

Association of 1-deoxy-sphingolipids with steatosis but not steatohepatitis nor fibrosis in non-alcoholic fatty liver disease

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Title page

Title: Association of 1-deoxy-sphingolipids with steatosis but not steatohepatitis nor fibrosis in Non-Alcoholic Fatty Liver Disease

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Authors: J. Weyler* [1,2], A. Verrijken* [2,3], T. Hornemann* [4,5], L. Vonghia[1,2], E. Dirinck[2,3], A. von Eckardstein[4], T. Vanwolleghem[1,2], P. Michielsen[1,2], F. Peiffer[2,3], A. Driessen[6], G. Hubens[6], B. Staels[8], S. Francque[# 1,2], L. Van Gaal[# 2,3]

* shared first authorship

[#] shared senior authorship

Affiliations:

[1] Department of Gastroenterology and Hepatology, Antwerp University Hospital, Edegem, Belgium

[2] Laboratory of Experimental Medicine and Paediatrics, University of Antwerp, Belgium

[3] Department of Endocrinology, Diabetology and Metabolic Diseases, Antwerp University Hospital, Edegem, Belgium

[4] Institute for Clinical Chemistry, University Hospital Zurich and Center for Integrative Human Physiology, University of Zurich, Switzerland

[5] Competence Center for Systems Physiology and Metabolic Diseases, University of Zurich, Switzerland

[6] Department of Pathology, Antwerp University Hospital, Edegem, Belgium

[7] Department of Abdominal Surgery, Antwerp University Hospital, Edegem, Belgium

[8] Univ. Lille, Inserm, CHU Lille, Institut Pasteur de Lille, U1011-EGID, F-59000 Lille, France

Corresponding author: prof. dr. Sven Francque

Department of Gastro-enterology and Hepatology

Wilrijkstraat 10

2650 Edegem

Belgium

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45

List of abbreviations

ALP	Alkaline phosphatase
ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
BMI	Body Mass Index
C_{16}-SA	C_{16}-sphinganine
C_{16}-SO	C_{16}-sphingosine
C_{17}-SO	C_{17}-sphingosine
C_{18}-SA	C_{18}-sphinganine
C_{18}-SO	C_{18}-sphingosine
C_{18}-phyto-SO	C_{18}-phyto-sphingosine
C_{19}-SO	C_{19}-sphingosine
C_{20}-SA	C_{20}-sphinganine
C_{20}-SO	C_{20}-sphingosine
CD	Confidence Distribution
CI	Confidence Interval
deoxy-SA	deoxy-sphinganine
deoxy-SO	deoxy-sphingosine
DSB	1-Deoxy Sphingoid Base
dSL	deoxy SphingoLipids
FLIP	Fatty Liver Inhibition of Progression
γGT	gamma Glutamyl Transpeptidase
HbA_{1c}	Haemoglobin A_{1c}
HDL-C	High Density Lipoprotein-Cholesterol

HOMA	Homeostatic Model Assessment
IFG	Impaired Fasting Glucose
IGT	Impaired Glucose Tolerance
IR	Insulin Resistance
LDL-C	Low Density Lipoprotein-Cholesterol
MetS	Metabolic Syndrome
NAFL	Non-Alcoholic Fatty Liver
NAFLD	Non-Alcoholic Fatty Liver Disease
NASH	Non-Alcoholic Steatohepatitis
NASH ≥ F2	Non-Alcoholic Steatohepatitis with significant fibrosis
OGTT	Oral Glucose Tolerance Test
PPAR	Peroxisome Proliferator-Activated Receptor
SA	Sphinganine
SA-dienine	Sphingadienine
SO	Sphingosine
S1P	Sphingosine-1-Phosphate
SphK	Sphingosine Kinase
SPT	Serine-Palmitoyl Transferase
TG	TriGlycerides
T2DM	Type 2 Diabetes Mellitus
WC	Waist Circumference

Abstract

Background: Non-Alcoholic Fatty Liver Disease (NAFLD) is the most important cause of chronic liver disease in the western world. In some individuals liver steatosis can be accompanied by inflammation and cell damage (Non-Alcoholic Steatohepatitis, NASH), and even liver fibrosis. Sphingolipids are a heterogeneous class of lipids and essential components of the plasma membrane and plasma lipoproteins. The atypical class of deoxy-sphingolipids have been implicated in the metabolic syndrome and type 2 diabetes.

Aim: To determine if circulating (deoxy)sphingolipids are associated with NAFLD and its different entities, steatosis, inflammatory changes (inflammation and ballooning) and fibrosis.

Methods: Sphingolipids were analysed by LC-MS after hydrolysing the N-acyl and O-linked headgroups in plasma of obese adults who underwent a liver biopsy in suspicion of NAFLD.

Results: 288 patients were included, liver status was as follows: 17.7% control, 16.3% NAFL (Non-Alcoholic Fatty Liver or isolated steatosis), 51.7% NASH without significant fibrosis and 14.3% NASH with significant fibrosis. There was no association between typical sphingolipids and NAFLD and its different entities. There was a statistically significant association between the presence of steatosis and the concentrations of deoxy-sphinganine (exp(B) 11.163 with CI [3.432, 36.306] and $p<0.001$) and deoxy-sphingosine (exp(B) 8.486 with CI [3.437, 20.949] and $p<0.001$). There was no association between these deoxy-sphingolipids and activity of the steatohepatitis, nor was there any association with fibrosis. Differences in deoxy-sphingolipids also correlated independently with the presence of the metabolic syndrome, but not diabetes.

Conclusion: Deoxy-sphingolipids are elevated in patients with steatosis compared to those without fatty liver, but not different between the different NAFLD subtypes, suggesting that deoxy-sphingolipid bases might be involved in steatogenesis, but not in the further progression of NAFLD to NASH nor in fibrogenesis.

Introduction

NAFLD

Non-Alcoholic Fatty Liver Disease (NAFLD) is the most important cause of chronic liver disease in the western world. In some individuals liver steatosis can be accompanied by liver cell damage, inflammation, and even liver fibrosis. The latter can lead to cirrhosis which can further result into end-stage liver disease[1]. The pathogenesis of NAFLD is still not completely elucidated. Presumably, the integrated action of numerous conditions acting in parallel (genetic predisposition, adipose tissue dysfunction, insulin resistance (IR), oxidative stress, lipotoxicity, gut dysbiosis…) results in NASH, giving rise to the multiparallel hit hypothesis[2]. Different studies have shown that fibrosis grade is the strongest predictor for hepatic- and extra-hepatic complications[3, 4], whereas steatohepatitis is considered the driving force of disease progression and adverse outcomes.

Sphingolipids

Sphingolipids are a heterogeneous class of bioactive lipids with a plethora of functions playing important roles in almost all major aspects of cell biology including metabolism, inflammation, autophagy and cell adhesion and migration[5]. Furthermore, they are major components of cell membranes and contribute to plasma lipoprotein formation. The enzyme Serine-Palmitoyl Transferase (SPT) catalyses the first and rate-limiting step in the *de novo* synthesis of sphingolipids, which converts L-serine and palmitoyl-CoA into 3-keto-sphinganine. The latter is converted into sphinganine (SA). SA is N-conjugated with another fatty acid to form dihydro-ceramide. When desaturated at C4 it forms ceramide, the building block for the more complex sphingolipids (Fig. 1)[6]. Ceramidase converts ceramide into sphingosine (SO), the most abundant long-chain sphingoid base in mammalian cells.

Sphingosine-1-Phosphate (S1P) is formed after phosphorylation of SO by two Sphingosine Kinases (SphK1 and -2)[7]. In plasma, SO is the most common sphingoid base, followed by sphingadienine (SA-dienine) and SA[8]. Ceramide and S1P have been implicated in key steps of the pathophysiology of NAFLD in animal models[7]. Data in humans are, however, scarce. L-serine and palmitoyl-CoA are the preferred substrates for SPT, however, other substrates can be used, including acyl-CoA with carbon chain lengths ranging from C12 to C18. Furthermore, SPT can use other amino acid substrates beside L-serine: L-alanine and to a certain extent L-glycine. The use of these alternative substrates generates a category of atypical sphingolipids: the 1-Deoxy Sphingolipids (dSLs) that lack the C1-OH group of the regular sphingolipids (Fig. 1). dSLs are not metabolised to form complex sphingolipids nor are they degraded by the regular sphingoid catabolism pathway[9]. The function of these dSLs is unknown. dSL levels were shown to be increased in plasma of patients with the Metabolic Syndrome (MetS) and Type 2 Diabetes Mellitus (T2DM)[9]. Furthermore, they could play a role in the pathogenesis of T2DM with the dSLs being cytotoxic for insulin producing cells as they induce senescence and multiple cell death pathways[10]. NAFLD, MetS and T2DM are closely related and originate from the same underlying pathological processes, such as IR, lipotoxicity, dyslipidaemia and chronic inflammation[11]. Since there is a correlation between the MetS, T2DM and (deoxy)sphingolipids, on the one hand, and a close relationship between the MetS, T2DM and NAFLD, on the other hand, we aimed at investigating whether :

1. circulating (1-deoxy) sphingoid bases are associated with NAFLD and its different components, steatosis, cell damage & inflammatory changes (lobular inflammation and ballooning) and fibrosis;

2. associations between deoxy-sphingolopids, on the one hand, and the MetS or T2DM, on the other hand, are driven by the presence of NAFLD and its different components.

Patients & Methods

Study participants

The protocol of patient selection has previously been described[12]. Briefly, patients visiting the obesity clinic of the Antwerp University Hospital were consecutively recruited. Every patient underwent a metabolic and hepatologic work-up both approved by the Ethics Committee of the Antwerp University Hospital (reference 6/25/125, Belgian registration number B30020071389) in order to screen for NAFLD. Subjects had to be 18 years or older. As T2DM harbours potential confounders, patients with a known history of diabetes were excluded. However, patients were included if diabetes was *newly* diagnosed. Sample collection for sphingolipid analysis was taken before any glucose lowering therapy was started. Patients with significant alcohol consumption were excluded. A liver biopsy was proposed if there was a suspicion of liver disease, these were mainly performed outside the setting of bariatric surgery in order to exclude severe liver abnormalities prior to surgery. Only patients who underwent a liver biopsy were included for this study. Biopsies deemed insufficient for adequate reading by the pathologist were excluded from further analysis. The different pathological features of NAFLD were scored according to both the NASH Clinical Research Network Scoring System and the Fatty Liver Inhibition of Progression (FLIP)-algorithms[13, 14]. The diagnosis of NASH required the combined presence of steatosis, ballooning and lobular inflammation. Based on liver histology, patients were classified in 4 distinct groups reflecting different stages of severity of the disease:

- *No-NAFLD*: no histological evidence for steatosis, inflammation, ballooning and fibrosis
- *Isolated steatosis* (or NAFL): histological evidence of steatosis with activity score (ballooning+inflammation) ≤1 and fibrosis stage ≤1
- *NASH without significant fibrosis (fibrosis stage (F)<2)*: histological evidence of steatosis with ballooning score ≥1, inflammation score ≥1 and fibrosis stage ≤1
- *NASH with significant fibrosis (≥F2) (so called "fibrotic NASH")*: histological evidence of steatosis with ballooning score ≥1, inflammation score ≥1 and fibrosis stage ≥2.

Patients with histological evidence of a liver disease other than NAFLD or patients with NAFLD but who could not be unequivocally categorised in one of the 4 groups were excluded.

Sphingolipid analysis

The protocol for analysing plasma sphingoid bases has previously been described[15]. Briefly, sphingoid bases were determined in fasting plasma samples. Sphingolipids in plasma are present in a broad variation of subspecies. They can be saturated and are usually acylated with another fatty acid. Furthermore, most of these sphingolipids are conjugated to an O-linked head group. To analyse the sphingoid backbones, the sphingolipids were subjected to a sequential acid and base hydrolysis. Acid hydrolysis specifically breaks the N-alkyl chain, whereas alkaline conditions lead to a release of the O-linked head group. The sphingoid bases were analysed using Liquid Chromatography/Mass Spectrometry. Analysed bases included C_{16}-sphinganine (C_{16}-SA), C_{16}-sphingosine (C_{16}-SO), C_{17}-sphingosine (C_{17}-SO), C_{18}-sphinganine (C_{18}-SA), C_{18}-sphingosine (C_{18}-SO), C_{19}-sphingosine (C_{19}-SO), C_{20}-sphinganine

211 (C_{20}-SA), C_{20}-sphingosine (C_{20}-SO), sphingadienine (SA-dienine), C_{18}-phyto-sphingosine (C_{18}-

212 phyto-SO), deoxy-sphinganine (deoxy-SA) and deoxy-sphingosine (deoxy-SO).

213

Statistical analysis

1. The role of sphingolipids in the different entities of NAFLD

Data were analysed with SPSS version 24. Means were compared by independent t testing or non-parametric tests when appropriate. To investigate the association of sphingoid bases with the different entities of NAFLD (steatosis, steatohepatitis and fibrosis), the following groups were compared: 'No-NAFLD' *vs.* 'NAFL' to investigate the association of the sphinghoid bases in steatosis; 'NAFL' *vs.* 'NASH <F2' to investigate the potential association of the sphingoid bases in steatohepatitis and eventually 'NASH <F2' *vs.* 'NASH ≥F2' to investigate the potential association of the sphingoid bases in fibrogenesis. Sphingoid bases that showed association with NAFLD (and its stages) in this first crude analysis were further analysed using binary logistic regression. Since NAFLD is closely related to the MetS and diabetes mellitus type 2, interaction with or confounding is possible. Furthermore, certain lipid lowering therapies can have a profound effects on NAFLD. Adjustment was done using metabolic markers (*e.g.* homeostasis model assessment (HOMA)-IR, presence of the MetS, presence of T2DM or Impaired Fasting Glucose (IFG) and the use of lipid lowering therapy) as covariates and/or interaction terms in a binary logistic regression.

2. *Deoxy-sphingoid bases and metabolic parameters*

To investigate whether the associations between deoxy-sphingolopids, on the one hand, and the MetS or T2DM, on the other hand, are driven by NAFLD, the possible confounding/interaction effect of NAFLD was checked using binary logistic regression analysis. Furthermore, correlations between deoxy-sphingolipids and other metabolic parameters (*e.g.* triglycerides and HOMA-IR) were checked. To investigate these correlations, Pearson's correlation coefficient was used when appropriate. If not, non-

237 parametric testing was performed: Kendall's tau-b was used if there were a great number of
238 tied ranks, in others cases Spearman's rho was used.

239

240

RESULTS

1. The role of sphingolipids in the different entities of NAFLD

288 consecutive patients with a complete dataset were included. Their liver phenotype was as follows: 17.7% No-NAFLD, 16.3% NAFL, 51.7% NASH <F2 and 14.3% NASH ≥F2. This is an obese study population with a mean BMI of 39.6 kg/m^2 (SD 6.4) (Table 1). Log transformation of the concentration of sphingoid bases was used in statistical analysis since the distribution of these bases was skewed (resulting in extreme exp (B) values).

1a. Sphingoid bases in No-NAFLD vs. NAFL

There were 47 patients with isolated liver steatosis and 51 (26.6%) patients with no histological evidence for the presence of NAFLD (No-NAFLD). There was a statistically significant difference in age, waist and HbA_{1c}, but there was no difference in BMI, presence of the Mets and presence of T2DM. There was no use of fibrates and statin use was not significantly different in both groups (Supplementary Table 1). There was a statistically significant association between the presence of steatosis and the concentrations of deoxy-SA (exp(B) 6.708 with CI [1.243, 36.195] and p 0.027) and deoxy-SO (exp(B) 4.851 with CI [1.315, 17.890] and 0.018). There was no interaction effect of metabolic markers in the association of steatosis and the sphingoid bases deoxy-SA and deoxy-SO, and statistical significance was maintained for the DSBs (deoxy-SA and deoxy-SO) even after correction for confounding (*e.g.* by metabolic markers).

1b. Sphingoid bases in NAFL vs. NASH <F2

149 patients had NASH, compared to 47 patient with isolated steatosis. There was a statistically significant difference in the presence of the MetS, HOMA-IR and liver enzymes (Supplementary Table 2). There was a statistically significant difference in plasma concentrations of C_{16}-SO, C_{17}-SO, C_{18}-SO and SA-dienine. Further analysis did, however, not show an association between NASH and C_{16}-SO (exp(B) 0.159 with CI [0.016, 1.595] and p=0.118) nor C_{17}-SO (exp(B) 0.106 with CI [0.008, 1.342] and p=0.083). There was a statistically significant association between NASH and C_{18}-SO (exp(B) 0.036 with CI [0.001, 0.991] and p 0.049) and SA-dienine (exp(B) 0.037 with [0.002, 0.725] and p 0.030). There was no interaction effect by the metabolic markers. However, after correction for the presence of the MetS there was a loss of the statistically significant association between NASH, on the one hand, and C_{18}-SO (exp(B) 0.043 with CI [0.001, 1.405] and p 0.077) and SA-dienine (exp(B) 0.062 with CI [0.003, 1.414] and p 0.081), on the other hand.

1c. Sphingoid bases in NASH <F2 vs. NASH ≥F2

41 patients with NASH had evidence of significant fibrosis (fibrotic NASH), compared to 149 (26.6%) NASH patients who had no significant fibrosis (Supplementary Table 3). There was a statistically significant association between fibrotic NASH and the concentrations of C_{19}-SO (exp(B) 0.157 with CI [0.025, 0.986] and p=0.048) and SA-dienine (exp(B) 0.013 with CI [0.001, 0.342] and p=0.009). There was no interaction between metabolic markers and the sphingoid bases C_{19}-SO and SA-dienine. After correcting for HOMA-IR, however, there was loss of statistical significance for the association between fibrotic NASH and C_{19}-SO (exp(B) after correction 0.203 with CI [0.028, 1.495] and p=0.118). Furthermore, there was a loss of

statistical significance for the association between fibrotic NASH and SA-dienine after correcting for gender (exp(B) after correction 0.048 with CI [0.001, 1.213] and p=0.065).

2. Deoxy-sphingoid bases and metabolic - parameters

Log transformation of the concentration of sphingoid bases was used in binary regression analysis since the distribution of these bases was skewed (resulting in extreme exp (B) values).

2a. Metabolic syndrome

There was a highly statistically significant association between MetS and the concentrations of deoxy-SA (exp(B) 29.934 with CI [8.457, 105.949] and p <0.001) and deoxy-S0 (exp(B) 29.024, with CI [9.796, 85.998] and p <0.001). There was no interaction with the metabolic markers (including T2DM) nor with NAFLD, nor was there loss of statistical significance after correction for the metabolic markers (including T2DM and HOMA-IR) nor NAFLD

2b. Type 2 diabetes mellitus

There was a highly statistically significant association between the presence of T2DM and the concentrations of deoxy-SA (exp(B) 14405.280 with CI [13.084, 15860091.190] and p 0.007) and deoxy-S0 (exp(B) 7.953 with CI [1.832, 34.515] and p 0.006). There was no interaction with metabolic markers (including MetS) nor with NAFLD. After correction for the MetS, however, there was a loss of a statistically significant association between T2DM and deoxy-SA (exp(B) after correction 1.964 with CI [0.275, 14.048] and p=0.501) and deoxy-SO (exp(B) after correction 2.138 with CI [0.400, 11.432] and p=0.374).

2c. Correlation with other metabolic parameters

307 There was a statistically significant correlation between the DSBs and triglycerides (Fig 2A
308 and 2B; Spearman's rho 0.553 and p <0.001 for deoxy-SA and Spearman's rho 0.581and p
309 <0.001 for deoxy-SO) and between deoxy-SA and IR. There was no statistically significant
310 correlation between deoxy-SO and IR (Fig 2C and 2D; Spearman's rho 0.1138 and p 0.021 for
311 deoxy-SA and Spearman's rho 0.104 and p 0.081 for deoxy-SO) (Table 2).

312

Discussion

Sphingolipids are essential components of the cell membrane and plasma lipoproteins. They play a complex role as signalling molecules in multiple processes and have been suggested to be of pathophysiological relevance in NASH. dSLs, an atypical category of sphingolipids, are generated when SPT uses L-alanine instead of L-serine. dSLs are neither metabolised to complex sphingolipids nor degraded by the sphingolipid catabolism pathways. There function is not clear, but they seem involved in T2DM and the MetS.

In a large, well characterised, prospectively screened cohort with ultimately histological diagnosis of NAFLD we observed a highly significant association between NAFLD and 1-deoxy sphingoid bases (DSBs). We found no statistically significant difference between the different stages of NAFLD, suggesting that dSLs are related to steatosis, but are not implicated in the further progression of NAFLD to NASH and not associated with fibrosis. Furthermore, this study confirms the association of deoxy-sphingolipids with the MetS but not diabetes.

In a study with 99 individuals, Bertea et al. found a statistical significant difference in plasma dSLs levels when comparing T2DM patients with control. Although there was a clear difference between these groups regarding gender, BMI and other metabolic markers, there was no correction performed, questioning these findings. Furthermore, they described a possible predictive role of dSLs for the presence of T2DM, although this case-control study did not lean itself to perform prediction analysis [9]. Also Othman et al investigated the role of dSLs in the MetS and T2DM. They found a significant difference between plasma levels of dSLs when comparing healthy volunteers with patients with the MetS or patients with T2DM, but found no difference between plasma levels of dSLs when comparing T2DM with

MetS. In this study, there was a statistically significant difference in BMI and waist circumference when comparing patients with T2DM to those with the Mets, and a (near statistically) significant difference when comparing triglycerides, but there was no correction performed [16].

Human data regarding the association between sphingolipids, more specifically, dSLs on the one hand, and the presence of NAFLD, on the other, is scarce. Gorden *et al* described a diverse panel of 20 plasma lipids, including the dSL 1-deoxy-ceramide, capable of distinguishing NAFLD from NASH in 88 patients with liver histology categorised as normal, steatotic, NASH, or cirrhotic. Although patients were selected on an "all-comers" approach where liver biopsy was obtained during surgery (gastric bypass, liver transplant, multi organ transplant, hernia repair…), the study population was obese with a mean fasting glucose > 100 mg/dL (other metabolic conditions were not mentioned). The possible effects of these conditions on the results was not assessed, meaning that an overestimation of the discriminate power of these lipids is possible [17]. Gai *et al.* reported a correlation between NAFLD and dSLs in 80 individuals, but failed to specify the contribution of these dSLs in the different entities of NAFLD. Furthermore, they did not provide a clear overview of the main (metabolic) characteristics of these patients and failed to check for the confounding or interactive effect of metabolic factors [18]. The role of dSLs in the MetS, T2DM and NAFLD is not fully understood, but they seem to play a role in the control of metabolism. A study by Zuellig *et al.* showed that dSLs compromised insulin secretion and triggered senescence and cell-death in insulin producing cells triggered by multiple pathways, including cytoskeletal remodelling, senescence, necrosis and apoptosis favouring hyperglycaemia. [10] Although we found an increasing presence of *de novo* T2DM with a more severe phenotype of NAFLD, there was no significant interaction between *de novo* T2DM nor IR and dSLs when

investigating the association with steatosis, nor was there loss of significance after correcting for T2DM or IR (data for T2DM not shown) suggesting that hyperglycaemia or IR *per se* is not a determinant of dSL formation in the pathophysiology of NAFLD. On the other hand, dSLs do not significantly differ between the 3 NAFLD categories whereas IR and impairment of glycaemic control worsen as disease worsens (despite comparable age and BMI). This observation hence does not support an important role for dSLs in impairment of glucose metabolism in NAFLD. Wei *et al.* also speculated that the formation of dSLs in T2DM is not caused by hyperglycaemia *per se*, but rather associated with metabolic changes in T2DM [19].

Another possible explanation in the association between dSLs and NAFLD is a shift in the utilization of alanine instead of serine by SPT. Mardinoglu *et al.* revealed a L-serine deficiency in NAFLD using a genome scale metabolic model of hepatocytes. As seen in literature, there was a significant association between T2DM and DBSs[9] and between the MetS and dSLs[16], which was also the case in our population. However, after correcting for the MetS, there is a loss of statistical significance in the association between dSLs and T2DM. Furthermore, this association between dSLs and MetS is not driven by the presence of NAFLD (since the lack of a significant interaction). Although it is not clear what exact role the DSBs play in steatosis, they seem to be associated with metabolic dysregulation, which might eventually lead to steatosis.

Sphingolipid metabolism has been proposed as a target for pharmacological therapy of NASH. Othman *et al* showed that fibrates have a DSBs lowering effect, independent of a triglyceride lowering effect [19]. It is not clear whether this is a peroxisome proliferator-activated receptor (PPAR) α or non-PPARα mediated effect[20]. Gai et al. described that the

activation of Farnesoid X Receptor reduces dSLs plasma levels in a high fat-fed mouse model and in 1-deoxysphinanine–treated mice [18]. Another group found that oral supplementation of serine suppressed the formation of dSLs in an hereditary sensory and autonomic neuropathy type 1 (HSAN1) animal model and in humans[21, 22]. Serine palmitoyltransferase long chain base subunit 1 and 2 mutations in SPT (as seen in HSAN1) induce a permanent shift in the substrate preference from L-serine to L-alanine[6]; dSLs formation in metabolic diseases is, however, not caused by a mutation in SPT but by a dysregulation in carbohydrate and fatty acid metabolism [19]. Our data are linking dSLs more to steatosis than to steatohepatitis and fibrosis (at least in a cross-sectional analysis). Steatosis is not *per se* harmful and can even be protective, questioning the therapeutic potential of modulating dSLs in the treatment of NASH. Nevertheless, reducing steatosis can be a target for NAFLD treatment, given the presumed role of lipotoxicity. The exact potential impact of modulating sphingolipid metabolism on NASH progression is hence to be determined.

In this study, patients were not included based on a *a priori* suspicion of liver disease .This selection procedure resulted in a series representing the whole spectrum of NAFLD, whereas retrospective series of biopsy proven NAFLD patients tend to be skewed towards more severe subtypes. On the other hand, analysis was performed cross-sectionally on a population that is predominantly Caucasian. A causal link between deoxy-sphingolipids and steatosis is therefore difficult to determine. Finally, the reported data only handles the sphingoid base concentrations. Although they reflect the total sphingoid base composition for all the individual sphingolipid subclasses, further investigation of these different subclasses is needed to provide a more detailed understanding of the pathological role of

406 the sphingolipidome in NAFLD and its potential role in the development of therapeutic
407 agents.

408 **Conclusion:** dSLs are increased in relation to steatosis independently of their correlation
409 with metabolic markers, but are not further increased when the spectrum of NAFLD
410 worsens, questioning their role as a therapeutic target for NASH.

411

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Tables

Table 1: Patients characteristics according to the histology-based classification

		TOTAL N=288 (100%)	No-NAFL N=51 (17.7%)	NAFL N=47 (16.3%)	NASH <F2 N=149 (51.7%)	NASH ≥F2 N=41 (14.3%)	p-value
General	**Age (years)**	**43.8 (±12.3)**	**38.1 (±10.6)**	**43.5 (±12.3)**	**45.4 (±12.4)**	**45.0 (±12.4)**	**0.003***
	Gender: men	**96 (33.3%)**	**8 (15.7%)**	**10 (21.3%)**	**53 (35.6%)**	**25 (61.0%)**	**<0.001*****
Anthropometry	**BMI (kg/m^2)**	**39.6 (±6.4)**	**38.2 (±4.9)**	**39.9 (±7.1)**	**38.9 (±6.0)**	**43.5 (±7.4)**	**0.001****
	Waist (cm)	**118.8 (±13.2)**	**111.93 (±10.4)**	**118.1 (±13.6)**	**117.9 (±11.3)**	**132.1 (±13.1)**	**<0.001****
Metabolic markers	**TG (mg/dL)**	**155.5 (±78.5)**	**137.2 (±66.6)**	**150.3 (±106.9)**	**157.1 (±72.8)**	**177.9 (±69.4)**	**0.009****
	LDL-C (mg/dL)	124.7 (±36.8)	121.5 (±43.9)	128.8 (±33.2)	123.4 (±35.7)	128.5 (±35.5)	0.674*
	HDL-C (mg/dL)	**48.9 (±13.7)**	**53.2 (±13.4)**	**51.3 (±15.9)**	**48.6 (±13.5)**	**41.9 (±9.3)**	**<0.001****
	Statin use (%)	30 (10.4)	2 (3.9)	3 (6.4)	22 (14.8)	3 (7.3)	0.087***
	HbA1c (%)	**5.7 (±0.5)**	**5.4 (0.3)**	**5.6 (±0.3)**	**5.6 (±0.4)**	**6.1 (±1.0)**	**<0.001****
	Glucose mg/dL	**86.2 (±22.4)**	**80.2 (±7.4)**	**82.4 (±11.0)**	**85.1 (±12.4)**	**102.1 (±49.8)**	**<0.001****
	HOMA-IR	**4.0 (±3.2)**	**3.1 (±3.2)**	**2.9 (±1.8)**	**3.8 (±2.4)**	**6.8 (±5.1)**	**<0.001****
	MetS	**155 (56.2%)**	**20 (40.8%)**	**17 (37.8%)**	**87 (61.3%)**	**31 (77.5%)**	**<0.001*****
	T2DM	**30 (10.5%)**	**1 (2.0%)**	**3 (6.5%)**	**16 (10.9%)**	**10 (24.4%)**	**0.004*****
Liver enzymes	**AST (U/L)**	**31.7 (±17.5)**	**24.4 (±8.0)**	**25.5 (±8.5)**	**30.7 (±11.8)**	**51.5 (±31.5)**	**<0.001****
	ALT (U/L)	**45.6 (±27.3)**	**31.0 (±11.3)**	**36.1 (±18.7)**	**45.8 (±21.6)**	**74.0 (±43.0)**	**<0.001****
Sphingoid bases	C_{16}-SA	0.469 (±0.221)	0.460 (±0.227)	0.441 (±0.159)	0.491 (±0.249)	0.432 (±0.156)	0.673**
	C_{16}-SO	15.491 (±5.313)	15.991 (±5.880)	16.584 (±4.753)	15.379 (±5.568)	14.026 (±3.828)	0.067**
	C_{17}-SO	**6.233 (±2.007)**	**6.533 (±2.365)**	**6.665 (±1.686)**	**6.180 (±2.053)**	**5.555 (±1.498)**	**0.018****
	C_{18}-SA	3.659 (±1.401)	3.599 (±1.588)	3.512 (±1.229)	3.726 (±1.459)	3.662 (±1.126)	0.822*
	C_{18}-SO	**84.161 (±20.682)**	**86.927 (±24.394)**	**89.051 (±17.530)**	**83.308 (±21.246)**	**78.050 (±14.864)**	**0.008****
	C_{18}-phytoSO	0.110 (±0.038)	0.115 (±0.052)	0.114 (±0.029)	0.110 (±0.037)	0.100 (±0.026)	0.135**
	C_{19}-SO	1.815 (±0.794)	1.875 (±0.839)	1.814 (±0.702)	1.871 (±0.858)	1.541 (±0.508)	0.200**
	C_{20}-SA	**0.024 (±0.108)**	**0.021 (±0.010)**	**0.024 (±0.012)**	**0.025 (±0.110)**	**0.025 (±0.009)**	**0.024****

	C_{20}-SO	0.195 (±0.068)	0.188 (±0.079)	0.199 (±0.060)	0.198 (±0.071)	0.190 (±0.052)	0.388**
	SA-diene	**31.376 (±9.161)**	**33.168 (±11.686)**	**33.966 (±7.963)**	**31.063 (±8.688)**	**27.315 (7.080)**	**<0.001****
	Deoxy-SA	0.074 (±0.042)	0.064 (±0.040)	0.077 (±0.042)	0.076 (±0.042)	0.077 (±0.040)	0.099**
	Deoxy-SO	**0.110 (±0.075)**	**0.088 (±0.075)**	**0.106 (±0.062)**	**0.117 (±0.077)**	**0.118 (±0.076)**	**0.019***

*No histological evidence of Non-Alcoholic Fatty Liver Disease (No-NAFLD), Non-Alcoholic Fatty Liver (NAFL), Non-Alcoholic Steatohepatitis with no significant fibrosis (NASH <F2), Non-Alcoholic Steatohepatitis with significant fibrosis (NASH ≥ F2), Body mass index (BMI), Triglycerides (TG), Low density lipoprotein cholesterol (LLD-C), High density lipoprotein cholesterol (HDL-C), Haemoglobin A_{1c} (HbA_{1c}), Homeostatic model assessment insulin resistance (HOMA-IR), Metabolic syndrome (MetS), aspartate aminotransferase (AST), alanine aminotransferase (ALT), C_{16}-Sphinganine (C_{16}-SA), C_{16}-Sphingosine (C_{16}-SO), C_{17}-Sphingosine (C_{17}-SO), C_{18}-Sphinganine (C_{18}-SA), C_{18}-Sphingosine (C_{18}-SO), C_{19}-Sphingosine (C_{19}-SO), C_{20}-Sphinganine (C_{20}-SA), C_{20}-Sphingosine (C_{20}-SO), Sphingadiene (SA-diene), C_{18}-phyto-sphingosine (C_{18}-phyto-SO), deoxy-sphinganine (deoxy-SA), deoxy-sphingosine (deoxy-SO). SD or percentage of the total group is presented between brackets. p-values: *ANOVA, **Kruskal-Wallis, ***Pearson Chi-square.*

Table 2: Correlation with metabolic and inflammatory parameters

	Triglycerides		HOMA-IR	
	Correlation coefficient*	p	Correlation coefficient*	p
Deoxy-SA	**0. 553**	**<0.001**	**0.138**	**0.021**
Deoxy-SO	**0.581**	**<0.001**	0.104	0.081

** Spearman's rho. Homeostatic model assessment insulin resistance (HOMA-IR), deoxy-sphinganine (deoxy-SA) deoxy-sphingosine (deoxy-SO)*

Supplementary materials

Supplementary Table 1: Patient characteristics for the comparison No-NAFLD vs. NAFL

		No-NAFLD N=51	NAFL N=47	p
General	**Age (years)**	**38.1 (±10.6)**	**43.5 (±12.3)**	**0.021****
	Gender: men	8 (15.7%)	10 (21.3%)	0.475***
Anthropometry	BMI (kg/m^2)	38.2 (±4.9)	39.9 (±7.1)	0.190**
	Waist (cm)	**111.93 (±10.4)**	**118.1 (±13.6)**	**0.007****
Metabolic markers	TG (mg/dL)	137.2 (±66.6)	150.3 (±106.9)	0.814*
	LDL-C (mg/dL)	121.5 (±43.9)	128.8 (±33.2)	0.369**
	HDL-C (mg/dL)	53.2 (±13.4)	51.3 (±15.9)	0.415*
	Statin use (%)	2 (3.9)	3 (6.4)	0.580***
	HbA1c (%)	**5.4 (0.3)**	**5.6 (±0.3)**	**<0.001***
	Glucose (mg/dL)	80.2 (±7.4)	82.4 (±11.0)	0.611*
	HOMA-IR	3.1 (±3.2)	2.9 (±1.8)	0.652*
	MetS	20 (40.8%)	17 (37.8%)	0.763***
	T2DM	1 (2.0%)	3 (6.5%)	0.259***
Liver enzymes	AST (U/L)	24.4 (±8.0)	25.5 (±8.5)	0.254*
	ALT (U/L)	31.0 (±11.3)	36.1 (±18.7)	0.248*
Imaging	**Steatosis on US**	**30 (61.2%)**	**40 (95.2%)**	**<0.001*****
Pathology	**Steatosis 0/1/2/3 (%)**	**100/0.0/0.0/0.0**	**0.0/80.9/17.0/2.1**	**<0.001*****
	Ballooning 0/1/2 (%)	**100/0.0/0.0**	**68.1/31.9/0.0**	**<0.001*****
	Inflammation 0/1/2 (%)	**100/0.0/0.0**	**72.3/27.3/0.0**	**<0.001*****
	Fibrosis 0/1/2/3/4 (%)	100/0.0/0.0/0.0/0.0	93.6/6.4/0.0/0.0/0.0	0.067***
Sphingoid bases	C_{16}-SA	0.460 (±0.227)	0.441 (±0.159)	0.890*
	C_{16}-SO	15.991 (±5.880)	16.584 (±4.753)	0.384*
	C_{17}-SO	6.533 (±2.365)	6.665 (±1.686)	0.384*
	C_{18}-SA	3.599 (±1.588)	3.512 (±1.229)	0.773*
	C_{18}-SO	86.927 (±24.394)	89.051 (±17.530)	0.328*
	C_{18}-phytoSO	0.115 (±0.052)	0.114 (±0.029)	0.526**
	C_{19}-SO	1.875 (±0.839)	1.814 (±0.702)	0.897**
	C_{20}-SA	0.021 (±0.010)	0.024 (±0.012)	0.236*
	C_{20}-SO	0.188 (±0.079)	0.199 (±0.060)	0.281**

	SA-diene	33.168 (±11.686)	33.966 (±7.963)	0.387**
	Deoxy-SA	**0.064 (±0.040)**	**0.077 (±0.042)**	**0.023****
	Deoxy-SO	**0.088 (±0.075)**	**0.106 (±0.062)**	**0.014****

* Mann-Whitney U, ** independent sample T-test, *** chi-square. Ballooning and Inflammation are graded according to SAF/Flip-algorithm. No histological evidence of Non-Alcoholic Fatty Liver Disease (no-NAFLD), Non-Alcoholic Fatty Liver (NAFL), Significant fibrosis (≥ F2), Body mass index (BMI), Triglycerides (TG), Low density lipoprotein cholesterol (LLD-C), High density lipoprotein cholesterol (HDL-C), Haemoglobin A_{1c} (HbA_{1c}), Homeostatic model assessment insulin resistance (HOMA-IR), Metabolic syndrome (MetS), aspartate aminotransferase (AST), alanine aminotransferase (ALT), C_{16}-Sphinganine (C_{16}-SA), C_{16}-Sphingosine (C_{16}-SO), C_{17}-Sphingosine (C_{17}-SO), C_{18}-Sphinganine (C_{18}-SA), C_{18}-Sphingosine (C_{18}-SO), C_{19}-Sphingosine (C_{19}-SO), C_{20}-Sphinganine (C_{20}-SA), C_{20}-Sphingosine (C_{20}-SO), Sphingadiene (SA-diene), C_{18}-phyto-sphingosine (C_{18}-phyto-SO), deoxy-sphinganine (deoxy-SA), deoxy-sphingosine (deoxy-SO). SD or percentage of the total group is presented between brackets.

Supplementary Table 2: patient characteristics for the comparison NAFL vs. NASH

		NAFL N=47	NASH <F2 N=149	p
General	Age (years)	43.5 (±12.3)	45.4 (±12.4)	0.350**
	Gender: men	10 (21.3%)	53 (35.6%)	0.067***
Anthropometry	BMI (kg/m^2)	39.9 (±7.1)	38.9 (±6.0)	0.486*
	Waist (cm)	118.1 (±13.6)	117.9 (±11.3)	0.952**
Metabolic markers	TG (mg/dL)	150.3 (±106.9)	157.1 (±72.8)	0.113*
	LDL-C (mg/dL)	128.8 (±33.2)	123.4 (±35.7)	0.373**
	HDL-C (mg/dL)	51.3 (±15.9)	48.6 (±13.5)	0.285*
	Statin use (%)	3 (6.4)	22 (14.8)	0.133***
	HbA1c (%)	5.6 (±0.3)	5.6 (±0.4)	0.514*
	Glucose (mg/dL)	82.4 (±11.0)	85.1 (±12.4)	0.151*
	HOMA-IR	**2.9 (±1.8)**	**3.8 (±2.4)**	**0.026***
	MetS	**17 (37.8%)**	**87 (61.3%)**	**0.006*****
	T2DM	3 (6.5%)	16 (10.9%)	0.386***
Liver enzymes	**AST (U/L)**	**25.5 (±8.5)**	**30.7 (±11.8)**	**0.005***
	ALT (U/L)	**36.1 (±18.7)**	**45.8 (±21.6)**	**0.001***
Imaging	Steatosis on US	40 (95.2%)	139 (96.5%)	0.699***
Pathology	**Steatosis 0/1/2/3 (%)**	**0.0/80.9/17.0/2.1**	**0.0/36.9/38.3/24.8**	**<0.001*****
	Ballooning 0/1/2 (%)	**68.1/31.9/0.0**	**0.0/50.3/49.7**	**<0.001*****
	Inflammation 0/1/2 (%)	**72.3/27.3/0.0**	**0.0/58.4/41.6**	**<0.001*****

	Fibrosis 0/1/2/3/4 (%)	**93.6/6.4/0.0/0.0/0.0**	**70.5/29.5/0.0/0.0/0.0**	**0.001*****
Sphingoid bases	C_{16}-SA	0.434 (±0.141)	0.478 (±0.233)	0.469*
	C_{16}-SO	**16.212 (±4.426)**	**15.087 (±5.262)**	**0.031***
	C_{17}-SO	**6.511 (±1.610)**	**6.045 (±1.960)**	**0.023***
	C_{18}-SA	3.464 (±1.238)	3.712 (±1.392)	0.689*
	C_{18}-SO	**86.492 (±17.300)**	**82.173 (±20.123)**	**0.005***
	C_{18}-phytoSO	0.111 (±0.029)	0.108 (±0.035)	0.229*
	C_{19}-SO	1.760 (±0.667)	1.800 (±0.806)	0.839**
	C_{20}-SA	0.023 (±0.011)	0.025 (±0.010)	0.315**
	C_{20}-SO	0.194 (±0.060)	0.196 (±0.067)	0.773**
	SA-diene	**32.700 (±7.803)**	**30.254 (±8.491)**	**0.028****
	Deoxy-SA	0.079 (±0.041)	0.077 (±0.042)	0.867**
	Deoxy-SO	0.115 (±0.075)	0.117 (±0.077)	0.441**

* Mann-Whitney U, ** independent sample T-test, *** chi-square. Ballooning and Inflammation are graded according to SAF/Flip-algorithm. Non-Alcoholic Steatohepatitis with no significant fibrosis (NASH <F2), Non-Alcoholic Steatohepatitis with significant fibrosis (NASH ≥ F2), Body mass index (BMI), Triglycerides (TG), Low density lipoprotein cholesterol (LLD-C), High density lipoprotein cholesterol (HDL-C), Haemoglobin A_{1c} (HbA_{1c}), Homeostatic model assessment insulin resistance (HOMA-IR), Metabolic syndrome (MetS), aspartate aminotransferase (AST), alanine aminotransferase (ALT), C_{16}-Sphinganine (C_{16}-SA), C_{16}-Sphingosine (C_{16}-SO), C_{17}-Sphingosine (C_{17}-SO), C_{18}-Sphinganine (C_{18}-SA), C_{18}-Sphingosine (C_{18}-SO), C_{19}-Sphingosine (C_{19}-SO), C_{20}-Sphinganine (C_{20}-SA), C_{20}-Sphingosine (C_{20}-SO), Sphingadiene (SA-diene), C_{18}-phyto-sphingosine (C_{18}-phyto-SO), deoxy-sphinganine (deoxy-SA), deoxy-sphingosine (deoxy-SO). SD or percentage of the total group is presented between brackets.

Supplementary Table 3: Patient characteristics for the comparison NASH <F2 vs. NASH ≥F2

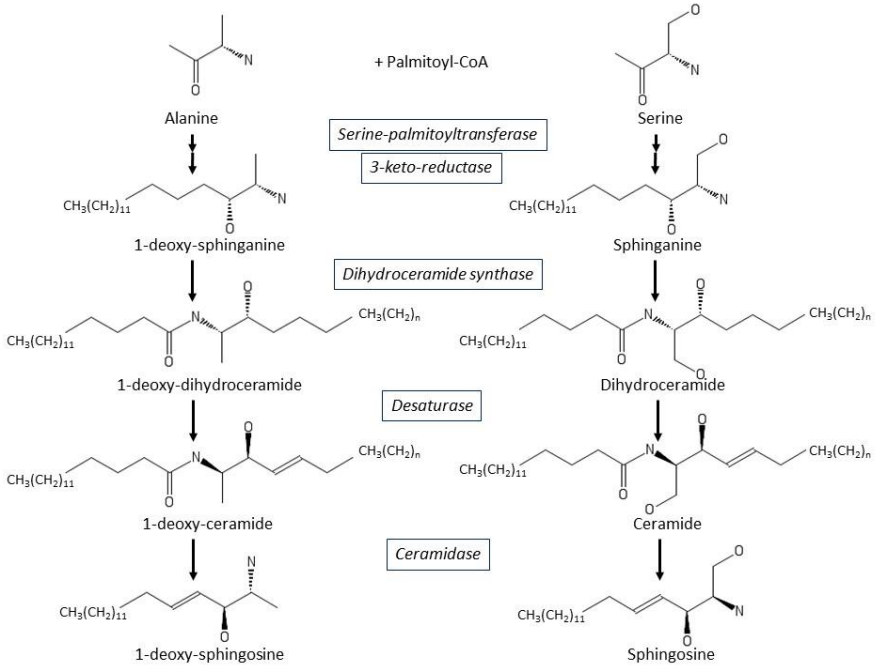
		NASH <F2 N=149	NASH ≥F2 N=41	p
General	Age (years)	45.4 (±12.4)	45.0 (±12.4)	0.855**
	Gender: men	**53 (35.6%)**	**25 (61.0%)**	**0.003*****
Anthropometry	**BMI (kg/m^2)**	**38.9 (±6.0)**	**43.5 (±7.4)**	**<0.001***
	Waist (cm)	**117.9 (±11.3)**	**132.1 (±13.1)**	**<0.001****
Metabolic markers	TG (mg/dL)	157.1 (±72.8)	177.9 (±69.4)	0.059*
	LDL-C (mg/dL)	123.4 (±35.7)	128.5 (±35.5)	0.420**
	HDL-C (mg/dL)	**48.6 (±13.5)**	**41.9 (±9.3)**	**0.002***
	Statin use (%)	22 (14.8)	3 (7.3)	0.212***
	HbA1c (%)	**5.6 (±0.4)**	**6.1 (±1.0)**	**<0.001***
	Glucose (mg/dL)	**85.1 (±12.4)**	**102.1 (±49.8)**	**<0.001***
	HOMA-IR	**3.8 (±2.4)**	**6.8 (±5.1)**	**<0.001***
	MetS	87 (61.3%)	31 (77.5%)	0.058***
	T2DM	**16 (10.9%)**	**10 (24.4%)**	**0.027*****
Liver enzymes	**AST (U/L)**	**30.7 (±11.8)**	**51.5 (±31.5)**	**<0.001***

	ALT (U/L)	**45.8 (±21.6)**	**74.0 (±43.0)**	**<0.001***
Imaging	Steatosis on US	139 (96.5%)	37 (±94.9)	0.632***
Pathology	**Steatosis 0/1/2/3 (%)**	**0.0/36.9/38.3/24.8**	**0.0/14.6/31.7/53.7**	**0.001*****
	Ballooning 0/1/2 (%)	0.0/50.3/49.7	0.0/51.2/48.8	0.920***
	Inflammation 0/1/2 (%)	**0.0/58.4/41.6**	**0.0/29.3/70.7**	**<0.001*****
	Fibrosis 0/1/2/3/4 (%)	**70.5/29.5/0.0/0.0/0.0**	**0.0/0.0/58.5/39.0/2.5**	**<0.001*****
Sphingoid bases	C_{16}-SA	0.491 (±0.249)	0.432 (±0.156)	0.375*
	C_{16}-SO	15.379 (±5.568)	14.026 (±3.838)	0.206**
	C_{17}-SO	6.180 (±2.053)	5.555 (±1.498)	0.083**
	C_{18}-SA	3.726 (±1.459)	3.662 (±1.126)	0.687*
	C_{18}-SO	83.308 (±21.246)	78.050 (±14.864)	0.250*
	C_{18}-phytoSO	0.110 (±0.037)	0.100 (±0.026)	0.126*
	C_{19}-SO	**1.871 (±0.858)**	**1.541 (±0.508)**	**0.046****
	C_{20}-SA	0.025 (±0.011)	0.026 (±0.009)	0.528**
	C_{20}-SO	0.198 (±0.071)	0.190 (±0.052)	0.958*
	SA-diene	**31.063 (±8.688)**	**27.316 (7.078)**	**0.008****
	Deoxy-SA	0.077 (±0.042)	0.077 (±0.040)	0.832**
	Deoxy-SO	0.117 (±0.078)	0.118 (±0.076)	0.895**

* Mann-Whitney U, ** independent sample T-test, *** chi-square. Ballooning and Inflammation are graded according to SAF/Flip-algorithm. Non-Alcoholic Steatohepatitis with no significant fibrosis (NASH <F2), Non-Alcoholic Steatohepatitis with significant fibrosis (NASH ≥ F2), Body mass index (BMI), Triglycerides (TG), Low density lipoprotein cholesterol (LLD-C), High density lipoprotein cholesterol (HDL-C), Haemoglobin A_{1c} (HbA_{1c}), Homeostatic model assessment insulin resistance (HOMA-IR), Metabolic syndrome (MetS), aspartate aminotransferase (AST), alanine aminotransferase (ALT), C_{16}-Sphinganine (C_{16}-SA), C_{16}-Sphingosine (C_{16}-SO), C_{17}-Sphingosine (C_{17}-SO), C_{18}-Sphinganine (C_{18}-SA), C_{18}-Sphingosine (C_{18}-SO), C_{19}-Sphingosine (C_{19}-SO), C_{20}-Sphinganine (C_{20}-SA), C_{20}-Sphingosine (C_{20}-SO), Sphingadiene (SA-diene), C_{18}-phyto-sphingosine (C_{18}-phyto-SO), deoxy-sphinganine (deoxy-SA), deoxy-sphingosine (deoxy-SO). SD or percentage of the total group is presented between brackets.

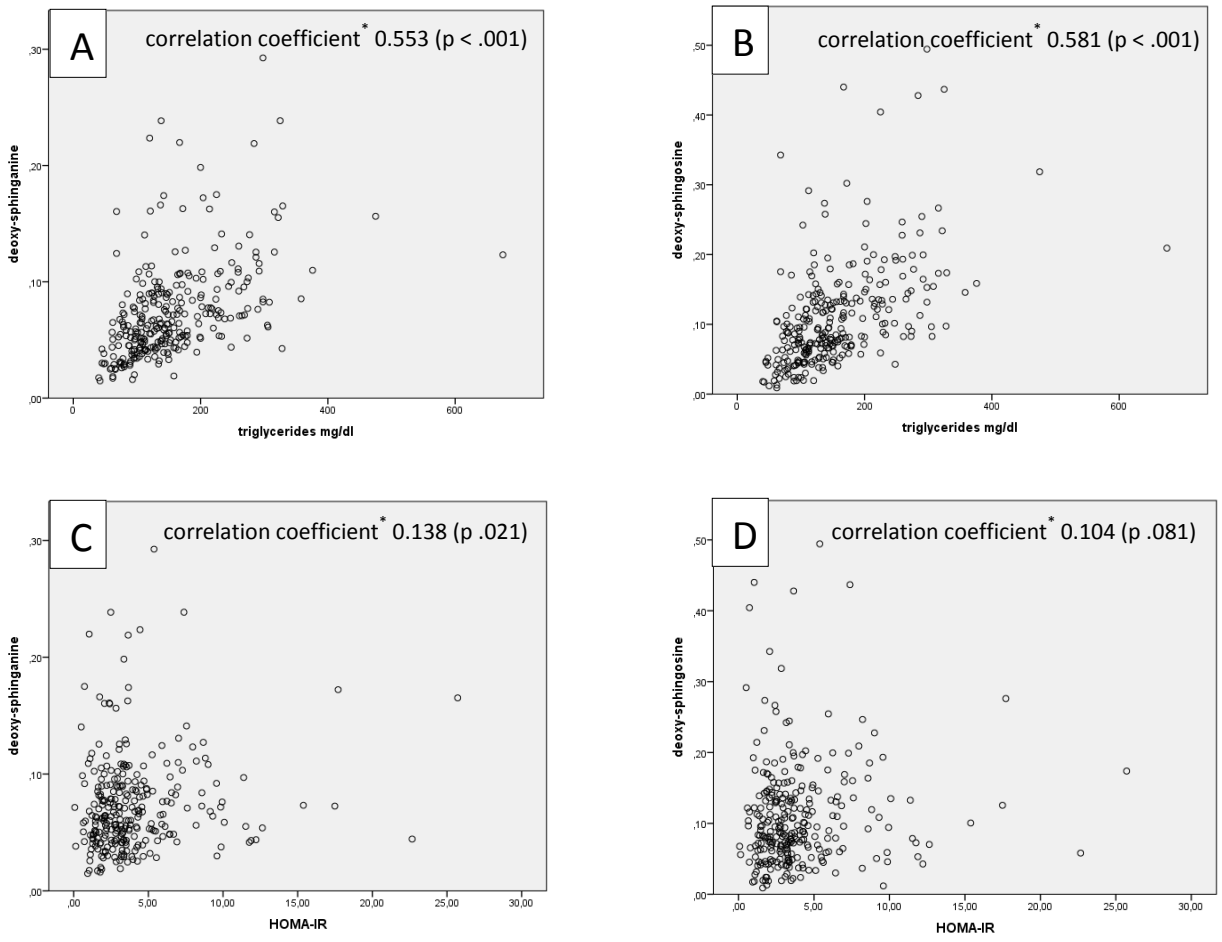
Legends to the Figures

Figure 1: Biosynthesis of sphingolipids and 1-deoxy-sphingolipids.



The enzyme serine-palmitoyltransferase (SPT) catalyses the first and rate-limiting step in the *de novo* synthesis of sphingolipids, which converts L-serine and palmitoyl-CoA into 3-keto-sphinganine. The latter is converted into sphinganine (SA). SA is N-conjugated with another fatty acid to form dihydro-ceramide. When desaturated at C4 it forms ceramide. Ceramidase converts ceramide into sphingosine.

Figure 2: Correlation of fasting plasma sphingoid bases and metabolic markers.



A: Correlation of fasting plasma deoxy- sphinganine and triglyceride levels; *B:* Correlation of fasting plasma deoxy- sphingosine and triglyceride levels; *C:* Correlation of fasting plasma deoxy-sphinganine concentrations and HOMA-IR; *D:* Correlation of fasting plasma deoxy- sphingosine concentrations and HOMA-IR. * Spearman's rho correlation coefficient and associated p-value