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Monitoring of asparagine depletion and anti-L-asparaginase antibodies in adult acute lymphoblastic leukemia treated in the pediatric-inspired GRAALL-2005 trial

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In childhood acute lymphoblastic leukemia (ALL), monitoring of asparagine depletion, asparaginase activity or anti-asparaginase antibodies (Abs) is crucial to appreciate the efficacy of L-asparaginase therapy^{1,2}. Full asparagine depletion³ and high asparaginase activity⁴ are both associated with improved outcomes in both children and adult ALL populations.

In children, the presence of Abs against *Escherichia coli* L-asparaginase has an adverse effect on treatment outcome if a switch toward erwinase (i.e., L-asparaginase from *Erwinia chrysantemi*) is not performed⁵. In adult, the incidence and clinical impact of anti-asparaginase Abs remain to be explored.

In the pediatric-inspired GRAALL trials, adult patients with Philadelphia-negative ALL were exposed to L-asparaginase during induction, consolidation, and delayed intensification. In a pilot study, performed in five French GRAALL centers, asparagine depletion and anti-asparaginase Abs were prospectively investigated in consecutive patients from these trials to determine the incidence of L-asparaginase inactivation.

The GRAALL-2005 and the LL-03 protocols have been previously reported. This phase-III trial aimed to evaluate the impact of high-dose cyclophosphamide during induction and of rituximab in patients with CD20-positive

ALL⁶. The LL-03 study evaluated the safety and efficacy of an ALL-type intensive chemotherapy in adult patients with lymphoblastic lymphomas (LL)⁷. The GRAALL-2005 and LL-03 trials shared the same chemotherapy backbone. During induction, *E. coli* L-asparaginase was administered at 6000 IU/m²/d intravenous (IV) on D8, D10, D12, then stopped for 8 days to avoid increased toxicity during cyclophosphamide and daunorubicine infusion, and finally resumed on D20, D22, D24, D26, and D28. Patients who failed to reach complete remission (CR) after induction received an idarubicin and high-dose cytarabine-based salvage regimen. Patients in CR received a consolidation course of six 2 weeks blocks including *E. coli* L-asparaginase (10,000 IU/m²/infusion) infused on day 3 of blocks 1/4 and on day 16 of blocks 2/5⁶. According to baseline and response criteria, patients in persistent CR received either an allogeneic stem cell transplantation (HSCT) or a late intensification similar to the induction chemotherapy followed by maintenance therapy⁸. In case of allergic reaction, *E. coli* L-asparaginase was switched for erwinase (each dose of *E. coli* L-asparaginase was replaced by one dose of erwinase: 12,000 IU/m² during late intensification, 20,000 IU/m² during consolidation).

Asparagine level and anti-asparaginase Abs were assessed on blood samples at D8, D13, D20, and D29 of induction and late intensification, as well as at the onset of consolidation blocks 1, 2, 4, and 5. Asparagine level was evaluated, after rapid freezing, by reversed-phase liquid chromatographic/tandem mass spectrometric method (full depletion if <2 μmol/L). Anti-asparaginase Abs were

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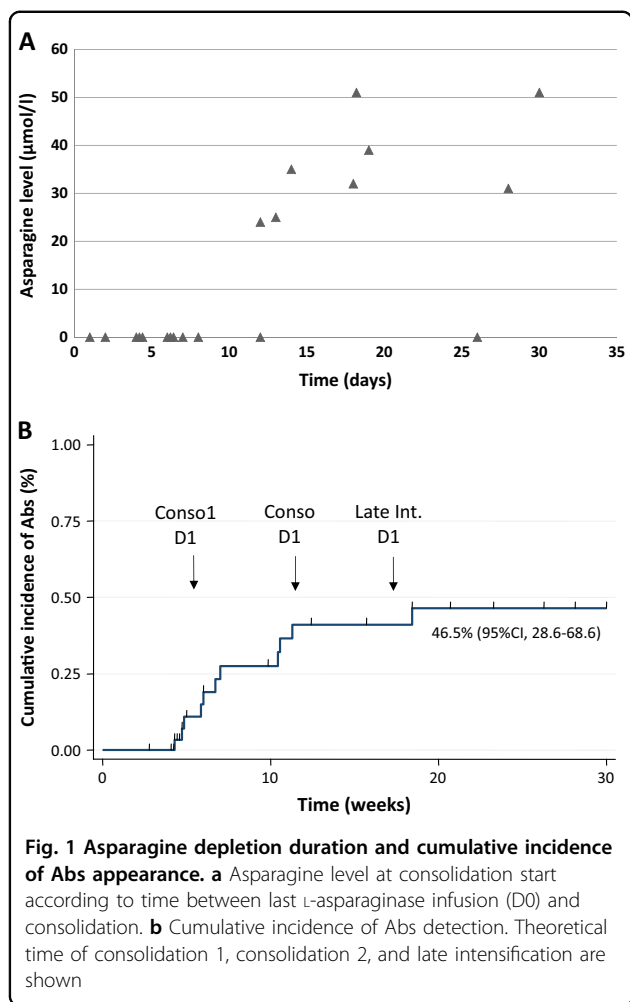
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encephalopathy. The cumulative incidence of Abs positivity, calculated considering relapse, death in first CR, and HSCT as competing events, was 46.5% (95% CI, 28.6–68.6) at 20 weeks (Fig. 1b). Unfortunately, the present pilot study was not powered and designed to address the question of the impact of Abs on relapse incidence. Indeed, only five relapses occurred, one after occurrence of Abs and four in patients without Abs.

We recently reported that patients with CD20-positive ALL who were randomized to receive rituximab had significantly less grade 3–4 hypersensitivity reactions to asparaginase (2/105 in the Rituximab group, 14/104 in the control group)⁹. In the present cohort, 5/31 patients received Rituximab. None of them had clinical signs of hypersensitivity and only 1/5 (20%) was detected with Abs. In contrast, 10/31 patients (32%) who were not exposed to Rituximab presented with hypersensitivity reaction and 13/31 (42%) developed Abs. Due to the small size of our cohort, none of these differences was statistically significant.

After numerous studies in childhood ALL, the present study is the first one to report asparaginase monitoring including anti-asparaginase Abs in adults. Despite the short half-life of L-asparaginase (8–30 h), we observed prolonged depletion in most patients. During induction, all patients who adequately received the L-asparaginase schedule were fully depleted in asparagine, especially at D20, 8 days after the previous asparaginase infusion, and up to 12 days after the last D28 infusion. Likewise, during consolidation phase, L-asparaginase depletion was observed up to 15 days after the previous infusion during consolidation blocks. This may be due in part to the sequential asparaginase infusions while measurement of half-life of L-asparaginase has been performed after one infusion only. However, a false depletion, due to the persistence of circulating asparaginase and post-sampling asparaginase activity cannot be ruled out despite careful freezing of samples. Monitoring asparaginase activity rather than asparagine depletion is currently recommended to appreciate the efficacy of asparaginase and will be evaluated prospectively in the GRAALL-2014 trial.

Due to the postponed sample analysis, physicians were not aware of Abs detection and/or absence of asparagine depletion during treatment. The cumulative incidence of Abs positivity at 20 weeks was 46.5%, in line with previous reports in children/adolescents^{10,11}. A strong correlation between the presence of Abs and clinical signs of hypersensitivity was observed and only 2/11 (18%) of patients with anti-asparaginase Abs had silent inactivation. However, such Abs are not always neutralizing and interpretation of their positivity is difficult in the absence of asparaginase activity monitoring. Although the study was not designed to address this question we should keep in mind that none of the patients receiving rituximab

experienced hypersensitivity reactions and only one had Abs positivity.

Despite the emergence of new therapies like inotuzumab ozogamicin and blinatumomab in B-cell ALL, asparaginase remain a major drug in the front-line treatment of ALL. The present study highlights the role of therapeutic drug monitoring in the management of adults with ALL, exposed early to L-asparaginase during chemotherapy. Further studies should explore the correlation between asparaginase residual activity and asparagine depletion to improve drug tolerance and efficacy. In the current GRAALL-2014 trial, a prospective monitoring of Abs is performed to switch from *E. coli* L-asparaginase to erwinase in case of clinical and/or silent allergy. The place of rituximab but also of other B-cell targeting therapies like inotuzumab ozogamicin or blinatumomab to decrease the risk of immunization should be further explored.

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Conflict of interest

EUSA Pharma then Jazz Pharmaceuticals provided research funding for asparagine dosage and anti-asparaginase Ab detection. M.H.-B. and N.B. received honoraria from Eusapharma. The remaining authors declare that they have no conflict of interest.

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