



HAL
open science

Reptiles in Guadeloupe (French West Indies) are a reservoir of major human *Salmonella enterica* serovars

Stéphanie Guyomard-Rabenirina, François-Xavier Weill, Simon Le Hello, Sylvaine Bastian, Franck Berger, Séverine Ferdinand, Pierre Legreneur, Cécile Loraux, Edith Malpote, Blandine Muanza, et al.

► To cite this version:

Stéphanie Guyomard-Rabenirina, François-Xavier Weill, Simon Le Hello, Sylvaine Bastian, Franck Berger, et al.. Reptiles in Guadeloupe (French West Indies) are a reservoir of major human *Salmonella enterica* serovars. PLoS ONE, 2019, 14 (7), pp.e0220145. 10.1371/journal.pone.0220145 . inserm-02273207

HAL Id: inserm-02273207

<https://inserm.hal.science/inserm-02273207>

Submitted on 28 Aug 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

RESEARCH ARTICLE

Reptiles in Guadeloupe (French West Indies) are a reservoir of major human *Salmonella enterica* serovars

Stéphanie Guyomard-Rabenirina^{1*}, François-Xavier Weill², Simon Le Hello², Sylvaine Bastian³, Franck Berger^{4,5}, Séverine Ferdinand¹, Pierre Legreneur⁶, Cécile Loraux³, Edith Malpote³, Blandine Muanza⁷, Vincent Richard⁸, Antoine Talarmin¹, Sébastien Breurec^{1,3,9}



1 Unité Transmission, Réservoir et Diversité des Pathogènes, Institut Pasteur de Guadeloupe, Les Abymes, France, **2** Unité des Bactéries pathogènes entériques, Centre National de Référence des *Escherichia coli*, *Shigella* et *Salmonella*, Institut Pasteur, Paris, France, **3** Laboratoire de Microbiologie clinique et environnementale, Centre Hospitalier Universitaire de Pointe-à-Pitre/les Abymes, Pointe-à-Pitre, France, **4** Service de Santé des Armées, Centre d'épidémiologie et de santé publique des armées, Marseille, France, **5** INSERM, IRD, Sciences Economiques et Sociales de la Santé et Traitement de l'Information Médicale, Université d'Aix Marseille, Marseille, France, **6** CRIS EA 647, Université de Lyon, Villeurbanne, France, **7** Service de Pédiatrie, Centre Hospitalier Universitaire de Pointe-à-Pitre/les Abymes, Pointe-à-Pitre, France, **8** Institut Pasteur de Nouvelle-Calédonie, Nouméa, Nouvelle-Calédonie, **9** Faculté de Médecine Hyacinthe Bastaraud, Université des Antilles, Pointe-à-Pitre, France

* sguyomard@pasteur-guadeloupe.fr

OPEN ACCESS

Citation: Guyomard-Rabenirina S, Weill F-X, Le Hello S, Bastian S, Berger F, Ferdinand S, et al. (2019) Reptiles in Guadeloupe (French West Indies) are a reservoir of major human *Salmonella enterica* serovars. PLoS ONE 14(7): e0220145. <https://doi.org/10.1371/journal.pone.0220145>

Editor: Jose A. Chabalgoity, Universidad de la Republica Uruguay, URUGUAY

Received: March 1, 2019

Accepted: July 9, 2019

Published: July 19, 2019

Copyright: © 2019 Guyomard-Rabenirina et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its Supporting Information files.

Funding: The Institut Pasteur was the funder of this study (<https://www.pasteur.fr/fr>). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Abstract

The epidemiology of human *Salmonella enterica* infections in Guadeloupe (French West Indies) appears to be specific, with a higher prevalence of the subspecies *enterica* serovars Panama and Arechavaleta (Panama and Arechavaleta) than in other regions. A study was performed in Guadeloupe to identify the reservoir of *Salmonella* serovars by comparing their distribution in warm- and cold-blooded animals and in humans living in Guadeloupe and mainland France. Furthermore, a case-control study was conducted in 2012–2013 to identify the main epidemiologic risk factors for *S. enterica* infection among children under 15 years of age. Between June 2011 and December 2014, feces from 426 reptiles (322 anoles, 69 iguanas and 35 geckos) and 50 frogs distributed throughout Guadeloupe and nearby islands were investigated. The frequency of *S. enterica* carriage was 15.0% (n = 64) in reptiles but varied by species. The only significant risk factor for *S. enterica* infection was a more frequent presence of frogs in the houses of cases than in those of controls (P = 0.042); however, isolates were not collected. Panama and Arechavaleta were the two serovars most often recovered between 2005 and 2014 from humans living in Guadeloupe (24.5% (n = 174) and 11.5% (n = 82), respectively), which is in contrast to the low prevalence in mainland France (0.4%). Their presence at low frequencies in wild reptiles (4.6% (n = 3) and 3.1% (n = 2), respectively) and pigs (7.5% (n = 5) and 1.5% (n = 1), respectively) suggests a broad host range, and humans may be infected by indirect or direct contact with animals. These serovars are probably poorly adapted to humans and therefore cause more severe infections. The unusual subspecies *houtenae* serovar 43:_{Z4,Z32}:- was a major subspecies in

wild reptiles (24.6%, $n = 16$) and humans (9.4%, $n = 67$) but was not recovered from warm-blooded animals, suggesting that reptiles plays a key role in human infection.

Introduction

All serovars of *Salmonella* belong to two species, *S. enterica* and *S. bongori*, although more than 99.5% of isolates are assigned to *S. enterica*. *S. enterica* comprises six subspecies: *enterica*, *salmamae*, *arizonae*, *diarizonae*, *houtanae*, and *indica*. Most cases of human illness arise from *enterica* subspecies. Serovars Typhi, Paratyphi A, Paratyphi B and Paratyphi C are grouped as typhoidal *Salmonella*, and other serovars are described as non-typhoidal *Salmonella* (NTS). Typhoidal *Salmonella* are human host-restricted bacteria that cause typhoid and paratyphoid fever, which are systemic diseases, whereas a large number of NTS serotypes are generally responsible for acute gastroenteritis. The incidence of NTS infections is lower in industrialized than in developing countries, at about 45 per 100 000 inhabitants [1]. Invasive infections caused by NTS, such as meningitis and septicemia, can occur, particularly among young children, the elderly, malaria-infected and malnourished children, and immunocompromised people [2].

NTS can be host-generalists, capable of infecting or colonizing a broad range of vertebrate species, or host-specialists, adapted or restricted to particular non-human species [2]. In industrialized countries, NTS are transmitted to humans predominantly through the consumption of commercially produced food contaminated with livestock feces (e.g. meat, dairy products, poultry, and eggs) [2]. Outbreaks and sporadic cases of NTS have also been reported after direct or indirect contact with reptiles [3,4], as NTS are commonly found in the digestive tracts of reptiles (crocodiles, lizards, snakes, turtles) and amphibians (frogs, newts) [5–7]. Approximately 1.4 million human cases of *Salmonella* infection occur each year in the USA, of which about 74,000 are a result of exposure to reptiles and amphibians [8]. Within the European Union, it is estimated that less than 1% of cases of human salmonellosis are associated with exposure to reptiles. Reptile-related salmonellosis has been associated with young age, a high rate of hospitalization, and invasive disease [9,10]. Of the *Salmonella* serovars, 40% have been cultured predominantly from reptiles and are rarely found in other cold- and warm-blooded animals, suggesting that human infections with these serovars are of reptile origin.

Guadeloupe, a French overseas territory in the Caribbean, is a very high-resource country according to the Human Development Index in 2013. Although few data are available on the epidemiology of *Salmonella* in humans in the Caribbean, infections appear to be specific. In Guadeloupe, Panama and Arechavaleta were the most prevalent serovars recovered from 171 infants and children infected with *S. enterica* who were seen at the university hospital in Pointe-à-Pitre (Guadeloupe) between 2010 and 2014. The two serovars represented 50% of all *Salmonella* isolates in that study [11]. Surprisingly, they have been rarely encountered in mainland France or in other regions of the world [12]. In addition, these serovars are significantly associated with bacteremia ($P < 0.001$) [11]. Four cases of Panama meningitis were recently described in exclusively breastfed infants in French Guiana, suggesting a specific reservoir [13]. In Guadeloupe, wild reptiles and amphibians (e.g. anoles, geckos, iguanas and frogs) are commonly found in and around houses. We therefore conducted a study in Guadeloupe to identify the reservoir of Panama and Arechavaleta by comparing the distribution of *Salmonella* serovars in warm- and cold-blooded animals and in humans. A matched case-control study

was also conducted to determine the main epidemiologic risk factors among children with *S. enterica* infection.

Material and methods

S. enterica isolates from human samples

S. enterica clinical isolates were received for serotyping between January 2004 and December 2014 by the French national reference center for *Escherichia coli*, *Shigella* and *Salmonella* (FNRC-ESS) (Institut Pasteur, Paris, France) from public and private clinical laboratories in Guadeloupe and mainland France. If more than one isolate with the same serovar was recovered from the same patient, only the first was included. Epidemiologic data (date and site of isolation, age and gender of the patient, history of travel abroad) were recorded when available.

S. enterica isolates from warm-blooded animals

Salmonella spp. were isolated from samples collected at poultry farms (droppings, dust, eggs, and poultry meat) and pig and beef farms (feces, carcasses, and meat) in Guadeloupe during 2010–2014 for sanitary inspections. Serotyping was performed at the Institut Pasteur of Guadeloupe, at the FNRC-ESS and at the French Agency for Food, Environmental and Occupational Health and Safety (ANSES). Data were compiled from the different databases.

***S. enterica* isolates from wild cold-blooded animals.** Between June 2011 and December 2014, a single cloacal swab was taken from 322 endemic anoles of 3 species and 11 sub-species at 85 sampling sites distributed throughout Guadeloupe and nearby islands (Les Saintes, Marie-Galante, La Désirade, and Petite-Terre) (S1 Table). Feces from iguanas living in colonies were collected at 10 sampling sites: 45 from the endemic *Iguana delicatissima* and 24 from the invasive *I. iguana* (S1 Table). Fifty frogs and 35 geckos at 8 and 12 sampling sites, respectively, were trapped and placed in sterile vials (S1 Table). Fecal droppings were collected within 24 h of capture. After sampling, all frogs and lizards were released at the capture sites. All samples were placed at +4°C immediately after sampling and were processed within 4 h.

All procedures were approved by the regional environment, planning and housing agencies and by the Guadeloupe National Park. The project was also approved by the Committee for Ethics in animal experiments of the French West Indies and Guyana (reference 69-2012-4). The care and use of animals were performed accordingly with the French Decree No 2013–118 of 1 February 2013 on the protection of animals, which meets the European Union Directive 2010/63 on the protection of animals used for experimental and other scientific purposes.

Samples were incubated in 9 ml of buffered peptone water for 16–20 h at 37°C. Three drops (75 µl) of pre-enrichment broth were inoculated onto modified semi-solid Rappaport-Vassiliadis (MSRV) agar and incubated for a maximum of 48 h. Positive MSRV spots were streaked onto a specific medium, xylose–lysine deoxycholate (XLD) agar, and incubated for up to 48 h. Presumptive *Salmonella* colonies (H₂S positive) on XLD agar were isolated and identified on API 20E test strips (bioMérieux, Marcy L’Etoile, France). Serotyping was performed on five colonies from each sample.

Serotyping of *S. enterica* isolates

Isolates were serotyped on the basis of somatic O and both phase 1 and phase 2 flagellar antigens in agglutination tests with antisera (Bio-Rad, Marnes-La-Coquette, France) as specified in the White-Kauffmann-Le Minor scheme [14].

Epidemiological study

A case–control study was carried out in 2012–2013. Cases were patients admitted to the pediatric department of the university hospital of Pointe-à-Pitre with an *S. enterica* infection (acute gastroenteritis or bacteremia). At the inclusion of each case, a trained scientist administered a standardized questionnaire by telephone with the parents to collect demographic, environmental and lifestyle information on the *S. enterica* infection; the same person selected age-matched (± 5 years) children without *S. enterica* infection or carriage and administered the same questionnaire to the parents.

The study protocol was approved by the French Advisory Committee on Information Processing in Material Research in the Field of Health (CCTIRS 11–40). Written informed consent to participate in the study was obtained from the parents of all children included in the case–control study.

Data analysis

Statistical analyses were performed with R software [15]. Conditional tests were used because assumptions of traditional parametric tests were not met, with small samples and non-normal distributions. Thus, resampling procedures were implemented with the “coin” package, which does not assume random sampling from well-defined populations. Resampling provides especially clear advantages when assumptions of traditional parametric tests are not met. In a two-way contingency table, inference was based on 9999 Monte-Carlo resampling. Statistical differences were considered significant for two-sided P values < 0.05 .

Results

Salmonella species, subspecies and serovars in humans in Guadeloupe

A total of 710 *S. enterica* isolates were collected between 2005 and 2014. Four subspecies were recovered: *enterica* ($n = 669$, 94.2%), *houtenae* ($n = 39$, 5.5%), *salamae* ($n = 1$), and *diarizonae* ($n = 1$). Of the 68 serovars found, the most prevalent were Panama (24.5%, 174/710), Arechavaleta (11.5%, 82/710), Enteritidis (9.4%, 67/710), 4,[5],12:i:- (monophasic variant of Typhimurium) (9.4%, 67/710), and Newport (7.2%, 51/710) (Table 1). Panama was the most frequently isolated serovar every year, except in 2011 and 2014. Arechavaleta ranked in the top five isolated serovars each year, except in 2006 and 2010. During the study period, one isolate of serovar Paratyphi B and 13 isolates (1.8%) of Typhi were isolated (Table 1).

Salmonella isolates were recovered mainly from stool (82.1%, 583/710) and blood (12.7%, 90/710) samples. Of the blood isolates, 79 were NTS, of which 48 (60.7%) were Panama ($n = 26$, 32.9%) or Arechavaleta ($n = 22$, 27.8%) serovars.

Salmonella species, subspecies, and serovars in humans in mainland France

A total of 87 305 *Salmonella* isolates from humans were investigated between 2005 and 2014. The most prevalent serovars were Typhimurium (40.5%), Enteritidis (9.6%), and 4,[5],12:i:- (9.4%) (Table 1). Panama was recovered only rarely ($n = 349$, 0.4%).

For the 180 Panama isolates collected between 2010 and 2014, information on travel abroad was available for 61 (33.9%) patients: 54 (88.5%) had traveled in the French West Indies or in South or Central America, and 7 (5.6%) had no history of travel.

Salmonella species, subspecies, and serovars in livestock in Guadeloupe

During the period 2010–2014, 386 *S. enterica* isolates were recovered: 319 (82.6%) from poultry, 60 from pigs (15.5%), and 7 from cattle ($n = 7$) (Table 2). A total of 37 serovars were

Table 1. Distribution of the 10 most frequent Salmonella serovars in humans in Guadeloupe and mainland France between 2005 and 2014.

Rank	Guadeloupe											Mainland France
	2005 n = 73	2006 n = 39	2007 n = 48	2008 n = 32	2009 n = 79	2010 n = 121	2011 n = 73	2012 n = 85	2013 n = 101	2014 n = 59	2005–2014 n = 710	2005–2014 n = 87 305
1	Panama (25%)	Panama (23.1%)	Panama (33.3%)	Panama (34.4%)	Panama (20.2%)	Panama (24%)	4,[5],12:i:- (20.5%)	Panama (27%)	Panama (30.7%)	Enteritidis (22%)	Panama (24.5%)	Typhimurium (40.5%)
2	Enteritidis (14%)	43:z ₄ ,z ₃₂ : ^{-a} (12.8%)	Arechavaleta (18.7%)	Arechavaleta (25%)	4,[5],12:i:- (16.5%)	Newport (13.2%)	Newport (13.2%)	4,[5],12:i:- (15.3%)	Arechavaleta (12.9%)	Panama (22%)	Arechavaleta (11.5%)	Enteritidis (9.6%)
3	Newport (8.2%)	Enteritidis (10.3%)	43:z ₄ ,z ₃₂ : ^{-a} (12.5%)	43:z ₄ ,z ₃₂ : ^{-a} (6.2%)	Arechavaleta (10.1%)	4,[5],12:i:- (10.7%)	Panama (11%)	Arechavaleta (14.1%)	Enteritidis (9.9%)	Arechavaleta (20.3%)	Enteritidis (9.4%)	4,[5],12:i:- (9.4%)
4	Arechavaleta (5.5%)	Typhimurium (7.7%)	Enteritidis (8.3%)	4,[5],12:i:- (6.2%)	Infantis (7.6%)	Typhimurium (9.9%)	Arechavaleta (11%)	Enteritidis (9.4%)	Newport (8.9%)	4,[5],12:i:- (6.8%)	4,[5],12:i:- (9.4%)	Infantis (2.0%)
5	Typhimurium (4.1%)	Infantis (7.7%)	Typhimurium (6.2%)	Typhi (6.2%)	Enteritidis (6.3%)	Infantis (8.3%)	Infantis (5.5%)	43:z ₄ ,z ₃₂ : ^{-a} (9.4%)	4,[5],12:i:- (6.9%)	Newport (6.8%)	Newport (7.2%)	Typhi (1.7%)
6	Agona (2.7%)	Manhattan (5.1%)	Newport (6.2%)	48:g,z ₅₁ : ^{-a} (3.1%)	Indiana (6.3%)	Arechavaleta (6.6%)	Enteritidis (5.5%)	Rubislaw (5.9%)	Infantis (4.9%)	Rubislaw (6.8%)	Infantis (5.3%)	Virchow (1.6%)
7	Infantis (2.7%)	Mississippi (5.1%)	Infantis (2.1%)	Bredeney (3.1%)	Rubislaw (5.1%)	Enteritidis (6.6%)	Rubislaw (4.1%)	Infantis (3.5%)	Braenderup (4.9%)	Infantis (5%)	43:z ₄ ,z ₃₂ : ^{-a} (4.9%)	Newport (1.6%)
8	Kottbus (2.7%)	Rubislaw (5.1%)	Braenderup (2.1%)	Infantis (3.1%)	Typhi (5.1%)	Rubislaw (5.8%)	Typhi (4.1%)	Newport (3.5%)	Rubislaw (3.9%)	43:z ₄ ,z ₃₂ : ^{-a} (3.4%)	Rubislaw (4.4%)	Derby (1.6%)
9	Manhattan (2.7%)	50:g,z ₅₁ : ^{-a} (2.6%)	Derby (2.1%)	Manhattan (3.1%)	43:z ₄ ,z ₃₂ : ^{-a} (3.8%)	43:z ₄ ,z ₃₂ : ^{-a} (4.1%)	Wелtevreden (3.7%)	Oranienburg (3.5%)	Typhimurium (2%)	Javiana (1.7%)	Typhimurium (4%)	Kentucky (1.5%)
10	Rubislaw (2.7%)	6,8:e,h:- (2.6%)	Javiana (2.1%)	Oranienburg (3.1%)	Newport (2.5%)	Agona (1.6%)	Typhimurium (2.7%)	Aberdeen (1.2%)	Uganda (2%)	Miami (1.7%)	Typhi (1.8%)	Hadar (0.8%)

^a *S. enterica* subsp. *houtenae*

<https://doi.org/10.1371/journal.pone.0220145.t001>

found; Newport (20.7%, 80/386), 4,[5],12:i:- (10.1%, 39/386), and Havana (8.8%, 34/386) were the most frequent. Newport (25.1%, 80/319) predominated in poultry, whereas 4,[5],12:i:- predominated in pigs and cattle (31.3%, 21/67). Panama and Arechavaleta were recovered only from pigs and at lower frequency, 7.5% (n = 5) and 1.5% (n = 1), respectively.

Salmonella species, sub-species, and serovars in cold-blooded animals in Guadeloupe

The frequency of *S. enterica* carriage in cloacal specimens from the 426 wild reptiles investigated was 15% (n = 64) but varied by species, from 0 in geckos (0/35), 11.2% (36/322) in anoles, to 40.5% (28/69) in iguanas (Table 3). No isolates were collected from frogs (0/50).

All the isolates belonged to the *enterica* species. Two subspecies were recovered: *enterica* (n = 48, 73.8%) and *houtenae* (n = 17, 26.2%). Ten serovars were found, eight in anoles and five in iguanas. The three most prevalent were Schwarzengrund (30.8%, 20/65), 43:z₄,z₃₂:^{-a} (*houtenae* subspecies) (24.6%, 16/65), and Pomona (13.8%, 9/65) (Table 3). *S.* 43:z₄,z₃₂:^{-a} predominated in anoles (44.4%, 16/37), whereas Schwarzengrund predominated in iguanas (71.4%, 20/28). Panama and Arechavaleta were found in 4.6% (n = 3) and 3.1% (n = 2) of specimens, respectively (Table 2).

Serotyping was performed on five colonies from each reptile, but only one anole contained two different serovars: one belonging to Newport and one to 43:z₄,z₃₂:^{-a} (*houtenae* subspecies).

Significant associations between prevalent serovars and location were found only for Pomona and Schwarzengrund. Pomona was found exclusively in anoles sampled on two islands (Marie-Galante and La Désirade), and Schwarzengrund was isolated from endemic iguanas on two islands (Petite-Terre and La Désirade) (83.3%, 20/24) but not in invasive iguanas sampled on the main island.

Epidemiological study

A total of 75 children were enrolled in the study (50 cases and 25 controls); 45 were boys. The mean age at inclusion was 33.2 months, and the age distribution was: 19 (25.3%) aged 0–11

Table 2. Distribution of the 10 most frequent Salmonella serovars in cold- and warm-blooded animals in Guadeloupe.

Rank	Cold-blooded animals									Warm-blooded animals								
	Anolis	n	(%)	Iguanas	n	%	Total	n	%	Poultry	n	%	Pigs and cattle	n	%	Total	n	%
1	43:z ₄ .z ₃₂ : ^a	16	44.4	Schwarzengrund	20	71.4	Schwarzengrund	20	30.8	Newport	80	25.1	4,[5],12:i:-	21	31.3	Newport	80	20.7
2	Pomona	9	25	Infantis	4	14.3	43:z ₄ .z ₃₂ : ^a	16	24.6	Havana	34	10.7	London	21	31.3	4,[5],12:i:-	39	10.1
3	Rubislaw	4	11.1	Panama	2	7.1	Pomona	9	13.8	Indiana	33	10.3	Uganda	7	10.4	Havana	34	8.8
4	Newport	3	8.3	Newport	1	3.6	Infantis	5	7.7	Infantis	30	9.4	Derby	6	9	Indiana	33	8.5
5	Arechavelata	2	5.5	Lesabymes (67:d:1,7)	1	3.6	Newport	4	6.1	4,[5],12:i:-	18	5.6	Panama	5	7.5	Infantis	30	7.8
6	Infantis	1	2.7				Rubislaw	4	6.1	Albany	15	4.7	Agona	2	3	London	22	5.7
7	1,44:-:- ^a	1	2.7				Panama	3	4.6	Uganda	15	4.7	Albany	1	1.5	Uganda	22	5.7
8	Panama	1	2.7				Arechavelata	2	3.1	Cerro	12	3.8	Anatum	1	1.5	Albany	16	4.1
9							1.44:-:- ^a	1	1.5	Enteritidis	10	3.1	Arechavelata	1	1.5	Cerro	12	3.1
10							Lesabymes (67:d:1,7)	1	1.5	Typhimurium	8	2.5	Enteritidis	1	1.5	Enteritidis	11	2.8

^a *S. enterica* subsp. *houtenae*

<https://doi.org/10.1371/journal.pone.0220145.t002>

months, 22 (29.3%) aged 12–23 months, 20 (26.7%) aged 24–59 months, and 14 (18.7%) aged 60–126 months.

Demographics, environmental characteristics and lifestyle information are summarized in Table 3. No significant difference was found between cases and controls in environment or lifestyle factors, except for a more frequent presence of amphibians in the houses of cases than in those of controls ($P = 0.042$) (Table 3).

Discussion

The two most frequent *Salmonella* serovars in humans living in Guadeloupe were Panama and Arechavelata. Panama was also the most prevalent serovar recovered from humans in two French overseas territories, Martinique in the Caribbean (35% of all isolates investigated between 1990 and 1994) and French Guiana in South America (11.7% in 2011) [12,16]. Although few data are available on the epidemiology of *Salmonella* infections in humans in this region, the Panama serovar appears to be highly prevalent. It was also the most prevalent serovar in humans in Colombia and Chile [17,18]. The prevalence is higher than those in other regions of the world, including mainland France, where it was found only rarely (0.4% of all isolates investigated, most collected from patients with a history of travel to either the French West Indies or South or Central America). To the best of our knowledge, the only cases of human infection with Arechavelata have been reported in New Zealand [19], but at a lower frequency than in Guadeloupe, with nine cases reported between 1997 and 2016. As in mainland France and more generally in Europe and other parts of the world, the monophasic variant of the serovar Typhimurium (4,[5],12:i:-) has predominated in Guadeloupe since the mid-2000s [20,21]. Nevertheless, although Typhimurium remains the most frequent serovar in mainland France, 4,[5],12:i:- replaced its biphasic variant in Guadeloupe after 2008. Unsurprisingly, 4,[5],12:i:- was the second most frequent serovar isolated from livestock in Guadeloupe during the study period, supporting the role of pigs and poultry in its transmission to

Table 3. Risk factors for *Salmonella enterica* infection.

Risk factor	Cases (N = 50)	Controls (N = 25)	Univariate analysis		
			P	Crude OR	(95% CI)
Age months, mean (standard deviation)	30.7 (27.8)	39.4 (33.4)	0.301		
Male sex	31 (62.0)	14 (56.0)	0.627	1.3	(0.4–3.8)
Way of life					
Live in the countryside	35 (70.0)	14 (56.0)	0.304	1.8	(0.6–5.5)
Live in an individual house	36 (72.0)	17 (68.0)	0.790	1.2	(0.4–3.8)
Live in a house with a garden	29 (58.0)	15 (60.0)	1.000	0.9	(0.3–2.7)
Presence of reptiles					
In the garden	30 (60.0)	14 (56.0)	0.806	1.2	(0.4–3.5)
In the house	36 (72.0)	15 (60.0)	0.307	1.7	(0.5–5.3)
Presence of amphibians					
In the garden	26 (52.0)	10 (40.0)	0.462	1.6	(0.6–4.9)
In the house	23 (46.0)	5 (20.0)	0.042	3.4	(1.0–13.8)
Presence of pets					
Dogs	8 (16.0)	4 (16.0)	1.000	1.0	(0.2–5.1)
Cats	9 (18.0)	6 (24.0)	0.553	0.7	(0.2–2.7)
Systematic handwashing before meals	31 (62.0)	19 (76.0)	0.301	1.9	(0.6–7.0)
Consumption of garden vegetables or fruits	27 (54.0)	11 (44.0)	0.469	1.5	(0.5–4.4)

<https://doi.org/10.1371/journal.pone.0220145.t003>

humans [22]. The other major serovars found in humans are commonly associated with human infections, except for 43:z₄,z₃₂- (*houtenae* subspecies). To our knowledge, this serovar was recovered in one case of osteomyelitis in a Taylor’s cantil pit viper but has not been isolated in humans[23]. As these unusual serovars were not (43:z₄,z₃₂-) or rarely (Panama and Arechavaleta) found in the warm-blooded animals sampled, we investigated a reptilian source of contamination in order to identify their reservoir.

The overall frequency of *S. enterica* carriage in the reptiles studied in Guadeloupe was 15%, which is in the lower range of the reported values (13–57%). *Salmonella* carriage rates differed by species in our study: no isolates were recovered from geckos, in agreement with the low prevalence reported in most studies [24,25]. The rate in anoles was 11.4%, between the two reported values (0 and 33%) [5,26], and *Salmonella* was found in 40.5% of iguanas tested, also in the middle of the range of reported values (26–98%) [12,27–29]. Several factors might explain the variation among studies in the recovery rates in reptiles. The frequency of carriage might differ by species, and each group is composed of several species. Carriage frequency also differed by habitat. Crowding of reptiles favors the transmission of NTS, as seen in the high frequency of Schwarzengrund in iguanas from the small islands Petite Terre (1.68 km²) and La Desirade (21.42 km²). In addition, reptiles are intermittent *Salmonella* shedders [30,31]. In most studies, including ours, specimens were taken only once, which results in underestimates of the rate of *Salmonella* carriage. Lack of a standard method for *Salmonella* isolation and differences in the sampling technique (cloacal swabbing of protected animals versus fresh fecal samples from sacrificed animals in other studies) are possible explanations [32].

The case–control study of the main epidemiologic risk factors of *Salmonella* infection showed, despite the small sample, that the presence of frogs in homes was significantly associated with [24,25] *Salmonella* infection. However, we were unable to isolate *Salmonella* from 50 frogs, perhaps because of selection bias, as we could not sample frogs from the houses of cases. Furthermore, an observer bias is possible since frogs are more easily observed in houses than anoles or geckos. Previous studies reported very low rates of carriage in frogs [24,33].

In contrast to the rates observed in other studies (< 50%), in our study most of the isolates recovered from reptiles were assigned to subspecies *enterica* (69%), which is commonly isolated from warm-blooded animals. Although the subspecies *salamae*, *arizonae*, *diarizonae* and *houtanae* are known to be harbored by reptiles, we recovered only *houtanae*.

The subspecies *houtanae* serovar 43:z₄,z₃₂- was prevalent in reptiles and humans in Guadeloupe but was not found in livestock, suggesting a strict reptilian origin of human infections caused by this uncommon serovar. The presence of Arechavaleta and Panama in anoles and iguanas in Guadeloupe also indicates a reptilian reservoir for both serovars. This is not surprising, as Panama has previously been isolated from frogs, toads, turtles, lizards, and snakes [12,34,35] and Arechavaleta from cane toads [36]. However, their host range is certainly much larger, as shown by its presence in pigs in Guadeloupe. Both serovars were also found in previous studies in warm-blooded animals; Panama was found in wild birds, pigs, poultry, and Indian mongooses [37–41] and Arechavaleta in dogs and Indian mongooses [36,39,42]. In Guadeloupe, wild reptiles and amphibians (e.g. anoles, geckos, iguanas and frogs) are commonly found in and around houses. Guadeloupe is also a highly anthropized island, suggesting that close promiscuity between warm- and cold-blooded animals is at the origin of inter-species transmission. Therefore, we hypothesize that Panama and Arechavaleta are transmitted to humans either by direct contact with animals, in particular reptiles, or indirectly, through the consumption of food contaminated with livestock or reptile feces.

The low prevalence of serovars Panama and Arechavaleta in cold- and warm-blooded animals contrasts with that observed in human *Salmonella* infections. Evolutionary models suggest that host-adapted *Salmonella* serovars, such as serovars Typhi and Paratyphi A in humans, tend to be of high virulence, causing higher mortality rates than those with a broad host range, such as serovars Typhimurium and Enteritidis [43]. These host-adapted *Salmonella* serovars can cause illness in all age groups, whereas those with a broad host range tend to be more frequently associated with disease in young animals than in adults, suggesting that they are not optimally adapted to cope with a fully mature immune system [43]. Finally, chronic carriage, which develops more frequently following systemic infections by that host-adapted *Salmonella* serovars increases transmissibility [43]. Serovars Panama and Arechavalata are lesser virulent than serovars Typhi and Paratyphi A, as illustrated by the fact that they cause lesser mortality rates (0 to 13% versus 12 to 32%) and that they cause disease primarily opportunistically [11,43–46]. The number of bacteremia cases associated with serovars Panama and Arechavalata were higher in infants and children than in adults during a 5 year-survey (January 2010 to December 2014) among all patients with *Salmonella* infection admitted to the emergency room at the University Hospital in Pointe-à-Pitre (31 cases versus one in a 87-year-old woman). In addition, no asymptomatic carriers or secondary cases were identified (unpublished data). Reptile-related salmonellosis is also known to lead to invasive disease in young age [9,10]. All these elements suggest that serovars Panama and Arechavaleta are probably poorly adapted to humans.

In conclusion, the data reported here add to understanding of the epidemiology of *Salmonella* in Guadeloupe and, by extension, in the Caribbean. Panama and Arechavaleta were the two serovars most often recovered in humans. Their presence in wild reptiles and pigs suggests a broad host range and that human infections may result from indirect or direct contact with animals.

Supporting information

S1 Table. Details of GPS coordinates of sampling locations of cold-blooded animals, number of positive animals and isolated serovars.

(XLSX)

Acknowledgments

We thank all the students and the technicians involved in this work at the Institut Pasteur of Guadeloupe, the University Hospital of Pointe-à-Pitre and the FNRC-ESS. We thank Nadia Babel for collecting data on livestock production.

Author Contributions

Conceptualization: Stéphanie Guyomard-Rabenirina, François-Xavier Weill, Simon Le Hello, Franck Berger, Edith Malpote, Antoine Talarmin, Sébastien Breurec.

Formal analysis: Stéphanie Guyomard-Rabenirina, Séverine Ferdinand, Vincent Richard, Antoine Talarmin, Sébastien Breurec.

Investigation: Stéphanie Guyomard-Rabenirina, Sylvaine Bastian, Pierre Legreneur, Cécile Loraux, Blandine Muanza, Antoine Talarmin, Sébastien Breurec.

Methodology: Stéphanie Guyomard-Rabenirina, François-Xavier Weill, Simon Le Hello, Franck Berger, Antoine Talarmin, Sébastien Breurec.

Project administration: Stéphanie Guyomard-Rabenirina, Franck Berger, Antoine Talarmin, Sébastien Breurec.

Supervision: Stéphanie Guyomard-Rabenirina.

Validation: Stéphanie Guyomard-Rabenirina, François-Xavier Weill, Simon Le Hello, Franck Berger, Séverine Ferdinand, Vincent Richard.

Writing – original draft: Stéphanie Guyomard-Rabenirina, Antoine Talarmin, Sébastien Breurec.

Writing – review & editing: François-Xavier Weill, Simon Le Hello, Sylvaine Bastian, Franck Berger, Séverine Ferdinand, Pierre Legreneur, Cécile Loraux, Edith Malpote, Blandine Muanza, Vincent Richard.

References

1. Mead PS, Slutsker L, Dietz V, McCaig LF, Bresee JS, Shapiro C, et al. Food-related illness and death in the United States. *Emerg Infect Dis.* 1999; 5: 607–625. <https://doi.org/10.3201/eid0505.990502> PMID: 10511517
2. Crump JA, Sjölund-Karlsson M, Gordon MA, Parry CM. Epidemiology, clinical presentation, laboratory diagnosis, antimicrobial resistance, and antimicrobial management of invasive *Salmonella* infections. *Clin Microbiol Rev.* 2015; 28: 901–937. <https://doi.org/10.1128/CMR.00002-15> PMID: 26180063
3. Mermin J, Hutwagner L, Vugia D, Shallow S, Daily P, Bender J, et al. Reptiles, amphibians, and human *Salmonella* infection: a population-based, case-control study. *Clin Infect Dis.* 2004; 38 Suppl 3: S253–61. <https://doi.org/10.1086/381594> PMID: 15095197
4. Team E, Bertrand S, Rimhanen-Finne R, Weill F-X, Rabsch W, Thornton L, et al. *Salmonella* infections associated with reptiles: The current situation in Europe. *Eurosurveillance.* 2008; 13: 11–16.
5. Kourany M, Myers CW, Schneider CR. Panamanian amphibians and reptiles as carriers of *Salmonella*. *Am J Trop Med Hyg.* 1970; 19: 632–638. <https://doi.org/10.4269/ajtmh.1970.19.632> PMID: 5425502
6. van der Walt ML, Huchzermeyer FW, Steyn HC. *Salmonella* isolated from crocodiles and other reptiles during the period 1985–1994 in South Africa. *Onderstepoort J Vet Res. South Africa;* 1997; 64: 277–283. PMID: 9551479
7. Clancy MM, Davis M, Valitutto MT, Nelson K, Sykes JM 4th. *Salmonella* infection and carriage in reptiles in a zoological collection. *J Am Vet Med Assoc. United States;* 2016; 248: 1050–1059. <https://doi.org/10.2460/javma.248.9.1050> PMID: 27074614
8. Hedberg C. Food-related illness and death in the United States. *Emerg Infect Dis.* 1999; 5: 840–842. <https://doi.org/10.3201/eid0506.990624> PMID: 10603229

9. Sauteur PMM, Relly C, Hug M, Wittenbrink MM, Berger C. Risk Factors for Invasive Reptile-Associated Salmonellosis in Children. *Vector-Borne Zoonotic Dis.* 2013; 13: 419–421. <https://doi.org/10.1089/vbz.2012.1133> PMID: 23473215
10. Murphy D, Oshin F. Reptile-associated salmonellosis in children aged under 5 years in South West England. *Arch Dis Child. England;* 2015; 100: 364–365. <https://doi.org/10.1136/archdischild-2014-306134> PMID: 25538189
11. Guyomard-Rabenirina S, Muanza B, Bastian S, Malpote E, Jestin P, Guerin M, et al. *Salmonella enterica* serovars Panama and Arechavaleta: Risk Factors for invasive non-typhoidal salmonella disease in Guadeloupe, French West Indies. *Am J Trop Med Hyg. The American Journal of Tropical Medicine and Hygiene;* 2018; 99: 584–589. <https://doi.org/10.4269/ajtmh.18-0192> PMID: 30014811
12. Gay N, Le Hello S, Weill FX, de Thoisy B, Berger F. *Salmonella* serotypes in reptiles and humans, French Guiana. *Vet Microbiol.* 2014; 170: 167–171. <https://doi.org/10.1016/j.vetmic.2014.01.024> PMID: 24560590
13. Elenga N, Cuadro E, Long L, Njuieyon F, Martin E, Kom-Tchameni R, et al. *Salmonella enterica* serovar Panama meningitis in exclusive breastfeeding infants: Report of 4 cases, clinical features and therapeutic challenges. *Medicine (Baltimore).* 2017; 96: e6665. <https://doi.org/10.1097/MD.0000000000006665> PMID: 28489741
14. Grimont PP, Weill FF-X. Antigenic formulae of the *Salmonella* serovars. *WHO Collab Cent Ref Res Salmonella.* 2007; 9: 1–166. <https://doi.org/10.1038/nature12153>
15. R Development Core Team. R: A Language and Environment for Statistical Computing [Internet]. Vienna, Austria: R Foundation for Statistical Computing; 2010. Available: <http://www.r-project.org/>
16. Olive C, Mansuy JM, Desbois N, Roche B, Cecile W, Saint-Aime C, et al. *Salmonella* Panama en Martinique: Aspects épidémiologiques et cliniques chez l'enfant hospitalisé. *Med Mal Infect.* 1996; 26: 590–593. [https://doi.org/10.1016/S0399-077X\(96\)80078-3](https://doi.org/10.1016/S0399-077X(96)80078-3)
17. Cordano AM, Virgilio R. Evolution of drug resistance in *Salmonella* Panama isolates in Chile. *Antimicrob Agents Chemother.* 1996; 40: 336–341. PMID: 8834876
18. Rodríguez EC, Díaz-Guevara P, Moreno J, Bautista A, Montano L, Realpe ME, et al. Laboratory surveillance of *Salmonella enterica* from human clinical cases in Colombia 2005–2011. *Enferm Infecc Microbiol Clin. Spain;* 2017; 35: 417–425. <https://doi.org/10.1016/j.eimc.2016.02.023> PMID: 27038678
19. The Institute of Environmental Science and Research Ltd (ESR). Human *Salmonella* Isolates, 2016 [Internet]. 2016. <https://doi.org/10.14121/j.cnki.1008-3855.2016.01.011>
20. Switt AI, Soyer Y, Warnick LD, Wiedmann M. Emergence, distribution, and molecular and phenotypic characteristics of *Salmonella enterica* serotype 4,5,12:i. *Foodborne Pathog Dis.* 2009; 6: 407–415. <https://doi.org/10.1089/fpd.2008.0213> PMID: 19292687
21. Arai N, Sekizuka T, Tamamura Y, Tanaka K, Barco L, Izumiya H, et al. Phylogenetic characterization of *Salmonella enterica* serovar Typhimurium and its monophasic variant isolated from food animals in Japan revealed replacement of major epidemic clones in the last 4 decades. *J Clin Microbiol.* 2018; 56: 1–14. <https://doi.org/10.1128/JCM.01758-17> PMID: 29491013
22. Bonardi S. *Salmonella* in the pork production chain and its impact on human health in the European Union. *Epidemiol Infect. Cambridge University Press;* 2017; 145: 1513–1526. <https://doi.org/10.1017/S095026881700036X> PMID: 28241896
23. Clancy MM, Newton AL, Sykes JCVP. Management of osteomyelitis caused by *Salmonella enterica* subsp. *houtenae* in a Taylor's Cantil (*Agkistrodon bilineatus taylori*) Using Amikacin Delivered Via Osmotic Pump. *J Zoo Wildl Med.* 2016; 47: 691–694. <https://doi.org/10.1638/2015-0207.1> PMID: 27468053
24. Gorski L, Jay-Russell MT, Liang AS, Walker S, Bengson Y, Govoni J, et al. Diversity of pulsed-field gel electrophoresis pulsotypes, serovars, and antibiotic resistance among *Salmonella* isolates from wild amphibians and reptiles in the California Central Coast. *Foodborne Pathog Dis.* 2013; 10: 540–548. <https://doi.org/10.1089/fpd.2012.1372> PMID: 23577627
25. Jiménez RR, Barquero-Calvo E, Abarca JG, Porras LP. *Salmonella* isolates in the introduced Asian house gecko (*Hemidactylus frenatus*) with emphasis on *Salmonella* Weltevreden, in two Regions in Costa Rica. *Vector borne zoonotic Dis.* 2015; 15: 550–5. <https://doi.org/10.1089/vbz.2015.1785> PMID: 26378974
26. Sumiyama D, Izumiya H, Kanazawa T, Murata K. *Salmonella* infection in Green Anoles (*Anolis carolinensis*), an invasive alien species on Chichi Island of the Ogasawara Archipelago in Japan. *J Vet Med Sci.* 2014; 76: 461–465. <https://doi.org/10.1292/jvms.13-0217> PMID: 24270852
27. Franco A, Hendriksen RS, Lorenzetti S, Onorati R, Gentile G, Dell'Omo G, et al. Characterization of *Salmonella* occurring at high prevalence in a population of the land *Iguana conolophus subcristatus* in Galápagos Islands, Ecuador. *PLoS One.* 2011; 6: 1–5. <https://doi.org/10.1371/journal.pone.0023147> PMID: 21853080

28. Lankau EW, Cruz Bedon L, Mackie RI. *Salmonella* strains isolated from Galápagos Iguanas show spatial structuring of serovar and genomic diversity. PLoS One. Public Library of Science; 2012; 7: e37302. <https://doi.org/10.1371/journal.pone.0037302> PMID: 22615968
29. Sylvester WRB, Amadi V, Pinckney R, Macpherson CNL, McKibben JS, Bruhl-Day R, et al. Prevalence, serovars and antimicrobial susceptibility of *Salmonella* spp. from wild and domestic green iguanas (*Iguana iguana*) in Grenada, West Indies. Zoonoses Public Health. Germany; 2014; 61: 436–441. <https://doi.org/10.1111/zph.12093> PMID: 24325463
30. Bauwens L, Vercammen F, Bertrand S, Collard JM, De Ceuster S. Isolation of *Salmonella* from environmental samples collected in the reptile department of Antwerp Zoo using different selective methods. J Appl Microbiol. 2006; 101: 284–289. <https://doi.org/10.1111/j.1365-2672.2006.02977.x> PMID: 16882135
31. Goupil BA, Trent AM, Bender J, Olsen KE, Morningstar BR, Wünschmann A. A longitudinal study of *Salmonella* from snakes used in a public outreach program. J Zoo Wildl Med. 2012; 43: 836–841. <https://doi.org/10.1638/2011-0281R1.1> PMID: 23272351
32. Pasmans F, Martel A, Boyen F, Vandekerchove D, Wybo I, Van Immerseel F, et al. Characterization of *Salmonella* isolates from captive lizards. Vet Microbiol. 2005; 110: 285–291. <https://doi.org/10.1016/j.vetmic.2005.07.008> PMID: 16153787
33. Parsons SK, Bull CM, Gordon DM. Low prevalence of *Salmonella enterica* in Australian wildlife. Environ Microbiol Rep. 2010; 2: 657–659. <https://doi.org/10.1111/j.1758-2229.2010.00152.x> PMID: 23766252
34. Nakadai A, Kuroki T, Kato Y, Suzuki R, Yamai S, Yaginuma C, et al. Prevalence of *Salmonella* spp. in pet reptiles in Japan. J Vet Med Sci. 2005; 67: 97–101. <https://doi.org/10.1292/jvms.67.97> PMID: 15699603
35. Ribas A, Poonlaphdecha S. Wild-caught and farm-reared amphibians are important reservoirs of *Salmonella*, a study in North-East Thailand. Zoonoses Public Health. 2017; 64: 106–110. <https://doi.org/10.1111/zph.12286> PMID: 27359101
36. Drake M, Amadi V, Zieger U, Johnson R, Hariharan H. Prevalence of *Salmonella* spp. in cane toads (*Bufo marinus*) from Grenada, West Indies, and their antimicrobial susceptibility. Zoonoses Public Health. 2013; 60: 437–441. <https://doi.org/10.1111/zph.12018> PMID: 23035820
37. Betancor L, Pereira M, Martinez A, Giossa G, Fookes M, Flores K, et al. Prevalence of *Salmonella enterica* in poultry and eggs in Uruguay during an epidemic due to *Salmonella enterica* serovar Enteritidis. J Clin Microbiol. 2010; 48: 2413–2423. <https://doi.org/10.1128/JCM.02137-09> PMID: 20484605
38. Kich JD, Coldebella A, Mores N, Nogueira MG, Cardoso M, Fraticcio PM, et al. Prevalence, distribution, and molecular characterization of *Salmonella* recovered from swine finishing herds and a slaughter facility in Santa Catarina, Brazil. Int J Food Microbiol. Netherlands; 2011; 151: 307–313. <https://doi.org/10.1016/j.ijfoodmicro.2011.09.024> PMID: 22024043
39. Miller S, Amadi V, Stone D, Johnson R, Hariharan H, Zieger U. Prevalence and antimicrobial susceptibility of *Salmonella* spp. in small Indian mongooses (*Herpestes auropunctatus*) in Grenada, West Indies. Comp Immunol Microbiol Infect Dis. Elsevier Ltd; 2014; 37: 205–210. <https://doi.org/10.1016/j.cimid.2014.05.003> PMID: 24906835
40. Tamang MD, Gurung M, Nam H-M, Moon DC, Kim S-R, Jang G-C, et al. Prevalence and characterization of *Salmonella* in pigs from conventional and organic farms and first report of S. serovar 1,4,[5],12:i:- from Korea. Vet Microbiol. Netherlands; 2015; 178: 119–124. <https://doi.org/10.1016/j.vetmic.2015.05.005> PMID: 25982261
41. Matias CAR, Pereira IA, Araújo M dos S de, Santos AFM, Lopes RP, Christakis S, et al. Characteristics of *Salmonella* spp. isolated from wild birds confiscated in illegal trade markets, Rio de Janeiro, Brazil. Biomed Res Int. 2015; 2016: 1–7. Available: <https://www.hindawi.com/journals/bmri/2016/3416864/0Afile:///C:/Users/Usuario/Downloads/3416864.pdf>
42. Seepersadsingh N, Adesiyun AA, Seebaransingh R. Prevalence and antimicrobial resistance of *Salmonella* spp. in non-diarrhoeic dogs in Trinidad. J Vet Med B Infect Dis Vet Public Health. 2004; 51: 337–42. <https://doi.org/10.1111/j.1439-0450.2004.00785.x> PMID: 15525361
43. Baumlér AJ, Tsolis EM, Ficht TA, Adams LG. Evolution of Host Adaptation in *Salmonella enterica*. Infect Immun. 1998; 66: 4579–4587. PMID: 9746553
44. Leeder FS. An epidemic of *Salmonella* Panama infections in infants. Ann N Y Acad Sci. 1944; 66: 54–60.
45. Yang Y-J, Huang M-C, Wang S-M, Wu J-J, Cheng C-P, Liu C-C. Analysis of risk factors for bacteremia in children with nontyphoidal *Salmonella* gastroenteritis. Eur J Clin Microbiol Infect Dis. 2002; 21: 290–293. <https://doi.org/10.1007/s10096-002-0715-3> PMID: 12072940
46. Wilkins EG, Roberts C. Extraintestinal salmonellosis. Epidemiol Infect. 1988; 100: 361–8. Available: <http://www.ncbi.nlm.nih.gov/pubmed/3378582%0Ahttp://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2249349> <https://doi.org/10.1017/s095026880006711x> PMID: 3378582