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Abstract: A series of 4-aminoquinoline-isoindoline-dione-isoniazid triads were synthesized and assessed for their anti-mycobacterial activities and cytotoxicity. Most of the synthesized compounds exhibited promising activities against the mc26230 strain of *M. tuberculosis* with MIC in the range of 3.125-12.5 µg/mL and were non-cytotoxic against Vero cells. The conjugates lacking either isoniazid or quinoline core in their structural framework failed to inhibit the growth of *M. tuberculosis*; thus, further strengthening the proposed design of triads in the present study.

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Professor Matthew Disney
Editor-in-chief
Bioorganic and Medicinal Chemistry Letters

Dear Professor Disney

Please find attached herewith the manuscript entitled **“Design and synthesis of 4-Aminoquinoline-isoindoline-dione-isoniazid triads as potential anti-mycobacterials”** for publication in *Bioorganic and Medicinal Chemistry Letters*. The manuscript describes the synthesis of Aminoquinoline-isoindoline-dione-isoniazid triads having isoniazid and quinoline core in their structural framework along with their anti-mycobacterial evaluation against mc²6230 strain of *M. tuberculosis* and evaluated for cytotoxicity on Vero cells.

Considering its focus, we firmly believe that this manuscript may be of interest for *Bioorganic and Medicinal Chemistry Letters* readership and hope it meets the quality requirements for publishing in your journal. All authors declare that they have participated in the study and that they have read, commented, and approved the final version of the paper. All authors state that the corresponding authors have full access to all data in the study and have final responsibility for the decision to submit for publication. There is no conflict of interest stated. This work has not been published and is not being considered for publication elsewhere.

Respectfully yours,
Vipin Kumar

Design and synthesis of 4-Aminoquinoline-isoindoline-dione-isoniazid triads as potential anti-mycobacterials

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Highlights

- A series of 4-aminoquinoline-isoindolinedione-isoniazid triads with anti-mycobacterial activities.
- The most active compounds exhibited MIC₉₉ 3.125 µg/mL against mc²6230 strain of *M. tuberculosis*.
- Docking studies were performed to delineate the mechanism of action.

Design and synthesis of 4-Aminoquinoline-isoindoline-dione-isoniazid triads as potential anti-mycobacterials

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Abstract: A series of 4-aminoquinoline-isoindoline-dione-isoniazid triads were synthesized and assessed for their anti-mycobacterial activities and cytotoxicity. Most of the synthesized compounds exhibited promising activities against the mc²6230 strain of *M. tuberculosis* with MIC in the range of 3.125-12.5 µg/mL and were non-cytotoxic against Vero cells. The conjugates lacking either isoniazid or quinoline core in their structural framework failed to inhibit the growth of *M. tuberculosis*; thus, further strengthening the proposed design of triads in the present study.

Keywords: 4-aminoquinoline-isoindoline-dione-isoniazid, Anti-mycobacterial activity, Cytotoxicity, Selectivity index, Molecular docking.

1. Introduction

Tuberculosis (TB), an infectious disease caused by the bacterium, *Mycobacterium tuberculosis*, has resulted in chronic granulomatous lesions in the lungs, which can be accompanied by extra-pulmonary infection foci in the skin, brain and lymph nodes [1]. The disease is the major cause of death after the Human Immune-deficiency Virus (HIV), accounting for 1945 deaths every day [2]. World Health Organization (WHO) estimated that at least 10 million people developed TB with 1.3 million deaths in 2017 [3]. Despite the availability of an important variety of drugs, TB remains a significant global health threat [4-5]. The foremost obstacle in the total eradication of TB is the development of resistance of *M. tuberculosis* to many existing drugs, which has led to the appearance of multidrug-resistance (MDR) and extensively drug-resistant (XDR) strains [6-11]. Also, TB and HIV co-infection is a major concern with nearly two-thirds

of the patients diagnosed with TB also being HIV-1 seropositive [12]. The re-designing and re-positioning of known anti-TB scaffolds is an effective strategy for the development of new frameworks with favorable pharmacological profiles such as high efficacy and low toxicity [13]. Moreover, this strategy can enable the rapid synthesis of highly effective compounds that are already clinically approved, ultimately shortening the drug commercialization pipeline.

Isoniazid (INH), a first-line anti-TB drug, is the most powerful agent used to treat *M. tuberculosis* infection since 1952 [14]. It is a pro-drug requiring activation *via* enzyme catalase-peroxidase encoded by *katG* to its active form [15]. The activated metabolite of INH inhibits the synthesis of mycolic acid, a vital component of the mycobacterial cell wall, by targeting the enoyl-ACP reductase InhA of the type II fatty acid synthase [16-18]. The decrease in catalase-peroxidase activity as a result of *katG* mutations is considered as the most common mechanism associated with INH resistance [19]. Among all the genes involved in INH resistance, excluding *katG*, mutations of InhA confer clinically relevant levels of resistance to INH. Multiple other genes such as *nat* coding for *N*-acetyltransferase (NAT) contribute to resistance to INH in *M. tuberculosis* before KatG activation. Furthermore, both NAT and KatG enzymes interact with the pro-drug directly [20]. Following INH metabolism in the liver, hydrazine metabolites (nitrogen-centered free radicals) are produced which generate highly reactive oxygen species and act as a stimulator of lipid peroxidation, resulting in cell death and hepatic necrosis [21-22]. As INH is the major therapeutic arsenal in the treatment of TB infection, continuous efforts are being made to develop new INH derivatives with greater activity, low cytotoxicity, and fewer side effects [23-24]. Several current reports show that the amalgamation of hydrophobic moieties into the basic structure of INH can enhance the penetration of the drug into the lipophilic cell wall of the bacterium [25]. Further, the *N*-acetyltrans-2 (NAT2) promoted inactivation of INH could also be avoided by functionalizing its hydrazine group [26].

Quinoline moiety is present in a variety of natural products and has diverse biological properties. Notably, quinoline appears as a central core in the recently developed anti-TB drugs such as TMC 207 or bedaquiline and some fluoroquinolones *viz.* gatifloxacin, and moxifloxacin (**Figure 1**) [27].

Recently, we introduced a functionalized isoindoline-1,3-diones into the alkyl chain of 4-aminoquinolines to access their anti-mycobacterial potency. The compounds were promising

candidates with a MIC of 5.9-6.25 $\mu\text{g/mL}$ against the mc²6230 strain of *M. tuberculosis* (**Figure 2**) [28-29]. Motivated by these results and in continuation [30], the present work is a logical extension and involves the synthesis and anti-mycobacterial evaluation of 4-aminoquinoline-isoniazid hybrids linked *via* isoindoline-1,3-diones. The length of the alkyl chain spacer between the pharmacophores along with the position of INH and 4-aminoquinoline around the isoindoline-1,3-dione were meticulously altered to study their influence on Structure-Activity Relationship.

2. Results and Discussion

2.1 Synthetic Chemistry:

For the synthesis of first series of desired 4-aminoquinoline-isoindoline-dione-isoniazid triads, **6a-e**, the reaction of substituted phthalic anhydride **1** with quinoline based diamines **2** was carried out in DMSO at 150 °C for 5 min, in a microwave synthesizer to result in 4-aminoquinoline-isoindoline-diones **5a-e**. EDC-HOBt promoted amide coupling between **5a-e** and isoniazid afforded the corresponding conjugates **6a-e** in moderate to good yields (**Scheme 1**).

Another set of 4-aminoquinoline-isoindoline-dione-isoniazid triads, **8a-g** were prepared *via* an initial microwave heating of substituted phthalic anhydride **1** with isoniazid **3** to result in isoniazid-isoindoline-dione **7**. Amide/Ester coupling between **7** and quinoline based diamine **2**/quinoline based alcohol **2'** afforded **8a-g** in good yields. A cycloalkyl *viz.* piperazinyl analogue **11** was also synthesized to determine the influence of alkyl chain on anti-mycobacterial activities. Further, in order to rationalize the anti-mycobacterial effect of quinoline core in the present series of triads, amide-tethered isoniazid-isoindoline-dione conjugates **13a-b** were synthesized *via* an initial treatment of **1** with amino acids **4** to afford **12** with subsequent treatment with isoniazid using standard coupling procedure. The structures of the synthesized 4-aminoquinoline-isoindoline-isoniazid conjugates were assigned based on the spectral and analytical evidence. For example, the compound **6a** showed a molecular ion peak at 515.1177 in its high-resolution mass spectrum (HRMS). The significant features of its ¹H NMR spectrum involved the presence of a triplet at δ 3.82 because of methylene (N-CH₂-); two doublets at 6.62 ($J=5.3$ Hz) and 7.76 ($J=2.2$ Hz) corresponding to quinoline protons along with a multiplet of isoindoline ring protons at 8.27-8.31. The presence of signals in its ¹³C NMR

spectrum at δ 164.6, 164.7, 167.7, and 167.8 corresponding to imide- and amide carbonyls, along with methylene carbons at 36.5 and 40.7 further validated the assigned structure.

2.2 *In vitro* anti-mycobacterial and cytotoxic evaluation:

The synthesized 4-aminoquinoline-isoniazid conjugates were assessed for their anti-mycobacterial activities against avirulent *M. tuberculosis* mc²6230 strain and the results are enlisted in **Table 1**. The cytotoxicity of all derivatives to mammalian Vero cells was also evaluated to establish whether the observed activities were due to their inherent anti-mycobacterial ability or due to cytotoxicity (**Table 1**). As evident, the compounds showed promising anti-mycobacterial activities, although they were not as active as INH. A closer examination of the results showed the activity dependence on both the length of the alkyl chain between the pharmacophores as well as the position of INH and 4-aminoquinoline around isoindoline core. Among the 4-aminoquinoline-isoindoline-1,3-diones **5a-e** (without INH-core), the compounds lack any anti-mycobacterial activity except **5c** and **5d** with weak activity but cytotoxicity. Inclusion of INH among these conjugates not only improved their anti-mycobacterial activity substantially but also decreased their cytotoxicity as seen in triads, **6a-e**. The improvement in anti-mycobacterial and cytotoxicity profile is apparent at longer chain lengths, as displayed by **6d** and **6e** with MIC₉₉ of 12.5 and 3.1 $\mu\text{g/mL}$ and IC_{50s} of 73 and 77 $\mu\text{g/mL}$, respectively.

Furthermore, interchanging the position of INH with quinoline diamines around isoindoline core in compound **8a-e** improved the anti-mycobacterial activities without much dependence on the alkyl chain length. The triad **8e** with an octyl spacer not only retained the anti-mycobacterial efficacy (MIC₉₉=3.1 $\mu\text{g/mL}$), it is also non-cytotoxic (IC₅₀ $\geq$ 100 $\mu\text{g/mL}$). The inclusion of a cyclic amine *viz.* piperazine instead of alkyl amine as in triad **11** also resulted in the good anti-mycobacterial activity.

The conjugates, **13a** and **13b** without a quinoline core, did not show any substantial anti-mycobacterial activity; thus, justifying the importance of the proposed design of 4-aminoquinoline-isoindoline-dione-isoniazid triads to elicit anti-mycobacterial activity and reduce cytotoxicity.

Relating the activity and cytotoxicity data of currently synthesized compounds (**6e**, **8e**, **11**) with our earlier report (**I-IV**) [28-31], the introduction of INH at C-5 of isoindoline ring not only

improved anti-mycobacterial activity but also resulted in the reduction of cytotoxicity (**Figure 2**). The general anti-TB SAR/cytotoxicity of the synthesized series of 4-aminoquinoline-isoindoline-isoniazid triads is elucidated in **Figure 3**.

2.3 Molecular docking studies and ADME properties prediction

Molecular docking studies were conducted using AutoDock to explore the potential binding interactions of selected compounds with *M. tuberculosis* enoyl-acyl carrier protein reductase (InhA; PDB ID: 4TZK). InhA is a prominent enzyme involved in the biosynthesis of mycolic acid for mycobacterial cell wall and the molecular target of isoniazid. Six inhibitors were docked into the active site of InhA, as shown in **Figures 4** and **5**, while the docking results are summarized in **Table 2**.

In consonant with the experimental results, the binding energies and receptor interactions of compounds **6e**, **8e** and **11** were superior to compounds **I**, **II** and **III** (**Table 2**). For instance, in compound **I** (**Figure 4a**), the chlorine atom was sandwiched between residues Phe149 and Met199 *via* hydrophobic interactions while the amino group of quinoline core formed a hydrogen bond interaction with the hydroxyl oxygen of Gly192. The isoindoline core also afforded alkyl, π - π stacking, π -alkyl, and π - σ interactions with Ile95, Phe41, Val65, Ile122, Phe149, and Met199. A similar binding profile was observed for the isoindoline core of compound **II** (**Figure 2**), i.e. π - π stacking with residue Phe41 as well as π - σ , π -alkyl, and alkyl interactions with Ile21, Val65, Ile122, Phe149, Ala198, Met147 and Ile95. The unit also featured a hydrogen bond interaction between the isoindoline and Ile95, while the fluorine substituent formed a halogen bond with Asp64. The docked complex of **III** (**Figure 3**) was established in the hydrophobic pocket created by hydrophobic interaction with the side chains of ten amino acid residues. Surprisingly, the oxygen atom of the isoindoline core formed a strong hydrogen bond with Tyr168. The interaction was absent in compounds **I** and **II**; hence, a potential platform for further structural modifications to enhance the anti-mycobacterial activity.

Furthermore, compound **6e**, **8e** and **11** binds in essentially the same manner within the active site of InhA. The compound **6e**-InhA complex is characterized by six hydrogen bond interactions between the amino acid residues of side chain and both the oxygen atom of isoniazid moiety and the amine linker. Besides, the quinoline core was anchored in the

hydrophobic pocket containing Val65, Ile95, Phe41, and Ile122, which suggested that the two-fold increase in potency might stem from additional interactions between the isoniazid moiety and the side chains of the active site residues. Furthermore, the isoniazid moiety in compound **8e** interacts with Gly192, Ala191 and Lys165 *via* hydrophobic and H-bonding interactions while the aminoquinoline group formed hydrophobic interactions with residue Val65, Leu63, Gly14 and Gly40, respectively. Other residues that made H-bond interactions include, Gly96, Ile95, Ile165, which also serve as motivators for the stabilization of the inhibitor within the catalytic pocket of the receptor. Moreover, the docked complex of compound **11** showed additional hydrogen bond interactions between the piperazine ring and the carbonyl oxygen of Gly96, which was accompanied by hydrophobic interaction with Ile16.

On the other hand, the molecular descriptors ADME properties were calculated for potent compounds using web-based software pkCSM (<http://biosig.unimelb.edu.au/pkcsm/prediction>) and SwissADME (<http://www.swissadme.ch/>). The results are presented in **Table 2**. The drug candidates with TPSA values of 140 Å or lower are expected to have good absorption. Accordingly, all the compounds were within this limit, i.e., <140 Å, which implies that these compounds fulfilled the optimal requirement for drug absorption.

3. Conclusion

In conclusion, a series of 4-aminoquinoline-isoindoline-dione-isoniazid triads have been synthesized with the target of reviewing their SAR against *M. tuberculosis*. Nearly all of the synthesized conjugates presented promising anti-TB activity and useful selectivity index. The conjugates without the quinoline core **13a/13b** and the INH nucleus **5a-e** failed to inhibit the growth of *M. tuberculosis* as was observed in our previous report on 1,8-naphthalimide-isoniazid hybrids [31], further validating the design of triads in the present case.

Electronic Supplementary Information: Material and methods, Synthesis and spectral data of all the compounds along with scanned ¹H and ¹³C NMR spectra of few representative compounds *viz.* **5a**, **6a**, **6b**, **7**, **8a**, **9**, **13a** and **11** are provided in the electronic supplementary information.

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Conflict of interest

The authors have declared no conflict of interest.

References:

1. (a) World Health Organization, Annual Report, 2017. <http://apps.who.int/iris/bitstream/10665/254762/1/978929022584-eng.pdf> (b) K. F. M. Pasqualoto, E. I. Ferreira, O. A. Santos-Filho, A. J. Hopfinger, J. Med. Chem. 47 (2004) 3755. (c) P. Modi, S. Patel, M. Chhabria, Bioorg. Chem. 87 (2019) 240.
2. “Get the Facts about TB Disease” TBFACTS.ORG, <http://www.tbfacts.org/symptomstb>.
3. World Health Organization (WHO). Global tuberculosis report. 2019. Available from: https://www.who.int/tb/publications/global_report/tb19_Exec_Sum_12Nov2019.pdf?ua=1
4. D. S. Reddy, M. Kongot, S. P. Netalkar, M. M. Kurjogi, R. Kumar, F. Avecilla, A. Kumar, Eur. J. Med. Chem. 150 (2018) 864.
5. A. Zumla, P. Nahid, S. T. Cole, Nat. Rev. Drug Discov. 12 (2013) 388.
6. T. Cohen, M. C. Becerra, M. B. Murray, Microb. Drug Resist. 10 (2004) 280.
7. J. Chen, S. Zhang, P. Cui, W. Shi, W. Zhang, Y. Zhang, J. Antimicrob. Chemother. 72 (2017) 3272.
8. A. V. Deun, A. K. Maug, V. Bola, R. Lebeke, M. A. Hossain, W. B. de Rijk, L. Rigouts, A. Gumusboga, G. Torrea, B. C. de Jong, J. Clin. Microbiol. 51 (2013) 2633.
9. R. E. Stanley, G. Blaha, R. L. Grodzicki, M. D. Strickler, T. A. Steitz, Nat. Struct. Mol. Biol. 17 (2010) 289.
10. C. Vilcheze, W. R. Jacobs, Microbiol. Spectr. 2 (2014) 0014.
11. X. Lu, J. Tang, S. Cui, B. Wan, S. G. Franzblau, T. Zhang, X. Zhang, K. Ding, Eur. J. Med. Chem., 125 (2017) 41.
12. W. Manosuthi, S. Wiboonchutikul, S. Sungkanuparph, AIDS Res. Ther. 13 (2016) 22.

13. K. Anil, A. Eric, L. Nacer, G. Jerome, A. Koen, *Nature* 469 (2011) 483.
14. V. H. Teixeira, C. Ventura, R. Leitão, C. Ràfols, E. Bosch, F. Martins, *et al.*, *Mol. Pharm.* 12 (2015) 898.
15. E. H. Robitzek, I. Selikoff. *J. Am Rev Tuberc* 65 (1952) 402.
16. A. Banerjee, E. Dubnau, A. Quemard, V. Balasubramanian, K. S. Um, T. Wilson, *et al.* *Science* 263 (1994) 227.
17. H. Marrakchi, G. Laneelle, A. Quemard. *Microbiology* 146 (2000) 289.
18. C. Vilchèze, F. Wang, M. Arai, [M. H. Hazbón](#), [R. Colangeli](#), [L. Kremer](#), [T. R. Weisbrod](#), [D. Alland](#), [J. C. Sacchettini](#), W. R. Jacobs Jr, *Nat. Med.* 12 (2006) 1027–1029.
19. Y. Zhang, B. Heym, B. Allen, D. Young, S. Cole, *Nature* 358 (1992) 591.
20. A. U. Nusrath, G. P. Doss C, T. Kumar, S. Swathi, A. R. Lakshmi, L. E. Hanna, J. *Global Antimicrobial Resistance* 11 (2017) 57.
21. E. Vavříkova, S. Polanc, M. Kocevar, K. Horvati, S. Bosze, J. Stolaríkova, K. Vavrova, J. Vinsova, *Eur. J. Med. Chem.* 46 (2011) 4937.
22. E. Vavříkova, S. Polanc, M. Kocevar, J. Kosmrlj, K. Horvati, S. Bosze, J. Stolaríkova, A. Imramovský, J. Vinsova, *Eur. J. Med. Chem.* 46 (2011) 5902.
23. K. Elumalai, M. A. Ali, M. Elumalai, K. Eluri, S. Srinivasan, *J. Acute Disease.* 2 (2013) 316.
24. L. Matei, C. Bleotu, I. Baciú, C. Draghici, P. Ionita, A. Paun, M.C. Chifiriuc, A. Sbarcea, I. Zarafu, *Bioorg. Med. Chem.* 21 (2013) 5355.
25. (a) J. Sandy, A. Mushtaq, A. Kawamura, J. Sinclair, E. Sim, M. Noble, *J. Mol. Biol.* 318 (2002) 1071-1083. (b) Yuan-Qiang Hu, S. Zhang, F. Zhao, C. Gao, Lian-Shun Feng, Zao-Sheng Lv, Z. Xu, X. Wu. *Eur. J. Med. Chem.* 133 (2017) 255.
26. D. Shingnapurkar, P. Dandawate, C. E. Anson, A. K. Powell, Z. Afrasiabi, E. Sinn, S. Pandit, S. K. Venkateswara, S. Franzblau, S. Padhye, *Bioorg. Med. Chem. Lett.* 22 (2012) 3172.
27. (a) J. C. Sundaramurthi, L. E. Hanna, S. Selvaraju, S. Brindha, J. Joel Gnanadoss, S. Vincent, H. Singh, S. Swaminathan, *Tuberculosis* 100 (2016) 69. (b) B. Villemagne, C. Crauste, M. Flipo, *Eur. J. Med. Chem.* 51 (2012) 1. (c) D. T. Hoagland, J. Liu, R. B. Lee, R.

- E. Lee, Adv. Drug Deliv. Rev. 102 (2016) 55. (d) G. Kumar, A. Sathe, V. S. Krishn, D. Sriram, S. M. Jachak, Eur. J. Med. Chem. 157 (2018) 1-13. (e) A. H. Diacon, A. Pym, M. Grobusch, R. Patientia, R. Rustomjee, L. Page-Shipp, C. Pistorius, R. Krause, M. Bogoshi, G. Churchyard, A. Venter, J. Allen, J. C. Palomino, T. De Marez, R. N. Veziris, M. Ibrahim, N. Lounis, T. Andries, V. Jarlier, PLoS ONE, 6 (2011) 17556. (f) J. Cohen, Science, 2013, 339, 130.
28. A. Rani, A. Viljoen, Sumanjit, L. Kremer, V. Kumar, ChemistrySelect, 2 (2017) 10782.
 29. A. Rani, A. Viljoen, M. D. Johansen, L. Kremer, V. Kumar. RSC Adv. 9 (2019) 8515.
 30. (a) Shalini, A. Viljoen, L. Kremer, V. Kumar, Bioorg. Med. Chem. Lett. 28 (2018) 1309. (b) Shalini, M. D. Johansen, L. Kremer, V. Kumar, Bioorg. Chem. 92 (2019) 103241. (c) A Singh, A Viljoen, L Kremer, V Kumar, ChemistrySelect 3 (29), 8511.
 31. Shalini, M. D. Johansen, L. Kremer, V. Kumar, Chem. Bio. Drug Design 94 (2019) 1300.

Captions:

Figure 1: Chemical structures of standard drug isoniazid (INH) and quinoline based anti-tubercular agents.

Figure 2: Comparison of activity against mc²6230 and cytotoxicity against Vero cells of **6e**, **8e** and **11** with previously reported scaffolds **I**, **II** and **III**.

Scheme 1: Synthesis of 4-aminoquinoline-isoindoline-dione-INH conjugates (**6a-e**), (**8a-e**), **11** and (**13a-b**).

Table 1: *In vitro* anti-mycobacterial activity of synthesized compounds against *M. tuberculosis* mc²6230 and cytotoxicity evaluation on mammalian Vero cells and their selectivity index (SI).

Figure 3: Structure-Activity Relationship (SAR) of both series of synthesized 4-aminoquinoline-isoindoline-dione-INH triads.

Figure 4: Binding interactions of compounds (a) **I** (b) **II** (c) **III**

Figure 5: Binding interactions of compounds (a) **6e** (b) **8e** (c) **11**

Table 2: Molecular descriptors and the binding energy of the potent compounds

Figure 1: Chemical structures of standard drug isoniazid (INH) and quinoline based anti-tubercular agents.

Figure 2: Comparison of activity against mc²6230 and cytotoxicity against Vero cells of **6e**, **8e** and **11** with previously reported scaffolds **I-IV** [28-29, 31]

Scheme 1: Synthesis of 4-aminoquinoline-isoindoline-dione-isoniazid conjugates.

Table 1: *In vitro* anti-mycobacterial activity (MIC₉₉) of synthesized compounds against mc²6230 strain of *M. tuberculosis* and cytotoxicity (IC₅₀) evaluation on mammalian Vero cells and their selectivity index (SI).

Code	Structure	% Yield	MIC ₉₉ (µg/mL)	IC ₅₀ (µg/mL)	SI value
5a		81	>200	>100	>0.5
5b		76	200	99	0.5
5c		77	100	17	0.2
5d		76	100	11	0.1

5e		79	200	>100	>0.5
6a		81	200	>100	>0.5
6b		76	25	63	2.5
6c		82	12.5	9	0.7
6d		78	12.5	73	5.8
6e		79	3.1	77	25
8a		79	3.1	33	10.6
8b		77	3.1	85	27
8c		74	12.5	8	0.6
8d		76	6.3	19	3
8e		72	3.1	>100	>31
8f		85	>200	>100	>0.5
8g		84	6.3	>100	>16
11		83	3.1	>100	>31

13a		83	100	>100	>1
13b		85	50	>100	>2
INH		-	0.01

Figure 3: Structure-Activity Relationship (SAR) of both series of synthesized 4-aminoquinoline-isoindoline-dione-INH triads.

Figure 4: Binding interactions of compounds (a) **I** (b) **II** (c) **III**

Figure 5: Binding interactions of compounds (a) **6e** (b) **8e** (c) **11**

Table 2: Molecular descriptors and the binding energy of the potent compounds

Compound	B.E (kcal/mol)	TPSA	logP	HBD	HBA	logS	Caco2	log K_p	logBB
I	-9.7	62.30	5.05	1	3	-5.5	0.465	-2.773	0.133
II	-9.9	91.40	2.77	2	5	-3.716	0.645	-2.816	-1.407
III	-10.2	65.54	4.35	3	3	-4.945	0.76	-2.877	0.231
6e	-10.4	133.39	4.42	3	6	-4.094	0.768	-2.735	-1.745
8e	-10.3	133.39	4.40	3	6	-4.324	1.65	-2.735	-1.544
11	-11.7	98.74	2.27	1	5	-4.044	0.692	-2.767	-1.129

TPSA - Topological Polar Surface Area (60 – 140); log BB - logarithm of predicted blood/brain barrier partition coefficient (-3.0-1.2); Caco-2 - cell membrane permeability (> 0.9 is high); HBA - number of hydrogen bond acceptors; HBD - number of hydrogen bond donors; log K_p - predicted skin permeability(SP) > -2.5 is low.

Design and synthesis of 4-Aminoquinoline-isoindoline-dione-isoniazid triads as potential anti-mycobacterials

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Contents

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2. Physical and spectral data of **5a-e**, **6a-e**, **8a-g**, **11** and **13a-b**
3. Copies of ¹H and ¹³C NMR spectra of **5a**, **6a**, **6b**, **7**, **8a**, **9**, **13a** and **11**

Experimental section

General

All starting materials and reagents were purchased from Sigma-Aldrich and Merck. Thin-layer chromatography was performed on a film of silica gel that contained a fluorescent indicator F₂₅₄ supported on an aluminum sheet (Merck). Melting points were determined by open capillary using a Stuart Digital Melting Point apparatus (SMP10) and are uncorrected. ¹H NMR spectra were recorded in DMSO-d₆ Jeol 400 (400 MHz) spectrometers while TMS is used as the internal standard. Microwave reactions were performed in an Anton Paar Monowave Initiator+ instrument using a sealed 10 mL process vials. Reaction times refer to irradiation time at the target temperature, not the total irradiation time. The temperature was measured with an IR sensor. Chemical shift values are specified as parts per million (ppm) downfield from TMS while coupling constant (*J*) values are in Hertz. Patterns of Splitting are designated as s: singlet, d: doublet, t: triplet, dd: double doublet, m: multiplet, ddd: doublet of doublet of doublet, and br: broad peak. ¹³C NMR spectra were recorded on Jeol 100 MHz spectrometers in DMSO-d₆. High-Resolution Mass Spectra (HRMS) were recorded on a Bruker-micrOTOF-Q II spectrometer using ESI as the ion source.

General Procedure for the synthesis of 4-aminoquinoline-isoindoline-isoniazid triad:

Synthesis of 2-(((7-chloroquinolin-4-yl)amino)alkyl)-1,3-dioxoisoindoline-5-carboxylic acid (5a-e):

An equimolar mixture of phthalic anhydride **1** (1 eq.) and 7-chloro-4-aminoquinoline-diamines **2** (1 eq.) was dissolved in a 2 mL of DMSO in a microwave glass vial. The reaction vial was irradiated to 150 °C for 5 min in microwave synthesizer. After completion of the reaction as evident from TLC, the vessel contents were poured in water (20 mL) and solid precipitates obtained were filtered and washed with cold water and recrystallized using absolute ethanol.

Synthesis of 2-(((7-chloroquinolin-4-yl)amino)alkyl)-N'-isonicotinoyl-1,3-dioxoisoindoline-5-carbohydrazide (6a-e):

1.0 equivalent of 4-aminoquinoline-isoindoline acids **5**, *N*-ethyl-*N*-dimethylaminopropyl carbodiimide (EDC) (1.1 eq.), hydroxybenzotriazole (HOBt) (1.2 eq.) and 4-diisopropylethylamine (DIPEA) (2 eq.) were mixed in minimum DMF, and the obtained mixture was stirred for 5 min. Isoniazid (1.1 eq.) was subsequently added to the reaction mixture, and the stirring was continued for 3 h at room temperature. The progress was monitored by thin-layer chromatography (TLC). DMF was then evaporated under reduced pressure using a rotary evaporator, and cold water (15 mL) was added, and solid precipitates obtained were filtered and washed with cold water. The crude product was recrystallized in absolute ethanol.

Synthesis of 2-(isonicotinamido)-1,3-dioxoisoindoline-5-carboxylic acid (7):

A mixture of phthalic anhydride **1** (1 eq.) and isoniazid **3** (1.1 eq.) was dissolved in a 2 mL of DMSO in a microwave glass vial. The reaction vial was then irradiated to 150 °C for 10 min in microwave synthesizer. After completion of the reaction as evident from TLC, the vessel contents were poured in water (20 mL) and solid precipitates obtained were filtered and washed with cold water and recrystallized using absolute ethanol.

Synthesis of N-(((7-chloroquinolin-4-yl)amino)alkyl)-2-(isonicotinamido)-1,3-dioxoisoindoline-5-carboxamide (8a-g):

A mixture of 2-(isonicotinamido)-1,3-dioxoisoindoline-5-carboxylic acid **7** (1.0 eq.), *N*-ethyl-*N*-dimethylaminopropyl carbodiimide (EDC) (1.1 eq.), hydroxybenzotriazole (HOBt) (1.2

eq.) and 4-diisopropylethylamine (DIPEA) (2 eq.) were dissolved in minimum DMF and was stirred for 5 min at room temperature. Then, quinoline-diamines **2**/quinoline-ethanolamines **2'** (1.1 eq.) was added to the reaction mixture, and the stirring was continued for 5 h. On completion, DMF was evaporated under vacuum using a rotary evaporator and cold water (15 mL) was added. The solid precipitates obtained were filtered and washed with cold water with subsequently re-crystallization in absolute ethanol.

*Synthesis of N-(5-(4-(7-chloroquinolin-4-yl)piperazin-1-yl)-1,3-dioxisoindolin-2-yl)isonicotinamide **11**:*

To a microwave reaction vial was added a solution of 4-fluoro-phthalic anhydride **1** (1.0 eq.) and Isoniazid (1.0 eq.) in 2mL of DMSO. After sealing, the vial was heated to 150 °C for 10 min in the microwave reactor. After the completion of the first step, as clear from TLC, 7-chloro-4-(piperazin-1-yl)quinoline **10** (1.0 mmol) was added in the same reaction vial. The reaction mixture was again heated at 180 °C for 15 min in the microwave, and the completion was determined using TLC. When all the reactants consumed, the contents were poured in water (20 mL) resulting in the precipitation of the desired product. The obtained product was filtered and recrystallized using absolute ethanol.

*Synthesis of N-phthaloylamino acids **12**:*

A mixture of phthalic anhydride **1** (1 eq.) and amino acids **4** (1 eq.) in a small aliquot of glacial acetic acid was placed in a 10 mL microwave glass vial and irradiated for 10 min. The reaction mixture on completion was treated with cold water to result in a white powder (**12**), which was filtered, dried and recrystallized from ethanol.

*Synthesis of N'-(2-(1,3-dioxisoindolin-2-yl)acetyl)isonicotinohydrazide/N'-(4-(1,3-dioxisoindolin-2-yl)benzoyl)isonicotinohydrazide (**13a-b**):*

1.0 equivalent of N-phthaloylamino acids, N-ethyl-N-dimethylaminopropyl carbodiimide (EDC) (1.1 eq.), hydroxybenzotriazole (HOBt) (1.2 eq.) and 4-Dimethylaminopyridine (DMAP) (0.2 eq.) were mixed in minimum DMF and the obtained mixture, was stirred for 5 min. Isoniazid (1.1 eq.) was added to the reaction mixture and the stirring was continued for 3 h at room temperature. DMF was evaporated using a rotary evaporator, and cold water (15 mL) was added. The solid precipitates thus obtained were filtered, washed with cold water and recrystallized in absolute ethanol.

Methods for assessment of the anti-tubercular activity of test compounds:

Bacterial strains and growth conditions: *M. tuberculosis* mc²6230 [1] was grown at 37°C in Middlebrook 7H9 supplemented with oleic-albumin-dextrose-catalase enrichment (OADC) and 109 µM of pantothenic acid (complete 7H9).

Drug susceptibility testing: The vulnerability of *M. tuberculosis* mc²6230 to the synthesized compounds was determined as reported previously [2]. Briefly, an exponentially growing culture was diluted in complete 7H9 to OD₆₀₀ = 0.01. The bacteria were then seeded in 100 µL volumes in all the wells of a 96-well plate except the first column of wells which contained 200 µL of the bacterial suspension. Compounds (stock concentration 10 mg/mL) were directly added to the wells of the first column to achieve a concentration of 200 µg/mL. Two-fold serial dilutions were then carried out by transferring 100 µL of bacterial suspension from the first column of wells to the second column, mixing, and repeating this procedure for each consecutive column. The plates were then placed in sealed plastic bags and incubated at 37°C. After 7 days of incubation, plates were visually inspected to determine the MIC, which was defined as the minimal concentration of compound at which no growth of bacteria was observed. In addition, 10 µL of resazurin (0.025% w/v) (Sigma, Germany) was added to each well and left to incubate for 24 h to determine metabolic activity of remaining bacteria. If the colour remains blue, bacteria are dead/metabolically inactive, while if there is a colour shift to bright pink, bacteria are alive and metabolically active (Graphical abstract).

Cytotoxicity assay

Cell viability was determined using Vero cells (ATCC, Sigma, Germany) grown in RPMI medium (Gibco, USA), supplemented with 10% decompemented fetal calf serum, under a 5% CO₂ atmosphere. Cells were seeded in 96-well plates at a density of 2×10^4 cells/well in 160 µL RPMI medium and incubated overnight at 37°C to allow cells to adhere. Compounds (dissolved in DMSO) were freshly diluted to appropriate concentrations in RPMI, to allow the addition of 20 µL volumes of the diluted compounds to the cells that resulted in serial dilutions ranging from 100 µg/mL to 0.78 µg/mL. The maximum final concentration of DMSO was 1 % (v/v), and no cytotoxic effect of DMSO was observed at this concentration. After 24 h incubation at 37°C, 10 µL of resazurin (0.025% w/v) (Sigma, Germany) was added to each well, and the cells were incubated for an additional 3 h at 37°C. Fluorescence was measured in a Polarstar Omega fluorometer using appropriate filters (540 nm excitation and 590 nm emission wavelength). Percentage survival was determined by dividing fluorescence values obtained in the compound containing wells by values obtained for control wells containing cells incubated with a dilution series of DMSO and multiplying this value by

100. The IC₅₀ is defined as the lowest concentration of compound tested at which at least 50% cell viability was observed.

References

1. V.K. Sambandamurthy, S. C. Derrick, T. Hsu, B. Chen, M. H. Larsen, K. V. Jalapathy, M. Chen, J. Kim, S. A. Porcelli, J. Chan, S. L. Morris, W. R. Jr. Jacobs, Vaccine. 24 (2006) 6309.
2. J. Ollinger, M. A. Bailey, G. C. Moraski, A. Casey, S. Florio, T. Alling, M. J. Miller, T. Parish, PLoS One, 8 (2013) e60531.

2-(2-((7-chloroquinolin-4-yl)amino)ethyl)-1,3-dioxoisindoline-5-carboxylic acid (5a): Pale yellow solid; mp 298-299°C; ¹H NMR (400 MHz, DMSO-d₆) δ 3.57-3.61 (m, 2H, -CH₂-); 3.80 (t, *J*=5.9 Hz, 2H, -CH₂-); 6.66 (d, *J*=5.6 Hz, 1H, Ar-H); 7.42 (dd, *J*=8.8 Hz, 2.2 Hz, 1H, Ar-H); 7.70 (t, *J*=6.0 Hz, 1H, NH-exchangeable with D₂O); 7.78 (d, *J*=2.3 Hz, 1H, Ar-H); 7.89-7.92 (m, 1H, Ar-H); 8.02 (d, *J*=9.0 Hz, 1H, Ar-H); 8.16 (d, *J*=1.2 Hz, 1H, Ar-H); 8.28-8.31 (m, 1H, Ar-H); 8.42 (d, *J*=5.6 Hz, 1H, Ar-H); ¹³C NMR (100 MHz, DMSO-d₆) δ 37.5, 43.3, 98.8, 117.3, 123.0, 123.2, 124.8, 125.4, 125.7, 134.6, 135.8, 137.5, 138.0, 146.2, 149.7, 151.1, 155.9, 165.5, 166.8, 167.6 HRMS Calcd for C₂₀H₁₄ClN₃O₄ [M+H]⁺ 396.0673 found 396.0686.

2-(3-((7-chloroquinolin-4-yl)amino)propyl)-1,3-dioxoisindoline-5-carboxylic acid (5b): Pale yellow solid; mp 294-295°C; ¹H NMR (400 MHz, DMSO-d₆) δ 1.97-2.03 (m, 2H, -CH₂-); 3.55-3.60 (m, 2H, -CH₂-); 3.79 (t, *J*=5.8 Hz, 2H, -CH₂-); 6.65 (d, *J*=5.7 Hz, 1H, Ar-H); 7.41(dd, *J*=8.9 Hz, 2.1 Hz, 1H, Ar-H); 7.69 (t, *J*=6.1 Hz, 1H, NH-exchangeable with D₂O); 7.76 (d, *J*=2.2 Hz, 1H, Ar-H); 7.87-7.91 (m, 1H, Ar-H); 8.00 (d, *J*=9.1 Hz, 1H, Ar-H); 8.14 (d, *J*=1.4 Hz, 1H, Ar-H); 8.25-8.29 (m, 1H, Ar-H); 8.41 (d, *J*=5.4 Hz, 1H, Ar-H); ¹³C NMR (100 MHz, DMSO-d₆) δ 28.5, 36.3, 43.1, 98.6, 117.2, 123.1, 123.3, 124.6, 125.3, 125.6, 134.4, 135.7, 137.4, 138.1, 146.0, 149.6, 151.0, 155.7, 165.3, 166.6, 167.5 HRMS Calcd for C₂₁H₁₆ClN₃O₄ [M+H]⁺ 410.0829 found 410.0836.

2-(4-((7-chloroquinolin-4-yl)amino)butyl)-1,3-dioxoisindoline-5-carboxylic acid (5c): Pale yellow solid; mp 285-286°C; ¹H NMR (400 MHz, DMSO-d₆) δ 1.62-1.65 (m, 4H, 2x-CH₂-); 3.50-3.53 (m, 2H, -CH₂-); 3.60 (t, *J*=5.7 Hz, 2H, -CH₂-); 6.64 (d, *J*=5.6 Hz, 1H, Ar-H); 7.40 (dd, *J*=8.8 Hz, 2.0 Hz, 1H, Ar-H); 7.68 (t, *J*=6.0 Hz, 1H, NH-exchangeable with D₂O); 7.77

(d, $J=2.1$ Hz, 1H, Ar-H); 7.86-7.90 (m, 1H, Ar-H); 8.01 (d, $J=9.0$ Hz, 1H, Ar-H); 8.15 (d, $J=1.3$ Hz, 1H, Ar-H); 8.26-8.30 (m, 1H, Ar-H); 8.42 (d, $J=5.4$ Hz, 1H, Ar-H); ^{13}C NMR (100 MHz, DMSO- d_6) δ 25.4, 25.5, 35.2, 42.5, 98.4, 117.1, 123.2, 123.4, 124.7, 125.2, 125.7, 134.5, 135.8, 137.3, 138.2, 146.0, 149.4, 151.1, 155.6, 165.3, 166.5, 167.4 HRMS Calcd for $\text{C}_{22}\text{H}_{18}\text{ClN}_3\text{O}_4$ $[\text{M}+\text{H}]^+$ 424.0986 found 424.0994.

2-(6-((7-chloroquinolin-4-yl)amino)hexyl)-1,3-dioxoisindoline-5-carboxylic acid (5d): Pale yellow solid; mp 272-273°C; ^1H NMR (400 MHz, DMSO- d_6) δ 1.26-1.41 (m, 4H, 2x- CH_2 -); 1.52-1.64 (m, 4H, 2x- CH_2 -); 3.44-3.51 (m, 2H, - CH_2 -); 3.58 (t, $J=5.8$ Hz, 2H, - CH_2 -); 6.63 (d, $J=5.6$ Hz, 1H, Ar-H); 7.40 (dd, $J=8.9$ Hz, 2.1 Hz, 1H, Ar-H); 7.65 (t, $J=6.0$ Hz, 1H, NH-exchangeable with D_2O); 7.76 (d, $J=2.0$ Hz, 1H, Ar-H); 7.86-7.91 (m, 1H, Ar-H); 8.01 (d, $J=9.1$ Hz, 1H, Ar-H); 8.14 (d, $J=1.4$ Hz, 1H, Ar-H); 8.24-8.30 (m, 1H, Ar-H); 8.41 (d, $J=5.3$ Hz, 1H, Ar-H); ^{13}C NMR (100 MHz, DMSO- d_6) δ 26.2, 26.3, 28.5, 28.6, 35.1, 42.4, 98.6, 117.3, 123.4, 123.6, 124.8, 125.2, 125.6, 134.4, 135.9, 137.4, 138.3, 146.1, 149.6, 151.2, 155.5, 165.3, 166.4, 167.5 HRMS Calcd for $\text{C}_{24}\text{H}_{22}\text{ClN}_3\text{O}_4$ $[\text{M}+\text{H}]^+$ 452.1299 found 452.1309.

2-(8-((7-chloroquinolin-4-yl)amino)octyl)-1,3-dioxoisindoline-5-carboxylic acid (5e): yellow solid; mp 245-246°C; ^1H NMR (400 MHz, DMSO- d_6) δ 1.23-1.35 (m, 8H, 4x- CH_2 -); 1.50-1.62 (m, 4H, 2x- CH_2 -); 3.28-3.47 (m, 2H, - CH_2 -); 3.56 (t, $J=5.9$ Hz, 2H, - CH_2 -); 6.65 (d, $J=5.5$ Hz, 1H, Ar-H); 7.41 (dd, $J=9.0$ Hz, 2.1 Hz, 1H, Ar-H); 7.63 (t, $J=6.0$ Hz, 1H, NH-exchangeable with D_2O); 7.74 (d, $J=2.1$ Hz, 1H, Ar-H); 7.83-7.90 (m, 1H, Ar-H); 8.00 (d, $J=9.0$ Hz, 1H, Ar-H); 8.15 (d, $J=1.2$ Hz, 1H, Ar-H); 8.22-8.29 (m, 1H, Ar-H); 8.40 (d, $J=5.4$ Hz, 1H, Ar-H); ^{13}C NMR (100 MHz, DMSO- d_6) δ 26.4, 26.7, 28.3, 28.6, 29.7, 29.8, 35.0, 42.3, 98.8, 117.2, 123.3, 123.7, 124.9, 125.1, 125.4, 134.2, 135.7, 137.1, 138.3, 146.2, 149.5, 151.3, 155.5, 165.2, 166.4, 167.4 HRMS Calcd for $\text{C}_{26}\text{H}_{26}\text{ClN}_3\text{O}_4$ $[\text{M}+\text{H}]^+$ 480.1612 found 480.1627.

*2-(2-((7-chloroquinolin-4-yl)amino)ethyl)-*N'*-isonicotinoyl-1,3-dioxoisindoline-5-carbohydrazide (6a)*: Yellow solid; mp 182-183°C; ^1H NMR (400 MHz, DMSO- d_6) δ 3.56-3.60 (m, 2H, - CH_2 -); 3.82 (t, $J=6.8$ Hz, 2H, - CH_2 -); 6.62 (d, $J=5.3$ Hz, 1H, Ar-H); 7.39 (dd, $J=9.1$ Hz, 2.1 Hz, 1H, Ar-H); 7.48 (t, $J=5.9$ Hz, 1H, NH-exchangeable with D_2O); 7.76 (d, $J=2.2$ Hz, 1H, Ar-H); 7.78-7.80 (m, 2H, Ar-H); 7.98-8.01 (m, 2H, Ar-H); 8.27-8.31 (m, 2H, Ar-H); 8.40 (s, 1H, Ar-H); 8.76 (d, $J=4.2$ Hz, 2H, Ar-H); 10.98 (s, 2H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, DMSO- d_6) δ 36.5, 40.7, 99.1, 118.1, 121.8, 121.9, 123.9,

124.3, 124.8, 128.0, 132.8, 133.9, 134.2, 135.0, 137.8, 139.7, 149.5, 150.3, 151.0, 152.4, 164.6, 164.7, 167.7, 167.8 HRMS Calcd for $C_{26}H_{19}ClN_6O_4$ $[M+H]^+$ 515.1156 found 515.1177.

2-(3-((7-chloroquinolin-4-yl)amino)propyl)-N'-isonicotinoyl-1,3-dioxoisoindoline-5-carbohydrazide (6b): Yellow solid; mp 175-176°C; 1H NMR (400 MHz, DMSO- d_6) δ 1.99-2.05 (m, 2H, $-CH_2-$); 3.31-3.38 (m, 2H, $-CH_2-$); 3.72 (t, $J=6.8$ Hz, 2H, $-CH_2-$); 6.44 (d, $J=5.5$ Hz, 1H, Ar-H); 7.23 (t, $J=5.5$ Hz, 1H, NH-exchangeable with D_2O); 7.39 (d, $J=9.1$ Hz, 1H, Ar-H); 7.73 (s, 1H, Ar-H); 7.79 (d, $J=5.6$ Hz, 2H, Ar-H); 7.96 (d, $J=8.1$ Hz, 1H, Ar-H); 8.17 (d, $J=9.1$ Hz, 1H, Ar-H); 8.24-8.35 (m, 2H, Ar-H); 8.35 (d, $J=5.4$ Hz, 1H, Ar-H); 8.76 (d, $J=5.2$ Hz, 2H, Ar-H); 10.91 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, DMSO- d_6) δ 27.1, 36.4, 40.6, 99.3, 117.9, 121.8, 121.9, 123.9, 124.60, 124.67, 127.8, 132.7, 134.0, 134.2, 135.0, 137.7, 139.8, 149.3, 150.6, 151.0, 152.2, 164.6, 164.8, 167.92, 167.93 HRMS Calcd for $C_{27}H_{21}ClN_6O_4$ $[M+H]^+$ 529.1313 found 529.1334.

2-(4-((7-chloroquinolin-4-yl)amino)butyl)-N'-isonicotinoyl-1,3-dioxoisoindoline-5-carbohydrazide (6c): Yellow solid; mp 162-163°C; 1H NMR (400 MHz, DMSO- d_6) δ 1.65-1.68 (m, 4H, 2x- $-CH_2-$); 3.29-3.35 (m, 2H, $-CH_2-$); 3.70 (t, $J=6.6$ Hz, 2H, $-CH_2-$); 6.42 (d, $J=5.4$ Hz, 1H, Ar-H); 7.21 (t, $J=5.5$ Hz, 1H, NH-exchangeable with D_2O); 7.37 (d, $J=9.0$ Hz, 1H, Ar-H); 7.71 (s, 1H, Ar-H); 7.78 (d, $J=5.7$ Hz, 2H, Ar-H); 7.94 (d, $J=8.2$ Hz, 1H, Ar-H); 8.15 (d, $J=9.0$ Hz, 1H, Ar-H); 8.21-8.32 (m, 2H, Ar-H); 8.34 (d, $J=5.5$ Hz, 1H, Ar-H); 8.77 (d, $J=5.2$ Hz, 2H, Ar-H); 10.89 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, DMSO- d_6) δ 25.6, 25.9, 35.9, 40.4, 99.2, 117.7, 121.6, 121.8, 123.6, 124.4, 124.8, 127.7, 132.6, 134.1, 134.4, 135.2, 137.8, 139.6, 149.2, 150.4, 151.1, 152.3, 164.6, 164.9, 167.5, 167.6 HRMS Calcd for $C_{28}H_{23}ClN_6O_4$ $[M+H]^+$ 543.1469 found 543.1487.

2-(6-((7-chloroquinolin-4-yl)amino)hexyl)-N'-isonicotinoyl-1,3-dioxoisoindoline-5-carbohydrazide (6d): Yellow solid; mp 158-159°C; 1H NMR (400 MHz, DMSO- d_6) δ 1.22-1.38 (m, 4H, 2x- $-CH_2-$); 1.50-1.61 (m, 4H, 2x- $-CH_2-$); 3.49-3.53 (m, 2H, $-CH_2-$); 3.68 (t, $J=6.7$ Hz, 2H, $-CH_2-$); 6.40 (d, $J=5.4$ Hz, 1H, Ar-H); 7.20 (t, $J=5.4$ Hz, 1H, NH-exchangeable with D_2O); 7.36 (d, $J=9.0$ Hz, 1H, Ar-H); 7.73 (s, 1H, Ar-H); 7.77 (d, $J=5.7$ Hz, 2H, Ar-H); 7.93 (d, $J=8.3$ Hz, 1H, Ar-H); 8.16 (d, $J=9.1$ Hz, 1H, Ar-H); 8.20-8.32 (m, 2H, Ar-H); 8.32 (d, $J=5.6$ Hz, 1H, Ar-H); 8.79 (d, $J=5.3$ Hz, 2H, Ar-H); 10.90 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, DMSO- d_6) δ 26.3, 26.7, 28.4, 28.6, 36.7, 40.1, 99.3, 117.7, 121.5, 121.8, 123.4, 124.5, 124.9, 127.6, 132.8, 134.3, 134.4, 135.5, 137.5,

139.8, 149.1, 150.5, 151.3, 152.4, 164.5, 164.9, 167.4, 167.7 HRMS Calcd for $C_{30}H_{27}ClN_6O_4$ $[M+H]^+$ 571.1782 found 571.1793.

2-(8-((7-chloroquinolin-4-yl)amino)octyl)-N'-isonicotinoyl-1,3-dioxoisindoline-5-carbohydrazide (6e): Yellow solid; mp 141-142°C; 1H NMR (400 MHz, DMSO- d_6) δ 1.20-1.36 (m, 8H, 4x- CH_2 -); 1.52-1.65 (m, 4H, 2x- CH_2 -); 3.29-3.45 (m, 2H, - CH_2 -); 3.59 (t, $J=6.5$ Hz, 2H, - CH_2 -); 6.39 (d, $J=5.5$ Hz, 1H, Ar-H); 7.19 (t, $J=5.4$ Hz, 1H, NH-exchangeable with D_2O); 7.34 (d, $J=8.9$ Hz, 1H, Ar-H); 7.72 (s, 1H, Ar-H); 7.75 (d, $J=5.6$ Hz, 2H, Ar-H); 7.94 (d, $J=8.3$ Hz, 1H, Ar-H); 8.14 (d, $J=9.0$ Hz, 1H, Ar-H); 8.22-8.32 (m, 2H, Ar-H); 8.34 (d, $J=5.5$ Hz, 1H, Ar-H); 8.78 (d, $J=5.4$ Hz, 2H, Ar-H); 10.90 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, DMSO- d_6) δ 26.4, 26.6, 28.7, 28.9, 29.5, 29.8, 35.9, 40.4, 99.1, 117.5, 121.7, 121.9, 123.4, 124.8, 125.1, 127.3, 132.7, 134.3, 134.6, 135.7, 137.6, 139.9, 149.0, 150.6, 151.2, 152.5, 164.4, 164.8, 167.3, 167.6 HRMS Calcd for $C_{32}H_{31}ClN_6O_4$ $[M+H]^+$ 599.2095 found 599.2082.

2-benzamido-1,3-dioxoisindoline-5-carboxylic acid (7): Light yellow solid; mp >300 °C; 1H NMR (400 MHz, DMSO- d_6) δ 7.83-7.84 (m, 2H, Ar-H); 8.10-8.12 (m, 1H, Ar-H); 8.32-8.33 (m, 1H, Ar-H); 8.42-8.44 (m, 1H, Ar-H); 8.81-8.82 (m, 2H, Ar-H); 11.78 (s, 1H, NH-exchangeable with D_2O); HRMS Calcd for $C_{16}H_{10}N_2O_5$ $[M+H]^+$ 311.0590 found 311.0598.

N-(2-((7-chloroquinolin-4-yl)amino)ethyl)-2-(isonicotinamido)-1,3-dioxoisindoline-5-carboxamide (8a): Brown solid; mp 186-187°C; 1H NMR (400 MHz, DMSO- d_6) δ 3.52-3.59 (m, 2H, - CH_2 -); 3.77-3.85 (m, 2H, - CH_2 -); 6.63 (d, $J=5.2$ Hz, 1H, Ar-H); 7.32-7.44 (m, 2H, Ar-H); 7.72-7.79 (m, 3H, Ar-H); 7.94-8.01 (m, 2H, Ar-H); 8.21-8.41 (m, 3H, Ar-H); 8.70-8.77 (m, 2H, Ar-H); 10.92 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, DMSO- d_6) δ 35.2, 41.6, 106.8, 118.6, 121.2, 122.1, 126.9, 127.2, 129.8, 130.9, 131.8, 136.3, 137.1, 139.6, 140.7, 142.7, 144.3, 151.0, 151.6, 156.7, 161.4, 162.3, 164.6, 172.8 HRMS Calcd for $C_{26}H_{19}ClN_6O_4$ $[M+H]^+$ 515.1156 found 515.1171.

N-(3-((7-chloroquinolin-4-yl)amino)propyl)-2-(isonicotinamido)-1,3-dioxoisindoline-5-carboxamide (8b): Brown solid; mp 181-182°C; 1H NMR (400 MHz, DMSO- d_6) δ 1.80-1.86 (m, 2H, - CH_2 -); 3.48-3.51 (m, 2H, - CH_2 -); 3.74-3.82 (m, 2H, - CH_2 -); 6.61 (d, $J=5.3$ Hz, 1H, Ar-H); 7.30-7.41 (m, 2H, Ar-H); 7.71-7.80 (m, 3H, Ar-H); 7.92-8.00 (m, 2H, Ar-H); 8.23-8.38 (m, 3H, Ar-H); 8.71-8.75 (m, 2H, Ar-H); 10.91 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, DMSO- d_6) δ 28.6, 35.1, 41.6, 105.5, 118.0, 121.3, 122.1, 126.6, 127.0,

129.4, 130.5, 131.5, 135.7, 136.8, 139.2, 140.4, 142.6, 144.4, 150.9, 151.6, 157.1, 161.4, 162.0, 164.2, 172.5 HRMS Calcd for C₂₇H₂₁ClN₆O₄ [M+H]⁺ 529.1313 found 529.1326.

N-(4-((7-chloroquinolin-4-yl)amino)butyl)-2-(isonicotinamido)-1,3-dioxoisindoline-5-carboxamide (**8c**): Brown solid; mp 178-179°C; ¹H NMR (400 MHz, DMSO-d₆) δ 1.50-1.61 (m, 2H, -CH₂-); 1.69-1.75 (m, 2H, -CH₂-); 3.35-3.39 (m, 2H, -CH₂-); 3.59-3.63 (m, 2H, -CH₂-); 6.60 (d, *J*=5.2 Hz, 1H, Ar-H); 7.32-7.40 (m, 2H, Ar-H); 7.70-7.79 (m, 3H, Ar-H); 7.91-8.01 (m, 2H, Ar-H); 8.24-8.37 (m, 3H, Ar-H); 8.70-8.74 (m, 2H, Ar-H); 10.92 (s, 1H, NH-exchangeable with D₂O); ¹³C NMR (100 MHz, DMSO-d₆) δ 26.3, 27.1, 37.5, 40.9, 105.4, 118.1, 121.4, 122.5, 126.4, 127.2, 129.4, 130.6, 131.2, 135.9, 136.7, 139.1, 140.3, 142.3, 144.5, 150.7, 151.5, 157.0, 161.2, 162.1, 164.1, 172.2 HRMS Calcd for C₂₈H₂₃ClN₆O₄ [M+H]⁺ 543.1469 found 543.1478.

N-(6-((7-chloroquinolin-4-yl)amino)hexyl)-2-(isonicotinamido)-1,3-dioxoisindoline-5-carboxamide (**8d**): Brown solid; mp 172-173°C; ¹H NMR (400 MHz, DMSO-d₆) δ 1.30-1.38 (m, 4H, 2x-CH₂-); 1.54-1.60 (m, 2H, -CH₂-); 1.65-1.70 (m, 2H, -CH₂-); 3.31-3.39 (m, 2H, -CH₂-); 3.56-3.59 (m, 2H, -CH₂-); 6.61 (d, *J*=5.3 Hz, 1H, Ar-H); 7.30-7.41 (m, 2H, Ar-H); 7.71-7.78 (m, 3H, Ar-H); 7.91-8.00 (m, 2H, Ar-H); 8.23-8.36 (m, 3H, Ar-H); 8.71-8.74 (m, 2H, Ar-H); 10.92 (s, 1H, NH-exchangeable with D₂O); ¹³C NMR (100 MHz, DMSO-d₆) δ 26.1, 27.3, 29.3, 29.7, 38.4, 40.8, 105.3, 118.0, 121.5, 122.4, 126.4, 127.1, 129.3, 130.7, 131.1, 135.8, 136.6, 139.0, 140.4, 142.5, 144.6, 150.8, 151.6, 157.1, 161.3, 162.0, 164.3, 172.4 HRMS Calcd for C₃₀H₂₇ClN₆O₄ [M+H]⁺ 571.1782 found 571.1789.

N-(8-((7-chloroquinolin-4-yl)amino)octyl)-2-(isonicotinamido)-1,3-dioxoisindoline-5-carboxamide (**8e**): Brown solid; mp 165-166°C; ¹H NMR (400 MHz, DMSO-d₆) δ 1.28-1.39 (m, 8H, 4x-CH₂-); 1.51-1.65 (m, 4H, 2x-CH₂-); 3.30-3.36 (m, 2H, -CH₂-); 3.54-3.58 (m, 2H, -CH₂-); 6.60 (d, *J*=5.4 Hz, 1H, Ar-H); 7.29-7.41 (m, 2H, Ar-H); 7.70-7.77 (m, 3H, Ar-H); 7.89-7.99 (m, 2H, Ar-H); 8.21-8.34 (m, 3H, Ar-H); 8.68-8.72 (m, 2H, Ar-H); 10.91 (s, 1H, NH-exchangeable with D₂O); ¹³C NMR (100 MHz, DMSO-d₆) δ 25.6, 27.3, 28.5, 28.7, 29.3, 29.7, 38.8, 41.6, 105.2, 118.1, 121.6, 122.3, 126.5, 127.0, 129.5, 130.4, 131.0, 135.9, 136.6, 139.0, 140.3, 142.6, 144.8, 150.7, 151.6, 157.0, 161.2, 162.4, 164.2, 172.3 HRMS Calcd for C₃₂H₃₁ClN₆O₄ [M+H]⁺ 599.2095 found 599.2103.

2-((7-chloroquinolin-4-yl)amino)ethyl-2-(isonicotinamido)-1,3-dioxoisindoline-5-carboxylate (**8f**): Brown solid; mp 162-163°C; ¹H NMR (400 MHz, DMSO-d₆) δ 3.68-3.72

(m, 2H, $-\text{CH}_2-$); 4.32 (t, $J=6.1$ Hz, 2H, $-\text{CH}_2-$); 6.45 (d, $J=5.3$ Hz, 1H, Ar-H); 6.52 (t, $J=5.8$ Hz, 1H, NH-exchangeable with D_2O); 7.35 (dd, $J=9.0$ Hz, 2.1 Hz, 1H, Ar-H); 7.74-7.79 (m, 3H, Ar-H); 7.85-7.93 (m, 2H, Ar-H); 8.20-8.26 (m, 2H, Ar-H); 8.53 (d, $J=5.5$ Hz, 1H, Ar-H); 8.72-8.78 (m, 2H, Ar-H); 11.01 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 42.2, 60.4, 101.6, 118.4, 121.4, 122.4, 125.0, 125.4, 126.6, 127.3, 132.4, 132.7, 134.7, 135.5, 136.8, 138.4, 147.9, 149.3, 150.1, 150.8, 164.2, 165.1, 165.4, 166.2 HRMS Calcd for $\text{C}_{26}\text{H}_{18}\text{ClN}_5\text{O}_5$ $[\text{M}+\text{H}]^+$ 516.0996 found 516.0987.

3-((7-chloroquinolin-4-yl)amino)propyl-2-(isonicotinamido)-1,3-dioxoisindoline-5-carboxylate (8g): Brown solid; mp 156-157°C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 1.96-2.01 (m, 2H, $-\text{CH}_2-$); 3.57-3.68 (m, 2H, $-\text{CH}_2-$); 4.12 (t, $J=6.2$ Hz, 2H, $-\text{CH}_2-$); 6.44 (d, $J=5.4$ Hz, 1H, Ar-H); 6.51 (t, $J=5.7$ Hz, 1H, NH-exchangeable with D_2O); 7.32 (dd, $J=8.9$ Hz, 2.0 Hz, 1H, Ar-H); 7.72-7.77 (m, 3H, Ar-H); 7.84-7.93 (m, 2H, Ar-H); 8.21-8.25 (m, 2H, Ar-H); 8.54 (d, $J=5.6$ Hz, 1H, Ar-H); 8.70-8.76 (m, 2H, Ar-H); 11.01 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 28.2, 42.5, 60.1, 101.5, 118.2, 121.4, 122.5, 125.1, 125.2, 126.7, 127.2, 132.3, 132.6, 134.7, 135.6, 136.5, 138.5, 147.8, 149.2, 150.2, 150.9, 164.1, 165.2, 165.3, 166.1 HRMS Calcd for $\text{C}_{27}\text{H}_{20}\text{ClN}_5\text{O}_5$ $[\text{M}+\text{H}]^+$ 530.1153 found 530.1166.

N-(5-fluoro-1,3-dioxoisindolin-2-yl)benzamide (9): White solid; mp 259-260°C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.74-7.79 (m, 1H, Ar-H); 7.82-7.84 (m, 2H, Ar-H); 7.92-7.94 (m, 1H, Ar-H); 8.06-8.09 (m, 1H, Ar-H); 8.80-8.82 (m, 2H, Ar-H); 11.73 (s, 1H, NH-exchangeable with D_2O); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 107.9, 114.5, 117.8, 121.9, 126.4, 132.9, 138.4, 151.2, 156.2, 164.6, 165.5, 166.2. HRMS Calcd for $\text{C}_{15}\text{H}_9\text{FN}_2\text{O}_3$ $[\text{M}+\text{H}]^+$ 285.0597 found 285.0585.

N-(5-(4-(7-chloroquinolin-4-yl)piperazin-1-yl)-1,3-dioxoisindolin-2-yl)isonicotinamide (11): Yellow solid; mp 152-153°C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 3.32-3.34 (m, 4H, 2x- CH_2-); 3.75-3.77 (m, 4H, 2x- CH_2-); 7.04 (d, $J=5.0$ Hz, 1H, Ar-H); 7.38 (dd, $J=8.6$ Hz, 2.4 Hz, 1H, Ar-H); 7.49 (d, $J=2.2$ Hz, 1H, Ar-H); 7.54-7.57 (m, 1H, Ar-H); 7.77-7.79 (m, 2H, Ar-H); 7.82-7.83 (m, 2H, Ar-H); 7.97 (d, $J=2.2$ Hz, 1H, Ar-H); 8.11 (d, $J=9.0$ Hz, 1H, Ar-H); 8.71 (d, $J=5.0$ Hz, 1H, Ar-H); 8.80-8.81 (m, 2H, Ar-H); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 47.1, 53.4, 108.9, 110.0, 117.0, 118.8, 121.9, 126.1, 126.4, 126.6, 128.6, 132.5, 134.2, 138.3, 150.1, 150.9, 151.3, 152.7, 155.8, 156.4, 164.4, 165.4, 166.1 HRMS Calcd for $\text{C}_{27}\text{H}_{21}\text{ClN}_6\text{O}_3$ $[\text{M}+\text{H}]^+$ 513.1364 found 513.1387.

N'-(2-(1,3-dioxoisindolin-2-yl)acetyl)isonicotinohydrazide (13a): White solid; mp 255-256°C; ¹H NMR (400 MHz, DMSO-d₆) δ 4.32 (s, 2H, -CH₂-); 7.71 (d, *J*=6.0 Hz, 1H, Ar-H); 7.82-7.91 (m, 4H, Ar-H); 8.71 (d, *J*=6.0 Hz, 1H, Ar-H); 10.48 (s, 1H, NH-exchangeable with D₂O); 10.77 (s, 1H, NH-exchangeable with D₂O); ¹³C NMR (100 MHz, DMSO-d₆) δ 39.4, 123.8, 123.9, 132.1, 135.2, 143.9, 147.1, 163.0, 166.2, 167.9 HRMS Calcd for C₁₆H₁₂N₄O₄ [M+H]⁺ 325.0859 found 325.0877.

N'-(2-(1,3-dioxoisindolin-2-yl)acetyl)isonicotinohydrazide (13b): White solid; mp 267-268°C; ¹H NMR (300 MHz, DMSO-d₆) δ 7.61 (s, 1H, Ar-H); 7.80-8.02 (m, 8H, Ar-H); 8.76 (s, 1H, Ar-H); 10.85 (s, 2H, NH-exchangeable with D₂O); HRMS Calcd for C₂₁H₁₄N₄O₄ [M+H]⁺ 387.1015 found 387.1026.

¹H NMR of 2-(2-((7-chloroquinolin-4-yl)amino)ethyl)-1,3-dioxoisindoline-5-carboxylic acid (5a):

¹³C NMR of 2-(2-((7-chloroquinolin-4-yl)amino)ethyl)-1,3-dioxoisindoline-5-carboxylic acid (5a):

¹H NMR of 2-(2-((7-chloroquinolin-4-yl)amino)ethyl)-N'-isonicotinoyl-1,3-dioxisoindoline-5-carbohydrazide (6a):

¹³C NMR of 2-(2-((7-chloroquinolin-4-yl)amino)ethyl)-N'-isonicotinoyl-1,3-dioxisoindoline-5-carbohydrazide (6a):

¹H NMR of 2-(3-((7-chloroquinolin-4-yl)amino)propyl)-N'-isonicotinoyl-1,3-dioxisoindoline-5-carbohydrazide (6b):

^{13}C NMR of 2-(3-((7-chloroquinolin-4-yl)amino)propyl)-N'-isonicotinoyl-1,3-dioxisoindoline-5-carbohydrazide (6b):

^1H NMR of 2-benzamido-1,3-dioxisoindoline-5-carboxylic acid (7):

^1H NMR of N-(2-((7-chloroquinolin-4-yl)amino)ethyl)-2-(isonicotinamido)-1,3-dioxisoindoline-5-carboxamide (8a):

^{13}C NMR of N-(2-((7-chloroquinolin-4-yl)amino)ethyl)-2-(isonicotinamido)-1,3-dioxisoindoline-5-carboxamide (8a):

¹H NMR of N-(5-fluoro-1,3-dioxoisindolin-2-yl)benzamide (9):

¹³C NMR of N-(5-fluoro-1,3-dioxoisindolin-2-yl)benzamide (9):

¹H NMR of N-(5-(4-(7-chloroquinolin-4-yl)piperazin-1-yl)-1,3-dioxoisindolin-2-yl)isonicotinamide (11):

^{13}C NMR of N-(5-(4-(7-chloroquinolin-4-yl)piperazin-1-yl)-1,3-dioxisoindolin-2-yl)isonicotinamide (11):

^1H NMR of N'-(2-(1,3-dioxisoindolin-2-yl)acetyl)isonicotinohydrazide (13a):

^{13}C NMR of N'-(2-(1,3-dioxisoindolin-2-yl)acetyl)isonicotinohydrazide (13a):

Declaration of interests

☒ The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.