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Fungicide Residues Exposure and β-amyloid Aggregation in a Mouse Model of Alzheimer’s Disease

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BACKGROUND: Pesticide residues have contaminated our environment and nutrition over the last century. Although these compounds are present at very low concentrations, their long-term effects on human health is of concern. The link between pesticide residues and Alzheimer’s disease is not clear and difficult to establish. To date, no in vivo experiments have yet modeled the impact of this chronic contamination on neurodegenerative disorders.

OBJECTIVES: We investigated the impact of fungicide residues on the pathological markers of Alzheimer’s disease in a transgenic mouse model.

METHODS: Transgenic (J20, hAPPsw/le) mice were chronically exposed to a cocktail of residues of cyprobidinil, mepanipyrim, and pyrimethanil at 0.1 μg/L in their drinking water for 9 months. We assessed the effects of fungicide residues on the pathological markers of the disease including Aβ aggregates, neuroinflammation, and neuronal loss. Then, we studied the dynamics of Aβ aggregation in vivo via a longitudinal study using two-photon microscopy. Finally, we investigated the molecular mechanisms involved in the production and clearance of Aβ peptides.

RESULTS: We found that a chronic exposure to three fungicide residues exacerbated aggregation, microgliosis, and neuronal loss. These fungicides also increased vascular amyloid aggregates reminiscent of cerebral amyloid angiopathy between 6 and 9 months of treatment. The mechanism of action revealed that fungicides promoted Aβ peptide fibril formation in vitro and involved an in vivo overexpression of the levels of the β-secretase–cleaving enzyme (BACE1) combined with impairment of Aβ clearance through neprylisin (NEP).

CONCLUSIONS: Chronic exposure of the J20 mouse model of Alzheimer’s disease to a cocktail of fungicides, at the regulatory concentration allowed in tap water (0.1 μg/L), strengthened the preexisting pathological markers: neuroinflammation, Aβ aggregation, and APP β-processing. We hypothesize prevention strategies toward pesticide long-term exposure may be an alternative to counterbalance the lack of treatment and to slow down the worldwide Alzheimer’s epidemic.

Introduction

Alzheimer’s disease is the most common form of dementia in elderly people and currently afflicts 50 million people worldwide, and this number is expected to double in the coming generation (OECD 2018). The etiology of Alzheimer’s disease has not yet been clarified, which may explain the lack of effective treatments. However, identifying environmental risk factors may represent an alternative strategy to reduce the number of cases. Indeed, projection studies of the incidence of Alzheimer’s disease between 2010 and 2050 have indicated that delaying the onset of Alzheimer’s disease by 1, 2, or 5 years could decrease its prevalence by about 10%, 20%, and 50%, respectively (Mura et al. 2010).

One of the features of Alzheimer’s disease is the accumulation and aggregation of amyloid-β peptides (Aβ) into amyloid plaques (Iwatsubo et al. 1994). Aβ peptides are produced by the sequential enzymatic processing of the amyloid precursor protein (APP) by the β- (BACE1) (Cai et al. 2001; Luo et al. 2001) and γ-secretases (Carroll and Li 2016) (for a review on APP processing, see O’Brien and Wong 2011). Impairments of enzymes such as insulin degrading enzyme (IDE) (Miners et al. 2006, 2009), neprylisin (NEP) (Miners et al. 2009; Zhao et al. 2007), and/or receptors such as the low-density lipoprotein receptor-related protein 1 (LRP1) (Kang et al. 2000; Shinohara et al. 2014) involved in the Aβ clearance pathways are major events reinforcing the disease. The causes leading to these deregulations have not yet been clearly identified. Our hypothesis is that environmental pollutants may corrupt APP metabolism or the Aβ aggregation process, which may lead to the onset or aggravation of Alzheimer’s disease.

Human epidemiological studies have shown that circulating levels of certain heavy metals are higher in patients with Alzheimer’s disease than in control patients (Fatrabadi et al. 2018; Xu et al. 2018). This suggests a potential role of heavy metals in Alzheimer’s disease, but it is not yet clear whether they interfere in the development or the progression of the disease. However, experimental studies have shown that incubation of SH-SY5Y cells with lead (5 to 100 μM) can induce an increase of APP expression and Aβ peptides secretion, associated with a decreased NEP expression (Chin-Chan et al. 2015; Huang et al. 2011). Furthermore, an acute injection of lead [27 mg/kg via the intraperitoneal (ip) route] in a mouse model of Alzheimer’s disease (APP V717F) has resulted in a lower expression of LRP1 in the choroid plexus than injection of vehicle (Gu et al. 2011). Mercury was shown to be involved in the disruption of Aβ peptide clearance pathways both in vivo [in rats: 2 mg/kg per day for 4 weeks by gavage (Kim et al. 2014)] and in vitro [in SH-SY5Y cells: 10 and 20 μM (Chin-Chan et al. 2015)]. Moreover, rats orally treated with aluminum (20 g/d, twice a week) through diet, from 6 months of age until the end of their life and who developed cognitive deficits, presented an
up-regulation of the APP gene and a greater APP accumulation in hippocampal and cortical neurons compared with cognitively intact control rats (Walton and Wang 2009). In vitro treatment with aluminum (50 μM) on rat cortical neurons revealed Aβ peptides conformational changes and enhanced aggregation (Kawahara et al. 2001). Regarding pesticides, occupational exposure of farmers to organophosphates has been associated with an increased risk to develop Alzheimer’s disease (Hayden et al. 2010). Moreover, in their clinical study Richardson et al. (2014) found a 4-fold higher level of dichlorodiphenylchloroethylene (DDE), a metabolite of the organochlorine insecticide dichlorodiphenyltrichloroethane (DDT), in the serum of Alzheimer’s disease patients compared with controls. Among these patients, the apolipoprotein E ε4 (APOE4) carriers with higher DDE levels had lower mini-mental state (MMS) scores (Richardson et al. 2014).

Although herbicides and insecticides have been highlighted for their role in Alzheimer’s disease, data on the impact of fungicides in neurological disorders have barely been reported. Fungicides are widely used for the cultivation and preservation of fresh fruits and vegetables. Because prevention campaigns encourage people to consume fruits and vegetables every day, we wonder if long-term exposure to fungicide residues, present in these foodstuffs, can affect human health. In this study, we focused our attention on three widely used fungicides: cyprodinil (cypro), mepanipyrim (mepa), and pyrimethanil (pyri), belonging to the anilinopyrimidine class. Many reports and publications have underlined the presence of anilinopyrimidine residues throughout our environment: in the air (ANSES 2010; Désert et al. 2018), in soils (Bermúdez-Cousso et al. 2007; Rial-Otero et al. 2004), and in water (ANSES 2013; Herrero-Herrández et al. 2016) (Table 1). Several French (ANSES 2011) and European (EFSA 2017b, 2018, 2019) reports have showed that residues of cypro, mepa, and pyri were detected in many foodstuffs, findings that have been confirmed by studies from independent nongovernmental organizations (Générations Futures 2013, 2016a; PAN Europe 2008) (Table 1). From an epidemiological standpoint, it is difficult to assess the impact of a long-term exposure to pesticide residues on neurodegenerative diseases such as Alzheimer’s disease. Indeed, the majority of Alzheimer’s disease cases are diagnosed in elderly populations, yet the exposure to risk factors occur years or decades before the first symptoms appear. However, one can hypothesize that chronic exposure to pesticide residues via our diet or drinking water may have a role in the etiology or aggravation of neurological diseases.

To test this hypothesis, we investigated the impact of fungicide residues on the J20 mouse model of Alzheimer’s disease (hAPPsw/Ind) (Mucke et al. 2000). Mice were chronically exposed to a cocktail of cypro, mepa, and pyri at the regulatory limit dose of 0.1 μg/L for each compound over 9 months through tap water. We analyzed different markers of Alzheimer’s disease such as soluble and aggregated Aβ peptides, astroglisis, microglisis, and neuronal loss. To better understand the dynamics of aggregation with the cocktail of fungicides, we performed a longitudinal study at 3, 6, and 9 months posttreatment using two-photon microscopy. To decipher the impact of these fungicides on either the amyloidogenic or the clearance pathways, we analyzed the main proteins involved (i.e., APP, BACE1, NEP, IDE, LRPI). The potential interaction of anilinopyrimidines with the different forms of Aβ was analyzed through ex vivo binding experiments and in vitro kinetics of amyloid fibril formation.

### Materials and Methods

#### Animals Used and Breeding

This project followed the specific French national guidelines on animal experimentation and well-being and was approved by the French National Ethics Committee for Animal Experimentation (APAIS no. 7357-20161025-18055782). Hemizygous hAPPsw/Ind transgenic mice (referred to as hAPPsw/Ind or also called J20 mice) (Mucke et al. 2000) were purchased from Jackson Laboratory.

| Table 1. Occurrences and Cmax of cyprodinil, mepanipyrim, and pyrimethanil residues in the European diet and environment. |
| --- | --- | --- | --- | --- |
| Elements analyzed | Cyprodinil | Mepanipyrim | Pyrimethanil | References |
| Fruits | 37.3% (n = 75) | 6.7% (n = 75) | 17.3% (n = 75) | ANSES 2011 |
| Vegetables | Cmax = 90 μg/kg | Cmax = 40 μg/kg | Cmax = 320 μg/kg | ANSES 2011 |
| Strawberries | 29.5% (n = 61) | 1.6% (n = 61) | 0% (n = 61) | Générations Futures 2013 |
| Table grapes | 16.00% (n = 1,128) | No data | 9.52% (n = 1,187) | EFSA 2017a |
| Strawberries | 34.49% (n = 1,206) | No data | 7.21% (n = 1,206) | EFSA 2018 |
| Pears | 11.09% (n = 1,199) | No data | 7.42% (n = 1,199) | EFSA 2019 |
| Austrian wine | 71.4% (n = 7) | No data | 85.7% (n = 7) | PAN Europe 2008 |
| French wine | 50% (n = 10) | No data | 90% (n = 10) | PAN Europe 2008 |
| German wine | 18.2% (n = 11) | No data | 63.6% (n = 11) | PAN Europe 2008 |
| Dust | 86.4% (n = 22) | 9.1% (n = 22) | 68.2% (n = 22) | Générations Futures 2016b |
| Hair | 14.3% (n = 7) | No data | 57.1% (n = 7) | Générations Futures 2017 |
| Atmosphere | 25% (n = 1,515) | No data | No data | ANSES 2010 |

Note: Cmax, maximal concentration; LOD, limit of detection; LOQ, limit of quantification.

*Occurrences were calculated according to the total number of samples analyzed and the number of samples presenting residues of pesticides in quantified concentrations (above the LOQ: >0.1 μg/kg). However, samples with pesticide residues below the LOD (0.03 μg/kg) were not reported in these studies because the reporting countries of the European Union did not compile the information consistently. So, samples were not taken into account when residues concentration was below the LOQ.*
This mouse model does not exhibit neurofibrillary aggregates in the CA1 area of the hippocampus but not in the CA3 area. The J20 mouse model overexpresses human APP containing the Swedish (K670N/M671L) and Indiana (V717F) mutations under the control of platelet-derived growth factor-beta (PDGF-β) promoter, leading to a neuronal expression of the transgene (Mucke et al. 2000). This model overproduces Aβ42 peptides. The appearance of aggregates is progressive, with robust plaques in the hippocampus and the cortex by 8–9 months of age (Mucke et al. 2000). This mouse model does not exhibit neurofibrillary tangles (Mucke et al. 2000). J20 mice exhibit a neuronal loss at 3, 6, and 9 months of age in the CA1 area of the hippocampus but not in the CA3 area (Wright et al. 2013). Regarding astrogliosis, a significant increase of glial fibrillary acidic protein (GFAP)-positive cells was observed in the hippocampus of J20 mice at 6 and 9 months of age compared with age-related WT mice (Wright et al. 2013).

Animals’ Treatment and Experiments

Cypro, mepa, and pyri were purchased from Sigma-Aldrich. Stock solutions were prepared in glass vials (Wheaton) at 1 mg/mL in pure dimethyl sulfoxide (DMSO) according to the supplier’s recommendations. Stock solutions were diluted 1,000 times in distilled water to a final concentration of 1 mg/L, filtered (0.5 μm), and stored in glass bottles (Duran-Schott) at 4°C until use. For animal treatment, stock solutions at 1 mg/L were further diluted 10,000 times in tap water to reach a final concentration of 0.1 μg/L. Animals’ treatment started after weaning (at postnatal day 30), and they were treated with a cocktail of the three fungicides at 0.1 μg/L each (cypro: 444 pM; mepa: 448 pM; and pyri: 502 pM) (Figure 1A), which corresponds to the regulatory maximal limit dose allowed in drinking water (Council of the European Union 1998). According to EU regulations, the total concentration of several pesticide residues in drinking water cannot exceed 0.5 μg/L (Council of the European Union 1998). In parallel, a solution of pure DMSO (100%) was diluted 1,000 times in distilled water to a final concentration of 0.1%, filtered (0.5 μm), and stored in a glass bottle at 4°C until use. To treat control vehicle animals, the stock solution of DMSO at 0.1% was further diluted 10,000 times in tap water (1 × 10⁻⁵ % corresponding to the equivalent volume of one fungicide). Thus, for three fungicides, the final percentage of DMSO is 3 × 10⁻³. Mice water bottles were changed once a week for all groups and prepared from stock solutions as described above. Each week, water levels were measured for each cage and pesticide consumption was estimated per mouse. In parallel, the body weight gain was measured every month.

For this project, a total of 33 J20 and 31 WT mice were treated with the cocktail of the three fungicides via the drinking water (referred to as T groups) for 9 months (35 weeks) after weaning (Figure 1B). Control groups (J20 n = 32 and WT n = 30) were treated with an equivalent volume of DMSO and referred to as CTR groups (Figure 1B). All these mice were generated from three independent breedings. Animals obtained from the first set of experiments, referred to as Study 1 (WT CTR: n = 27; WT T: n = 28; J20 CTR: n = 25; and J20 T: n = 24), were used for biochemical analyses (left hemispheres for Aβ42 assay) and for immunohistochemistry [right hemispheres for thioloavin T (ThT) staining and detection of fungicides] (Figure 1C, Study 1). A second breeding, called Study 2 (WT CTR: n = 3; WT T: n = 3; J20 CTR: n = 4; and J20 T: n = 4) was done to perform a-labeling with the anti-GFAP, anti-ionized calcium binding adaptor molecule 1 (IBA1), and anti-neuronal nuclei (NeuN) antibodies; b) for interleukins (ILs) and cytokines time resolved-fluorescence resonance energy transfer (TR-FRET, also called homogeneous time resolved fluorescence or HTRF® assays); c) quantitative polymerase chain reaction (qPCR) analyses; and d) western blotting (Figure 1C, Study 2). Mice from a third experiment, called Study 3 (J20 CTR: n = 3 and J20 T: n = 5), were used in a longitudinal study of appearance of plaques using two-photon microscopy at 3, 6, and 9 months posttreatment (Figure 1D). In addition, six 10-month-old J20 mice (without any treatment) were used for ex vivo interaction assays of fungicides with amyloid plaques. Detailed protocols of these experiments are described below.

Tissue Collection and Preparation

After 9 months of treatment, mice from Studies 1 and 2 were anesthetized [16% ketamine, 4% xylazine in sodium chloride (NaCl) 0.9%] and sacrificed by a cardiac perfusion of phosphate-buffered saline (PBS), to remove blood from tissues. Brains were removed and cut along the sagittal axis: The left hemispheres were frozen directly in liquid nitrogen and stored at −80°C (for biochemical analyses), and the right hemispheres were fixed in a commercial parafomaldehyde solution (AntigenFix, Diaphat) for 48 h at 4°C (for immunohistochemistry). Right hemispheres were rinsed with PBS and incubated in a 30% sucrose solution for 4 d at 4°C. Brains were then included in optimum cutting temperature compound (Tissue-Tek® O.C.T.; Sakura Finetek), quickly frozen in acetone chilled on dry ice, and conserved at −80°C until use.

Ex Vivo Interaction of Cypro, Mepa, and Pyri with Amyloid Plaques on J20 Brain Tissue Sections

Frozen brains included in OCT from 6 J20 mice, without any treatment, were mounted on a cryostat (Leica) and 25-μm coronal sections were performed and conserved at −20°C in an antifreeze solution (25% glycerol and 25% ethylene glycol in 0.1 M phosphate buffer) until use. Three to six brain sections per mouse were rinsed, mounted on Superfrost™ Plus glass slides (Microm), and then dried overnight at room temperature (RT). Prior to amyloid labeling, tissue sections were incubated for 10 min with a 70% formic acid solution for antigen retrieval. Sections were then washed in PBS and incubated 1.5 h at RT with a solution containing 2 mM of cypro, or mepa, or pyri or a cocktail of the three fungicides (2 mM each). Sections were incubated 1 h at RT with a blocking solution (3% goat serum, 0.3% Triton X100 in PBS) and then 2 h at RT with a mouse anti-β-amyloid 1-16 conjugated to AlexaFluor® 488 (1:100, clone 6E10, Ozyme; referred to as 6E10-Alexa488), followed by the primary antibodies recognizing cypro (Esteve-Turrillas et al. 2012), mepa (Esteve-Turrillas et al. 2013), or pyri (Mecedor et al. 2012). Then, the sections were incubated with a goat anti-mouse Cy3 antibody (Jackson ImmunoResearch) for 2 h at RT, and sections were counterstained with 2-(4-aminophenyl)-1H-indole-6-carboxamidine (DAPI; 1:50,000 in PBS) (Molecular Probes). To test the...
specificity of the anti-fungicide antibodies, control brain tissue sections were incubated either with PBS or with an equivalent volume of DMSO (1%) corresponding to the final concentration present in the cocktail of fungicides used. Then, control slides were incubated either without any anti-fungicide antibodies or with secondary antibody Cy3 alone, or anti-fungicide antibodies (anti-cypro, -mepa, and -pyri) alone or in cockail. All slides were revealed with a goat anti-mouse Cy3 antibody as described above, excepted for the control slide without any fungicide antibodies. All images were taken with a Leica DM2500 microscope (Leica Microsystems) (magnification: ×20 and ×40).

**Kinetics of Fibril Formation of Human Aβ1-42 Peptides in Presence of a Cocktail of Cypro, Mepa and Pyri**

Synthetic human Aβ1-42 peptides were purchased from the ERI Amyloid Laboratory LLC. Peptides were maintained in a monomeric state using the protocol described by Serra-Batiste et al. (2016). Briefly, Aβ peptides were dissolved in a 6.8 M guanidine thiocyanate solution (Sigma-Aldrich) at a concentration of 8.5 mg/mL. The solution was then sonicated for 5 min at 52°C and diluted with ultrapure water to reach a final concentration of 5 mg/mL of Aβ peptides and 4 M of guanidine thiocyanate. The solution was centrifuged at 10,000 x g for 6 min at 4°C. The collected supernatant was filtered [polysyndetene fluoride (PVDF); 0.45 μm] and then injected into a Superdex™ 75 Increase 10/300 GL column (GE Heathcare Life Science) previously equilibrated with 10 mM sodium phosphate buffer at pH 7.4. Purification was performed with a 0.5-mL/min flow to collect the peak attributed to monomeric Aβ peptides. Then, their concentration was determined with a NanoDrop™ 8,000 spectrophotometer (Thermo Scientific), and aliquots of peptides were stored at −20°C. The Aβ stock solution was diluted to 30 μM in a 10 mM sodium phosphate buffer at pH 7.4 and left to aggregate in low-binding Eppendorf tubes with a final volume of 600 μL. Experiments with a cocktail of the three fungicides were conducted under the same conditions, except that fungicides were added from a stock solution to get a 1:1 or 1:15 M ratio (Aβ:fungicides), with a final concentration of DMSO of 1% (vol/vol) for both conditions. To keep the conditions identical in the control tubes, an equivalent volume of DMSO of 1% (vol/vol) was added to the three fungicides were conducted under the same conditions, except that fungicides were added from a stock solution to get a 1:1 or 1:15 M ratio (Aβ:fungicides), with a final concentration of DMSO of 1% (vol/vol) for both conditions. To keep the conditions identical in the control tubes, an equivalent volume of DMSO of 1% (vol/vol) was added to the control tubes, an equivalent volume of DMSO of 1% (vol/vol) was added to the control tubes.
and mixed with 14 µL of 124 mM Glycine-sodium hydroxide (GlyNaOH) buffer at pH 8.3 and 6 µL of 100 µM of ThT, in a 96-well plate of black polystyrene with clear bottoms and a polystyrene glycol coating (Thermofisher Scientific). ThT fluorescence of each sample was measured ($\lambda_{ex} = 445$ nm and $\lambda_{em} = 485$ nm) in a Fluoroskan Ascent™ microplate fluorimeter (Thermofisher Scientific). Aggregation profiles were generated with GraphPad Prism software (version 5.0; GraphPad), and each kinetic trace described previously (Lafon et al. 2018). Brie

**Aggregate Staining and Quantification**

For this experiment, the number of brains analyzed per group was as follows: WT CTR: $n = 4$; WT T: $n = 4$; J20 CTR: $n = 5$; and J20 T $n = 8$ (Figure 1C, Study 1). Floating brain tissue sections were incubated with ThT, a fluorescent dye, used to stain amyloid plaques. Sections were first mounted on Superfrost™ Plus glass slides, dried overnight at RT, incubated with a ThT solution at 0.01% (31 µM) for 10 min (Sigma-Aldrich), rinsed with an 80% ethanol solution and counterstained with DAPI (1:50,000) (Molecular Probes). All sections were visualized under a Leica DM2500 microscope (Leica Microsystems) (magnifications: ×20 and ×40 for enlargements). Images were analyzed with Fiji software (version 2.0; National Institutes of Health). Hemi-brain tissue sections were used to quantify the number and the surface area of amyloid aggregates in the hippocampus and the cortex. Eight to 10 sections were used per animal, and all the values obtained were pooled to get the total number and total surface area of aggregates for each region analyzed. These data were then normalized to the animal’s total surface area of hippocampus or cortex of the section quantified. In parallel, the distribution of amyloid plaques within the different regions of the hippocampus corresponding to dentate gyrus (DG), CA1, CA2, and CA3 was done. Two independent quantifications of amyloid aggregates were performed blindly, and the same trends were observed for the two sets of values obtained. Only one quantification was kept for statistical analyses.

**Aβ1–42 Assay**

For this experiment, the number of brains analyzed per group was as follows: WT CTR: $n = 4$; WT T: $n = 4$; J20 CTR: $n = 5$; and J20 T $n = 8$ (Figure 1C, Study 1). Hemi-brains were homogenized as described previously (Lafon et al. 2018). Briefly, frozen left hemispheres were homogenized in 20% wt/vol in a homogenization buffer [140 mM potassium chloride (KCl), 10 mM sodium hydrogencarbonate (Na2HPO4), 1.7 mM monopotassium phosphate (KH2PO4), and 1 mM ethylenediaminetetraacetic acid (EDTA)] containing protease inhibitors (Complete Ultra, Roche) and phosphatases inhibitors (PhosStop™, Sigma). Homogenization was performed in microbead-containing tubes on a Riboysor apparatus (Biorad). Samples were aliquoted and immediately frozen at −80°C until use. For the Aβ1–42 assay, 2% sodium dodecyl sulfate (SDS) was added to brain homogenates before ultracentrifugation at 196,000×g (Beckman Optima TL100 centrifuge) for 30 min at 4°C. Supernatants obtained were used to quantify soluble fractions of human Aβ1–42 peptides with an enzyme-linked immunosorbent assay kit (Invitrogen). Results were normalized according to their protein concentration determined by the bicinchoninic acid (BCA) method protein assay (Pierce Biotechnology).

**Detection by Immunohistochemistry of Cyto, Mopa, and Pyri on Brain Tissue Sections of J20 Mice Treated with a Cocktail of Fungicides for 9 Months**

For this experiment, the number of brains analyzed per group was as follows: J20 CTR: $n = 7$ and J20 T: $n = 8$ (Figure 1C, Study 1). Three to six floating brain tissue sections per mouse were rinsed, mounted on Superfrost™ Plus glass slides, and then dried overnight at RT. Prior to amyloid labeling, brain sections were incubated for 10 min with a 70% formic acid solution for antigen retrieval. Sections were then incubated 1 h at RT with a blocking solution (3% goat serum, 0.3% Triton X100 in PBS) and incubated overnight at 4°C with the 6E10-Alexa488 antibody (1:1,000). Then, sections were washed and incubated with the antibodies anti-cyto, anti-mopa, or anti-pyri, alone or in cocktail, for 2 h at RT, followed by a 2-h incubation with a goat anti-mouse Cy3 antibody (1:1,000). Sections were counterstained with DAPI (1:50,000 in PBS). Control tissue sections were incubated either with PBS or the secondary antibody Cy3 alone. Images were taken with a Leica DM2500 microscope (magnification ×20).

**Immunohistochemistry and Quantifications**

For this experiment, the number of brains analyzed per group was as follows: WT CTR: $n = 3$; WT T: $n = 3$; J20 CTR: $n = 4$; and J20 T $n = 4$ (Figure 1C, Study 2). Neuroinflammation was assessed with a mouse anti-GFAP antibody (1:1,000; Sigma-Aldrich) for astroglia or with a rabbit polyclonal anti-IAB1 antibody (1:750; Wako Chemicals) for microglia. Neurons were analyzed with a mouse anti-NeuN antibody (1:500, clone A60; Merck-Millipore). Briefly, brain tissue sections were blocked 1 h at RT and incubated with one of the primary antibodies overnight at 4°C. Then, sections were washed and incubated with secondary antibodies, a goat anti-mouse Cy3 (1:1,000) or a goat anti-rabbit Alexa488 (1:1,000) for 2 h at RT. Nuclei were stained with DAPI (1:50,000). Images were taken with a Leica DM2500 microscope (magnifications: ×20 and ×40) and quantifications were performed with Fiji software in the DG, CA1, CA2, and CA3.

Astrogliosis was evaluated by quantifying the number of astrocytes (proliferation), the intensity of the labeling (expression of GFAP) and the area labeled (hypertrophy). To avoid saturation of the GFAP signal, all hippocampi were previously visualized on the microscope using the LAS software (version 4.10; Leica Microsystems) tool named show under- and over-exposure in the viewer in order to find the best image acquisition settings (intensity and gain) without overexposure of the labeling. The same acquisition settings were kept for all the conditions and hippocampi studied (intensity: 355.9 ms; gain: 10.1 ×; and 7: 1.53). The intensity and the area of the labeling were measured on 8-bit images using the threshold tool of the FIJI software. The same threshold parameters were used for all images.

The total number of microglial cells and the activated subset were quantified as described previously by Mansuy et al. (2018). To decipher the subset of activated microglial cells from the quiescent ones, quantifications were performed based on their morphological differences. The ramified IBA1-positive cells with small soma and long processes were considered to be quiescent cells, whereas IBA1-positive cells with enlarged soma, short processes, and decreased branching were considered to be activated microglial cells (ameboid).

The number of NeuN-positive neurons was quantified in the four different regions of the hippocampus, presented above, and was normalized by the surface area of the granular cell layers analyzed. All the results are presented as mean ± SEM.

**Interleukins and Cytokines Assays by HTRF®**

For this experiment, the number of brains analyzed per group was as follows: WT CTR: $n = 3$; WT T: $n = 3$; J20 CTR: $n = 4$; and J20 T $n = 4$ (Figure 1C, Study 2). Two percent SDS was added to brain homogenates prepared as described above, which were then centrifuged at 14,000 rpm, for 20 min at 4°C. IL1β,
IL6, and tumor necrosis factor α (TNFα) levels were determined using HTRF® assay kits (Cisbio). According to the manufacturer’s protocol, 16 μL of supernatant were mixed with 2 μL of donor antibody coupled to Eu3⁺-cryptate and 2 μL of acceptor antibody coupled to d2, both antibodies recognizing IL1β, IL6, or TNFα. The mix was incubated 1 h (IL6 and TNFα) or overnight (IL1β) at 25°C. Standard curves of each kit were made in the same homogenization buffer used for samples, containing SDS. The FRET signal was measured using the d2 acceptor emission (665 nm) and the Eu3⁺-cryptate donor emission (620 nm) with a FRET signal was measured using the d2 acceptor emission (665 nm) and the Eu3⁺-cryptate donor emission (620 nm) with a

mRNA Extraction and Real-Time qPCR

For this experiment, the number of brains analyzed per group was as follows: WT CTR: n = 3; WT T: n = 3; J20 CTR: n = 4; and J20 T: n = 4 (Figure 1C, Study 2). Total mRNA was extracted from brain homogenates with the Nucleospin RNA extraction kit (Macherey Nagel), according to the manufacturer’s instructions. RNA quality and concentrations were measured in a NanoDrop™ 8,000 spectrophotometer (Thermo Scientific). mRNA was reverse-transcribed with a high-capacity circular-DNA reverse transcription kit (Applied Biosystems, ThermoFisher Scientific) and amplified by real-time PCR (RT PCR) with a Light Cycler detection system (Roche) according to the manufacturer’s instructions. RT-qPCR was performed in collaboration with the qPCR Haut Débit qPCR platform (University of Montpellier, France) following the Minimum Information for Publication of Quantitative Real-Time PCR Experiments (MIQE) guidelines (Bustin et al. 2009), and β-actin was used as a reference gene. The ΔCt value of the reference and target genes was quantified as described previously (Nasri et al. 2016). For each target gene tested, experiments were performed three times independently and in triplicate. Primers (Eurogentec) used for RT-qPCR are listed in Table S1.

Statistical Analyses

All data were initially analyzed on GraphPad Prism software (version 5.0; GraphPad) using Shapiro-Wilk’s normality test. When data sets were normally distributed (p > 0.05), statistical analyses were performed using parametric tests: multiple t-test (Holm-Sidak’s correction), the student’s t-test, the linear correlations, or the two-way analysis of variance (ANOVA; Holm-Sidak’s post hoc analysis). However, when data sets were not normally distributed (p < 0.05) statistical analyses were performed using a nonparametric test: Mann-Whitney U-test. A probability of 0.05 has been defined as a significant difference for all statistical analyses.

Results

Ex Vivo Binding of Cypro, Mepa, and Pyri to Amyloid Plaques and Their Effects on the Kinetics of Aβ1-42 Fibril Formation in Vitro

We first evaluated whether cypro, mepa, and pyri could bind to amyloid plaques ex vivo. Brain tissue sections from 10-month-old J20 mice were incubated with 2 mM of these fungicides, either alone or in cocktail, for 1.5 h at RT. Primary antibodies against cypro (Estève-Turrillas et al. 2012), mepa (Estève-Turrillas et al. 2013), and pyri (Mercader et al. 2012) were used to detect them (Figure 2A, column 3). Amyloid aggregates were revealed using a 6E10 antibody conjugated to Alexa488 (Figure 2A, column 2). Remarkably, the merge column showed that fungicides, either incubated alone or in cocktail, bound to amyloid plaques ex vivo (Figure 2A 2–6). There was no nonspecific staining in controls when using an equivalent volume of DMSO (1%) corresponding to the cocktail condition (Figure 2A 1). Given that these antibodies

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were used for the first time for histological studies, further controls are presented in Figure S1.

Because anilinopyrimidines bound to amyloid plaques ex vivo, we wondered whether they could modify the kinetic profile of Aβ1–42 fibrils in vitro. Aβ1–42 monomers were obtained by size exclusion chromatography to exclude oligomeric species as described by Serra-Batiste et al. (2016). For the kinetics of amyloid fibril formation, Aβ1–42 monomers were mixed with the three anilinopyrimidines at a molar ratio of 1:1 (Aβ:fungicides) (Figure 2B, triangles) or of 1:15 (Figure 2C, squares). Control conditions were performed with an equivalent volume of DMSO (1%; Figure 2B, crosses, and 2C, circles). Tubes were incubated at 25°C and fibril formation was monitored by ThT, once a day. We found that the presence of fungicides dramatically modified the kinetic profile by accelerating the formation of Aβ1–42 fibrils at both ratios. These processes were shifted through a substantial decrease of the lag phase and an increased level of ThT fluorescence at the plateau phase compared with control conditions (Figure 2B,C).

**Body Weight, Water Consumption, and Pesticide Ingestion of Mice Treated with the Cocktail of Fungicides or with DMSO**

Our aim was to determine the impact of those three fungicides on different markers of Alzheimer’s disease in vivo. First, we assessed the impact of the cocktail of the fungicides on different physical parameters. From the weaning of mice until the end of the treatment 9 months later (35 weeks), neither WT nor J20 mice exhibited a modification of body weight gain upon treatment for either females (see Figure S2A) or males (see Figure S2B). Water consumption was estimated per mouse and per week for each group. The treatment with fungicides did not modify the drinking behavior for either females (see Figure S2C) or males (see Figure S2D) compared with their respective DMSO-treated mice (CTR). However, when comparing WT and J20 mice treated with the cocktail of fungicides, we noticed that transgenic mice drank more water than their WT littermates, for both females (see Figure S3A, 23.46 ± 0.38 mL/week for WT T vs. 27.88 ± 0.90 for J20 T, p < 0.001) and males (see Figure S3B, 25.18 ± 0.44 mL/week for WT T vs. 34.64 ± 0.60 for J20 T, p < 0.001). Estimation of fungicides ingestion per mouse and per week revealed that J20 female mice treated with the cocktail of fungicides ingested a significantly higher level of pesticides than their WT littermates (see Figure S3C, 2.418 ± 0.034 ng of fungicides for WT T vs. 2.788 ± 0.09 for J20 T, p = 0.0023). A similar difference in fungicide ingestion was observed between WT and J20 males (see Figure S3D, 2.518 ± 0.044 ng for WT T vs. 3.387 ± 0.061 for J20 T, p < 0.001).

**In Vivo Effect of Fungicides on Soluble and Aggregated Aβ1–42 in J20 Mice**

WT and J20 mice were treated with 0.1 μg/L of cypro, mepa, and pyri administered as a cocktail through drinking water (referred to as T) (Figure 1, Study 1). Control groups of WT and J20 mice were treated with an equivalent volume of DMSO (3 × 10−5%, referred to as CTR) (Figure 1, Study 1). Treatment started after weaning of mice and lasted 9 months, so mice were sacrificed at 10 months of age. The impact of the cocktail of fungicides on amyloid aggregates was carried out by histological analyses using ThT staining (Figure 3). First, we quantified the number and the size of aggregates in the hippocampus and the cortex, taking into account sex differences. Results showed no differences in the number or size of amyloid plaques between males and females after 9 months of treatment (see Figure S4; p > 0.05); therefore, results were pooled for statistical analyses. As expected, WT mice treated either with DMSO (CTR) or with the cocktail of fungicides (T) did not exhibit ThT-positive aggregates at this stage [Figure 3A (left panel) and 3B–E]. Treatment of J20 mice with the cocktail of fungicides (J20 T) for 9 months significantly increased the number of ThT-positive aggregates [Figure 3A (right panel)] by 26% and 53% in the hippocampus and the cortex, respectively (Figure 3B,D); hippocampus: 421 ± 29.4 aggregates/cm² for J20 CTR vs. 530 ± 23.7 for J20 T, p = 0.003 and cortex: 173.9 ± 14.6 for J20 CTR vs. 265.5 ± 23.1 for J20 T, p < 0.001). The treatment with fungicides also significantly modified the surface area of ThT-positive plaques by 42% in the hippocampus (Figure 3C; 0.279 ± 0.026 A.U. for J20 CTR vs. 0.397 ± 0.025 for J20 T, p = 0.002) as well as by 80% in the cortex (Figure 3E; 0.108 ± 0.01 for J20 CTR vs. 0.194 ± 0.018 for J20 T, p < 0.001). Distribution of aggregates in the different areas of the hippocampus (DG, CA1, CA2, and CA3) revealed the higher number of plaques in J20 T mice to be essentially located in the CA1 region (Figure 3F; 108.1 ± 12.1 aggregates/cm² for J20 CTR vs. 172.7 ± 26.8 for J20 T, p = 0.04). The same effect in the CA1 area was noticed for the surface area of ThT-positive aggregates (Figure 3G; 0.075 ± 0.007 A.U. for J20 CTR vs. 0.135 ± 0.019 for J20 T, p = 0.03). Soluble human Aβ1–42 levels were assayed in supernatants from ultracentrifuged brain homogenates. J20 treated mice exhibited almost three times more soluble Aβ1–42 compared with the control DMSO-treated mice (Figure 3H; 0.465 ± 0.212 pg/mg of proteins for J20 CTR vs. 1.323 ± 0.140 for J20 T, p = 0.02).

**Attempts to Detect Cypro, MePA, and Pyri in Situ in J20 Mice Brain Tissue Sections Treated for 9 Months with a Cocktail of Fungicides**

We believed that after a long-term exposure of fungicide those compounds could potentially accumulate in brains, especially in amyloid aggregates, based on our ex vivo and in vitro binding results. To evaluate this, brain tissue sections from J20 mice treated (J20 T, n = 7) or not (J20 CTR, n = 8) with 0.1 μg/L of fungicides during 9 months were analyzed (Figure 1C, Study 1). Tissues were first labeled with the 6E10-Alexa488 antibody to detect amyloid aggregates, and then, anti-fungicide antibodies (anti-cypro, -mepa, -pyri) were used to reveal their potential accumulation in Aβ plaques. As a control for the specificity of the fungicide labeling, DMSO-treated J20 brain tissue sections were used. As expected, no labeling was detected in the red channel (see Figure S5A-F, column 3). Regarding brain tissue sections from J20 mice treated with the cocktail of fungicides, antibodies raised against cypro, mepa, and pyri were not able to detect the presence of the fungicides in the amyloid aggregates in situ (see Figure S6A-F).

**Effect of the Cocktail Treatment on Neuroinflammation Processes**

**Astrogliosis analyses.** To determine the impact of the chronic pesticide exposure on the activation of astrocytes, we performed an immunohistochemistry evaluation on brain tissue sections with an anti-GFAP antibody and analyzed the different areas of the hippocampus (DG, CA1, CA2, and CA3) (Figure 4A). We quantified the intensity of the labeling reflecting GFAP expression and the surface area of the labeling to measure the hypertrophy of astrocytes (Figure 4B–E). The number of GFAP-positive cells is presented in Figure S7. Regarding J20 mice, treatment with fungicides significantly up-regulated GFAP expression in the CA1 area, reflecting an astrogliosis [Figure 4C (left panel); 1.749 ± 285.8 A.U. for J20 CTR vs. 3.456 ± 478.1 for J20 T, p = 0.030]. A tendency of up-regulation of GFAP was observed in the DG although it was not statistically significant [Figure 4B (left panel)]. In addition, in J20 T mice, both DG and CA1 exhibited...
**Figure 2.** (A) *Ex vivo* co-localization of cyprodinil, mepanipyrim, and pyrimethanil with Aβ aggregates on J20 brain tissue sections and (B,C) kinetics of Aβ1-42 fibril formation with a cocktail of the three fungicides. (A) Brain tissue sections from 10-month-old J20 mice were incubated for 90 min at room temperature with 2 mM of either cyprodinil (A2), mepanipyrim (A3), pyrimethanil (A4), a cocktail of the three fungicides (A5-6), or an equivalent volume of DMSO (1%) as a control (A1). Sections were labeled with an anti-human β-amyloid 6E10-Alexa488 and anti-cypro, anti-mepa, or anti-pyri antibodies. Aβ aggregates are seen in the green channel (column 2) and the different fungicides in the red channel (column 3). No red fluorescence was observed in the control condition (A1). The Merge column showed the co-localization of the fungicides with amyloid aggregates, and higher enlargements are presented in the panel A6 for the cocktail. For each condition, three to six sections were analyzed and independent experiments were performed at least three to six times. Images selected are representative of the labeling observed in different regions of the hippocampus (A1,3: dentate gyrus; A4: near dentate gyrus; A2,5-6: near CA1 area). Scale bars: 100 µM (A1-5: magnification ×20) and 50 µM (A6: magnification ×40). (B) Effect of a cocktail of fungicides on the kinetics of Aβ fibril formation. Human Aβ1-42 monomers (30 µM) were incubated at a molar ratio 1:1 with a cocktail of cypro, mepa, and pyri (30 µM each) (triangles) or with an equivalent volume of DMSO (1%) (crosses). ThT fluorescence intensity was measured once a day (in A.U.). For each condition, independent experiments were performed in duplicate. The different values obtained were plotted following a sigmoidal fit and representative curves of duplicates were presented. (C) Effect of a cocktail of fungicides on the kinetics of Aβ fibrils formation with a molar ratio 1:15 (30 µM Aβ:750 µM of each fungicide) (squares). Control conditions were performed with an equivalent volume of DMSO (1%) (circles). ThT fluorescence intensity was measured once a day (in A.U.). For each condition, independent experiments were performed in duplicate. The different values obtained were plotted following a sigmoidal fit and representative curves of duplicates were presented. Note: DAPI, 2-(4-aminophenyl)-1H-indole-6-carboxamidine; DMSO, dimethyl sulfoxide; ThT, thioflavin T.
Figure 3. (A) Thioflavin T (ThT) staining of brain sections of WT and J20 mice treated with the cocktail of fungicides or with DMSO for 9 months, (B,C,F,G) quantification of aggregates in the hippocampus, (D,E) in the cortex and (H) human soluble Aβ1–42 assay. (A) Representative images of amyloid plaques stained with ThT (in green) in the hippocampus and in the cortex of WT and J20 mice treated with the cocktail of fungicides (0.1 μg/L each) (T) or with an equivalent volume of DMSO (3 × 10⁻⁵%) (CTR) (Scale bar: 100 μm, magnification ×20). Inserts are enlargements of representative aggregates (scale bar: 50 μm, magnification ×40). (B–E) Quantifications of the number (B,D) and the surface area (C,E) of ThT-positive aggregates in WT and J20 mice (T and CTR), normalized by the total surface area of hippocampus or cortex analyzed. Aggregates were quantified using Fiji software. The number of aggregates are expressed as mean ± SEM/cm² and their surface area as mean ± SEM × 1,000 (without units). Statistical differences were calculated using a two-way ANOVA followed by a Holm-Sidak’s post hoc analysis. (F,G) The number (F) and surface area (G) of aggregates in the hippocampus were quantified in the different areas: dentate gyrus (DG), CA1, CA2, and CA3 for the J20 mice (T and CTR). Values are expressed as mean ± SEM and statistical differences were calculated using a multiple t-test followed by a Holm-Sidak’s post hoc analysis. For all histological analyses, 8–10 brain tissue sections per animal were used (A–G). (H) Soluble forms of human Aβ1–42 in WT and J20 mice treated with the fungicides (T) or with DMSO (CTR) were quantified on supernatants of ultracentrifuged brain homogenates by ELISA. Results were normalized by protein concentration and are expressed as mean ± SEM (pg of Aβ1–42/mg of proteins). Statistical differences were obtained using a two-way ANOVA followed by a Holm-Sidak’s post hoc analysis. For all statistical tests a probability of 0.05 was defined as a significant difference (*p < 0.05, **p < 0.01, ***p < 0.001). n = 4–8 mice per group issued from Study 1. Note: ANOVA, analysis of variance; CTR, control; DAPI, 2-(4-amidinophenyl)-1H-indole-6-carboxamidine; DMSO, dimethyl sulfoxide; ELISA, enzyme-linked immunosorbent assay; T, treatment; WT, wild type.
A hyper trophy of astrocytes upon treatment [Figure 4B,C (right panels)] (in DG: 47.3 ± 8.6 μm²/cell for J20 CTR vs. WT CTR) for J20 T mice, p = 0.047; in CA1: 39.4 ± 5.9 for J20 CTR vs. 82.9 ± 11.6 for J20 T, p = 0.028). Regarding the number of astrocytes, treatment had an impact only on WT mice in the CA2 region (see Figure S6C: 375.7 ± 99.9 astrocytes/mm² for WT CTR vs. 651.1 ± 34.1 for WT T, p = 0.027), but was not associated with an upregulation of GFAP expression or a hypertrophy of astrocytes (Figure 4D).

Microgliosis analyses. To study microglial cells, we performed histological analyses using the constitutive marker, IBA1 (Figure 5A). Labeling in the hippocampus was quantified as the total number of IBA1-positive cells and the ameboid ones in the DG, CA1, CA2, and CA3 areas (Figure 5B–E). For J20 mice, the total number of microglial cells was greater upon treatment with fungicides than in mice treated with vehicle in all regions analyzed (Figure 5B–E (left panels)); DG: 266.7 ± 9.9 for J20 CTR vs. 518.1 ± 47.2 for J20 T, p < 0.001; CA1: 148.4 ± 11.8 for J20 CTR vs. 217.9 ± 7.5 for J20 T, p = 0.0036; CA2: 150.7 ± 8.2 for J20 CTR vs. 268.3 ± 25.8, p = 0.0029 and CA3: 193.5 ± 5.3 for J20 CTR vs. 272.6 ± 23.9 for J20 T, p = 0.014]. The treatment also resulted in a greater number of activated microglial cells in all regions apart from CA1 (DG: 95.17 ± 11.01 activated microglia/mm² for J20 CTR vs. 178.9 ± 21.07 for J20 T, p = 0.006; CA2: 43.97 ± 3.3 for J20 CTR vs. 75.36 ± 8.09, p = 0.019; and CA3: 44.77 ± 4.34 for J20 CTR vs. 76.34 ± 6.87 for J20 T, p = 0.006). Analysis of microglial cells in WT mice did not reveal any difference upon treatment with the cocktail of fungicides in the hippocampus (Figure 5A–E).

Interleukins and cytokines secretion. Because brain inflammatory processes involve the secretion of pro-inflammatory interleukins and cytokines, we next measured levels of IL1β (IL1β), IL6, and TNF-α in brain homogenates by HTRF® (Figure 6). Brains from J20 mice treated with fungicides exhibited a 1.6-fold higher level of IL1β (Figure 6A; 10.74 ± 1.18 pg/mg for J20 CTR vs. 17.72 ± 1.15 for J20 T, p = 0.093) and a nearly 3-fold higher level of IL6 (Figure 6B; 0.39 ± 0.05 pg/mg for J20 CTR vs. 1.09 ± 0.15 pg/mg for J20 T, p = 0.0429). However, TNF-α levels were unchanged in the J20 mice treated with the cocktail of fungicides compared with J20 CTR mice (Figure 6C). There were no differences in WT mice treated with or without the fungicide cocktail (Figure 6A,C) apart from a slight nonsignificant increase in IL6 levels (Figure 6B).

Effect of the Chronic Fungicide Treatment on the Hippocampal Neuronal Loss

We wondered whether the treatment with the cocktail of fungicides could also impact hippocampal neuronal loss. To this aim, we labeled brain tissue sections with an anti-NeuN antibody to study undamaged neurons in the four different areas of the hippocampus (DG, CA1, CA2, and CA3) (Figure 7). Remarkably, in WT animals, analyses revealed that treatment with the cocktail of fungicides increased the number of neurons in the CA1 and CA3 areas (Figure 7C,E) [means ± standard error means (SEMs) and statistical analyses for WT mice are presented in Table S2]. Interestingly, J20 mice exhibited a significant decrease of the number of neurons in the CA3 region compared with DMSO-treated animals (Figure 7E; 2.386 ± 120.7 neurons/mm² of granular cell layers for J20 CTR vs. 1.978 ± 71.21 for J20 T, p = 0.0258) (means ± SEMs and statistical analyses for J20 mice are presented in Table S3).
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Longitudinal Study of Appearance of Aggregates in the Superficial Cortex of J20 Mice Chronically Exposed to Fungicides for 9 Months

To clarify the impact of the chronic exposure to low doses of fungicides on the progression of the amyloid burden, we studied the dynamics of aggregation in vivo using a longitudinal approach by two-photon microscopy. Because the J20 mouse model is also known for its cerebral amyloid angiopathy (CAA) phenotype (Kimbrough et al. 2015), we took advantage of this characteristic to monitor amyloid plaque formation on the superficial cortex over time on the same animals. In this study (Figure 1D), J20 mice treated with the cocktail of fungicides (n = 5) or with DMSO (n = 3) underwent three imaging sessions at 3, 6, and 9 months of treatment (Figures 8 and 9). Mice were injected at each time point with Methoxy-XO4, an amyloid dye (in blue but artificially colored in red) and with dextran-FITC to stain blood vessels (in green). As expected, no Methoxy-XO4-positive aggregates were detected at 3 months on the superficial cortex in both groups, given that at this age no vascular aggregates were detected (Kimbrough et al. 2015) (Figure 8A,B). In J20 CTR mice, we observed an accumulation of Aβ peptides both around blood vessels (arrows) and in the parenchyma (circles) at 6 and 9 months (Figure 8A). Strikingly, J20 mice treated with the cocktail of fungicides exhibited an enhanced perivascular aggregation (arrows) at 9 months (Figure 8A, arrows). To visualize the evolution of perivascular aggregation, we performed an analysis of variance using a two-way ANOVA followed by a Holm-Sidak’s post hoc analysis (α = 0.05). Note: ANOVA, analysis of variance; CTR, control; DMSO, dimethyl sulfoxide; HTRF®, homogeneous time resolved fluorescence; IL, interleukin; T, treatment; TNFα, tumor necrosis factor α; WT, wild type.

Figure 5. (A) Microglial labeling of WT and J20 mice treated with 0.1 µg/L of a cocktail of fungicides or with DMSO for 9 months and quantification of the number of total microglial cells and the activated subset in the (B) dentate gyrus (DG), (C) CA1, (D) CA2, and (E) CA3 areas. Animals were treated with a cocktail of fungicides (0.1 µg/L each) (T) or with an equivalent volume of DMSO (3 × 10⁻⁵%/v/v) (CTR) for 9 months. The numbers of total and activated (ameboid) microglial cells per mm² were quantified for the DG (B), CA1 (C), CA2 (D), and CA3 (E) regions. For all immunohistochemistry analyses, 8–10 brain tissue sections per animal were used. Values are expressed as mean ± SEM and statistical differences were measured using a two-way ANOVA followed by a Holm-Sidak’s post hoc analysis (α = 0.05, *p < 0.05, **p < 0.01, ***p < 0.001). Scale bar: 100 µm (magnification × 40). Inserts are representative images of the microglial cells status in the different groups (scale bar: 50 µm, magnification × 40). n = 3–4 animals per group issued from Study 2. Note: ANOVA, analysis of variance; CTR, control; DMSO, dimethyl sulfoxide; T, treatment; WT, wild type.

Figure 6. (A) IL1β, (B) IL6, and (C) TNFα levels in brain homogenates from WT and J20 mice treated with fungicides or with DMSO for 9 months. (A) IL1β, (B) IL6, and (C) TNFα levels were measured in brain homogenates of WT and J20 mice treated with the cocktail of fungicides (0.1 µg/L each) (T) or with 3 × 10⁻⁵ % of DMSO (CTR) for 9 months. Interleukin and cytokine levels were assayed using HTRF® kits and results were normalized by their protein concentration (BCA). Three to four different brains were analyzed in triplicate for each group issued from Study 2. Results are presented as mean ± SEM (pg/mg of proteins) and statistical differences were calculated using a two-way ANOVA followed by a Holm-Sidak’s post hoc analysis (α = 0.05, *p < 0.05, ***p < 0.001). Note: ANOVA, analysis of variance; CTR, control; DMSO, dimethyl sulfoxide; HTRF®, homogeneous time resolved fluorescence; IL, interleukin; T, treatment; TNFα, tumor necrosis factor α; WT, wild type.
months compared with their control littermates treated with DMSO (Figure 8B). We quantified the total number of aggregates as well as their distribution in the parenchyma and around blood vessels. Analyses revealed that the number of parenchymal aggregates were not impacted by the cocktail of fungicides (Figure 8D), whereas vascular aggregates were about three times higher in J20 T mice compared with J20 CTR at 9 months posttreatment (Figure 8E; 63.67 ± 12.91 for J20 CTR vs. 183.4 ± 52.66 for J20 T, p = 0.031). When counting the number of aggregates, we could see the fusion of some plaques between 6 and 9 months in J20 T mice, as shown by arrowheads in Figure 8B, leading to an under-quantification of their number. To overcome this bias, we performed a 3D reconstruction using Imaris software (version 8.0; Bitplane) on the stacks of images acquired at 3, 6, and 9 months of treatment to quantify the volume of aggregates (Figure 9A). In these 3D images, we found the main characteristic of CAA, which is the presence of vascular aggregates on leptomeningeal and penetrating vessels surrounded by some parenchymal aggregates (Figure 9B). We performed a quantification of the total volume of aggregates as well as the distribution between the parenchymal and vascular ones, normalized by the total volume of the stacks. Analyses showed a significant 3-fold increase of the total volume of aggregates at 9 months of treatment (Figure 9C; 60.1 ± 4.68 A.U. for J20 CTR vs. 173.9 ± 49.51 for J20 T, p = 0.028), mainly due to an 18-fold increase of the volume of the vascular aggregates (Figure 9E; 6.86 ± 1.15 A.U. for J20 CTR vs. 125.6 ± 60.06 for J20 T, p = 0.023). During this experiment, vascular aggregates were all found to be located on the blood vessel walls, except for one aggregate that was found inside a blood vessel (see Figure S8). We then performed a linear correlation between the volume of vascular and parenchymal aggregates using all the data collected at 3, 6, and 9 months posttreatment. For each mouse, three values were presented in the graph, with a total of 24 points for eight representative images of the granular cell layers (scale bar: 50 μm, magnification ×40). n = 3–4 mice per group issued from Study 2. Note: ANOVA, analysis of variance; CTR, control; DMSO, dimethyl sulfoxide; NeuN, neuronal nuclei; T, treatment; WT, wild type.
of vascular aggregates [Figure 9F (squares); $y = 7.566x + 17.19$, $r = 0.742, p = 0.014$].

**Impact of the Chronic Fungicide Treatment on Gene and Protein Expression of Protagonists Involved in the Production or Clearance of Aβ Peptides**

Because amyloid plaques and soluble forms of Aβ$_{1-42}$ were increased upon fungicide treatment (Figure 3B–E,H), we addressed whether cypro, mepa, and pyri could modulate either the production or the clearance of Aβ peptides. To this aim, we studied the gene and protein expression of the main protagonists of the amyloid cascade. First, total mRNA was extracted from brain homogenates of J20 mice treated with a cocktail of fungicides (T, $n = 4$) or with an equivalent volume of DMSO (CTR, $n = 4$) (Figure 1C, Study 2). The same procedure was performed for their WT littermates (CTR, $n = 3$; T, $n = 3$) (Figure 1C, Study 2). We analyzed mRNA expression of endogenous *App* (human), *Bace1*, *Ide*, *Nep*, and *Lrp1* by RT-qPCR (see Figure S9A–F). No modifications of mRNA expression were observed for all the genes tested between fungicide-treated and DMSO-control.
mice. In parallel, we analyzed the expression of proteins corresponding to the genes cited above by immunoblotting. Analyses revealed a substantial increase of BACE1 expression in J20 mice treated with the cocktail of fungicides compared with the J20 CTR mice (Figure 10C; 0.762 ± 0.076 for J20 CTR vs. 1.323 ± 0.211 for J20 T, \( p = 0.046 \)). In addition, NEP expression, one of the main clearance enzymes, was expressed in J20 mice treated with fungicides during 9 months at a level 50% of that of control-treated mice (Figure 10E; 0.274 ± 0.032 for J20 CTR vs. 0.141 ± 0.024 for J20 T, \( p = 0.017 \)).

**Discussion**

Pesticides are widely used to preserve fields and harvests from pests. Their massive use since the 20th century has led to the...
contamination of all environmental media (water, soil, air), leading to residues in our diet. Although these compounds are present at very low concentrations (from nanograms to micrograms) in our environment (Table 1), the long-term effect of multiple pesticide residues on human health is subject to intense debate and concerns. Toxicological regulations are mainly based on high-dose effects and minimize the consequences of these traces. According to European institutions, pesticide residues can be consumed over a lifetime.
without presenting a significant risk for human health (Council of the European Union 1998; EFSA 2017b, 2018, 2019). However, a study by Lukowicz et al. (2018) has demonstrated the impact of these chemical traces on human health. They showed that chronic exposure to low doses of pollutants occurring throughout life and the pathology. However, Richardson et al. (2014) showed that DDE accumulated in the brains of 11 individuals with Alzheimer’s disease although this herbicide has been prohibited since the 1970s. Furthermore, acute injections of chlorpyrifos (50 mg/kg) or paraquat (10 mg/kg twice a week for 3 weeks) to Tg2576 (hAPPsw) mice induced an increased production of Aβ peptides in the brain (Chen et al. 2012; Salazar et al. 2011). However, the doses and the route of treatment used in these in vivo studies are not relevant to the typical consumer’s exposure to pesticides. To address the effect of pesticide residues in Alzheimer’s disease, we used the J20 mouse model that had been chronically exposed to a cocktail of three fungicides (cypro, mepa, and pyri) through drinking water. Those residues are frequently detected in the European diet (Table 1). Mice were treated with 0.1 μg/L of each fungicide, leading to a final concentration of 0.3 μg/L in water. According to EU regulations, 0.1 μg/L corresponds to the maximal dose allowed for one pesticide in tap water and cannot exceed 0.5 μg/L for the presence of multiple residues (Council of the European Union 1998).

**Limits of Detection and Quantification of Cypro, Mepa, and Pyri**

The limits of detection (LOD) and quantification (LOQ) of these fungicides in water were respectively 0.019 and 0.063 μg/L for cypro [solid phase extraction–gas chromatography–mass spectrometry (SPE-GC-MS; Climent et al. 2018)], 0.03 and 0.11 μg/L for mepa [single drop micro extraction (SDME)–GC-MS; Araujo et al. 2013], and 0.026 and 0.088 μg/L for pyri (SPE-GC-MS; Climent et al. 2018). In the present study, the dose used to treat WT and J20 animals was 0.1 μg/L for each fungicide, ranging from 444 to 502 pM. To detect such doses in an animal’s brain, these fungicides should accumulate in this tissue and the detection of the nonmetabolized fungicides seems to be the limiting factor. Indeed, reports on mammalian toxicokinetics (absorption, distribution, excretion, and metabolism) of the three fungicides showed that these compounds were highly absorbed (80–95%) within 48 h and then highly excreted (EFSA 2006a, 2006b, 2017b). For example, cypro was extensively metabolized (EFSA 2006a) and up to 28 metabolites could be generated (FAO 2003). This compound was mainly excreted via urine (52–63%) and feces (33–45%), and only 3–8% of cypro was excreted unchanged in the feces (EFSA 2006a). The biodistribution of cypro in rats showed that there was no evidence of accumulation of this compound and <1% of the dose administered was found in tissues after 7 d of treatment (EFSA 2006a). The toxicokinetics of mepa and pyri were similar to those of cypro (EFSA 2006b, 2017b). In addition, toxicological reports did not indicate any data regarding the presence of these fungicides in brains of different animals studied (e.g., cows, goats, rats, hens) (EFSA 2006a, 2006b, 2017b; FAO 2003, 2007). The presence of cypro, mepa, and pyri within amyloid plaques on brain tissue sections of J20 mice treated for 9 months with DMSO or with the cocktail of fungicides was assessed using anti-fungicide antibodies (see Figures S5 and S6). However, none of these fungicides could be detected in the amyloid plaques. The absence of labeling in this experiment could be explained either by the fact that these fungicides may not accumulate in brains and/or that compounds may be extensively metabolized. In addition, the anti-fungicide antibodies may not recognize metabolites. By contrast, the *ex vivo*
suggest that the three fungicides may interfere directly in the aggregation of these fungicides, their nonaccumulation in tissues and their extensive rate of metabolism, the remaining concentration of the unchanged compounds may be barely detectable (i.e., <LOD) after a chronic treatment at 0.1 µg/L. In addition, the high number of possible metabolites for each compound 28 for cypro (FAO 2007), 4 for mepa (EFSA 2011), and 19 for pyri (FAO 2007) may further dilute the signal and increase the difficulties of detecting those compounds in brains.

**Plaque Formation in the Hippocampus and the Cortex of J20 Mice Treated with the Fungicide Cocktail**

Our results showed that a 9-month exposure to residues of cypro, mepa, and pyri (0.1 µg/L of each compound), mimicking a 20-y exposure in humans, induced a substantial increase in the number of amyloid plaques of about 26% and 53% in the hippocampus and the cortex of J20 mice, respectively (Figure 3B,D). Similarly, the treatment incremented the surface area of amyloid aggregates as well (hippocampus, ~42%; cortex, ~80%; Figure 3CE). These results suggest that the three fungicides may interfere directly in the aggregation process, as suggested by our *in vitro* studies (Figure 2). As expected, WT mice did not exhibit ThT-positive aggregates [Figure 3A (left panel) and 3B-E], which was consistent with the literature as mouse Aβ aggregates are only visible from 15 months of age (Ahlemeyer et al. 2018). Moreover, our two-photon analysis revealed that the dynamics of Aβ aggregation in presence of fungicides occurred only between 6 and 9 months of treatment (Figures 8 and 9). Thus, it seems that the increase of amyloid aggregates in these conditions is dependent on preexisting amyloid plaques, possibly explaining the absence of ThT-positive aggregates in WT mice treated 9 months with fungicides. Remarkably, we showed that fungicides co-localized with amyloid plaques of J20 brain tissue sections after incubation, either alone or in cocktail, reflecting some affinity of these fungicides toward amyloid fibrils (Figure 2A). In addition, kinetics of amyloid fibril formation in presence of the three fungicides drastically reduced the lag phase and increased by more than 50% the ThT fluorescence intensity at the plateau, suggesting that the fibrils formed may have different structural characteristics (Figure 2BC). After 100 h of incubation of Aβ1-42 peptides with 30 µM of each fungicide (ratio 1:1) or with 750 µM of each fungicide (ratio 1:15), the kinetics reached the plateau phase, whereas the control curves had only begun their elongation phase, suggesting that these compounds promote a much faster nucleation than the control (Figure 2BC). Our *in vitro* studies demonstrated a pro-aggregating effect of these fungicides. Remarkably, this process also occurred in J20 mice treated with concentrations as low as 444–502 PM (0.1 µg/L) (Figures 2 and 3). Brain homogenates analyses also showed that J20 treated mice presented almost three times more soluble human Aβ1-42 levels than the control group (Figure 3H). Because the kinetics of fibril formation started nearly immediately in the presence of fungicides (Figure 2BC), we hypothesize that this increased soluble human Aβ1-42 fractions found in vivo may be composed mainly of soluble oligomeric species, rather than monomers. The substantial increase of Aβ species upon treatment may be related to the higher levels of BACE1 (Figure 10C) and lower levels of NEP proteins (Figure 10E). We speculate that the higher levels of BACE1 may increase β-cleavage of APP and lower levels of NEP may result in a decrease in clearance of amyloid peptides, thus facilitating accumulation of Aβ species. Our results are in agreement with a recent study showing the effect of six triazine herbicides on β- and γ-secretases associated with an overproduction of Aβ1-42 peptides on inducible pluripotent stem cells (iPSCs) derived from patients with familial Alzheimer’s disease (Portelius et al. 2016). Regarding WT mice, no differences were observed in protein expression levels for the different protagonists involved in the amyloid cascade (Figure 10A–F), possibly explaining why we did not observe the presence of amyloid plaques in WT mice after 9 months of fungicide treatment (Figure 3A-E).

**Cerebral Amyloid Angiopathy upon Fungicide Cocktail Treatment**

Because we did not know when the pro-aggregative effect of these fungicides started in the J20 treated mice, we carried out a longitudinal study at 3, 6, and 9 months by two-photon microscopy (Figures 8 and 9). Analyses of the different time points revealed that treatment of J20 mice with fungicides a) increased amyloid plaque formation between 6 and 9 months, b) caused a substantial augmentation of vascular amyloid plaques reminiscent to cerebral amyloid angiopathy (CAA), and c) had no effect on parenchymal aggregates of the superficial cortex. In addition, in the J20 fungicide-treated group, the highlighting of the low responders and the high responders subpopulations implied an interindividual susceptibility to pesticide residues exposure toward CAA (Figure 9F). We thus can hypothesize that this sensitivity toward low doses of fungicides may also exist in humans. Analysis of the 3D reconstructed images also showed that vascular aggregates were placed on the blood vessel walls and may not cross them, except for one aggregate that was found inside a blood vessel at 6 months of treatment and disappeared at 9 months (see Figure S8). These results suggest that the enhanced CAA observed in J20 treated mice could also be related to defects in the peri-arterial interstitial fluid drainage pathway to eliminate the surplus of aggregates (Yoon and AhnJo 2012). This striking effect on the CAA is likely linked to the disruption of the blood–brain barrier (BBB) in J20 mice (Kimbrough et al. 2015), likely facilitating the entrance of cypro, mepa, and pyri into the central nervous system (CNS). A toxicological report showed that oral treatment of a cow with pyri (0.4 mg/kg body weight per day for 7 d) resulted in the detection of 34 µg of metabolites in the blood (FAO 2007). Results were similar for the biodistribution of pyri metabolites when treatment was applied to rats (FAO 2007). Altogether, these toxicological data demonstrate that these fungicides and their metabolites circulate in the blood and thus can reach the brain. In addition, studies on J20 mice showed that CAA deposits disrupted the normal contact of astrocytic end-feet with vessels, thus damaging the BBB (Kimbrough et al. 2015). This disruption is an open window for pesticides to reach CNS and strengthen CAA. Our results are in accordance with the permeability of the BBB of the J20 model. Previous studies showed that β-amyloid deposits formed a physical cast, making blood vessels more rigid and unable to constrict and dilate as do vessels free of amyloid and leading to vascular pathologies (Kimbrough et al. 2015). In this context, one can hypothesize that the chronic exposure to those fungicide residues of patients already having a permeability of the BBB and an altered gliovascular unit could result in an aggravation of vascular deposits. These deposits could lead ultimately to CAA-associated intracerebral hemorrhages, a known comorbidity of Alzheimer’s disease that often leads to cognitive impairments (Shi et al. 2000).

**Neuroinflammation Processes upon Fungicide Treatment in J20 Treated Mice**

Neuroinflammation processes are one of the hallmarks of Alzheimer’s disease and is associated with the presence of both activated astrocytes and microglia. GFAP is the typical marker used to analyze the
activation of astrocytes and their morphological changes (Eng et al. 1992). IBA1 allows the study of changes in microglial cells from a ramified state (quiescent) to an activated state (ameoboid) (Ito et al. 1998). A previous study revealed that J20 mice exhibited a peak of astrogliosis in the hippocampus at 6 months of age while activated microglial cells increased over time (Wright et al. 2013). Our results showed that J20 mice treated with fungicides exhibited higher astrogliosis (DG, CA1, Figure 4), as well as higher activated microglial cells in the different regions of the hippocampus upon 9 months of fungicide treatment (Figure 5). In a study of 40 postmortem brains of patients with Alzheimer’s disease, Serrano-Pozo et al. (2011) found more activated microglia near amyloid plaques (<50 μm) than distant to the plaques. Authors speculated that this finding may imply that amyloid plaques could be involved in the activation of microglial cells (Serrano-Pozo et al. 2011). Augmentation of amyloid aggregates in the hippocampus of J20 mice treated with fungicides (Figure 3) may participate in the increased number of ameboid plaques observed in the DG (Figure 5A, row 1). Moreover, we also measured a greater number of the pro-inflammatory interleukins IL1β and IL6 in the brains of J20 mice treated with fungicides compared with J20 mice treated with vehicle (Figure 6A–B). These two interleukins have been shown to contribute to neuronal death. Indeed, IL1β was shown to induce neuronal cell death when recombinant IL1β treatment (0.01–100 ng/mL) was applied in a glial–neuronal coculture extracted from rat embryos (Thorton et al. 2006). By contrast, IL6 had a dual effect: In physiological conditions, IL6 was involved in adult neurogenesis (Deverman and Patterson 2009) as well as in cell survival (Hama et al. 1989); however, in pathological conditions, IL6 can contribute to neuron cell death, associated with other factors. A study showed that cortical neurons extracted from rat embryos, treated with IL6 (5 ng/mL), can have an effect only when combined with Aβ33–35 peptides (20 μM), by increasing cell damage (LDH assay) and by decreasing cell survival (MTT assay) (Qiu and Grool 2003). The higher levels of IL1β and IL6 detected in the J20 mice treated with fungicides may explain the neuronal cell death observed in the CA3 area of these mice (Figure 7E). In addition, activation of IL1β, IL6, and TNFz receptors has been described to activate NFkB and AP-1 transcription factors (Figure 6 of Cronin et al. 2016; Jain et al. 2014; Shi and Sun 2018). Interestingly, NFkB and AP-1 can activate the transcription of BACE1 (Chami and Checler 2012), which could explain, in part, the increased expression of the β-secretase in the J20 mice treated with fungicides but not in their WT littermates (Figure 10C). So we could imagine a vicious circle where the fungicides could induce more amyloid aggregates, which in turn would produce more inflammatory processes. We can then hypothesize that the activation of microglia and astrocytes could lead to a secretion of pro-inflammatory interleukins, emphasizing BACE1 expression and ultimately leading to an increased production of Aβ42 peptides. Thus, in the presence of the fungicides, Aβ42 peptides may aggregate faster, producing higher and larger amyloid plaques and thus exacerbating, again, inflammatory processes.

Conclusions

To our knowledge, our study is the first one demonstrating the effect of very low doses of fungicides in the aggravation of Alzheimer’s disease markers in a transgenic mouse model reminiscent of familial cases. Because the fungicides we studied exacerbated amyloid fibril formation, amyloid plaques, and inflammatory processes, we can suspect that they may also corrupt normal brain aging and aggravate the markers of sporadic Alzheimer’s disease cases. For this purpose, future studies will be carried out on a mouse model expressing WT human APP to decipher whether treatment with fungicides could also have a causal effect in Alzheimer’s disease. According to the EU, such low doses in water (0.1 μg/L) have been selected to “ensure that water intended for human consumption can be consumed safely on a lifelong basis and thus represent a high level of health protection” (Council of the European Union 1998). However, our results showed that these doses, on the long term, are deleterious in a transgenic mouse model of Alzheimer’s disease. Cypro, mepa, and pyri could be considered Alzheimerogen pollutants (Cam et al. 2018; Portelius et al. 2016) at the subnanomolar regulatory levels (0.1 μg/L) because they accelerated fibrils formation and promoted the progression of disease’s markers in the J20 mouse model. We hypothesize that prevention strategies toward pesticide long-term exposure may be an alternative to counterbalance the lack of treatment and to slow down the worldwide Alzheimer’s epidemic.

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P.A.L. and V.P. conceived the study, participated in its design and interpretation and wrote the manuscript. P.A.L., Y.W., J.T., M.A.L., L.S.P., M.M., C.D., and V.P. performed the experiments and analyzed data. J.V.M. provided antibodies against cyprodinil, mepanipyrim, and pyrimethanil. N.M.F., C.D., and L.G. provided protocols and advice for histological studies. F.J., J.T., L.S.P., and J.L. provided constructive criticisms for manuscript. All authors read and approved the final manuscript.

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