanford University School of Medicine, Department of Medicine, Division of Oncology, Stanford, CA, USA; ¹¹⁴Memorial Sloan-Kettering Cancer Center, Clinical Genetics Research Lab, Department of Cancer Biology and Genetics, New York, NY, USA; ¹¹⁵University of California at Los Angeles, David Geffen School of Medicine, Department of Obstetrics and Gynecology, Los Angeles, CA, USA; ¹¹⁶Cedars-Sinai Medical Center, Women's Cancer Program at the Samuel Oschin Comprehensive Cancer Institute, Los Angeles, CA, USA; ¹¹⁷Department of Gynecology and Obstetrics, Medical Faculty and University Hospital Carl Gustav Carus, Technische Universität Dresden, Dresden, Germany; ¹¹⁸Odense University Hospital, Department of Clinical Genetics, Odense, Denmark; ¹¹⁹Cancer Genetics Centre, Hong Kong Hereditary Breast Cancer Family Registry, Happy Valley, Hong Kong; ¹²⁰The University of Hong Kong, Department of Surgery, Pok Fu Lam, Hong Kong; ¹²¹Hong Kong Sanatorium and Hospital, Department of Surgery, Happy Valley, Hong Kong; ¹²²Lyon University, UMR CNRS 5558, Lyon, France; ¹²³Karolinska Institutet, Department of Oncology, Stockholm, Sweden; ¹²⁴Magee-Womens Hospital, University of Pittsburgh School of Medicine, Pittsburgh, PA, USA; ¹²⁵University of Munich, Campus Großhadern, Department of Gynecology and Obstetrics, Munich, Germany; ¹²⁶Roswell Park Cancer Institute, NRG Oncology, Statistics and Data Management Center, Buffalo, NY, USA; ¹²⁷Veneto Institute of Oncology IOV - IRCCS, Immunology and Molecular Oncology Unit, Padua, Italy; ¹²⁸Memorial Sloan-Kettering Cancer Center, Clinical Genetics Service, Department of Medicine, New York, NY, USA; ¹²⁹University of Toronto, Department of Laboratory Medicine and Pathobiology, Toronto, ON, Canada; ¹³⁰University Health Network, Laboratory Medicine Program, Toronto, ON, Canada; ¹³¹University of Helsinki, Department of Obstetrics and Gynecology, Helsinki University Hospital, Helsinki, Finland; ¹³²University Hospital Düsseldorf, Heinrich-Heine University Düsseldorf, Department of Gynecology and Obstetrics, Düsseldorf, Germany; ¹³³Rigshospitalet, Copenhagen University Hospital, Center for Genomic Medicine, Copenhagen, Denmark; ¹³⁴Latvian Biomedical Research and Study Centre, Riga, Latvia; ¹³⁵The University of Chicago, Center for Clinical Cancer Genetics, Chicago, IL, USA; ¹³⁶Campus Virchow Klinikum, Charité, Institute of Human Genetics, Berlin, Germany; ¹³⁷University of Florence, Department of Experimental and Clinical Biomedical Sciences 'Mario Serio', Medical Genetics Unit, Florence, Italy; ¹³⁸Seoul National University College of Medicine, Department of Preventive Medicine, Seoul, Korea; ¹³⁹Seoul National University Graduate School, Department of Biomedical Sciences, Seoul, Korea; ¹⁴⁰Seoul National University, Cancer Research Institute, Seoul, Korea; ¹⁴¹Aalborg University Hospital, Molecular Diagnostics, Aalborg, Denmark; ¹⁴²Aalborg University Hospital, Clinical Cancer Research Center, Aalborg, Denmark; ¹⁴³Aalborg University, Department of Clinical Medicine, Aalborg, Denmark; ¹⁴⁴Portuguese Oncology Institute, Department of Genetics, Porto, Portugal; ¹⁴⁵FOM - the FIRG Institute of Molecular Oncology, Genome Diagnostics Program, Milan, Italy; ¹⁴⁶Medical University of Vienna, Dept of OB/GYN and Comprehensive Cancer Center, Vienna, Austria; ¹⁴⁷IDIBELL (Bellvitge Biomedical Research Institute), Catalan Institute of Oncology, ProCURE, Barcelona, Spain; ¹⁴⁸Fondazione IRCCS Istituto Nazionale dei Tumori (INT), Unit of Molecular Bases of Genetic Risk and Genetic Testing, Department of Research, Milan, Italy; ¹⁴⁹Klinikum rechts der Isar der Technischen Universität München, Department of Gynaecology and Obstetrics, Munich, Germany; ¹⁵⁰University of NSW Sydney, School of Women's and Children's Health, Faculty of Medicine, Sydney, NSW, Australia; ¹⁵¹The Kinghorn Cancer Centre, Garvan Institute of Medical Research, Sydney, NSW, Australia; ¹⁵²University of NSW Sydney, Adult Cancer Program, Lowy Cancer Research Centre, Sydney, NSW, Australia; ¹⁵³Karolinska Institutet, Clinical Genetics, Stockholm, Sweden; ¹⁵⁴Carmel Medical Center and Technion Faculty of Medicine, Clalit National Cancer Control Center, Haifa, Israel; ¹⁵⁵Yale School of Medicine, Chronic Disease Epidemiology, New Haven, CT, USA; ¹⁵⁶Region of Southern Denmark, Vejle Hospital, Department of Clinical Genetics, Vejle, Denmark; ¹⁵⁷Wexner Medical Center, The Ohio State University, Department of Gynecology and Obstetrics, Columbus, OH, USA; ¹⁵⁸Institut de Cancérologie Strasbourg Europe, ICANS, Strasbourg, France; ¹⁵⁹Université de Strasbourg, Laboratoire d'ImmunoRhumatologie Moléculaire, Plateforme GENOMAX, INSERM UMR_S 1109, LabEx TRANSPLANTEX, Fédération de Médecine Translationnelle de Strasbourg (FMTS), Faculté de Médecine, Strasbourg, France; ¹⁶⁰The Ohio State University, Clinical Cancer Genetics Program, Division of Human Genetics, Department of Internal Medicine, The Comprehensive Cancer Center, Columbus, OH, USA; ¹⁶¹University of Kansas Medical Center, Department of Internal Medicine, Division of Medical Oncology,

ARTICLE

Westwood, KS, USA; ¹⁶²City of Hope, Clinical Cancer Genomics, Duarte, CA, USA; ¹⁶³Monash University, Precision Medicine, School of Clinical Sciences at Monash Health, Clayton, VIC, Australia; ¹⁶⁴The University of Melbourne, Department of Clinical Pathology, Melbourne, VIC, Australia; ¹⁶⁵Cancer Council Victoria, Cancer Epidemiology Division, Melbourne, VIC, Australia; ¹⁶⁶Hannover Medical School, Institute of Human Genetics, Hannover, Germany; ¹⁶⁷INSERM U830, Department of Tumour Biology, Paris, France; ¹⁶⁸Université Paris Descartes, Paris, France; ¹⁶⁹University Hospital Heidelberg, Institute of Human Genetics, Heidelberg, Germany; ¹⁷⁰University of Porto, Biomedical Sciences Institute (ICBAS), Porto, Portugal; ¹⁷¹Cancer Research Malaysia, Breast Cancer Research Programme, Subang Jaya, Selangor, Malaysia; ¹⁷²University of Malaya, Department of Surgery, Faculty of Medicine, Kuala Lumpur, Malaysia; ¹⁷³Magee-Womens Hospital, University of Pittsburgh School of Medicine, Department of Medicine, Pittsburgh, PA, USA; ¹⁷⁴The Ohio State University, Department of Cancer Biology and Genetics, Columbus, OH, USA; ¹⁷⁵University Of Melbourne, Department of Medicine, Melbourne, VIC, Australia; ¹⁷⁶Beth Israel Deaconess Medical Center, Department of Medical Oncology, Boston, MA, USA; ¹⁷⁷Centro de Investigación en Red de Enfermedades Raras (CIBERER), Madrid, Spain; ¹⁷⁸Fundación Pública Galega de Medicina Xenómica, Santiago de Compostela, Spain; ¹⁷⁹Instituto de Investigación Sanitaria de Santiago de Compostela (IDIS), Complejo Hospitalario Universitario de Santiago, SERGAS, Santiago de Compostela, Spain; ¹⁸⁰University Hospital Ulm, Department of Gynaecology and Obstetrics, Ulm, Germany; ¹⁸¹Mayo Clinic, Department of Oncology, Rochester, MN, USA; ¹⁸²Faculty of Medicine and University Hospital Cologne, University of Cologne, Center for Molecular Medicine Cologne (CMMC), Cologne, Germany; ¹⁸³Centre for Cancer Genetic Epidemiology, Department of Oncology, University of Cambridge, Cambridge, UK.

GEMO Study Collaborators

Pascaline Berthet²², Chrystelle Colas⁴², Marie-Agnès Collonge-Rame⁴³, Capucine Delnatte⁴⁴, Laurence Faivre⁴⁵, Sophie Giraud⁴⁶, Christine Lasset⁴⁷, Véronique Mari⁴⁸, Noura Mebirouk⁸, Emmanuelle Mouret-Fourme⁴², Hélène Schuster⁴⁹ and Dominique Stoppa-Lyonnet⁴²

EMBRACE Collaborators

Julian Adlard³, Munaza Ahmed⁴, Antonis Antoniou¹, Daniel Barrowdale¹, Paul Brennan³¹, Carole Brewer³², Jackie Cook⁵⁰, Rosemarie Davidson⁵¹, Douglas Easton^{1,183}, Ros Eeles⁵², D. Gareth Evans^{53,54}, Debra Frost¹, Helen Hanson⁴⁵, Louise Izatt⁵⁵, Kai-ren Ong⁵⁶, Lucy Side⁵⁷, Aoife O'Shaughnessy-Kirwan⁵⁸, Marc Tischkowitz^{59,60} and Lisa Walker⁶¹

kConFab Investigators

Georgia Chenevix-Trench¹⁰⁰, Kelly-Anne Phillips^{37,38,101,102} and Amanda Spurdle¹⁰⁰

HEBON Investigators

Marinus Blok²⁵, Peter Devilee⁶³, Frans Hogervorst⁹³, Maartje Hooning⁹⁴, Marco Koudijs¹⁰³, Arjen Mensenkamp¹⁰⁴, Hanne Meijers-Heijboer¹⁰⁵, Matti Rookus² and Klaartje van Engelen¹⁰⁶

GENEPSO Investigators

Nadine Andrieu^{6,7,8,9} and Catherine Noguès¹⁰⁷