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1 **Distribution of *Kingella kingae* capsular serotypes in France assessed by a multiplex**
2 **PCR on osteoarticular samples**

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18 **Key words:** *Kingella kingae*, capsule, typing, epidemiology, pathophysiology

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21 *Kingella kingae* is the leading pathogen of osteoarticular infection (OAI) in <4-year-
22 old children in different countries where improved culture methods and nucleic acid
23 amplification assays are routinely employed (1). The oropharynx is recognized as the portal of
24 entry for *K. kingae*, which can penetrate the bloodstream and invade distant organs, facilitated
25 by several virulence factors such as a recently described polysaccharide capsule (2). On an
26 international collection of 150 invasive and carriage isolates from 10 countries, Porsch et al.
27 (2) have described four capsule types using a multiplex polymerase chain reaction (PCR)
28 approach. Over 95% of invasive isolates in the collection were type 'a' or type 'b', while
29 capsule type 'c' and type 'd' were more commonly observed among carriage isolates (2, 3).
30 This distribution was described on *K. kingae* isolates; however *K. kingae* is a fastidious
31 bacterium and whether the strains harboring a capsule type 'a' or 'b' can be more easily
32 cultivated from clinical samples is unknown. Such difference may lead to biased
33 epidemiological results. Moreover, among this collection, 30 invasive isolates were from
34 France, mainly isolated from 2010 to 2013 (n=24/30, range 1972-2013). To our knowledge,
35 no more recent data are available yet, and whether the epidemiology has changed remains to
36 be determined.

37 We aimed to describe the *K. kingae* capsule serotypes using a multiplex PCR protocol, as
38 recently described (4), on a fraction of our collection of osteoarticular samples, which were
39 positive with the *K. kingae* specific real-time PCR, used as routine in our laboratory (5).

40 From July 2013 to April 2018, we found 115 *K. kingae*-positive osteoarticular samples from
41 105 patients (1 to 3 per patients). Samples origins were: joint fluid (95/105; 90.5%), synovial
42 biopsy (8/105; 7.6%), bone abscess (1/105; 0.9%), and sub-cutaneous collection (1/105;
43 0.9%). Samples origins were: joint fluid (95/105; 90.5%), synovial biopsy (8/105; 7.6%),
44 bone abscess (1/105; 0.9%), and sub-cutaneous collection (1/105; 0.9%). Capsular PCR
45 allowed identifying unambiguously a capsule type in all of these 115 samples (Figure 1).

46 Among the 105 patients, the PCR results showed 71 (67.6%) capsules 'a', 33 (31.4%)
47 capsules 'b', 1 (0.9%) capsule 'c', and no capsule 'd'. When multiple samples were available
48 for one patient, the results were identical for each sample. There was no significant variation
49 between distribution of capsules 'a' and 'b' along the study period or by age or whether the
50 culture was positive or negative (Table 1).

51 In a large collection of *K. kingae*-positive osteoarticular samples in France, we
52 demonstrated that our capsular PCR is able to identify the capsular serotypes of invasive *K.*
53 *kingae* in skeletal system specimens. Knowing that *rtxA/B* and *cpn60* PCRs cannot
54 discriminate *K. kingae* from *K. negevensis* and *Simonsiella muelleri*, respectively, it may be
55 suggested that adding the capsular PCR to the molecular diagnosis may increase its specificity
56 (4). We observed that 99% of *K. kingae* OAI involved a strain harboring a capsule type 'a' or
57 'b', which is similar to that observed in the Israeli and the international collections of invasive
58 isolates (2, 3). Our results strengthen the hypothesis that the type 'a' and type 'b' capsules
59 may have enhanced pathogenic properties. Polysaccharide capsules are a common target for
60 development of vaccine, such as pneumococcal vaccine; the implication of capsules 'a' and
61 'b' in *K. kingae* invasive infections could be of high interest for the development of a *K.*
62 *kingae* vaccine. On the other hand, very few data on the distribution of capsule serotypes
63 among carriage isolates are available yet. In a recent study it was described that types 'a' and
64 'b' were predominant in a small population of healthy children carrying *K. kingae* in France
65 (4). A better characterization of the distribution of *K. kingae* capsular serotypes among
66 healthy carriers would lead to better understand the role played by the capsule in the potential
67 of the organism to cause invasive infection.

68 **References**

- 69 1. Yagupsky P. 2015. *Kingella kingae*: carriage, transmission, and disease. Clin
70 Microbiol Rev 28:54-79.

- 71 2. Porsch EA, Starr KF, Yagupsky P, St Geme JW, 3rd. 2017. The Type a and Type b
72 Polysaccharide Capsules Predominate in an International Collection of Invasive
73 *Kingella kingae* Isolates. mSphere 2: e00060-17
- 74 3. Starr KF, Porsch EA, Seed PC, Heiss C, Naran R, Forsberg LS, Amit U, Yagupsky P,
75 Azadi P, St Geme JW, 3rd. 2016. *Kingella kingae* Expresses Four Structurally Distinct
76 Polysaccharide Capsules That Differ in Their Correlation with Invasive Disease. PLoS
77 Pathog 12:e1005944.
- 78 4. Basmaci R, Deschamps K, Levy C, Mathy V, Corrad F, Thollot F, Bechet S, Sobral
79 E, Bidet P, Cohen R, Bonacorsi S. 2018. Prevalence of *Kingella kingae* oropharyngeal
80 carriage and predominance of type a and type b polysaccharide capsules among
81 French young children. Clin Microbiol Infect in press.
- 82 5. Ilharreborde B, Bidet P, Lorrot M, Even J, Mariani-Kurkdjian P, Liguori S, Vitoux C,
83 Lefevre Y, Doit C, Fitoussi F, Pennecot G, Bingen E, Mazda K, Bonacorsi S. 2009.
84 New Real-Time PCR-Based Method for *Kingella kingae* DNA Detection: Application
85 to Samples Collected from 89 Children with Acute Arthritis. J Clin Microbiol
86 47:1837-1841.

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88 **Legend to Figure**

89 **Figure 1. Gel electrophoresis of the *Kingella kingae* multiplex capsular PCR in**
90 **osteoarticular samples.** The molecular weight is related to the capsule serotype: ‘a’, ‘b’, ‘c’
91 and ‘d’ types are identified from the highest to the lowest molecular weight, respectively (lane
92 4). Capsule type ‘a’ was identified in 4 samples (lanes 1, 6, 7 and 8), capsule type ‘b’ was
93 identified in 2 samples (lanes 3 and 5), and capsule type ‘c’ was identified in 1 sample (lane
94 2). Lane 9, sterile water; Lane 10, DNA ladder.

1 **Table 1. Distribution of capsule types among 105 patients with *K. kingae* osteoarticular**
 2 **infection**

	All (n=105)	Capsule 'a' (n=71)	Capsule 'b' (n=33)	Capsule 'c' (n=1)
Year				
2013	24	19 (79.2)	5 (20.8)	0
2014	13	8 (61.5)	5 (38.5)	0
2015	0	-	-	-
2016	19	10 (52.6)	9 (47.4)	0
2017	39	26 (66.7)	12 (30.8)	1 (2.6)
2018	10	8 (80.0)	2 (20.0)	0
Age				
<1 year	30	20 (66.7)	10 (33.3)	0
1 year	60	41 (68.3)	18 (30.0)	1 (1.7)
2 years	12	7 (58.3)	5 (41.7)	0
3 years	2	2 (100.0)	0	0
4 years	1	1 (100.0)	0	0
Culture				
Positive	11	7 (63.6)	4 (33.4)	0
Negative	94	64 (68.1)	29 (30.9)	1 (1.0)

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