

**Emerging Challenges of Preclinical Models of  
Anti-tumor Immunotherapeutic Strategies Utilizing  
V $\gamma$ 9V $\delta$ 2 T Cells**

Noémie Joalland, Emmanuel Scotet

► **To cite this version:**

Noémie Joalland, Emmanuel Scotet. Emerging Challenges of Preclinical Models of Anti-tumor Immunotherapeutic Strategies Utilizing V $\gamma$ 9V $\delta$ 2 T Cells. *Frontiers in Immunology*, Frontiers, 2020, 11, pp.992. 10.3389/fimmu.2020.00992 . inserm-02908442

**HAL Id: inserm-02908442**

**<https://www.hal.inserm.fr/inserm-02908442>**

Submitted on 29 Jul 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



# Emerging Challenges of Preclinical Models of Anti-tumor Immunotherapeutic Strategies Utilizing V $\gamma$ 9V $\delta$ 2 T Cells

Noémie Joalland<sup>1,2</sup> and Emmanuel Scotet<sup>1,2\*</sup>

<sup>1</sup> Université de Nantes, INSERM, CNRS, CRCINA, Nantes, France, <sup>2</sup> LabEx IGO "Immunotherapy, Graft, Oncology", Nantes, France

## OPEN ACCESS

### Edited by:

Dieter Kabelitz,  
University of Kiel, Germany

### Reviewed by:

Serena Meraviglia,  
University of Palermo, Italy  
Thomas H. Winkler,  
University of Erlangen  
Nuremberg, Germany

### \*Correspondence:

Emmanuel Scotet  
emmanuel.scotet@inserm.fr

### Specialty section:

This article was submitted to  
T Cell Biology,  
a section of the journal  
Frontiers in Immunology

**Received:** 14 February 2020

**Accepted:** 27 April 2020

**Published:** 22 May 2020

### Citation:

Joalland N and Scotet E (2020)  
Emerging Challenges of Preclinical  
Models of Anti-tumor  
Immunotherapeutic Strategies Utilizing  
V $\gamma$ 9V $\delta$ 2 T Cells.  
Front. Immunol. 11:992.  
doi: 10.3389/fimmu.2020.00992

Despite recent advances, the eradication of cancers still represents a challenge which justifies the exploration of additional therapeutic strategies such as immunotherapies, including adoptive cell transfers. Human peripheral V $\gamma$ 9V $\delta$ 2 T cells, which constitute a major transitional immunity lymphocyte subset, represent attractive candidates because of their broad and efficient anti-tumor functions, as well as their lack of alloreactivity and easy handling. V $\gamma$ 9V $\delta$ 2 T cells act like immune cell stress sensors that can, in a tightly controlled manner but through yet incompletely understood mechanisms, detect subtle changes of levels of phosphorylated metabolites of isoprenoid synthesis pathways. Consequently, various anti-tumor immunotherapeutic strategies have been proposed to enhance their reactivity and cytotoxicity, as well as to reduce the deleterious events. In this review, we expose these advances based on different strategies and their validation in preclinical models. Importantly, we next discuss advantages and limits of each approach, by highlighting the importance of the use of relevant preclinical model for evaluation of safety and efficacy. Finally, we propose novel perspectives and strategies that should be explored using these models for therapeutic improvements.

**Keywords:** human V $\gamma$ 9V $\delta$ 2 T lymphocytes, cancer, functions, immunotherapy, preclinical models

## BIOLOGY AND ANTI-TUMOR FUNCTIONS OF HUMAN V $\gamma$ 9V $\delta$ 2 T CELLS

The complex immune system orchestrates with molecular and cellular components that act concomitantly or sequentially to sense and to eliminate potential exogenous and endogenous threats. In vertebrates, the innate immunity is the first germline-encoded and older evolutionary defense strategy that contributes to a rapid, but poorly specific, reactivity that also remains essential for recruiting immune cells and establishing a physical and chemical barrier. A major hallmark of the adaptive immunity, that is associated to immunological memory and next takes place after this initial process, is the extreme specificity against particular antigenic structures. This slower, but highly potent, immune reactivity axis is made up of the two main T and B lymphocyte effector subsets. Recently, a growing class of additional cellular contributors, that express either TCR (*T Cell Receptor*) or BCR (*B Cell Receptor*) molecules and share phenotypical and functional characteristics from both systems has been identified. They have been differentially named as transitional immunity, unconventional or innate-like effectors. Most well-described cell subsets

that fall at this interface between innate and adaptive immunities are NKT (*Natural Killer T*), MAIT (*Mucosal Associated Invariant T*) and  $\gamma\delta$  T cells (1). This latter T cell subset expresses a heterodimeric TCR composed of  $\gamma$  and  $\delta$  chains, named as opposed to  $\alpha$  and  $\beta$  chains, associated to the CD3 signaling complex (2, 3). It is important to note that, although  $\gamma\delta$  T cell subsets are present in most vertebrates, there is little conservation of  $\gamma$  and  $\delta$  TCR chains and reactivities between species. This has been evidenced by genetic studies indicating a high heterogeneity between murine and primate TCR genes (4, 5).

For the sake of clarity, we propose to focus this present review on human  $\gamma\delta$  T cell biology, and more precisely, the human V $\gamma$ 9V $\delta$ 2 T cell subset, as well as their therapeutic targeting in preclinical cancer models. The  $\gamma\delta$  TCR is composed from a limited number of  $\gamma$  and  $\delta$  chains (<15 V $\gamma$  and V $\delta$  gene segments) but this low combinatorial diversity is efficiently counterbalanced by both an elevated junctional diversity and pairing assembly restrictions to finally produce an extremely diverse  $\gamma\delta$  TCR. In humans,  $\gamma\delta$  T cells are distributed into four major subsets, which have been identified according to the expression of  $\delta$  chain segments, as V $\delta$ 1<sup>+</sup>, V $\delta$ 2<sup>+</sup>, V $\delta$ 3<sup>+</sup>, and V $\delta$ 5<sup>+</sup> populations (6). Following ontogeny, most  $\gamma\delta$  T cell subsets naturally exhibit a preferential tropism for particular tissues/organs, through yet unclear mechanisms. As a typical example of this preferential localization, human V $\gamma$ 9V $\delta$ 2 T cells [using the nomenclature of Lefranc & Rabbitts (7)] constitute a major subset in adult peripheral blood (8, 9). V $\gamma$ 9V $\delta$ 2 T cells represent prototypical immune sensors of cellular stress activated under various pathological contexts, such as infections and cancer. The species-specific antigenic activation of V $\gamma$ 9V $\delta$ 2 T cells is a contact- and a TCR-dependent process. It is important to note that this activation is not restricted by either conventional MHC class I or II molecules, thus theoretically minimizing the risk of alloreactive reactions for allogeneic V $\gamma$ 9V $\delta$ 2 T cell-therapies (e.g., GVHD, *graft vs. host disease*). This antigenic activation process implicates mandatory small phosphorylated carbohydrate metabolites (hereafter called phosphoantigens, PAg), such as IPP (*Isopentenyl PyroPhosphate*), which endogenous (MEV mammalian mevalonate pathway), or exogenous (MEP microbial pathway) expression might be altered in some pathological contexts (e.g., cancer, infections). Recent studies have shown that PAg levels are sensed through mechanisms involving both BTN2A1 and BTN3A1 butyrophilins (BTN) expressed by target cells (10–12). Accordingly, pharmacological compounds that inhibit the synthesis (i.e., statins) or the degradation (i.e., aminobisphosphonates, alkylamines) of PAg in mammalian cells can block, or induce, the antigenic activation of V $\gamma$ 9V $\delta$ 2 T cells, respectively (13, 14). Importantly, this *Self*-reactive nature of V $\gamma$ 9V $\delta$ 2 T cells needs to be tightly regulated by a set of various molecules [see (15) and (16), for recent reviews], such as adhesion molecules (i.e., CD54), activating (i.e., NKG2D) or inhibiting (i.e., CD94/NKG2A), NKR (*Natural Killer Receptors*), FcR (*Fc Receptor*) (i.e., Fc $\gamma$ RIIIA/CD16), Nectin/Nectin-like (i.e., CD226), TLR (*Toll-like Receptor*) (i.e., TLR4), cytokine receptors

(i.e., IL (*interleukin*)-15R, IL-21R), and immune checkpoint inhibitors (i.e., PD-1, *programmed cell death protein 1*). Hence, numerous V $\delta$ 2 T cell dysfunctions in cancer indications (e.g., hypo-reactivity, exhaustion) have been associated to altered expression profiles of these molecules. In this exquisite sensing process, V $\gamma$ 9V $\delta$ 2 T cells integrate activatory and inhibitory signals to rapidly deliver strong functional responses such as proliferation, cytotoxicity (through perforin/granzyme-, TRAIL (*TNF-related apoptosis inducing ligand*)-, CD95-pathways) and cytokines (i.e., TNF (*Tumor Necrosis Factor*)- $\alpha$ , IFN (*Interferon*)- $\gamma$ )/chemokines (i.e., CCL3, *Chemokine (C-C motif) ligand 3*)/anti-microbial factors (i.e., granulysin)/epithelial growth factors (i.e., KGF, *Keratinocyte Growth Factor*) release. These latter functions are also linked to their capacity to help other immune effectors and induce the maturation of antigen-presenting cells, including themselves (9). Finally, as for most lymphocyte subsets, the migration of V $\gamma$ 9V $\delta$ 2 T cells is tightly controlled by a set of chemoattractant factors, such as chemokine receptors. Their expression regulates V $\gamma$ 9V $\delta$ 2 T cell trafficking during physiological and inflammatory conditions (e.g., CCR5, *Chemokine (C-C motif) Receptor 5*), a process that is proposed to be of particular importance for tumor addressing and infiltration *in vivo*.

For a long time, a set of compelling *in vitro* studies evidenced the natural reactivity of human V $\gamma$ 9V $\delta$ 2 T cells against a broad range of human tumor cell lines and normal cells infected by a variety of viruses, parasites and bacteria (17–19). With respect to transformed cells, the range of cell lines recognized by V $\gamma$ 9V $\delta$ 2 T cells, initially thought to be primarily restricted to hematopoietic tumors (20, 21), was next extended to several solid tumors, such as renal and colon carcinomas (22–24). Importantly, this vision has been next modified by the availability of aminobisphosphonates (e.g., pamidronate, zoledronate) and synthetic PAg (e.g., BrHPP, *BromoHydrin Pyrophosphate*) that can further help sensitizing a broad variety of cells to V $\gamma$ 9V $\delta$ 2 T cell sensing and elimination. Like for most  $\gamma\delta$  T cell subsets, *in vitro* studies showed that V $\gamma$ 9V $\delta$ 2 T cells are able to directly kill target cells and express pro-inflammatory cytokines that can be also involved in the clearance of tumor cells (25, 26). Altogether, these *in vitro* observations supported a natural implication of V $\gamma$ 9V $\delta$ 2 T cells in protective anti-tumor immunity. Based on initial results indicating an altered tumor growth control in TCR  $\delta^{\text{neg}}$  mice (27), several *in vivo* studies showed that transferred allogeneic V $\gamma$ 9V $\delta$ 2 T cells can reach and infiltrate tumor site and display a strong anti-tumor activity as evidenced by significant clinical benefits (e.g., survival, tumor growth) (28, 29). The implication of V $\gamma$ 9V $\delta$ 2 T cells in the anti-tumor immune reactivity is supported by the fact that infiltrating  $\gamma\delta$  T cells are considered as a favorable cancer prognosis marker for several cancers (30, 31), V $\delta$ 2 T cells infiltrating tumors were detected in various types of cancer. However, their precise physiological role might vary from one condition to another, mainly due to the heterogeneity of the tumor microenvironment which can modulate their functions as well as their functional plasticity (30, 31).

## RATIONALE FOR HARNESSING V $\gamma$ 9V $\delta$ 2 T CELLS IN CANCER IMMUNOTHERAPY

Human V $\gamma$ 9V $\delta$ 2 T cells should be considered as attractive immune effectors of high therapeutic potential for the main following reasons:

1. Inter-individual conservation and elevated frequency in the peripheral blood of human adults;
2. Antigenic specificity linked to cell stress-associated molecules whose expression is frequently dysregulated in cancer cells;
3. Clinical-grade synthetic agonist molecules, such as aminobisphosphonates and PAg, that specifically induce activation, expansion and sensitization of human tumor cells;
4. Simple handling and elevated in/ex vivo expansion index;
5. Absence of alloreactivity (no MHC class I/II restrictions);
6. Capacity to reach and infiltrate tumors;
7. Direct and indirect cytotoxic activities against tumor cells, through the secretion of lytic molecules and pro-inflammatory cytokines.

## SUCCESSSES AND LIMITATIONS OF V $\gamma$ 9V $\delta$ 2 T CELL CANCER IMMUNOTHERAPIES

Several types of immunotherapies that aim at helping the immune system to better react against tumor cells, are used to treat cancer. They include immune checkpoint inhibitors, monoclonal antibodies and immune cell therapy. In this latter category, active and passive immunotherapies are distinguished, according to the approaches developed for inducing V $\gamma$ 9V $\delta$ 2 T cell activation and expansion.

Regarding active immunotherapies, several strategies have been considered to obtain *in vivo* activation of V $\gamma$ 9V $\delta$ 2 T cell effectors induced following administration(s) of specific clinical-grade agonist molecules, such as PAg or aminobisphosphonates, together with pro-proliferating cytokines (e.g., IL-2) (32, 33). These approaches originated from initial observations describing increased frequencies of peripheral V $\gamma$ 9V $\delta$ 2 T cells in hematological cancer patients treated with pamidronate (34). In patients with non-Hodgkin's lymphoma or multiple myeloma, systemic administrations of both pamidronate with IL-2 were tolerated by patients and induced expansions of endogenous peripheral V $\gamma$ 9V $\delta$ 2 T cells, accompanied by partial remissions of cancer in some patients (35). Next, this strategy was applied to solid tumors (i.e., non-hormonal prostate cancer) and showed that activation of V $\gamma$ 9V $\delta$ 2 T cells *in vivo* was associated with the development of a pro-inflammatory (IFN- $\gamma$ ) responses (36). Following these first encouraging results, several clinical trials have been conducted in patients with renal cell carcinoma or bone metastases deriving from breast or prostate cancers (32, 33). These studies have demonstrated therapeutic responses such as stabilized diseases and partial remissions in some patients (37–39). More recently, the efficacy of this strategy was improved in patients with malignant hemopathies receiving haploidentical donor lymphocyte infusion (40). Importantly, the majority of

treated patients in these trials experienced mild side effects (i.e., flu-like syndrome), likely associated to IL-2, thus confirming the reduced toxicity of this strategy. To further improve its specificity, synthetic PAg compounds have been produced at a clinical grade (i.e., BrHPP) and tested *in vivo*. In metastatic renal cell carcinoma patients who received repeated infusions of BrHPP and IL-2, potent V $\gamma$ 9V $\delta$ 2 T cell expansions with moderate clinical activities were observed (41). These phase I/0 clinical assays showed a satisfactory feasibility of this approach, with a reduced toxicity (mainly due to IL-2) and excellent V $\gamma$ 9V $\delta$ 2 T cell expansion rates, but modest clinical efficacies, that might have been complicated by the bad clinical condition of cancer patients. Further studies, including assays in monkeys, next pointed out potential issues such as the progressive exhaustion of V $\gamma$ 9V $\delta$ 2 T cells (i.e., tachyphylaxis), altered immune status, activation contexts and tissue homing, that should be solved to improve the therapeutic efficacy of this strategy in cancer treatment (42).

Passive immunotherapies, which are based on *ex vivo* PBL (Peripheral Blood Lymphocytes)-V $\gamma$ 9V $\delta$ 2 T cell expansions and cell transfer(s) have been tested in renal carcinoma patients with reduced toxicity by low therapeutic efficacy (43). Next, several clinical trials have been carried out in patients with circulating or solid cancers, who have been treated by adoptive transfer of autologous V $\gamma$ 9V $\delta$ 2 T cells, amplified *ex vivo*, associated with IL-2 and zoledronate (32, 33). In general, these studies indicated a good feasibility but a low, although promising, therapeutic efficacy of this therapy as evidenced by some partial/complete cancer remissions. Clinical trials of adoptive transfer of allogeneic V $\gamma$ 9V $\delta$ 2 T cells are currently underway. Recently, a case report of a patient with a stage IV cholangiocarcinoma showing recurrent mediastinal lymph node metastasis after liver transplantation was published. This patient received consecutive infusions of allogeneic V $\gamma$ 9V $\delta$ 2 T cells which have been expanded from PBMC (Peripheral Blood Mononuclear Cells) of a healthy donor. No adverse effects were detected and a significant clinical response, with no detectable peritoneal lymph node metastasis, was reported at the end of treatment (44). Improved strategies for redirecting immune cell effectors to target cells expressing  $\gamma\delta$  T cell tumor antigens are being generated and tested for therapy (45). Thanks to the significant advance of genetic engineering and transduction approaches, the efficient expression of natural or optimized TCR (e.g.,  $\gamma\delta$  TCR) can be realized in lymphocytes (46). The clinical-grade production of these engineered effectors [e.g., CAR (Chimeric Antigen Receptor)  $\gamma\delta$  T cells] has recently been validated and is currently being tested in patients with leukemia or multiple myeloma (33, 46).

Though promising (e.g., feasibility, safety), the results of these clinical trials, though exploratory or phase 0/I only, stressed the urgent need of developing new V $\gamma$ 9V $\delta$ 2 T cell-immunotherapies (e.g., combined therapies) and optimizing their use (e.g., clinical positioning) to significantly improve their clinical efficacy in circulating and solid oncological indications. The achievement of this ambitious goal should require the use of robust physiological preclinical models *in vivo* for assessing both the feasibility and efficacy parameters of these strategies.

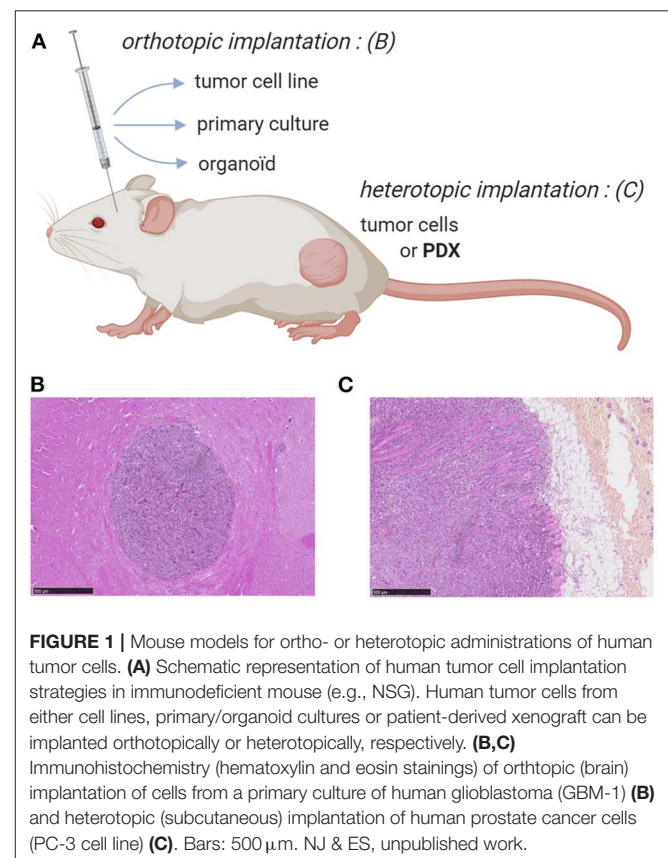
## IMPLEMENTATION OF PRECLINICAL CANCER MODELS AND V $\gamma$ 9V $\delta$ 2 IMMUNOTHERAPIES

A substantial number of preclinical *in vivo* cancer models, including, for some of them, V $\gamma$ 9V $\delta$ 2 T cell immunotherapies, have been carried out using heterotopic mouse models established with subcutaneous injections of cultured human tumor cell lines. Among various advantages, these approaches allow a fast and easy monitoring of tumor growth by physical measurement of tumor volume (47). However, these fast growing models, also generally used for non-V $\gamma$ 9V $\delta$ 2 T cell therapies, rarely faithfully recapitulate the complexity and heterogeneity of human oncological disease (e.g., origin of tumor cells, nature of the environment), also because of the large number of cells implanted in this particular localization (48, 49). Another problem is linked to their limited ability to disseminate (e.g., metastasis). To address this problem, it is important to orthotopically implant tumor cells (i.e., corresponding anatomical position) and, if possible, in a minimal quantity to further mimic the first stages of tumor development and its natural dissemination, according to the organ of origin (Figure 1). Importantly, the growing tumor should be characterized using various approaches such as imaging, histology or analysis of the phenotype of tumor cells to accurately determine the relevance of this model *in vivo* (50–52).

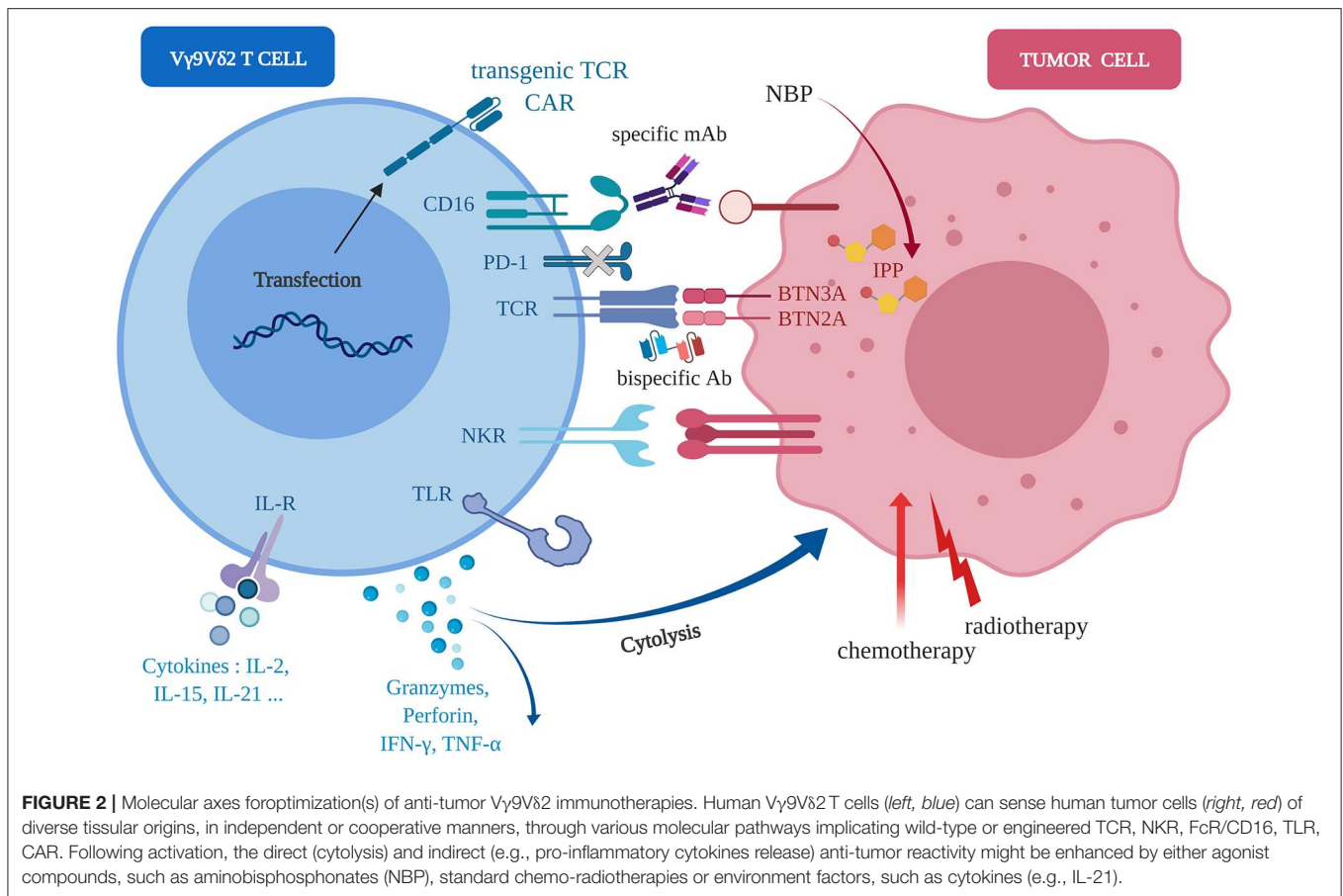
An indisputable tool for creating preclinical murine models representative of human pathology, not to mention therapies subsequently used, is the graft of explanted tumor samples from cancer patients. Two main strategies have been proposed: (i) the administration of dissociated and cultured tumor cells or (ii) the implantation of tumor fragments (hereafter called PDX, *patient-derived xenograft*) (Figure 1). These human tumors can heterotopically or orthotopically engrafted in immunodeficient mice, such as NSG (*NOD.Cg-PrkdcscidIl2rgtm1Wjl/SzJl*), with engraftment rates (40–95%) related to the presence of stroma. In the case of dissociated and cultured primary tumor cells, all these preparation steps might affect their phenotype and promote the biased growth of particular tumor variants. This issue can be addressed using tumor fragments (53) but the implantation of such large PDX is rarely possible in orthotopic because of size or surgical issues (e.g., intracerebral glioblastoma) (Figure 1). The engraftment and the growth of PDX often take 2–4 months, which can vary by tumor type, implant location, and the strain of immunodeficient mice utilized. This long duration parameter, further complicated by the necessity to divide and to re-implant small fragments of grafted PDX, strongly limits the use of PDX models and its interest for establishing robust, homogeneous and reliable tumor mouse models (i.e., progressive loss of tumor heterogeneity and replacement of the human microenvironment disappears by a murine stroma) (53).

In the absence of relevant and robust *in vitro* tumor models, including the promising, but currently developed

spheroid/organoid/3D tumor systems, the development of novel therapies still requires assessment steps in preclinical animal models before being proposed for therapy. Their use is expected to help predict selected therapeutic effects and analyze selected parameters such as feasibility, side effects and toxicity. In particular with regard to human V $\gamma$ 9V $\delta$ 2 T cells, it is not possible to use syngeneic murine models due to the lack of counterpart of this lymphocyte subset in mice and its species specificity (4, 5). Mostly due to an elevated cost of care, non-human primates, which also contain PAg-reactive  $\gamma\delta$  T cells, are ill-developed for cancer therapy studies. Highly immunodeficient mice, such as nude or NSG mice, represent the most relevant strains of choice for developing human cancer and immunotherapy models. However, the downside is that most of these immunodeficient mouse models are not relevant for analyzing the contribution of the tumor environment and immune system components to the anti-tumor efficacy of immunotherapies. Rebuilding human immune system in mice remains possible, thanks to the systemic injection of human PBMC in irradiated mice (54). However, this approach might generate strong xenogeneic reaction against the host, which limits the relevance and the operational time window for using these humanized animal models. In addition, the management and reproducibility of these sophisticated models remains complicated. Right now, the mostly developed method



**FIGURE 1** | Mouse models for ortho- or heterotopic administrations of human tumor cells. **(A)** Schematic representation of human tumor cell implantation strategies in immunodeficient mouse (e.g., NSG). Human tumor cells from either cell lines, primary/organoid cultures or patient-derived xenograft can be implanted orthotopically or heterotopically, respectively. **(B,C)** Immunohistochemistry (hematoxylin and eosin stainings) of orthotopic (brain) implantation of cells from a primary culture of human glioblastoma (GBM-1) **(B)** and heterotopic (subcutaneous) implantation of human prostate cancer cells (PC-3 cell line) **(C)**. Bars: 500  $\mu$ m. NJ & ES, unpublished work.



of humanization remains the injection/grafting of human tumor cells into immunodeficient mice.

## PATHS FOR THE DEVELOPMENT OF IMPROVED V $\gamma$ 9V $\delta$ 2 IMMUNOTHERAPIES USING PRECLINICAL MODELS

Various preclinical strategies are currently developed to assess the therapeutic potential of novel optimized V $\gamma$ 9V $\delta$ 2 immunotherapies targeting various pathways (which are summarized in **Figure 2**). In particular, several research and clinical groups aim at proposing immunotherapeutic strategies to enhance the reactivity and the cytotoxic activity of human V $\gamma$ 9V $\delta$ 2 T cells against tumor cells (16). Of note, the routes for treatment administration in preclinical models should be carefully selected. Most clinical treatments are either orally or systemically administered which might significantly contribute to an increased toxicity or allow therapeutic efficiency (e.g., low specificity). An increasing number of studies evidenced the benefits of local administrations for limiting the systemic toxicity while increasing therapeutic doses (55, 56). These principles should be applied, if clinically possible, for the adoptive transfer(s) of V $\gamma$ 9V $\delta$ 2 T cells.

Following an initial assessment *in vitro*, both orthotopic and heterotopic preclinical models of human tumors from various types of tissues, have been implemented in mice and used for the following V $\gamma$ 9V $\delta$ 2 T cell immunotherapy optimizations (**Figure 2**):

- Natural reactivity of allogeneic V $\gamma$ 9V $\delta$ 2 T cells against some tumor cell types. For example, some human mesenchymal GBM cells naturally overexpress NKG2D ligands that induce a natural V $\gamma$ 9V $\delta$ 2 T cell reactivity leading to the elimination of tumor cells *in vivo* (57);
- Transfer(s) of human V $\gamma$ 9V $\delta$ 2 T cells combined to agonist compounds (e.g., aminobisphosphonate compounds) administration(s) to further increase the recognition of tumor cells by V $\gamma$ 9V $\delta$ 2 T cells (47). These therapeutic components can be injected either systemically (e.g., intravenously) or close to the tumor sites (for solid tumors) in orthotopic models (50, 52);
- Transfer(s) of human V $\gamma$ 9V $\delta$ 2 T cells combined to radio-chemotherapies. This strategy relies on the observations that these first-line standard treatments are proposed for most cancer indications and could promote cell stress events that could increase the reactivity of V $\gamma$ 9V $\delta$ 2 T cells against human tumor cells (58–60);

- Transfer(s) of human V $\gamma$ 9V $\delta$ 2 T cells combined to antibodies directed against inhibitory immune checkpoint molecules (e.g., PD-1) (61) or ADCC (*Antibody-Dependent Cellular Cytotoxicity*)-triggering molecules (e.g., Fc $\gamma$ RIIIA/CD16) (62, 63);
- Bispecific antibodies which are generated under various molecular formats and have a dual specificity for a selected tumor antigen (targeting) and for the V $\gamma$ 9V $\delta$ 2 TCR (activation) (64).
- Agonist antibodies targeting V $\gamma$ 9V $\delta$ 2 T cell activation molecules (e.g., ectodomain of BTN3A1) to induce a PAg-independent, but TCR-specific, reactivity of V $\gamma$ 9V $\delta$ 2 T cells against tumor cells (65);
- Selected cytokines that can enhance the cytotoxic potential of V $\gamma$ 9V $\delta$ 2 T cells, such as IL-21 (51, 66), or boost their proliferation during *ex vivo* amplification before adoptive transfer, such as IL-15 associated to Vitamin C (67).

## CONCLUDING REMARKS

In terms of phenotype and functions, the human V $\gamma$ 9V $\delta$ 2 T cell subset represents a unique and highly attractive T cell population for designing efficient cancer immunotherapies. However, initial clinical trials targeting this subset yielded mixed results with both encouraging (e.g., good feasibility, weak toxicity) and disappointing observations (e.g., modest clinical efficacy), that could be attributed to inappropriately designed strategies or incorrect therapeutic settings. Nonetheless, these studies clearly met on the urgent need for proposing improved therapies, an objective that should be achieved through the development of more relevant and physiological preclinical models. Although first *in vivo* models have been proposed decades ago, with a major bias toward rodents (ie. mouse), their use for designing V $\gamma$ 9V $\delta$ 2 T cell-immunotherapies has

long been severely hampered by the strict species-specific restrictions of this primate subset. However, the research in this field widely accelerated in the past years thanks to the emergence and development of immunodeficient murine strains as well as improved grafting, mouse humanization and high-dimension detection/“big data” analysis approaches. Importantly, animal-free *in vitro* models (e.g., spheroids, organoids) are currently developed and should also be used soon in this process. Altogether, these elements should accelerate the entry of this research field within a novel dimension, which should be evidenced by an increased number of successful immunotherapeutic trials in cancer patients.

## AUTHOR CONTRIBUTIONS

NJ and ES contributed to the writing process, prepared the manuscript, and approved the final version.

## FUNDING

This work was supported by grants from INSERM, CNRS, Université de Nantes, Fondation pour la Recherche Médicale (FRM DEQ20170839118), Ligue Contre le Cancer AO GO2019 (Côtes d’Armor, Loire-Atlantique), Association pour la Recherche contre le Cancer (PJA20191209404). This work was realized in the context of the LabEX IGO program, supported by the National Research Agency Investissements d’Avenir via the program ANR-11-LABX-0016-01.

## ACKNOWLEDGMENTS

The authors thank the staff of the cellular and tissular imaging core (MicroPICell) and the animal facilities (UTE) from SFR F. Bonamy, Nantes University, for their help and expert assistance.

## REFERENCES

1. Dranoff G. Cytokines in cancer pathogenesis and cancer therapy. *Nat Rev Cancer*. (2004) 4:11–22. doi: 10.1038/nrc1252
2. Brenner MB, McLean J, Dialynas DP, Strominger JL, Smith JA, Owen FL, et al. Identification of a putative second T-cell receptor. *Nature*. (1986) 322:145–9. doi: 10.1038/322145a0
3. Saito H, Kranz DM, Takagaki Y, Hayday AC, Eisen HN, Tonegawa S. Complete primary structure of a heterodimeric T-cell receptor deduced from cDNA sequences. *Nature*. (1984) 309:757–62. doi: 10.1038/309757a0
4. Kazen AR, Adams EJ. Evolution of the VD and J gene segments used in the primate  $\gamma\delta$  T-cell receptor reveals a dichotomy of conservation and diversity. *Proc Natl Acad Sci USA*. (2011) 108:E332–40. doi: 10.1073/pnas.1105105108
5. Sturm E, Bontrop RE, Vreugdenhil RJ, Otting N, Bolhuis RL. T-cell receptor gamma/delta: comparison of gene configurations and function between humans and chimpanzees. *Immunogenetics*. (1992) 36:294–301. doi: 10.1007/BF00215657
6. Zhao Y, Niu C, Cui J. Gamma-delta ( $\gamma\delta$ ) T cells: friend or foe in cancer development? *J Transl Med*. (2018) 16:3. doi: 10.1186/s12967-018-1491-x
7. Lefranc MP, Rabbitts TH. A nomenclature to fit the organization of the human T-cell receptor  $\gamma$  and  $\delta$  genes. *Res Immunol*. (1990) 141:615–8. doi: 10.1016/0923-2494(90)90068-A
8. Pauza CD, Cairo C. Evolution and function of the TCR Vgamma9 chain repertoire: it's good to be public. *Cell Immunol*. (2015) 296:22–30. doi: 10.1016/j.cellimm.2015.02.010
9. Vantourout P, Hayday A. Six-of-the-best: unique contributions of  $\gamma\delta$  T cells to immunology. *Nat Rev Immunol*. (2013) 13:88–100. doi: 10.1038/nri3384
10. Rigau M, Ostrouska S, Fulford TS, Johnson DN, Woods K, Ruan Z, et al. Butyrophilin 2A1 is essential for phosphoantigen reactivity by  $\gamma\delta$  T cells. *Science*. (2020) 367:eaay5516. doi: 10.1126/science.aay5516
11. Karunakaran MM, Willcox CR, Salim M, Paletta D, Fichtner AS, Noll A, et al. Butyrophilin-2A1 directly binds germline-encoded regions of the V $\gamma$ 9V $\delta$ 2 TCR and is essential for phosphoantigen sensing. *Immunity*. (2020) 52:487–98.e6. doi: 10.1016/j.immuni.2020.02.014
12. Boutin L, Scotet E. Towards deciphering the hidden mechanisms that contribute to the antigenic activation process of human V $\gamma$ 9V $\delta$ 2 T cells. *Front Immunol*. (2018) 9:828. doi: 10.3389/fimmu.2018.00828
13. Coppens I. Targeting lipid biosynthesis and salvage in apicomplexan parasites for improved chemotherapies. *Nat Rev Microbiol*. (2013) 11:823–35. doi: 10.1038/nrmicro3139
14. Thompson K, Rogers MJ. Statins prevent bisphosphonate-induced  $\gamma\delta$ -T-cell proliferation and activation in vitro. *J Bone Miner Res Off J Am Soc Bone Miner Res*. (2004) 19:278–88. doi: 10.1359/JBMR.0301230

15. Ribeiro ST, Ribot JC, Silva-Santos B. Five layers of receptor signaling in  $\gamma\delta$  T-cell differentiation and activation. *Front Immunol.* (2015) 6:15. doi: 10.3389/fimmu.2015.00015
16. Hoeres T, Smetak M, Pretscher D, Wilhelm M. Improving the efficiency of V $\gamma$ 9V $\delta$ 2 T-cell immunotherapy in cancer. *Front Immunol.* (2018) 9:800. doi: 10.3389/fimmu.2018.00800
17. Gober H-J, Kistowska M, Angman L, Jenö P, Mori L, De Libero G. Human T cell receptor  $\gamma\delta$  cells recognize endogenous mevalonate metabolites in tumor cells. *J Exp Med.* (2003) 197:163–8. doi: 10.1084/jem.20021500
18. Harly C, Peigné C-M, Scotet E. Molecules and mechanisms implicated in the peculiar antigenic activation process of human V $\gamma$ 9V $\delta$ 2 T cells. *Front Immunol.* (2014) 5:657. doi: 10.3389/fimmu.2014.00657
19. Silva-Santos B, Serre K, Norell H.  $\gamma\delta$  T cells in cancer. *Nat Rev Immunol.* (2015) 15:683–91. doi: 10.1038/nri3904
20. Kunzmann V, Bauer E, Feurle J, Weissinger F, Tony HP, Wilhelm M. Stimulation of gammadelta T cells by aminobisphosphonates and induction of antiplasma cell activity in multiple myeloma. *Blood.* (2000) 96:384–92. doi: 10.1182/blood.V96.2.384.013k07\_384\_392
21. Sturm E, Braakman E, Fisch P, Vreugdenhil RJ, Sondel P, Bolhuis RL. Human V $\gamma$ 9-V $\delta$ 2 T cell receptor-gamma delta lymphocytes show specificity to Daudi Burkitt's lymphoma cells. *J Immunol Baltim Md.* (1990) 145:3202–8.
22. Kobayashi H, Tanaka Y, Yagi J, Toma H, Uchiyama T. Gamma/delta T cells provide innate immunity against renal cell carcinoma. *Cancer Immunol Immunother CII.* (2001) 50:115–24. doi: 10.1007/s002620100173
23. Viey E, Fromont G, Escudier B, Morel Y, Da Rocha S, Chouaib S, et al. Phosphostim-activated  $\gamma\delta$  T cells kill autologous metastatic renal cell carcinoma. *J Immunol Baltim Md 1950.* (2005) 174:1338–47. doi: 10.4049/jimmunol.174.3.1338
24. Corvaisier M, Moreau-Aubry A, Diez E, Bennouna J, Mosnier J-F, Scotet E, et al. V $\gamma$ 9V $\delta$ 2 T cell response to colon carcinoma cells. *J Immunol Baltim Md 1950.* (2005) 175:5481–8. doi: 10.4049/jimmunol.175.8.5481
25. Dunne MR, Mangan BA, Madrigal-Estebas L, Doherty DG. Preferential Th1 cytokine profile of phosphoantigen-stimulated human V $\gamma$ 9V $\delta$ 2 T cells. *Mediators Inflamm.* (2010) 2010:704941. doi: 10.1155/2010/704941
26. García VE, Sieling PA, Gong J, Barnes PF, Uyemura K, Tanaka Y, et al. Single-cell cytokine analysis of gamma delta T cell responses to nonpeptide mycobacterial antigens. *J Immunol Baltim Md 1950.* (1997) 159:1328–35.
27. Girardi M, Glusac E, Filler RB, Roberts SJ, Propperova I, Lewis J, et al. The distinct contributions of murine T cell receptor (TCR) $\gamma\delta$ + and TCR $\alpha\beta$ + T cells to different stages of chemically induced skin cancer. *J Exp Med.* (2003) 198:747–55. doi: 10.1084/jem.20021282
28. Kabelitz D, Wesch D, Pitters E, Zöller M. Characterization of tumor reactivity of human V $\gamma$ 9V $\delta$ 2 $\gamma\delta$  T cells in vitro and in SCID mice in vivo. *J Immunol Baltim Md 1950.* (2004) 173:6767–76. doi: 10.4049/jimmunol.173.11.6767
29. Zheng BJ, Chan KW, Im S, Chua D, Sham JS, Tin PC, et al. Anti-tumor effects of human peripheral  $\gamma\delta$  T cells in a mouse tumor model. *Int J Cancer.* (2001) 92:421–5. doi: 10.1002/ijc.1198
30. Gentles AJ, Newman AM, Liu CL, Bratman SV, Feng W, Kim D, et al. The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nat Med.* (2015) 21:938–45. doi: 10.1038/nm.3909
31. Lo Presti E, Dieli F, Meraviglia S. Tumor-infiltrating  $\gamma\delta$  T lymphocytes: pathogenic role, clinical significance, and differential programming in the tumor microenvironment. *Front Immunol.* (2014) 5:607. doi: 10.3389/fimmu.2014.00607
32. Fournié J-J, Sicard H, Poupot M, Bezombes C, Blanc A, Romagné F, et al. What lessons can be learned from  $\gamma\delta$  T cell-based cancer immunotherapy trials? *Cell Mol Immunol.* (2013) 10:35–41. doi: 10.1038/cmi.2012.39
33. Sebastyen Z, Prinz I, Déchanet-Merville J, Silva-Santos B, Kuball J. Translating gammadelta ( $\gamma\delta$ ) T cells and their receptors into cancer cell therapies. *Nat Rev Drug Discov.* (2019) 19:169–84. doi: 10.1038/s41573-019-0038-z
34. Kunzmann V, Bauer E, Wilhelm M.  $\gamma/\delta$  T-cell stimulation by pamidronate. *N Engl J Med.* (1999) 340:737–8. doi: 10.1056/NEJM199903043400914
35. Wilhelm M, Kunzmann V, Eckstein S, Reimer P, Weissinger F, Ruediger T, et al.  $\gamma\delta$  T cells for immune therapy of patients with lymphoid malignancies. *Blood.* (2003) 102:200–6. doi: 10.1182/blood-2002-12-3665
36. Dieli F, Gebbia N, Poccia F, Caccamo N, Montesano C, Fulfaro F, et al. Induction of  $\gamma\delta$  T-lymphocyte effector functions by bisphosphonate zoledronic acid in cancer patients in vivo. *Blood.* (2003) 102:2310–1. doi: 10.1182/blood-2003-05-1655
37. Dieli F, Vermijlen D, Fulfaro F, Caccamo N, Meraviglia S, Cicero G, et al. Targeting human  $\gamma\delta$  T cells with zoledronate and interleukin-2 for immunotherapy of hormone-refractory prostate cancer. *Cancer Res.* (2007) 67:7450–7. doi: 10.1158/0008-5472.CAN-07-0199
38. Lang JM, Kaikobad MR, Wallace M, Staab MJ, Horvath DL, Wilding G, et al. Pilot trial of interleukin-2 and zoledronic acid to augment  $\gamma\delta$  T cells as treatment for patients with refractory renal cell carcinoma. *Cancer Immunol Immunother CII.* (2011) 60:1447–60. doi: 10.1007/s00262-011-1049-8
39. Meraviglia S, Eberl M, Vermijlen D, Todaro M, Buccheri S, Cicero G, et al. In vivo manipulation of V $\gamma$ 9V $\delta$ 2 T cells with zoledronate and low-dose interleukin-2 for immunotherapy of advanced breast cancer patients. *Clin Exp Immunol.* (2010) 161:290–7. doi: 10.1111/j.1365-2249.2010.04167.x
40. Wilhelm M, Smetak M, Schaefer-Eckart K, Kimmel B, Birkmann J, Einsele H, et al. Successful adoptive transfer and in vivo expansion of haploidentical  $\gamma\delta$  T cells. *J Transl Med.* (2014) 12:45. doi: 10.1186/1479-5876-12-45
41. Bennouna J, Levy V, Sicard H, Senellart H, Audrain M, Huret S, et al. Phase I study of bromohydrin pyrophosphate (BrHPP, IPH 1101), a V $\gamma$ 9V $\delta$ 2 T lymphocyte agonist in patients with solid tumors. *Cancer Immunol Immunother CII.* (2010) 59:1521–30. doi: 10.1007/s00262-010-0879-0
42. Sicard H, Ingoure S, Luciani B, Serraz C, Fournié J-J, Bonneville M, et al. In vivo immunomanipulation of V $\gamma$ 9V $\delta$ 2 T cells with a synthetic phosphoantigen in a preclinical nonhuman primate model. *J Immunol Baltim Md 1950.* (2005) 175:5471–80. doi: 10.4049/jimmunol.175.8.5471
43. Kobayashi H, Tanaka Y, Yagi J, Osaka Y, Nakazawa H, Uchiyama T, et al. Safety profile and anti-tumor effects of adoptive immunotherapy using gamma-delta T cells against advanced renal cell carcinoma: a pilot study. *Cancer Immunol Immunother CII.* (2007) 56:469–76. doi: 10.1007/s00262-006-0199-6
44. Alnaggar M, Xu Y, Li J, He J, Chen J, Li M, et al. Allogenic V $\gamma$ 9V $\delta$ 2 T cell as new potential immunotherapy drug for solid tumor: a case study for cholangiocarcinoma. *J Immunother Cancer.* (2019) 7:36. doi: 10.1186/s40425-019-0501-8
45. Fisher JP, Heuveljans J, Yan M, Gustafsson K, Anderson J.  $\gamma\delta$  T cells for cancer immunotherapy: a systematic review of clinical trials. *Oncoimmunology.* (2014) 3:e27572. doi: 10.4161/onci.27572
46. Straetemans T, Kierkels GJJ, Doorn R, Jansen K, Heijhuus S, Dos Santos JM, et al. GMP-grade manufacturing of T cells engineered to express a defined  $\gamma\delta$ TCR. *Front Immunol.* (2018) 9:1062. doi: 10.3389/fimmu.2018.01062
47. Santolaria T, Robard M, Léger A, Catros V, Bonneville M, Scotet E. Repeated systemic administrations of both aminobisphosphonates and human V $\gamma$ 9V $\delta$ 2 T cells efficiently control tumor development in vivo. *J Immunol Baltim Md 1950.* (2013) 191:1993–2000. doi: 10.4049/jimmunol.1300255
48. Perrin S. Preclinical research: make mouse studies work. *Nature.* (2014) 507:423–5. doi: 10.1038/507423a
49. Wege AK. Humanized mouse models for the preclinical assessment of cancer immunotherapy. *BioDrugs Clin Immunother Biopharm Gene Ther.* (2018) 32:245–66. doi: 10.1007/s40259-018-0275-4
50. Jarry U, Chauvin C, Joalland N, Léger A, Minault S, Robard M, et al. Stereotoxic administrations of allogeneic human V $\gamma$ 9V $\delta$ 2 T cells efficiently control the development of human glioblastoma brain tumors. *Oncoimmunology.* (2016) 5:e1168554. doi: 10.1080/2162402X.2016.1168554
51. Joalland N, Chauvin C, Oliver L, Vallette FM, Pecqueur C, Jarry U, et al. IL-21 increases the reactivity of allogeneic human V $\gamma$ 9V $\delta$ 2 T cells against primary glioblastoma tumors. *J Immunother Hagerstown Md 1997.* (2018) 41:224–31. doi: 10.1097/CJI.0000000000000225
52. Joalland N, Lafrance L, Oullier T, Marionneau-Lambot S, Lousouarn D, Jarry U, et al. Combined chemotherapy and allogeneic human V $\gamma$ 9V $\delta$ 2 T lymphocyte-immunotherapies efficiently control the development of human epithelial ovarian cancer cells in vivo. *Oncoimmunology.* (2019) 8:e1649971. doi: 10.1080/2162402X.2019.1649971
53. Jung J, Seol HS, Chang S. The generation and application of patient-derived xenograft model for cancer research. *Cancer Res Treat Off J Korean Cancer Assoc.* (2018) 50:1–10. doi: 10.4143/crt.2017.307



54. Morton JJ, Bird G, Refaeli Y, Jimeno A. Humanized mouse xenograft models: narrowing the tumor-microenvironment gap. *Cancer Res.* (2016) 76:6153–8. doi: 10.1158/0008-5472.CAN-16-1260
55. Pauza CD, Liou M-L, Lahusen T, Xiao L, Lapidus RG, Cairo C, et al. Gamma delta T cell therapy for cancer: it is good to be local. *Front Immunol.* (2018) 9:1305. doi: 10.3389/fimmu.2018.01305
56. Yuasa T, Sato K, Ashihara E, Takeuchi M, Maita S, Tsuchiya N, et al. Intravesical administration of  $\gamma\delta$  T cells successfully prevents the growth of bladder cancer in the murine model. *Cancer Immunol Immunother CII.* (2009) 58:493–502. doi: 10.1007/s00262-008-0571-9
57. Chauvin C, Joalland N, Perroteau J, Jarry U, Lafrance L, Willem C, et al. NKG2D controls natural reactivity of V $\gamma$ 9V $\delta$ 2 T lymphocytes against mesenchymal glioblastoma cells. *Clin Cancer Res Off J Am Assoc Cancer Res.* (2019) 25:7218–28. doi: 10.1158/1078-0432.CCR-19-0375
58. Chitadze G, Lettau M, Luecke S, Wang T, Janssen O, Fürst D, et al. NKG2D- and T-cell receptor-dependent lysis of malignant glioma cell lines by human  $\gamma\delta$  T cells: modulation by temozolomide and A disintegrin and metalloproteases 10 and 17 inhibitors. *Oncoimmunology.* (2016) 5:e1093276. doi: 10.1080/2162402X.2015.1093276
59. Todaro M, Orlando V, Cicero G, Caccamo N, Meraviglia S, Stassi G, et al. Chemotherapy sensitizes colon cancer initiating cells to V $\gamma$ 9V $\delta$ 2 T cell-mediated cytotoxicity. *PLoS ONE.* (2013) 8:e65145. doi: 10.1371/journal.pone.0065145
60. Zoine JT, Knight KA, Fleischer LC, Sutton KS, Goldsmith KC, Doering CB, et al. Ex vivo expanded patient-derived  $\gamma\delta$  T-cell immunotherapy enhances neuroblastoma tumor regression in a murine model. *Oncoimmunology.* (2019) 8:1593804. doi: 10.1080/2162402X.2019.1593804
61. Rossi C, Gravelle P, Decaup E, Bordenave J, Poupot M, Tosolini M, et al. Boosting  $\gamma\delta$  T cell-mediated antibody-dependent cellular cytotoxicity by PD-1 blockade in follicular lymphoma. *Oncoimmunology.* (2019) 8:1554175. doi: 10.1080/2162402X.2018.1554175
62. Capietto A-H, Martinet L, Fournié J-J. Stimulated  $\gamma\delta$  T cells increase the in vivo efficacy of trastuzumab in HER-2+ breast cancer. *J Immunol Baltim Md 1950.* (2011) 187:1031–8. doi: 10.4049/jimmunol.1100681
63. Gertner-Dardenne J, Bonnafous C, Bezombes C, Capietto A-H, Scaglione V, Ingoure S, et al. Bromohydrin pyrophosphate enhances antibody-dependent cell-mediated cytotoxicity induced by therapeutic antibodies. *Blood.* (2009) 113:4875–84. doi: 10.1182/blood-2008-08-172296
64. de Bruin RCG, Veluchamy JB, Loughheed SM, Schneiders FL, Lopez-Lastra S, Lameris R, et al. A bispecific nanobody approach to leverage the potent and widely applicable tumor cytolytic capacity of V $\gamma$ 9V $\delta$ 2-T cells. *Oncoimmunology.* (2017) 7:e1375641. doi: 10.1080/2162402X.2017.1375641
65. Benyamine A, Le Roy A, Mamessier E, Gertner-Dardenne J, Castanier C, Orlanducci F, et al. BTN3A molecules considerably improve V $\gamma$ 9V $\delta$ 2 T cells-based immunotherapy in acute myeloid leukemia. *Oncoimmunology.* (2016) 5:e1146843. doi: 10.1080/2162402X.2016.1146843
66. Thedrez A, Harly C, Morice A, Salot S, Bonneville M, Scotet E. IL-21-mediated potentiation of antitumor cytolytic and proinflammatory responses of human V $\gamma$ 9V $\delta$ 2 T cells for adoptive immunotherapy. *J Immunol Baltim Md 1950.* (2009) 182:3423–31. doi: 10.4049/jimmunol.0803068
67. Kouakanou L, Xu Y, Peters C, He J, Wu Y, Yin Z, et al. Vitamin C promotes the proliferation and effector functions of human  $\gamma\delta$  T cells. *Cell Mol Immunol.* (2019) 17:462–73. doi: 10.1038/s41423-019-0247-8

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Joalland and Scotet. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.