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Apolipoprotein E promotes subretinal mononuclear phagocyte survival and chronic inflammation in age-related macular degeneration

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Abstract

Physiologically, the retinal pigment epithelium (RPE) expresses immunosuppressive signals such as FAS ligand (FASL), which prevents the accumulation of leukocytes in the subretinal space. Age-related macular degeneration (AMD) is associated with a breakdown of the subretinal immunosuppressive environment and chronic accumulation of mononuclear phagocytes (MPs). We show that subretinal MPs in AMD patients accumulate on the RPE and express high levels of APOE. MPs of *Cx3cr1*^{-/-} mice that develop MP accumulation on the RPE, photoreceptor degeneration, and increased choroidal neovascularization similarly express high levels of APOE. *ApoE* deletion in *Cx3cr1*^{-/-} mice prevents pathogenic age- and stress-induced subretinal MP accumulation. We demonstrate that increased APOE levels induce IL-6 in MPs via the activation of the TLR2-CD14-dependent innate immunity receptor cluster. IL-6 in turn represses RPE FasL expression and prolongs subretinal MP survival. This mechanism may account, in part, for the MP accumulation observed in *Cx3cr1*^{-/-} mice. Our results underline the inflammatory role of APOE in sterile inflammation in the immunosuppressive subretinal space. They provide rationale for the implication of IL-6 in AMD and open avenues toward therapies inhibiting pathogenic chronic inflammation in late AMD.

Keywords age-related macular degeneration; apolipoprotein E; interleukin 6; mononuclear phagocyte; neuroinflammation

Subject Categories Immunology; Neuroscience

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Introduction

Age-related macular degeneration (AMD) is the leading cause of irreversible blindness in the industrialized world (Klein *et al*, 2004). Early and intermediate AMD is characterized by sizeable (> 125 μm) deposits of lipoproteinaceous debris, called large drusen, which are located in the Bruch's membrane (BM) and are partially covered by the retinal pigment epithelium (RPE) (Sarks, 1976). The presence of large drusen is an important risk factor for late AMD (Klein *et al*, 2004). There are two clinical forms of late AMD: wet AMD, which is defined by choroidal neovascularization (CNV), and geographic atrophy (GA), which is characterized by an extending lesion of both the retinal pigment epithelium (RPE) and photoreceptors (Sarks, 1976).

Mononuclear phagocytes (MP) comprise a family of cells that include microglial cells (MC), monocytes (Mo), and macrophages (Mφ) among others (Chow *et al*, 2011). Physiologically, MCs are present only in the inner retina. A number of neuron- and glial-derived factors (e.g., CX3CL1, CD200L) repress their activation (Galea *et al*, 2007). The subretinal space, located between the RPE and the photoreceptor outer segments (POS), is a zone of immune privilege mediated by immunosuppressive RPE signals, including leukocyte-suppressing FasL (CD95L) (Griffith *et al*, 1995; Streilein *et al*, 2002). In healthy subjects, the subretinal space is physiologically devoid of all leukocytes, including MPs, but in both advanced forms of AMD, wet AMD and geographic atrophy (GA), MPs accumulate in the subretinal space (Oh *et al*, 1999; Gupta *et al*, 2003; Combadiere *et al*, 2007; Sennlaub *et al*, 2013). They are in close contact with the RPE in CNV and in the vicinity of the RPE lesion in GA (Oh *et al*, 1999; Gupta *et al*, 2003; Combadiere *et al*, 2007;

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Sennlaub *et al*, 2013). It is generally admitted that MPs play an important role in CNV (Sakurai *et al*, 2003; Tsutsumi *et al*, 2003). Recent evidence strongly suggests that CCL2/CCR2-dependent subretinal MP accumulation is an important contributing factor in photoreceptor degeneration in models of photo-oxidative stress (Rutar *et al*, 2012; Suzuki *et al*, 2012), in the *Abca4*^{-/-}*Rdh8*^{-/-} mouse Stargardt/AMD model (Kohno *et al*, 2013), in the carboxy-ethylpyrrole immunization-induced AMD model (Cruz-Guilloty *et al*, 2013), and in the age- and light-induced photoreceptor degeneration of *Cx3cr1*^{-/-} mice (Combadiere *et al*, 2007; Sennlaub *et al*, 2013). The reasons for the breakdown of subretinal immune suppression and accumulation of MPs in AMD remain unknown.

Apolipoprotein E (APOE) plays a crucial role in lipid transport (Mahley & Rall, 2000). APOE is expressed in the liver (Linton *et al*, 1995), is a major component of high-density lipoproteins (HDL) of the blood (Mahley & Rall, 2000), and is the main lipoprotein of the brain and the retina (Mahley & Rall, 2000; Anderson *et al*, 2001). APOE is also strongly expressed in MCs (Nakai *et al*, 1996; Peri & Nusslein-Volhard, 2008) and Mφs (Basu *et al*, 1982; Rosenfeld *et al*, 1993) and promotes macrophage lipid efflux (Matsuura *et al*, 2006). In conjunction with APOA-I, APOE plays a crucial role in reverse cholesterol transport (Mahley & Rall, 2000). It also binds lipopolysaccharide (LPS) (Berbee *et al*, 2005) and amyloid-β (Zhao *et al*, 2009) and inhibits their activation of Toll-like receptors (TLR) and the induction of inflammatory mediators similar to APOA-I (Guo *et al*, 2004; Ali *et al*, 2005). TLR2 and TLR4 signaling necessitates their recruitment to the CD14-dependent innate immunity receptor cluster, located in cholesterol-rich membrane domains called lipid rafts (Schmitz & Orso, 2002). Interestingly, APOA-I and APOE can also extract membrane cholesterol from lipid rafts, activate the innate immunity receptor cluster in the absence of TLR ligands, and induce several inflammatory cytokines including IL-6 (shown for APOA-I) (Smoak *et al*, 2010). Recently, it has been shown that increased IL-6 levels are associated with AMD incidence (Klein *et al*, 2014) and with late AMD (Seddon *et al*, 2005; Klein *et al*, 2008). It is currently not clear how APOE and IL-6 participate in AMD pathogenesis.

In humans, APOE exists in three isoforms (APOE2, APOE3, and APOE4) arising from two cysteine–arginine interchanges at residues 112 and 158. Homozygous APOE2-allele carriers are at increased risk for developing late AMD (recently confirmed in 20,000 subjects (McKay *et al*, 2011)) and are protected against Alzheimer's disease (AD), while the APOE4 allele protects against AMD and is a risk factor for AD (Mahley & Rall, 2000).

The risk-conferring property of the APOE2 allele might be due to a difference in quantity or result from functional differences of the APOE2 protein structure. APOE concentrations in plasma and cerebro-spinal fluid (CSF) of APOE2 humans and transgenic humanized mice are significantly higher than in APOE3 carriers (Mooijaart *et al*, 2006; Riddell *et al*, 2008). This difference is due to APOE2's decreased ability to bind and be cleared via the LDL receptor (Mahley & Rall, 2000). Consistently increased APOE immunostaining in eyes with AMD would also suggest that APOE concentrations are locally increased in AMD (Klaver *et al*, 1998; Anderson *et al*, 2001).

On the other hand, *ApoE*^{-/-} mice that are fed a high-fat diet develop lipid accumulations in BM, which are proposed as being similar to early AMD (Ong *et al*, 2001). APOE4 transgenic mice on high-fat diet develop similar deposits (Malek *et al*, 2005) even

though the APOE concentration observed in APOE4 transgenic mice is only marginally decreased in their plasma (Riddell *et al*, 2008) and similar in the CSF (Riddell *et al*, 2008) and retina (Malek *et al*, 2005) compared to APOE3 mice. The structural changes in the APOE4 protein, however, lead to diminished association with HDL (Dong & Weisgraber, 1996) and impaired reverse cholesterol transport (Heeren *et al*, 2004). Low APOE concentrations or impaired reverse cholesterol transport could thereby hinder efficient lipid evacuation from the RPE to the choroid and lead to drusen development. (Mahley & Rall, 2000; Mooijaart *et al*, 2006; Riddell *et al*, 2008; Malek *et al*, 2005). This hypothesis is, however, in contradiction with the APOE accumulation observed in AMD donor eyes (Klaver *et al*, 1998; Anderson *et al*, 2001) and the protective effect of the APOE4 allele in AMD (McKay *et al*, 2011).

In the present article, we show that subretinal MPs in AMD and in subretinal inflammation observed in *Cx3cr1*^{-/-} mice strongly express APOE. We demonstrate that APOE prolongs subretinal MP survival and is necessary for subretinal MP accumulation in *Cx3cr1*^{-/-} mice. We demonstrate that increased APOE induces IL-6 in MPs, via the activation of the TLR2-CD14-dependent innate immune receptor cluster. IL-6 in turn represses RPE FasL expression, prolongs subretinal MP survival, and promotes chronic subretinal inflammation. Our data suggest that CD14 or IL-6 inhibition can help reestablish RPE immune-suppressive function and inhibit pathogenic inflammation in late AMD.

Results

Subretinal MPs accumulate on the RPE in the vicinity of atrophic lesions and large drusen

Physiologically, the subretinal space does not contain significant numbers of MPs, possibly due in part to immunosuppressive RPE signals (Streilein *et al*, 2002). In late AMD, immunohistochemical studies on sections revealed the presence of subretinal MPs on RPE cells adjacent to the lesions of atrophic AMD (Gupta *et al*, 2003; Sennlaub *et al*, 2013) and MPs are found in subretinal neovascular membranes (Oh *et al*, 1999). Because the small, dispersed MPs are difficult to detect on sections, we performed MP marker IBA-1 immunohistochemistry on healthy and diseased macular RPE/choroidal flatmounts (IBA-1 green fluorescence, RPE autofluorescence visible as orange due to its autofluorescence in the red and green channel). Confocal microscopy confirmed that subretinal IBA-1⁺ MPs are only very occasionally observed in healthy age-matched donor central RPE (Fig 1A). Within the atrophic lesions of GA patients where the RPE has disappeared, MPs were numerous, but were also invariably observed on the apical side of the RPE adjacent to the lesions (Fig 1B). Furthermore, IBA-1⁺ cells were detected on the RPE adjacent to large drusen (> 125 μm), visible under the dissecting microscope as pale lesions after removal of the retina (Fig 1C, inset) and as dome-shaped protrusions under the confocal microscope (Fig 1C, oblique projection of a Z-stack; Fig 1D, orthogonal Z-stack projection). Double-labeling on the subretinal side of the overlying retina (to avoid masking by RPE autofluorescence) shows that subretinal IBA-1⁺ MPs (Fig 1E green fluorescence) also express the pan-MP marker CD18 (Fig 1F red fluorescence, Fig 1G merge). IBA-1⁺ MPs in close contact with the RPE (Fig 1H, lateral

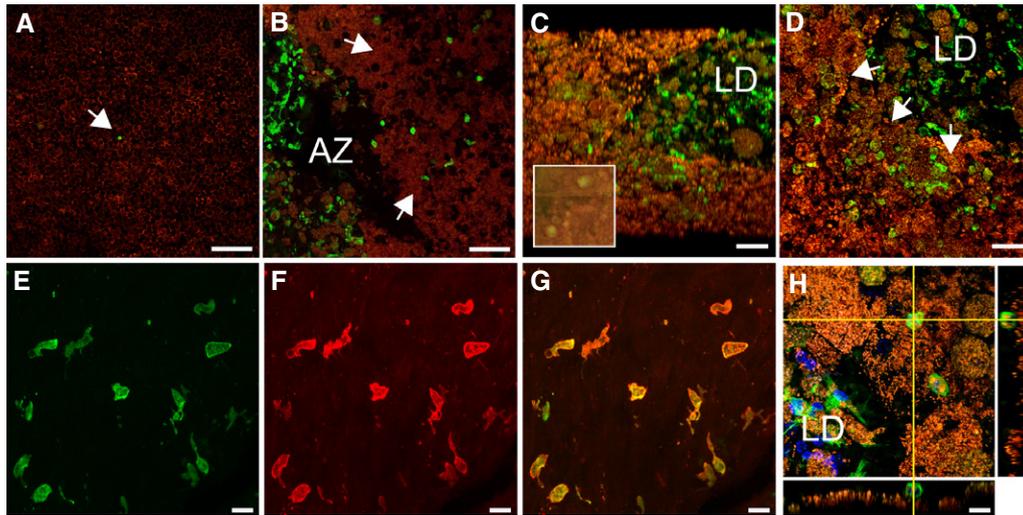


Figure 1. Subretinal MPs accumulate on the retinal pigment epithelium in AMD.

A–D Confocal microscopy of IBA-1 (green staining) immunohistochemistry of RPE flatmounts (RPE autofluorescence visible as orange due to its autofluorescence in the red and green channel) from a healthy donor (A), a geographic atrophy lesion (B), and large drusen (C and D). (A, B, D): orthogonal Z-stack projection; (C): oblique Z-stack projection and dissecting microscope appearance of postmortem large drusen after removal of the overlying retina (inset).

E–G Double-labeling on the subretinal side of the retina (to avoid masking by RPE autofluorescence) of IBA-1⁺ (E, green fluorescence) and CD18 (F, red fluorescence; G, merge).

H Orthogonal and lateral Z-stack of a subretinal IBA-1⁺ (green fluorescence) MPs adjacent to the RPE (orange autofluorescence) in the vicinity of a large drusen.

Data information: Controls omitting the primary antibody showed no staining apart from the autofluorescence. AZ: atrophic zone; LD: large drusen. Scale bar: 50 μ m (A–D); 10 μ m (E–H).

Z-stack projections) were observed in the vicinity of all examined large drusen and atrophic zones. Interestingly, we also detected IBA-1⁺ MPs within the large drusen, confirming our previous immunohistochemical detection of CCR2⁺ MPs on paraffin sections within large drusen (Sennlaub *et al*, 2013). HLA-DR and CD68-positive MP dendrites have previously been observed in smaller, dome-shaped drusen (Hageman *et al*, 2001). A detailed analysis of MPs within drusen of different sizes is ongoing in our laboratory, but beyond the scope of this study.

These observations considered together confirm the presence of subretinal MPs in AMD (Penfold *et al*, 1985; Gupta *et al*, 2003; Sennlaub *et al*, 2013) and illustrate their presence in contact with the RPE around large drusen and GA lesions. They are very rare in healthy donors. This further suggests that RPE-mediated immunosuppression is impaired in intermediate AMD (large drusen) and late AMD (GA).

Subretinal MPs express APOE

MPs have been reported to express APOE at high levels (Basu *et al*, 1982; Rosenfeld *et al*, 1993; Nakai *et al*, 1996; Peri & Nusslein-Volhard, 2008). Immunohistochemistry of APOE (Fig 2A, red) and IBA-1 (Fig 2B, green; Fig 2C, merge) on paraffin sections of human tonsils, which we used as a positive control, confirmed that IBA-1⁺ MPs can strongly express APOE. Similarly, on retinal flatmounts of donor eyes with large drusen, APOE (Fig 2D, red) staining was observed in and around subretinal IBA-1⁺ MPs (Fig 2E, green; Fig 2F, merge). The double-labeling was performed on the subretinal side of retinas to avoid masking by the RPE autofluorescence. We next performed APOE staining on paraffin sections of controls and

donor eyes with geographic atrophy lesions. We used a substrate revealing method (alkaline phosphatase/Fast Red) that is visible in bright field to circumvent confusion with RPE autofluorescence. In sections from control eyes, the APOE signal was concentrated at the basal portion of the RPE (Fig 2G red signal, arrows). In donor eyes with GA, adjacent to the atrophic area, a strong APOE signal was observed in the RPE, but it was less restricted to the basal aspect than in controls (Fig 2H APOE red signal, arrowheads). Additionally, APOE immunostaining was observed in cells adjacent to the RPE (Fig 2H APOE red signal, arrows). Double-labeling with IBA-1 identified these cells as subretinal IBA-1⁺ MPs (Fig 2I IBA-1 green signal, arrows). Omitting the APOE antibody and following the same experimental protocol did not produce any significant staining (Fig 2H inset).

Taken together, our results show that, in addition to the RPE, subretinal MPs in AMD patients strongly express APOE in a manner similar to other inflammatory settings (e.g., atherosclerotic lesions (Rosenfeld *et al*, 1993)).

APOE promotes subretinal MP accumulation in Cx3cr1^{GFP/GFP} mice

In the eye, CX3CL1 is constitutively expressed as a transmembrane protein in inner retinal neurons (Silverman *et al*, 2003; Zieger *et al*, 2014) and provides a tonic inhibitory signal to CX3CR1 bearing retinal MCs that keeps these cells in a quiescent surveillance mode under physiological conditions (Combadiere *et al*, 2007; Ransohoff, 2009). *Cx3cr1* deficiency in mice leads to a strong increase of subretinal MP accumulation with age, after light challenge or laser injury (Combadiere *et al*, 2007; Raoul *et al*, 2008; Ma *et al*, 2009), in

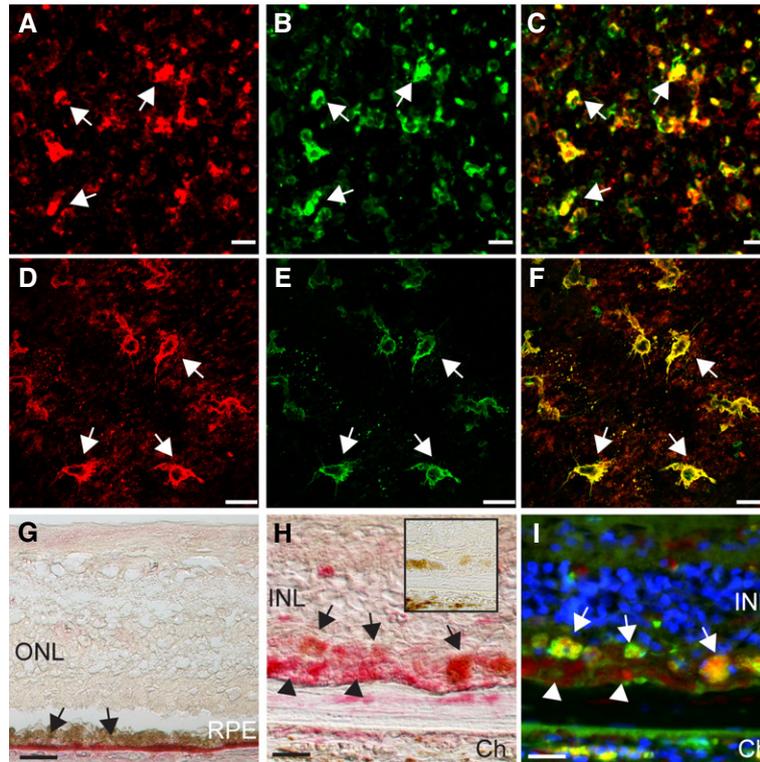


Figure 2. Subretinal MPs express APOE.

A–C APOE (A) and IBA-1 (B, merge in C) double-labeling on paraffin sections of surgical specimens of human tonsils.

D–F APOE (D) and IBA-1 (E, merge in F) double-labeling on the subretinal aspect of a large drusen overlaying retina (to avoid specific signal masking by the RPE orange autofluorescence).

G, H APOE (red staining) immunohistochemistry of control donor tissue (G) and adjacent to GA lesion (H).

I IBA-1 (green staining, RPE far-red autofluorescence) double-labeling of the GA lesion shown in (H). Representative images from five donor eyes with large drusen, five donor eyes with GA, and three healthy donors between 70 and 89 years old.

Data information: Controls omitting the primary antibody showed no staining apart from the autofluorescence (H inset). ONL: outer nuclear layer; INL: inner nuclear layer; Ch: choroid. Scale bar: 10 μm .

diabetes (Kezic *et al*, 2013) and in a paraquat-induced retinopathy model (Chen *et al*, 2013). *Cx3cr1*^{GFP/GFP} mice do not develop drusen and RPE atrophy, but do model MP accumulation on the RPE, as well as the associated photoreceptor degeneration and excessive CNV observed in AMD (Combadiere *et al*, 2007; Sennlaub *et al*, 2013).

We first evaluated APOE localization in 12-month-old *Cx3cr1*^{GFP/GFP} mice that present subretinal MP accumulation (Sennlaub *et al*, 2013). Immunohistochemical localization of APOE on retinal sections of both 12-month-old wild-type and *Cx3cr1*^{GFP/GFP} mice revealed APOE localization mainly in the RPE and inner retina as previously described (Anderson *et al*, 2001) (Supplementary Fig S1). Additionally, we detected a strong signal in cells apposed to the RPE on retinal sections and the subretinal side of retinal flatmounts in aged *Cx3cr1*^{GFP/GFP} mice (arrow, Fig 3A and B, red) that were identified as IBA-1-expressing MPs (Fig 3C, green), similar to AMD patients (Fig 1).

Subretinal MPs are derived from both Mos and MCs (Sennlaub *et al*, 2013). To evaluate whether *Cx3cr1*^{GFP/GFP} MPs differ in their *ApoE* expression, we first studied WT- and *Cx3cr1*^{GFP/GFP}-Mo (prepared from bone marrow) cultured for 24 h in contact with the photoreceptor outer segment (POS) of an overlaying retinal explant

to simulate MP differentiation in the subretinal space. *ApoE* mRNA was expressed at significantly higher levels in *Cx3cr1*^{GFP/GFP}-Mos in the presence of POS of an overlaying retinal explant (Fig 3D). Similarly, significantly increased amounts of *ApoE* mRNA were also observed in FACS-sorted MCs freshly extracted from adult *Cx3cr1*^{GFP/GFP} brain (Fig 3E). The expression of *ApoE* mRNA was also significantly higher in *Cx3cr1*^{GFP/GFP} peritoneal M ϕ s (prepared from thioglycollate-elicited peritonitis) when compared to WT-M ϕ s cultured for 24 h in the presence of CX3CL1 (Fig 3F). Western blot analysis of equivalent amounts of supernatant protein from peritoneal M ϕ s also showed increased APOE secretion (Fig 3G) in the *Cx3cr1*^{GFP/GFP} samples when compared to the soluble Mer receptor tyrosine kinase that is released constitutively from cultured macrophages (Sather *et al*, 2007) and which served here as a loading control.

To evaluate the role of APOE in subretinal MP accumulation, we analyzed *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice. Quantification of subretinal IBA-1⁺ MPs on retinal and RPE/choroidal flatmounts of 12-month-old *Cx3cr1*^{GFP/GFP} (Fig 3H) and *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice (Fig 2I) showed that the significant age-dependent subretinal MP accumulation observed in *Cx3cr1*^{GFP/GFP} mice was nearly completely inhibited in *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice (Fig 3J). Similarly, *Cx3cr1*^{GFP/GFP}

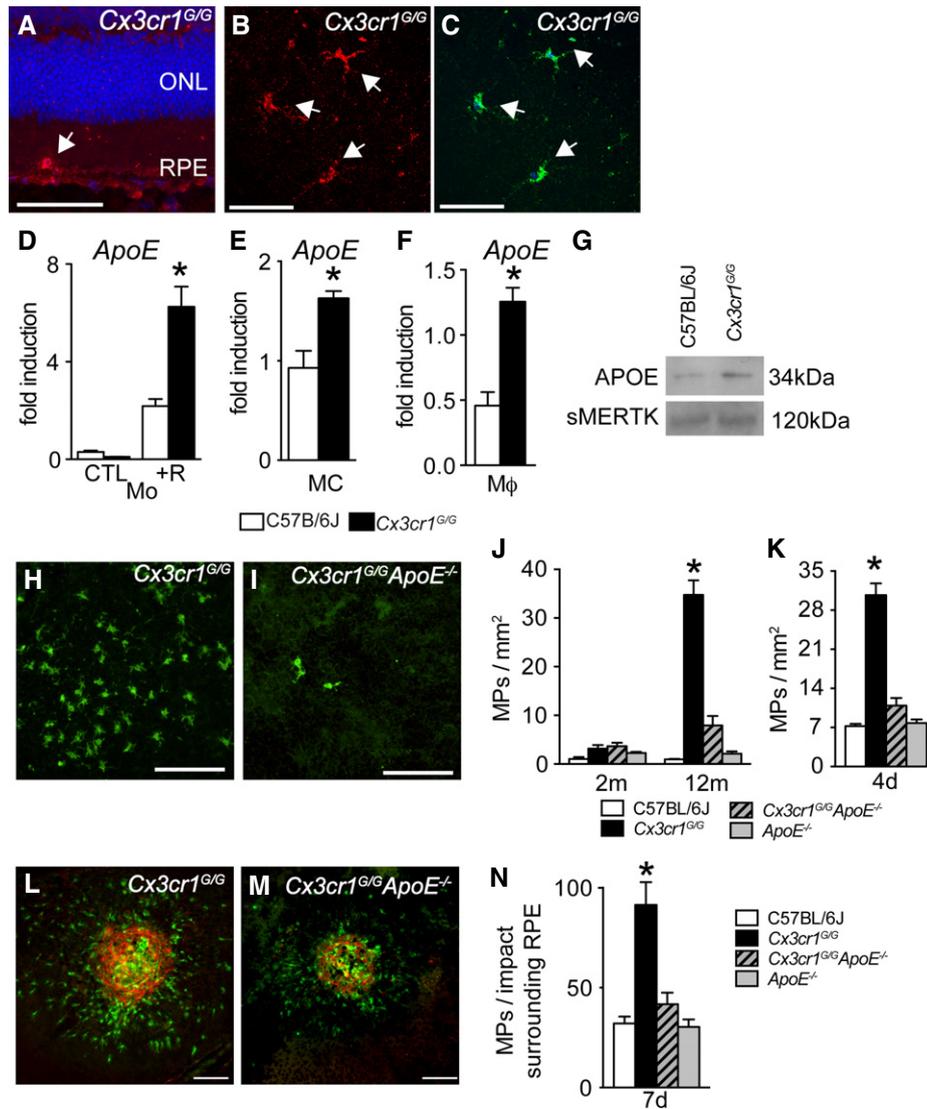


Figure 3. APOE promotes subretinal MP accumulation in *Cx3cr1^{GFP/GFP}* mice.

- A–C Immunohistochemistry of APOE (red) and IBA-1 (green) on a section (A, blue Hoechst) and the subretinal side of a retinal flatmount (B and C) from a 12-month-old *Cx3cr1^{GFP/GFP}* mouse (representative of 3 independent experiments, experiments omitting the primary antibody immunostaining served as negative controls). ONL: outer nuclear layer; RPE: retinal pigment epithelium.
- D Quantitative RT-PCR of *ApoE* mRNA normalized with S26 mRNA of C57BL/6J and *Cx3cr1^{GFP/GFP}* monocytes cultured for 24 h in contact with POS of an overlaying retinal explant ($n = 4$ /group per experiment; Mann–Whitney *U*-test, $*P = 0.0286$, the experiment was repeated twice with similar results).
- E Quantitative RT-PCR of *ApoE* mRNA normalized with S26 mRNA of C57BL/6J and *Cx3cr1^{GFP/GFP}* FACS-sorted microglial cells, freshly extracted from adult brain ($n = 4–5$ /group; Mann–Whitney *U*-test, C57BL/6J versus *Cx3cr1^{GFP/GFP}* $*P = 0.0159$).
- F Quantitative RT-PCR of *ApoE* mRNA normalized with S26 mRNA of C57BL/6J and *Cx3cr1^{GFP/GFP}* peritoneal macrophages cultured for 24 h with CX3CL1. ($n = 4$ /group per experiment; Mann–Whitney *U*-test, $*P = 0.0286$. The experiment was repeated twice with similar results).
- G APOE Western blot analysis of equivalent amounts of supernatant protein from CX3CL1-exposed C57BL/6J and *Cx3cr1^{GFP/GFP}* peritoneal macrophages at 24 h. Soluble Mer receptor tyrosine kinase that is released constitutively from cultured macrophages served as a loading control. The experiment was repeated twice with similar results.
- H, I 12-month-old IBA-1 stained RPE flatmounts of *Cx3cr1^{GFP/GFP}* (H) and *Cx3cr1^{GFP/GFP}* *ApoE^{-/-}* (I) mice.
- J Quantification of subretinal IBA-1⁺ mononuclear phagocytes in 2-month-old (left) and 12-month-old (right) mice of the indicated strains ($n = 9–20$ /group ANOVA/Dunnett test at 12 months: *Cx3cr1^{GFP/GFP}* versus any other group $*P < 0.0001$; Mann–Whitney *U*-test at 12 months of *Cx3cr1^{GFP/GFP}* versus *Cx3cr1^{GFP/GFP}* *ApoE^{-/-}* $*P < 0.0001$).
- K Quantification of subretinal IBA-1⁺ mononuclear phagocytes after 4 days of light challenge of 2-month-old mice of the indicated strains ($n = 10–25$ /group ANOVA/Dunnett test: *Cx3cr1^{GFP/GFP}* versus any other group $*P < 0.0001$; Mann–Whitney *U*-test of *Cx3cr1^{GFP/GFP}* versus *Cx3cr1^{GFP/GFP}* *ApoE^{-/-}* $*P < 0.0001$).
- L, M IBA-1- (green) and CD102- (red) stained RPE flatmounts 7 days after laser injury of 2-month-old *Cx3cr1^{GFP/GFP}* (E) and *Cx3cr1^{GFP/GFP}* *ApoE^{-/-}* (F) mice.
- N Quantification of subretinal IBA-1⁺ mononuclear phagocytes on the RPE counted on the RPE at a distance of 0–500 μ m to CD102⁺ CNV 7 days after the laser injury of 2-month-old mice of the indicated strains ($n = 8–10$ eyes/group ANOVA/Dunnett test: *Cx3cr1^{GFP/GFP}* versus any other group $*P < 0.0001$; Mann–Whitney *U*-test of *Cx3cr1^{GFP/GFP}* versus *Cx3cr1^{GFP/GFP}* *ApoE^{-/-}* $*P < 0.0001$).

Data information: +R: cultured with an overlaying retinal explant. Scale bars, 50 μ m.

ApoE^{-/-} mice were significantly protected against the subretinal MP accumulation observed in *Cx3cr1*^{GFP/GFP} mice after 4 days of light challenge (Fig 3K). It should be noted that the intensity of the light challenge model used herein is sufficient to induce subretinal inflammation in the *Cx3cr1*^{-/-} mice but does not cause significant subretinal inflammation nor degeneration in WT mice (Sennlaub *et al*, 2013). Moreover, 7 days after a laser impact, subretinal IBA-1⁺ MPs (green staining) counted on the RPE at a distance of 0–500 μm to CD102⁺ CNV (red staining) in *Cx3cr1*^{GFP/GFP} (Fig 3L) and *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice (Fig 3M) were significantly inhibited in *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice (Fig 2N). Additionally, *ApoE* deletion also significantly inhibited the age-dependent photoreceptor degeneration and exaggerated CNV observed in *Cx3cr1*^{GFP/GFP} mice (Supplementary Figs S2 and S3).

C57BL/6 mice are inbred and can carry *Pde6b*^{rd1} (retinal degeneration 1), *Crb1*^{rd8} (retinal degeneration 8), and *Gnat2*^{cpfl3} (Cone photoreceptor function loss 3) mutations relatively commonly (Chang *et al*, 2013). These mutations can lead to subretinal inflammation secondary to primary retinal degeneration (Luhmann *et al*, 2012). In our experiments, all mice strains used tested negative for these three mutations. Furthermore, subretinal MP accumulation in 12-month-old *Cx3cr1*^{+GFP} and *Cx3cr1*^{GFP/GFP} littermates of *Cx3cr1*^{+GFP} breeders showed no evidence of influence from an unknown contributor gene specific to the *Cx3cr1*^{GFP/GFP} mouse line (Supplementary Fig S4). *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice were generated twice with independently purchased *Cx3cr1*^{GFP/GFP} and *ApoE*^{-/-} mice (once at the Laboratoire Immunité et Infection and once at the Institut de la Vision), and both *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice strain generations were protected against the subretinal MP accumulation observed in the two *Cx3cr1*^{GFP/GFP} mouse strains of the two sites. Taken together, these results make it highly unlikely that the MP accumulation in *Cx3cr1*^{GFP/GFP} mice and the protection in *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice are due to genes other than *Cx3cr1* and *ApoE*.

In summary, we show that APOE is robustly expressed in subretinal MPs, more strongly expressed in *Cx3cr1*^{GFP/GFP} MPs, and that *ApoE* deletion very significantly inhibited the age-, light-, and laser-induced accumulation of subretinal MPs observed in *Cx3cr1*-deficient mice.

APOE inhibits subretinal MP clearance

The reasons for which subretinal MPs accumulate in *Cx3cr1*-deficient mice are not fully understood. Theoretically, the numbers of subretinal MPs are determined by (i) recruitment, (ii) *in situ* proliferation, (iii) migration (egress), and/or (iv) apoptotic clearance. We previously showed that *Cx3cr1*^{GFP/GFP} MPs overexpress CCL2, which in turn leads to increased CCR2⁺ Mos recruitment from the blood. This in part explains the accumulation of MPs in *Cx3cr1*-deficient mice (Sennlaub *et al*, 2013). Local injections of the traceable nucleotide EdU in light-challenged *Cx3cr1*^{GFP/GFP} mice failed to be incorporated in subretinal MPs, suggesting that *in situ* proliferation does not significantly contribute to the accumulation (Supplementary material of (Sennlaub *et al*, 2013)). To evaluate whether subretinal MPs egress from the subretinal space or undergo apoptosis, we adoptively transferred 12,000 CFSE-stained WT and *Cx3cr1*^{GFP/GFP} thioglycollate-elicited peritoneal cells (containing 70% Mφs) in the subretinal space of WT mice (Fig 4A, 12 h) and

counted the number of F4/80-expressing Mφs that co-stained for CFSE on RPE and retinal flatmounts once retinal detachment had subsided (8–12 h). Quantifications show that injected CFSE⁺ Mφs of both genotypes were cleared from the subretinal space over a period of 4 days, but that *Cx3cr1*^{GFP/GFP}-Mφ clearance was significantly slower and that *Cx3cr1*^{GFP/GFP}-Mφs subsisted in significantly higher numbers at one and two days (Fig 4B). We detected no signs of egress from the subretinal space in WT or *Cx3cr1*^{GFP/GFP}-Mφ-injected animals, as no CFSE⁺ cells were observed in the inner retina, choroid, blood, local lymph nodes, lung, liver, or spleen by histology or cytometry (data not shown). However, the nuclei of a large number of subretinal CFSE⁺ Mφs of both genotypes were TUNEL⁺ (Fig 4C, TUNEL stained (red) CFSE⁺ (green) WT-Mφs) and displayed signs of apoptosis (Fig 4C, inset: pyknotic and fragmented nuclei). These results suggest that subretinal Mφ clearance is predominantly mediated by apoptosis, in accordance with observations of inflammation resolution in peripheral tissue (Gautier *et al*, 2013) and in particular with leukocyte clearance in the context of the subretinal immunosuppressive environment (Streilein *et al*, 2002).

Cytometric quantification of CFSE⁺F4/80⁺CD11b⁺-Mφs in eye cell suspensions from injected eyes confirmed that *Cx3cr1*^{GFP/GFP}-Mφs were present in significantly higher numbers when compared to WT-Mφs 1 day after adoptive transfer (Fig 4D, the GFP fluorescence did not interfere with the several log stronger CFSE signal in the cytometric analysis). The CFSE fluorescence intensity of the F4/80⁺CD11b⁺-Mφs in the cytometric analysis was strong and homogeneous (Fig 4D), suggesting that CFSE uptake by host cells (which leads to variable CFSE intensities) or proliferation (which leads to cell populations with halved CFSE fluorescence intensity) did not occur to a significant degree. Furthermore, WT- and *Cx3cr1*^{GFP/GFP}-Mφs did not reveal differences in proliferation *in vitro* (Supplementary Fig S5), suggesting that fast proliferation of *Cx3cr1*^{GFP/GFP}-Mφs does not account for the observed difference in the adoptive transfer experiments.

We next evaluated whether the observed differences were specific to peritoneal Mφs or shared by MPs of other origins. We adoptively transferred CFSE-labeled, magnetic-bead-sorted bone marrow-derived Mos (~95% pure, Fig 4E) and CD11b FACS-sorted brain MCs (~95% pure, Fig 4F) from WT and *Cx3cr1*^{GFP/GFP} mice into the subretinal space of WT mice. As with peritoneal Mφs, *Cx3cr1*-deficient MPs of both origins were significantly greater in number when counted on retinal and RPE/choroidal flatmounts 1 day after the injections.

To evaluate whether MP APOE expression influences the rate of subretinal MP clearance, we adoptively transferred *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-}-Mφs into WT recipients. Strikingly, the increased resistance to subretinal clearance of *Cx3cr1*^{GFP/GFP}-Mφs was completely eliminated with *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-}-Mφs (Fig 4G). Furthermore, exogenous lipid-free APOE3, the predominant human APOE isoform, was sufficient to increase resistance to subretinal clearance when added to WT-CFSE⁺-Mφs (Fig 4H).

Taken together, our results show that *Cx3cr1*-deficient MPs of all origins studied (peritoneum, bone marrow, and brain) are more resistant to subretinal clearance. We show that this increase of resistance to clearance is APOE dependent and that local, recombinant APOE is sufficient to inhibit WT-Mφs elimination from the subretinal space.

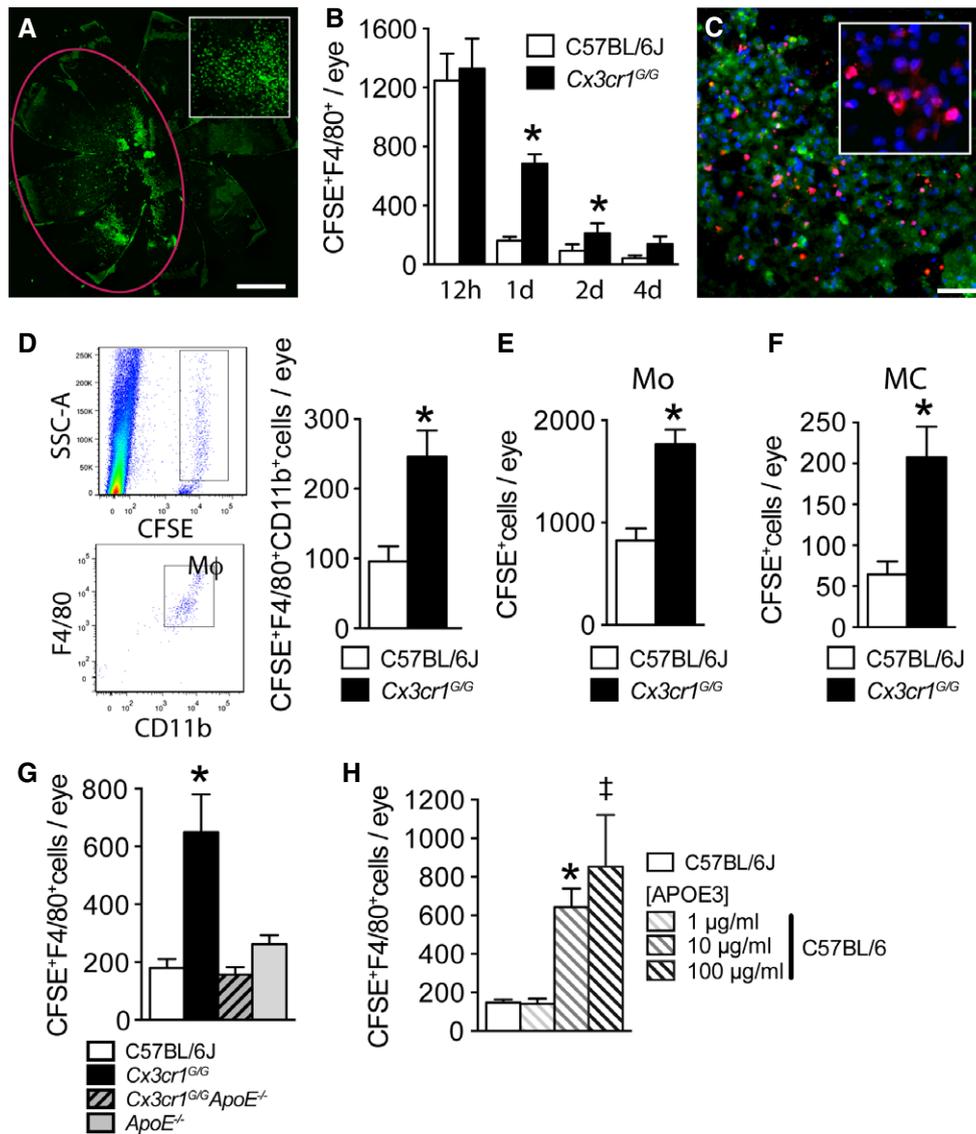


Figure 4. APOE inhibits subretinal MP clearance.

- A** Representative image of a RPE flatmount 12 h after the subretinal injection (red marking) of 4 μ l PBS with 12,000 CFSE-stained thioglycollate-elicited peritoneal cells that contain 70% macrophages (inset close-up view).
- B** Quantifications of CFSE⁺F4/80⁺ macrophages at different time points after subretinal injections of C57BL/6J and *Cx3cr1*^{GFP/GFP} CFSE⁺ macrophages ($n = 5$ /per group (12 h) and $n = 6$ /per group thereafter; Mann–Whitney *U*-test, C57BL/6J versus *Cx3cr1*^{GFP/GFP}: 1 day $n = 20$ /group * $P < 0.0001$; 2 day $n = 6$ /group * $P = 0.0317$).
- C** Representative image of TUNEL/Hoechst double-staining 12 h after subretinal injection of C57BL/6J CFSE⁺ macrophages (experiment repeated three times, inset close-up view).
- D** Representative cytometry images of SSC-A/CFSE and CD11b/F4/80 gated analysis of eye cell suspensions prepared 24 h after the injection of *Cx3cr1*^{GFP/GFP} CFSE⁺ macrophages and cytometric quantification of eye cell suspensions at 24 h after the injection of C57BL/6J and *Cx3cr1*^{GFP/GFP} macrophages into C57BL/6J ($n = 16$ –20/group; Mann–Whitney *U*-test, * $P = 0.0024$).
- E** Quantification of subretinal CFSE⁺ cells on RPE and retinal flatmounts 24 h after subretinal injections of CFSE⁺ magnetic-bead-sorted bone marrow-derived monocytes (Mo) from C57BL/6J and *Cx3cr1*^{GFP/GFP} mice into C57BL/6J mice ($n = 8$ –12/group; Mann–Whitney *U*-test, * $P = 0.0006$).
- F** Quantification of subretinal CFSE⁺ cells on RPE and retinal flatmounts 24 h after subretinal injections of CFSE⁺CD11b FACS-sorted brain microglial cells from C57BL/6J and *Cx3cr1*^{GFP/GFP} mice into C57BL/6J mice ($n = 9$ –12/group; Mann–Whitney *U*-test, * $P = 0.0087$).
- G** Quantification of subretinal CFSE⁺F4/80⁺ macrophages on RPE and retinal flatmounts 24 h after subretinal injections of CFSE⁺ macrophages from C57BL/6J, *Cx3cr1*^{GFP/GFP}, *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-}, and *ApoE*^{-/-} mice into C57BL/6J mice ($n = 8$ –12/group; one-way ANOVA/Dunnett test of *Cx3cr1*^{GFP/GFP} versus any other group * $P \leq 0.0001$; Mann–Whitney *U*-test, *Cx3cr1*^{GFP/GFP} versus *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} * $P = 0.0006$).
- H** Quantification of subretinal CFSE⁺F4/80⁺ macrophages on RPE and retinal flatmounts 24 h after subretinal injections of C57BL/6J CFSE⁺ macrophages into C57BL/6J and with exogenously added APOE3 at 1, 10, or 100 μ g/ml calculated intraocular concentrations ($n = 6$ –7/group; one-way ANOVA/Dunnett test: C57BL/6J versus 10 μ g * $P = 0.0488$; C57BL/6J versus 100 μ g † $P = 0.006$. Mann–Whitney *U*-test: C57BL/6J versus 10 μ g * $P = 0.0012$; C57BL/6J versus 100 μ g † $P = 0.0013$).

Data information: All primary cells were prepared from male mice; all recipient C57BL/6J mice were male. Mo: monocytes; MC: microglial cells; M ϕ : macrophages; SSC-A: side scatter detector A. Scale bars: 1 mm (A); 50 μ m (C).

FAS-FASL signaling mediates subretinal MP clearance

The RPE constitutively expresses FASL (CD95L), which in part mediates its immunosuppressiveness (Wenkel & Streilein, 2000). To test whether an alteration of the RPE immunosuppressive environment is associated with subretinal *Cx3cr1^{GFP/GFP}*-MP accumulation, we first

analyzed FasL expression *in vivo*. *FasL* mRNA was similarly expressed in 2-month-old WT and *Cx3cr1^{GFP/GFP}* mouse RPE/choroidal plexus, before significant subretinal MP accumulation occurred in *Cx3cr1^{GFP/GFP}* mice. In contrast, 12-month-old *Cx3cr1^{GFP/GFP}* mice and 2-month-old light-challenged *Cx3cr1^{GFP/GFP}* mice with subretinal MP accumulation expressed significantly less *FasL* mRNA (Fig 5A,

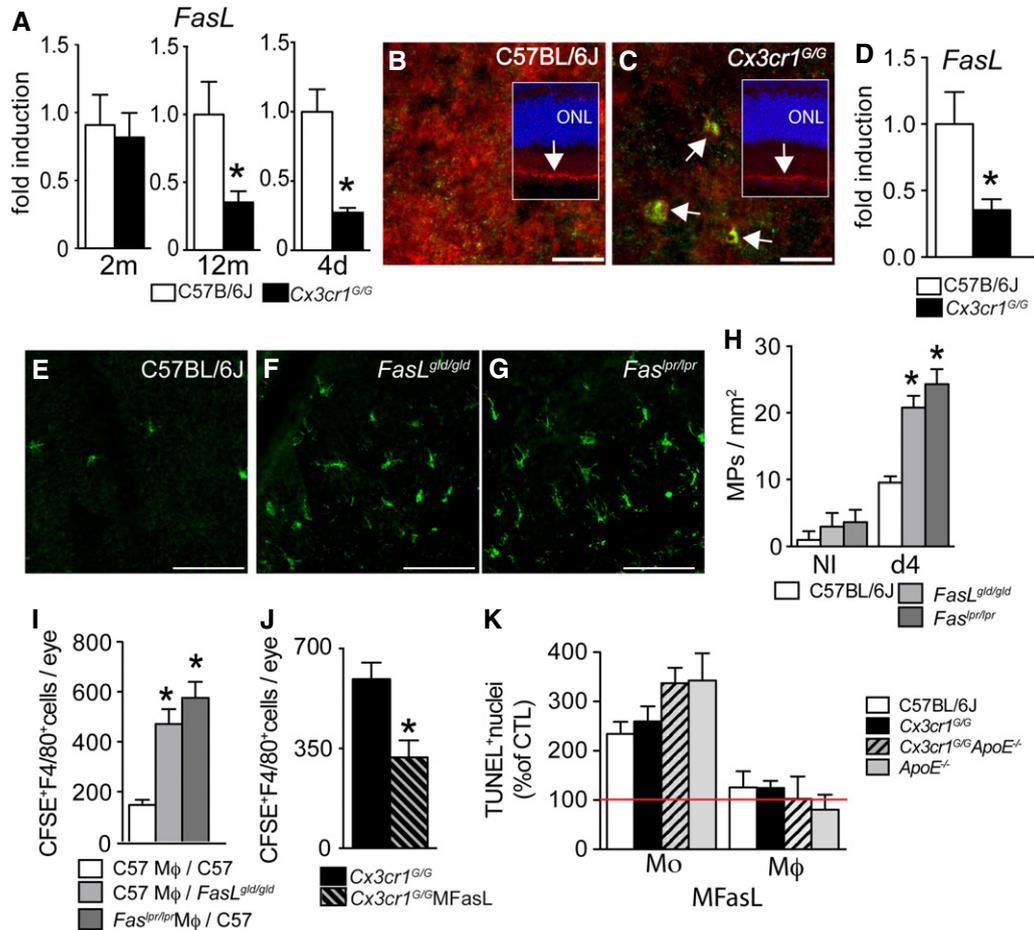


Figure 5. FAS-FASL signaling mediates subretinal MP clearance.

- A** Quantitative RT-PCR of *FasL* mRNA normalized with β -actin mRNA of 2-month-, 12-month-, and 2-month-old mice after 4 days of light challenge of C57BL/6 and *Cx3cr1^{GFP/GFP}* mouse RPE/choroid plexus (aging: $n = 5-6$ /group; Mann-Whitney *U*-test: 12 months $*P = 0.0129$; 4 days light challenge: $n = 7-8$ /group; $*P = 0.029$).
- B, C** Immunohistochemistry of FASL (red) and IBA-1 (green) of a RPE flatmounts and sections (insets, Hoechst staining blue) from a 12-month-old WT (B) and *Cx3cr1^{GFP/GFP}* (C) mouse (representative of three independent experiments, immunostainings omitting the primary antibody served as negative controls).
- D** Quantitative RT-PCR of *FasL* mRNA normalized with β -actin mRNA of 12-month-old C57BL/6 RPE/choroid plexus extracts 3 h after subretinal injection of C57BL/6j macrophages or *Cx3cr1^{GFP/GFP}* macrophages ($n = 6$ per group), Mann-Whitney *U*-test, $*P = 0.0087$.
- E-G** IBA-1-stained RPE flatmounts of 2-month-old C57BL/6j, *FasL^{gld/gld}*, and *Fas^{lpr/lpr}* mice after 4 days of light challenge.
- H** Quantification of subretinal IBA-1⁺ mononuclear phagocytes in control (left) and 4-day light-challenged (right) 2-month-old mice of the indicated strains ($n = 6-10$ /group ANOVA/Dunnett test at 4-day light challenge: C57BL/6j versus *FasL^{gld/gld}* and C57BL/6j versus *Fas^{lpr/lpr}* both $*P < 0.0001$; Mann-Whitney *U*-test at 4-day light challenge: C57BL/6j versus *FasL^{gld/gld}* $*P < 0.0001$; C57BL/6j versus *Fas^{lpr/lpr}* $*P < 0.0001$).
- I** Quantification of subretinal CFSE⁺F4/80⁺ macrophages on RPE and retinal flatmounts 24 h after subretinal injection of C57BL/6j CFSE⁺ macrophages into C57BL/6j and *FasL^{gld/gld}* mice and *Fas^{lpr/lpr}* macrophages into C57BL/6j mice ($n = 10-15$ /group, one-way ANOVA/Dunnett test: C57BL/6j macrophages inj. into C57BL/6j mice versus C57BL/6j macrophages inj. into *FasL^{gld/gld}* mice $*P = 0.0002$; C57BL/6j macrophages inj. into C57BL/6j mice versus *Fas^{lpr/lpr}* macrophages inj. into C57BL/6j mice $*P < 0.0001$. Mann-Whitney *U*-test: C57BL/6j macrophages inj. into C57BL/6j mice versus C57BL/6j macrophages inj. into *FasL^{gld/gld}* mice $*P < 0.0001$; C57BL/6j macrophages inj. into C57BL/6j mice versus *Fas^{lpr/lpr}* macrophages inj. into C57BL/6j mice $*P < 0.0001$).
- J** Quantification of subretinal CFSE⁺F4/80⁺ macrophages on RPE and retinal flatmounts 24 h after subretinal injection of *Cx3cr1^{GFP/GFP}* CFSE⁺ macrophages into C57BL/6j with or without the Fas agonist MegaFasL (calculated intraocular concentrations 1 ng/ml; $n = 7-8$; Mann-Whitney *U*-test: $*P = 0.014$).
- K** *In vitro* MegaFasL-induced apoptosis of monocytes and macrophages of the indicated genotypes cultured for 24 h. TUNEL⁺ nucleus quantification expressed as percentage of non-MegaFasL-exposed control.

Data information: All primary cells were prepared from male mice; all recipient C57BL/6j or *FasL^{gld/gld}* mice were male. Mo: monocytes; M Φ : macrophages. Scale bars: 20 μ m (B, C); 50 μ m (E-G).

RT-PCR) as compared to WT. Immunohistochemistry on retinal sections and RPE flatmounts of WT and *Cx3cr1^{GFP/GFP}* mice (Fig 5B and C, FasL, red; IBA-1, green) seemed to confirm the diminished FasL expression in the RPE of *Cx3cr1^{GFP/GFP}* mice at 12 months. These results might suggest that *Cx3cr1*-deficient MPs somehow inhibit RPE *FasL* transcription. Indeed, when we injected *Cx3cr1^{GFP/GFP}*-Mφs into the subretinal space of WT mice, *FasL* transcription on RPE/choroidal extracts was significantly inhibited after 3 h, when compared to WT-Mφs-injected eyes (Fig 5D, RT-PCR).

To evaluate whether FAS-FASL signaling participates in MP clearance, we first compared subretinal MP numbers in light-challenged WT (Fig 5E), FasL-defective (*FasL^{gld/gld}* mice, Fig 5F), and Fas-defective (*Fas^{lpr/lpr}* mice, Fig 5G) mice (*FasL^{gld/gld}* and *Fas^{lpr/lpr}* mice develop lymphadenopathy and systemic autoimmune disease with age, making it difficult to evaluate age-dependent MP accumulation at 12 months). Quantification of subretinal IBA-1⁺ MPs on retinal and RPE/choroidal flatmounts revealed a significant increase of subretinal MPs in 2-month-old *FasL^{gld/gld}* and *Fas^{lpr/lpr}* mice induced by 4 days of light challenge (Fig 5H), similar to that of *Cx3cr1^{GFP/GFP}* mice (Fig 3). Moreover, adoptive transfer experiments in which we subretinally injected thioglycollate-elicited WT-CFSE⁺-Mφs into WT or FasL-defective mice (*FasL^{gld/gld}* mice), and Fas-defective CFSE⁺ Mφs (prepared from thioglycollate-elicited peritonitis of *Fas^{lpr/lpr}* mice) into WT mice, revealed that subretinal CFSE⁺ F4/80⁺-Mφs were significantly greater in number 24 h after the injection when FAS or FASL function was impaired (Fig 5I) and comparable to the phenotype observed in *Cx3cr1^{GFP/GFP}* Mφs (Fig 3). In addition, co-administration of FAS agonist MegaFasL (Greaney et al, 2006) to *Cx3cr1^{GFP/GFP}* Mφs efficiently compensated for the observed FasL downregulation (Fig 5D) and significantly reduced the number of subretinal *Cx3cr1^{GFP/GFP}* CFSE⁺ F4/80⁺-Mφs after adoptive transfer (Fig 5J).

To test whether differences in the susceptibility to FasL-induced MP death might contribute to the protective effect of *ApoE* deletion in subretinal MP accumulation, we exposed Mos and thioglycollate-elicited Mφs from the different mouse strains to MegaFasL and quantified TUNEL⁺ cells at 24 h *in vitro* (Fig 5K). Our results confirm previous reports that FasL is sufficient to induce Mos apoptosis *in vitro* in comparison with Mφs, which are rather resistant to FasL-induced apoptosis *in vitro* (Um et al, 1996; Kiener et al, 1997; Park et al, 2003). We did not observe a difference between wild-type and *Cx3cr1^{GFP/GFP}* cells of either Mos or Mφs, but a tendency toward increased susceptibility in Mos of both *Cx3cr1^{GFP/GFP}* *ApoE^{-/-}* and *ApoE^{-/-}* cells, which might contribute toward the differences in clearance observed *in vivo* (Fig 3). These results also highlight that FasL acts along with other factors *in vivo* to induce Mφ apoptosis in the subretinal space, as the effect of MegaFasL on subretinal clearance of adoptively transferred peritoneal Mφs (Fig 5J) was much stronger than MegaFasL-induced apoptosis *in vitro* (Fig 5K). Similarly, a synergistic effect of FasL with other RPE-derived factors has been suggested by the observations that FasL is necessary to eliminate T cells and Mφs and prevent RPE allograft rejection (Wenkel & Streilein, 2000), but FasL over-expression alone in an allograft, such as a Langerhans cell graft, is not sufficient to prevent rejection (Kang et al, 1997).

Taken together, our data show that FAS-FASL signaling is implicated in subretinal MP clearance, that subretinal *Cx3cr1^{GFP/GFP}* MPs

are associated with a downregulation of RPE FasL expression, and that substitution by MegaFasL restores, in part, the clearance of subretinal *Cx3cr1^{GFP/GFP}* MPs.

APOE promotes subretinal macrophage survival via IL-6

Our results show that the increased survival of *Cx3cr1^{GFP/GFP}*-Mφs is associated with diminished RPE *FasL* transcription, which plays a role in subretinal MP clearance (Fig 4). Furthermore, APOE, over-expressed by *Cx3cr1^{GFP/GFP}*-Mφs or exogenously added to WT-Mφs, increases subretinal Mφs survival (Fig 3). However, subretinal injections of recombinant APOE without Mφs (at a concentration that increases subretinal MP survival (Fig 3H) and compared to PBS-injected eyes) did not replicate the observed *FasL* downregulation in RPE/choroid (Fig 6A, RT-PCR 3 h after injection), which we observed after adoptive transfer of *Cx3cr1^{GFP/GFP}*-Mφs (Fig 5D). These results suggest that APOE does not directly influence RPE *FasL* expression. However, recombinant IL-6, shown to downregulate *FasL* expression in lymphocytes (Ayroldi et al, 1998), was sufficient to significantly inhibit RPE *FasL* transcription in this experimental setting (Fig 6A). APOE and APOA-I can both activate the CD14-dependent innate immunity receptor cluster that contains TLR-2 and TLR-4 in the absence of TLR ligands (Smoak et al, 2010). This activation has been shown to induce IL-6, among other cytokines, in the case of APOA-I. Similarly, when we incubated WT-peritoneal-Mφs with recombinant lipid-free APOE3 for 24 h, IL-6 was very significantly induced (Fig 6B). The LPS inhibitor polymyxin B did not inhibit the induction, while 90-min heat denaturation abolished the induction, confirming that LPS contamination of APOE3 is not accountable for the effect, as shown for APOA-I using multiple approaches (Smoak et al, 2010). As previously shown for APOA-I, this induction was largely CD14 and TLR2 dependent, as neutralizing antibodies inhibited this effect, when compared to a control IgG (Fig 6B). Correspondingly, *Cx3cr1^{GFP/GFP}*-Mφs expressed significantly higher amounts of *IL-6* mRNA when compared to WT-Mφs cultured for 24 h. This effect was significantly inhibited in *Cx3cr1^{GFP/GFP}* *ApoE^{-/-}*-Mφs (Fig 6C), confirming the involvement of APOE. Although *IL-6* was not detectable by RT-PCR in whole-eye mRNA extracts *in vivo*, *IL-6* staining (Fig 6D, red) was reproducibly detected in subretinal IBA-1⁺-MPs (Fig 6E, green) adjacent to the phalloidin⁺ RPE (Fig 6E, blue, orthogonal, and lateral Z-stack projections of a confocal microscopy picture stack) of 12-month-old *Cx3cr1^{GFP/GFP}* mice and light-challenged *Cx3cr1^{GFP/GFP}* mice (data not shown).

In addition, recombinant IL-6 added to WT-CFSE⁺ Mφs more than doubled the number of subretinal CFSE⁺ F4/80⁺-Mφs (Fig 6F) and an IL-6-blocking antibody significantly decreased subretinal *Cx3cr1^{GFP/GFP}* CFSE⁺ F4/80⁺-Mφs (Fig 6G) 24 h after injection when compared to their controls.

When taken together, the results presented in Figs 2–5 suggest the following mechanism: *Cx3cr1^{GFP/GFP}* MPs express increased amounts of APOE. APOE induces the expression of IL-6 in MPs, which in turn downregulates *FasL* transcription in the RPE. The diminished FasL expression participates in the increased survival time of subretinal *Cx3cr1^{GFP/GFP}* MPs (Fig 6H).

Finally, to test whether CD14-dependent IL-6 induction does indeed participate in subretinal MP accumulation *in vivo*, we injected control IgG (Fig 6I), an IL-6- (Fig 6J) or CD14-neutralizing

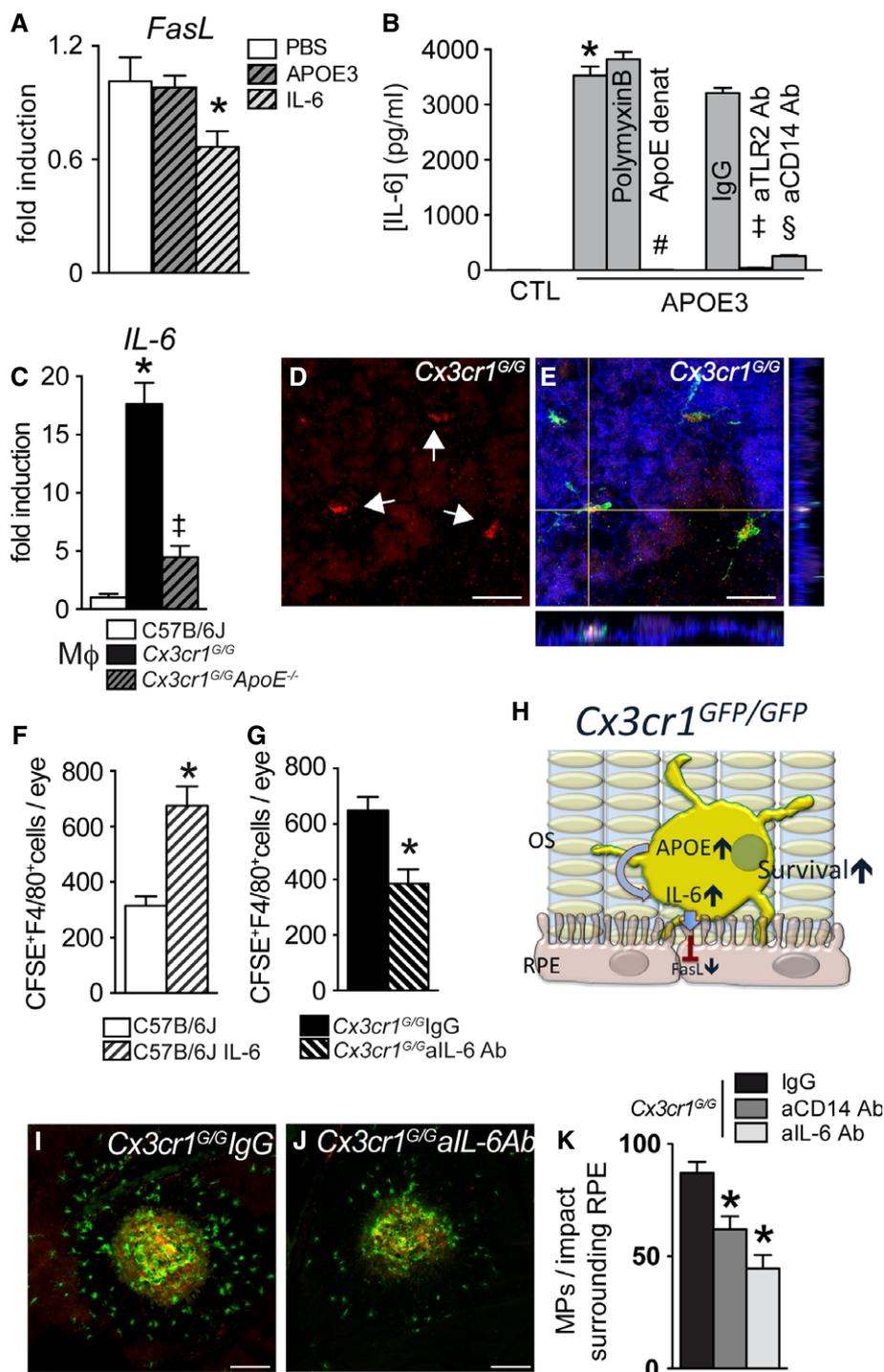


Figure 6.

antibody into the vitreous of *Cx3cr1^{GFP/GFP}* mice and induced subretinal inflammation with a laser injury (which also facilitates antibody penetration into the subretinal space). Our results show that the accumulation of subretinal IBA-1⁺ MPs (green staining) observed on the RPE adjacent to CD102⁺ CNV (red staining), 7 days after a laser impact, was significantly inhibited when CD14 or IL-6 was neutralized, as was the associated CNV (Supplementary Fig S6).

Discussion

MPs are activated and accumulate in wet AMD (Oh *et al*, 1999) and around the atrophic lesions of GA adjacent to the RPE (Combiadiere *et al*, 2007; Sennlaub *et al*, 2013). Our findings of numerous subretinal MPs in and around large drusen and GA lesions illustrate the significant focal inflammation that is present in AMD. The close contact of subretinal MPs and the RPE suggests that the

Figure 6. APOE promotes subretinal macrophage survival via IL-6.

- A Quantitative RT-PCR of *FasL* mRNA normalized with β -actin mRNA of 12-month-old C57BL/6 RPE/choroid plexus 3 h after subretinal injection of PBS (PBS; $n = 36$), IL-6 ($n = 17$), and APOE3 ($n = 21$); calculated intraocular concentrations: 5 ng/ml and 10 μ g/ml, respectively; one-way ANOVA/Dunnnett *post hoc* test PBS versus IL-6 * $P = 0.038$; t-test PBS versus IL-6 * $P = 0.043$.
- B Mouse IL-6 ELISA of supernatants from C57BL/6j peritoneal macrophages incubated for 24 h in control medium, lipid-free APOE3 (5 μ g/ml), APOE3 (5 μ g/ml), and polymyxin B (25 μ g/ml), heat-denatured APOE3 (dAPOE3, 5 μ g/ml), APOE3 (5 μ g/ml), and rat IgG1 isotype control (IgG, 100 μ g/ml) or APOE3 (5 μ g/ml) and rat anti-CD14 antibody (aCD14 Ab, 100 μ g/ml) or APOE3 (5 μ g/ml) and rat anti-TLR2 antibody (aTLR2 Ab, 100 μ g/ml) ($n = 5$ –6/group; one-way ANOVA/Bonferroni multi-comparison tests: APOE3 versus CTL * $P < 0.0001$; dAPOE3 versus APOE3 * $P < 0.0001$; APOE3 IgG versus CTL $P < 0.0001$; APOE3 IgG versus APOE3 aCD14 Ab $\S P < 0.0001$; APOE3 IgG versus APOE3 aTLR2 Ab ** $P < 0.0001$. Mann-Whitney *U*-test: APOE3 versus CTL * $P = 0.0043$; dAPOE3 versus APOE3 # $P = 0.0117$; APOE3 IgG versus CTL $P = 0.0080$; APOE3 IgG versus APOE3 aCD14 Ab $\S P = 0.0117$; APOE3 IgG versus APOE3 aTLR2 Ab ** $P = 0.0079$. The experiment was repeated twice with similar results).
- C Quantitative RT-PCR of *IL-6* mRNA normalized with *S26* mRNA of C57BL/6j and *Cx3cr1^{GFP/GFP}*, and *Cx3cr1^{GFP/GFP} ApoE^{-/-}* peritoneal macrophages cultured for 24 h with CX3CL1 ($n = 5$ per group, one-way ANOVA/Bonferroni *post hoc* multi-comparison tests: C57BL/6j versus *Cx3cr1^{GFP/GFP}* * $P < 0.0001$ and *Cx3cr1^{GFP/GFP}* versus *Cx3cr1^{GFP/GFP} ApoE^{-/-}* * $P < 0.0001$. Mann-Whitney *U*-test: C57BL/6j versus *Cx3cr1^{GFP/GFP}* * $P = 0.0079$ and *Cx3cr1^{GFP/GFP}* versus *Cx3cr1^{GFP/GFP} ApoE^{-/-}* * $P = 0.0079$. The experiment was repeated twice with similar results).
- D, E Orthogonal and lateral Z-stack projection of IL-6 (D, red), IBA-1⁺ (E, green), and phalloidin (E, blue) of IBA⁺ mononuclear phagocytes adjacent to the phalloidin⁺ RPE of a retinal flatmount from a 12-month-old *Cx3cr1^{GFP/GFP}* mouse. (Representative of three independent experiments, immunostainings omitting the primary antibody served as negative controls).
- F Quantification of subretinal CFSE⁺F4/80⁺ macrophages on RPE and retinal flatmounts 24 h after subretinal injection of C57BL/6j CFSE⁺ macrophages into C57BL/6j with or without IL-6 ($n = 15$ –20/group; calculated intraocular concentrations of 5 ng/ml. Mann-Whitney *U*-test: * $P < 0.0001$).
- G Quantification of subretinal CFSE⁺F4/80⁺ macrophages on RPE and retinal flatmounts 24 h after subretinal injection of *Cx3cr1^{GFP/GFP}*CFSE⁺M ϕ s into C57BL/6j with control (IgG, $n = 16$) or anti-IL-6 antibody (aIL-6 Ab, $n = 8$; calculated intraocular concentrations 5 μ g/ml; per group. Mann-Whitney *U*-test: * $P = 0.0036$).
- H Graphical summary.
- I, J 7 day laser-injured IBA-1 (green) and CD102 (red) double-stained RPE flatmounts of control IgG- (I) and anti-IL-6-treated (J) *Cx3cr1^{GFP/GFP}* mice.
- K Quantification of subretinal IBA-1⁺ mononuclear phagocytes/impact localized on the lesion surrounding RPE of *Cx3cr1^{GFP/GFP}* mice treated with control IgG, IL-6-, or CD14-blocking antibodies (calculated intraocular concentration 5 μ g/ml; $n = 13$ –14/group, one-way ANOVA/Dunnnett's *post hoc* tests of IgG versus any other group * $P < 0.001$. Mann-Whitney *U*-test IgG versus anti-IL-6 * $P = 0.0021$; IgG versus anti CD14 * $P = 0.0028$).

Data information: All primary cells were prepared from male mice; all recipient C57BL/6j mice were male; ONL: outer nuclear layer; OS: outer segments; RPE: retinal pigment epithelium. Scale bars: 20 μ m (D, E); 50 μ m (I, J).

immunosuppressive properties of the RPE (Streilein *et al*, 2002) are altered in the disease. We show that subretinal MPs, in addition to the RPE, express high levels of APOE similar to MPs in other inflammatory conditions (Rosenfeld *et al*, 1993).

Using *Cx3cr1^{GFP/GFP}* mice as a model of subretinal inflammation, we show that increased expression of APOE, observed in *Cx3cr1^{GFP/GFP}* MPs, is associated with significant acute (laser- and light-induced) and chronic (age-dependent) subretinal MP accumulation. The observation that *Cx3cr1*-deficient M ϕ s are cleared less efficiently from the subretinal space of WT mice suggests that the *in vivo* accumulation is at least in part due to the capacity of *Cx3cr1*-deficient M ϕ s to inhibit the RPE immunosuppression and prolong their survival when compared to WT-M ϕ s. Using *Cx3cr1^{GFP/GFP} ApoE^{-/-}*-M ϕ s and recombinant APOE, we show that this capacity is APOE dependent. We show that APOE increases the expression of IL-6 in a CD14-dependent manner, likely activating the innate immunity receptor cluster, as shown for APOA-I (Smoak *et al*, 2010). While our adoptive transfer experiments suggest that MP-derived APOE is sufficient to induce this effect (all experiments were conducted with C57BL/6J recipients), we do not exclude that RPE-derived APOE participates in the effect *in vivo*, in particular in human AMD where RPE APOE expression was strong. IL-6 in turn inhibited MP elimination, as a CD14- and IL-6-blocking antibody partly reversed the elimination deficit of adoptively transferred M ϕ s and inhibited subretinal MP accumulation in a model of laser-induced inflammation in *Cx3cr1^{GFP/GFP}* mice. While we observe that extracellular recombinant APOE can induce IL-6, we do not exclude the possibility that intracellular APOE participates in the increased IL-6 secretion observed in *Cx3cr1^{GFP/GFP}* MPs. IL-6 secreted from the subretinal MPs in turn downregulates *FasL* transcription in RPE. Using light-induced subretinal inflammation

and adoptive transfer experiments and *FasL^{gld}* and *Fas^{lpr}* mice, we show that FASL/FAS signaling takes part in MP elimination. Indeed, a pharmacological FAS agonist was able to partly reverse the elimination deficit of adoptively transferred *Cx3cr1*-deficient M ϕ s, similar to the IL-6-blocking antibody. Interestingly, *ApoE*-deficient monocytes, but not macrophages, also showed a tendency toward increased susceptibility to FASL-induced apoptosis *in vitro*, which might contribute toward the differences in clearance observed *in vivo*.

The accumulation of subretinal MPs is likely the result of MP recruitment that exceeds the MP elimination rate. In the present study, we have concentrated on MP elimination, but we do not suggest that increased recruitment is less important. Indeed, APOE like APOA-I also increases CCL2 expression in MPs, and we have previously shown the role of CCL2 in subretinal MP accumulation (Sennlaub *et al*, 2013). Also, our study focused on FasL expression, but other RPE-derived signals likely play a role in MP elimination and non-transcriptional interactions such as proteolytic cleavage of FASL probably participate in the mechanism.

The cross talk of APOE and APOA-I, the reverse cholesterol transport (RCT), and innate immunity are complex. On the one hand, excess of cholesterol and cholesterol crystals can activate Toll-like receptors (TLR) and APOE and APOA-I can bind and neutralize TLR ligands and inhibit the induction of inflammatory cytokines (Azzam & Fessler, 2012). On the other hand, Smoak *et al* demonstrated that APOE and APOA-I are capable of triggering TLR2/4 signaling in the absence of TLR ligands, possibly by extracting cholesterol from lipid rafts in which the CD14-containing innate immunity receptor cluster (necessary for TLR2/4 signaling) is located (Smoak *et al*, 2010). Our results using a CD14-blocking antibody and a recent report that subretinal MP accumulation is TLR4

dependent in a retinal degeneration model (Kohno *et al*, 2013) back the notion that TLR signaling is involved in the alteration of the immunosuppressive environment. The involvement of reverse cholesterol transport (RCT) in AMD might also be supported by the recent observation that APOA-I levels are elevated in the vitreous of AMD patients (Koss *et al*, 2014). Furthermore, a polymorphism of the ATP binding cassette transporter 1 (ABCA1, associated with low HDL and therefore possibly impaired RCT) has recently been shown to be protective against advanced AMD (Chen *et al*, 2010). Moreover, IL-6 levels are associated with early AMD incidence (Klein *et al*, 2014) and late AMD (Seddon *et al*, 2005; Klein *et al*, 2008) and might participate in the weakening of RPE immunosuppression in AMD.

In vitro, the RPE has been shown to secrete APOE preferentially to its basal side (Ishida *et al*, 2004). Our immunohistochemical localization to the basal side of the RPE in healthy human donor tissue might suggest that this is also the case *in vivo*. Interestingly, this polarization of the APOE signal was lost in GA donor tissue, where a strong APOE signal was observed throughout the RPE cells adjacent to the lesion. Apical APOE release from RPE into the subretinal space might participate in increasing its subretinal concentration in addition to APOE released from subretinal MPs.

It has been suggested that a lack of APOE participates in both the accumulation of lipids and the drusen formation (Ong *et al*, 2001; Malek *et al*, 2005; Johnson *et al*, 2011). Indeed, *ApoE*^{-/-} and *APOE4* mice fed on a high-fat diet develop lipid accumulations in the Bruch's membrane, which has been proposed as similar to early AMD (Ong *et al*, 2001; Malek *et al*, 2005). While these observations might apply to early AMD, they are unlikely to play a role in late AMD in which increased APOE immunoreactivity is observed (Klaver *et al*, 1998; Anderson *et al*, 2001) and in which the *APOE4*-allele plays a protective role (McKay *et al*, 2011).

Taken together, the results of our study describe a new cellular and molecular mechanism that may participate in the weakened subretinal immunosuppression and chronic inflammation of AMD. Although we have detected this mechanism in mice, increased APOA-I levels in the vitreous of AMD patients (Koss *et al*, 2014) and elevated IL-6 levels with AMD suggest that a similar mechanism is at work in human AMD patients. In the future, the inhibition of IL-6 and/or the restoration of RPE immunosuppression might be employed to control pathologic subretinal inflammation in AMD.

Materials and Methods

APOE, IBA-1, and CD18 immunohistochemistry on donor samples

Donor eyes with a known history of AMD and controls were collected through the Minnesota Lions Eye bank. Informed consent was obtained for all donor eyes by the Minnesota Lions Eye bank, and the experiments conformed to the principles set out in the WMA Declaration of Helsinki. Postmortem fundus photographs were taken, and the posterior segment was fixed 4 h in 4% PFA, transported in PBS, dissected, imbedded in paraffin, and sectioned (five control maculae; five GA donor maculae). Donors gave informed consent in accordance with the eye bank's ethics committee. Five tonsillectomy surgical samples, removed for recurrent

acute tonsillitis, were recuperated from tonsillectomies at the Fondation Rothschild and then fixed and sectioned in the same way. For flatmount immunohistochemistry, donor eyes with visible atrophic areas (five eyes), visible large drusen on RPE flatmounts (five eyes), and controls (three eyes) were dissected into approximately 5 × 5 mm tissue parts, and immunohistochemistry was performed on submerged samples. APOE (M068-3 mouse anti-human, citrate buffer heat antigen retrieval for paraffin sections, MBL), IBA-1 (rabbit anti-human, citrate buffer heat antigen retrieval, Wako Chemicals), and CD18 (MCA503, rat anti-human, citrate buffer heat antigen retrieval, AbD Serotec) immunohistochemical analyses were performed using appropriate fluorescent or alkaline phosphatase-coupled secondary antibodies (Molecular Probe) using a Fast Red substrate kit (Sigma).

Animals

Wild-type and *Cx3cr1*^{GFP/GFP}, *ApoE*^{-/-}, *Fas*^{lpr}, and *FasL*^{gld} were purchased (Charles River Laboratories, Jackson laboratories), and *Cx3cr1*^{GFP/GFP} *ApoE*^{-/-} mice were generated. All mice were negative for the *Crb1*^{rd8}, *Pde6b*^{rd1}, and *Gnat2*^{cpfl3} mutations. Mice were housed in the animal facility under specific pathogen-free condition, in a 12/12 h light/dark (100–500 lx) cycle with water and normal diet food available *ad libitum*. All experimental protocols and procedures were approved by the local animal care ethics committee 'Comité d'éthique en expérimentation animale Charles Darwin' (N° p3/2008/54).

Light challenge and laser injury model

Two- to four-month-old mice were adapted to darkness for 6 h, and pupils dilated and exposed to green LED light (starting at 2AM, 4,500 lx, JP Vezon equipments) for 4 days as previously described (Sennlaub *et al*, 2013). Laser coagulations were performed with a 532-nm ophthalmological laser mounted on an operating microscope (Vitra Laser, 532 nm, 450 mW, 50 ms and 250 μm). Intravitreal injections of 2 μl of PBS, isotype control rat IgG1, rat anti-mouse IL-6 (R&D Systems), and rat anti-mouse CD14 (BD Biosciences) were performed using glass capillaries (Eppendorf) and a microinjector. The 2 μl solution of the antibodies was injected at 50 μg/ml, corresponding to an intraocular concentration of 5 μg/ml assuming their dilution by approximately 1/10 in the intra-ocular volume.

Immunohistochemistry, MP quantification, and histology

Human and murine RPE and retinal flatmounts and human and murine sections were stained and quantified as previously described (Sennlaub *et al*, 2013) using polyclonal goat anti-human APOE (Millipore), polyclonal rabbit anti-IBA-1 (Wako), polyclonal rabbit anti-rat FASL (Millipore), monoclonal rat anti-mouse IL-6 (R&D Systems), AlexaFluor 555 phalloidin (Mol probes), and rat anti-mouse CD102 (clone 3C4, BD Biosciences Pharmingen) appropriate secondary antibodies and counterstained with Hoechst if indicated. Preparations were observed with fluorescence microscope (DM5500, Leica) or a FV1000 (Olympus) confocal microscope. Histology of mice eyes and photoreceptor quantification were performed as previously described (Sennlaub *et al*, 2013).

Cell preparations and cell culture

Resident and thioglycollate-elicited peritoneal cells, peritoneal macrophages, bone marrow-derived monocytes, brain microglial cell, and photoreceptor outer segment (POS) isolation, and MP-retinal explant co-cultures (all in serum-free X-VIVO 15 medium) were performed as previously described (Sennlaub *et al*, 2013). In specific experiments, cells were stimulated with recombinant human CX3CL1 or APOE3 (5 µg/ml, Leinco Technologies), APOE3 (5 µg/ml) with polymyxin B (25 µg/ml, Calbiochem), heat-denatured APOE3 (5 µg/ml, 95°C, 90 min), rat anti-IgG isotype control (100 µg/ml, R&D), rat anti-mouse CD14 (100 µg/ml, R&D), mouse anti-mouse TLR2 (100 µg/ml, Invivogen), and POS prepared as previously described (Molday *et al*, 1987). For *in vitro* apoptosis experiments, 100,000 Mos or Mφs of the different genotypes were cultured for 24 h with or without MegaFasL (1 ng/ml, AdipoGen). TUNEL staining (*In Situ* Cell Death Detection Kit, Roche Diagnostics) was performed according to the manufacturer's instructions; TUNEL⁺ and Hoechst⁺ nuclei were counted automatically using the Array Scan (Thermo Fischer).

Subretinal mononuclear phagocyte cell clearance

Thioglycollate-elicited peritoneal cells (containing 70% Mφs), bone marrow-derived monocytes (~95% pure), and brain microglial cell (~95% pure) were labeled in 10 µM CFSE (Life Technologies). Cells were washed and resuspended in PBS. 12,000 cells (4 µl) were injected into the subretinal space of anesthetized 2-month-old mice using a microinjector and glass microcapillaries (Eppendorf). A hole was pierced with the glass capillary prior to the subretinal injection to avoid intra-ocular pressure increase and to allow retinal detachment with 4 µl of solution. The subretinal injection was verified by funduscopy. In specific experiments, Mφs were co-injected with rhAPOE (APOE3, Leinco Technologies), rmIL-6, rat anti-mouse IL-6, rat anti-mouse CD14, the isotype control rat IgG1 (R&D Systems), or MegaFasL (AdipoGen). Indicated intraocular concentrations were calculated as a dilution of 10× of the injected solution, as the injected 4 µl corresponds to approximately 1/10 of the intraocular volume. Eyes were enucleated after 24 h, fixed in 4% PFA, and flatmounted. The flatmounts were double-labeled with anti-F4/80 antibody to identify CFSE⁺F4/80⁺Mφs and counted on the subretinal aspect of the retinal flatmount and the RPE/choroid flatmount of each eye. Eyes with subretinal hemorrhages were discarded. Double-labeled MPs in subretinal space were quantified on RPE flatmounts and the subretinal side of retinal flatmounts.

Flow cytometry

Cytometry was performed as previously described (Camelo *et al*, 2012), using anti-CD11b PE, anti F4/80 Pacific Blue, or APC, streptavidin APC (all from AbD Serotec). Acquisition was performed on LSRII cytometer (BD Biosciences), and data were analyzed with FlowJo 7.9.

Western blot, reverse transcription and real-time polymerase chain reaction, and ELISA

WB analysis was performed using a polyclonal goat anti-ApoE (Millipore) and a polyclonal goat anti-Mertk (R&D) as previously described (Houssier *et al*, 2008). RT-PCRs using SYBR Green (Life

The paper explained

Problem

Physiologically, the subretinal space, located between the retinal pigment epithelium (RPE) and the photoreceptors, is a zone of immune privilege mediated in part by immunosuppressive RPE signals that induce apoptosis of leukocytes. In healthy subjects, this space is therefore devoid of all leukocytes, including mononuclear phagocytes (MP), a family of cells that include microglial cells, monocytes, and macrophages among others. Nevertheless, MPs accumulate subretinally in contact with the RPE in age-related macular degeneration (AMD) and likely participate in neurodegeneration and neovascularization in the disease. The reasons for the breakdown of subretinal immunosuppression and accumulation of MPs in AMD remain unknown.

Results

This study shows that subretinal MPs in AMD express high amounts of apolipoprotein E (APOE), a lipoprotein that plays a crucial role in reverse cholesterol transport. Using *Cx3cr1*^{-/-} mice that accumulate subretinal MPs with age and after light- or laser-induced stress, we demonstrate that APOE prolongs subretinal MP survival and is necessary for subretinal MP accumulation. We demonstrate that increased APOE induces IL-6 in MPs, via the activation of the TLR2-CD14-dependent innate immune receptor cluster. IL-6 in turn represses RPE FasL expression, prolongs subretinal MP survival, and promotes chronic subretinal inflammation.

Impact

Taken together, the results of our study describe a new cellular and molecular mechanism that explains the weakening of subretinal immunosuppression and consequently participates in chronic inflammation in AMD. Although detected in mice, the association of the APOE isoform 2, which is associated with higher APOE concentrations and elevated IL-6 serum levels in AMD, suggests that a similar mechanism is at work in human AMD patients. In future, the inhibition of IL-6 and the restoration of RPE immunosuppression could be used to control pathologic subretinal inflammation in AMD.

Technologies) and ELISAs using mouse IL-6 DuoSet (R&D Systems) were performed as previously described (Sennlaub *et al*, 2013).

Terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) on flatmounts

4% PFA-fixed retinal flatmounts were post-fixed in frozen methanol/acetic acid (2:1) for 30 min and washed in PBS. Flatmounts were incubated overnight at 4°C with the terminal transferase and the supplied buffer (*In Situ* Cell Death Detection Kit, Roche Diagnostics). Flatmounts were then incubated at 37°C for 90 min, and the reaction was stopped by washing with PBS. Nuclei were counterstained with Hoechst (Sigma-Aldrich). Flatmounts images were captured with a DM5500 microscope (Leica).

Statistical analysis

Sample sizes for our experiments were determined according to our previous studies and a pilot study concerning subretinal MP injections. The pilot study revealed that severe hemorrhage secondary to subretinal injection interferes with MP clearance and was used as exclusion criteria. Graph Pad Prism 6 (GraphPad Software) was used for data analysis and graphic representation. All

values are reported as mean \pm SEM. Statistical analysis was performed by one-way ANOVA followed by Bonferroni or Dunnett's post-test (multiple comparison) or Mann–Whitney *U*-test (2-group experiments) for comparison among means depending on the experimental design. The *n* and *P*-values are indicated in the figure legends.

Supplementary information for this article is available online: <http://embomolmed.embopress.org>

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Author contributions

OL designed and performed experiments, analyzed data, and helped write the paper; BC, SL, WR, SJH, ED, and MH designed and performed experiments; MP, J-AS, A-PB, and CC collected samples and analyzed data; XG performed experiments, analyzed data, and helped write the paper; and FS designed the study, performed experiments, analyzed data, and wrote the paper.

Conflict of interest

The authors declare that they have no conflict of interest.

References

- Ali K, Middleton M, Pure E, Rader DJ (2005) Apolipoprotein E suppresses the type I inflammatory response in vivo. *Circ Res* 97: 922–927
- Anderson DH, Ozaki S, Nealon M, Neitz J, Mullins RF, Hageman GS, Johnson LV (2001) Local cellular sources of apolipoprotein E in the human retina and retinal pigmented epithelium: implications for the process of drusen formation. *Am J Ophthalmol* 131: 767–781
- Ayrolidi E, Zollo O, Cannarile L, D'Adamo F, Grohmann U, Delfino DV, Riccardi C (1998) Interleukin-6 (IL-6) prevents activation-induced cell death: IL-2-independent inhibition of Fas/fasL expression and cell death. *Blood* 92: 4212–4219
- Azzam KM, Fessler MB (2012) Crosstalk between reverse cholesterol transport and innate immunity. *Trends Endocrinol Metab* 23: 169–178
- Basu SK, Ho YK, Brown MS, Bilheimer DW, Anderson RG, Goldstein JL (1982) Biochemical and genetic studies of the apoprotein E secreted by mouse macrophages and human monocytes. *J Biol Chem* 257: 9788–9795
- Berbee JF, Havekes LM, Rensen PC (2005) Apolipoproteins modulate the inflammatory response to lipopolysaccharide. *J Endotoxin Res* 11: 97–103
- Camelo S, Raoul W, Lavalette S, Calippe B, Cristofaro B, Levy O, Houssier M, Sulpice E, Jonet L, Klein C et al (2012) Delta-like 4 inhibits choroidal neovascularization despite opposing effects on vascular endothelium and macrophages. *Angiogenesis* 15: 609–622
- Chang B, Hurd R, Wang J, Nishina P (2013) Survey of common eye diseases in laboratory mouse strains. *Invest Ophthalmol Vis Sci* 54: 4974–4981
- Chen W, Stambolian D, Edwards AO, Branham KE, Othman M, Jakobsdottir J, Tosakulwong N, Pericak-Vance MA, Campochiaro PA, Klein ML et al (2010) Genetic variants near TIMP3 and high-density lipoprotein-associated loci influence susceptibility to age-related macular degeneration. *Proc Natl Acad Sci USA* 107: 7401–7406.
- Chen M, Luo C, Penalva R, Xu H (2013) Paraquat-induced retinal degeneration is exaggerated in CX3CR1-deficient mice and is associated with increased retinal inflammation. *Invest Ophthalmol Vis Sci* 54: 682–690
- Chow A, Brown BD, Merad M (2011) Studying the mononuclear phagocyte system in the molecular age. *Nat Rev Immunol* 11: 788–798
- Combadiere C, Feumi C, Raoul W, Keller N, Rodero M, Pezard A, Lavalette S, Houssier M, Jonet L, Picard E et al (2007) CX3CR1-dependent subretinal microglia cell accumulation is associated with cardinal features of age-related macular degeneration. *J Clin Invest* 117: 2920–2928
- Cruz-Guilloty F, Saeed AM, Echegaray JJ, Duffort S, Ballmick A, Tan Y, Betancourt M, Viteri E, Ramkellawan GC, Ewald E et al (2013) Infiltration of proinflammatory m1 macrophages into the outer retina precedes damage in a mouse model of age-related macular degeneration. *Int J Inflamm* 2013: 503725
- Dong LM, Weisgraber KH (1996) Human apolipoprotein E4 domain interaction. Arginine 61 and glutamic acid 255 interact to direct the preference for very low density lipoproteins. *J Biol Chem* 271: 19053–19057
- Galea I, Bechmann I, Perry VH (2007) What is immune privilege (not)? *Trends Immunol* 28: 12–18
- Gautier EL, Ivanov S, Lesnik P, Randolph GJ (2013) Local apoptosis mediates clearance of macrophages from resolving inflammation in mice. *Blood* 122: 2714–2722
- Greaney P, Nahimana A, Lagopoulos L, Etter AL, Aubry D, Attinger A, Beltraminelli N, Huni B, Bassi I, Sordat B et al (2006) A Fas agonist induces high levels of apoptosis in haematological malignancies. *Leuk Res* 30: 415–426
- Griffith TS, Brunner T, Fletcher SM, Green DR, Ferguson TA (1995) Fas ligand-induced apoptosis as a mechanism of immune privilege. *Science* 270: 1189–1192
- Guo L, LaDu MJ, Van Eldik LJ (2004) A dual role for apolipoprotein e in neuroinflammation: anti- and pro-inflammatory activity. *J Mol Neurosci* 23: 205–212
- Gupta N, Brown KE, Milam AH (2003) Activated microglia in human retinitis pigmentosa, late-onset retinal degeneration, and age-related macular degeneration. *Exp Eye Res* 76: 463–471
- Hageman GS, Luthert PJ, Victor Chong NH, Johnson LV, Anderson DH, Mullins RF (2001) An integrated hypothesis that considers drusen as biomarkers of immune-mediated processes at the RPE-Bruch's membrane interface in aging and age-related macular degeneration. *Prog Retin Eye Res* 20: 705–732
- Heeren J, Grewal T, Laatsch A, Becker N, Rinninger F, Rye KA, Beisiegel U (2004) Impaired recycling of apolipoprotein E4 is associated with intracellular cholesterol accumulation. *J Biol Chem* 279: 55483–55492
- Houssier M, Raoul W, Lavalette S, Keller N, Guillonneau X, Baragatti B, Jonet L, Jeanny JC, Behar-Cohen F, Coceani F et al (2008) CD36 deficiency leads to choroidal involution via COX2 down-regulation in rodents. *PLoS Med* 5: e39
- Ishida BY, Bailey KR, Duncan KG, Chalkley RJ, Burlingame AL, Kane JP, Schwartz DM (2004) Regulated expression of apolipoprotein E by human retinal pigment epithelial cells. *J Lipid Res* 45: 263–271
- Johnson LV, Forest DL, Banna CD, Radeke CM, Maloney MA, Hu J, Spencer CN, Walker AM, Tsie MS, Bok D et al (2011) Cell culture model that mimics

- drusen formation and triggers complement activation associated with age-related macular degeneration. *Proc Natl Acad Sci USA* 108: 18277–18282
- Kang SM, Schneider DB, Lin Z, Hanahan D, Dichek DA, Stock PG, Baekkeskov S (1997) Fas ligand expression in islets of Langerhans does not confer immune privilege and instead targets them for rapid destruction. *Nat Med* 3: 738–743
- Kezic JM, Chen X, Rakoczy EP, McMenamin PG (2013) The Effects of Age and Cx3cr1 Deficiency on Retinal Microglia in the Ins2Akita Diabetic Mouse. *Invest Ophthalmol Vis Sci* 54: 854–863
- Kiener PA, Davis PM, Starling GC, Mehlin C, Klebanoff SJ, Ledbetter JA, Liles WC (1997) Differential induction of apoptosis by Fas-Fas ligand interactions in human monocytes and macrophages. *J Exp Med* 185: 1511–1516
- Klaver CC, Kliffen M, van Duijn CM, Hofman A, Cruts M, Grobbee DE, van Broeckhoven C, de Jong PT (1998) Genetic association of apolipoprotein E with age-related macular degeneration. *Am J Hum Genet* 63: 200–206
- Klein R, Peto T, Bird A, Vannewkirk MR (2004) The epidemiology of age-related macular degeneration. *Am J Ophthalmol* 137: 486–495
- Klein R, Knudtson MD, Klein BE, Wong TY, Cotch MF, Liu K, Cheng CY, Burke GL, Saad MF, Jacobs DR Jr et al (2008) Inflammation, complement factor h, and age-related macular degeneration: the Multi-ethnic Study of Atherosclerosis. *Ophthalmology* 115: 1742–1749
- Klein R, Myers CE, Cruickshanks KJ, Gangnon RE, Danforth LG, Sivakumaran TA, Iyengar SK, Tsai MY, Klein BE (2014) Markers of inflammation, oxidative stress, and endothelial dysfunction and the 20-year cumulative incidence of early age-related macular degeneration: the Beaver Dam Eye Study. *JAMA Ophthalmol* 132: 446–455
- Kohno H, Chen Y, Kevany BM, Pearlman E, Miyagi M, Maeda T, Palczewski K, Maeda A (2013) Photoreceptor Proteins Initiate Microglial Activation via Toll-like Receptor 4 in Retinal Degeneration Mediated by All-trans-retinal. *J Biol Chem* 288: 15326–15341
- Koss MJ, Hoffmann J, Nguyen N, Pfister M, Mischak H, Mullen W, Husi H, Rejdak R, Koch F, Jankowski J et al (2014) Proteomics of vitreous humor of patients with exudative age-related macular degeneration. *PLoS ONE* 9: e96895
- Linton MF, Atkinson JB, Fazio S (1995) Prevention of atherosclerosis in apolipoprotein E-deficient mice by bone marrow transplantation. *Science* 267: 1034–1037
- Luhmann UF, Lange CA, Robbie S, Munro PM, Cowing JA, Armer HE, Luong V, Carvalho LS, MacLaren RE, Fitzke FW et al (2012) Differential modulation of retinal degeneration by Ccl2 and Cx3cr1 chemokine signalling. *PLoS ONE* 7: e35551
- Ma W, Zhao L, Fontainhas AM, Fariss RN, Wong WT (2009) Microglia in the mouse retina alter the structure and function of retinal pigmented epithelial cells: a potential cellular interaction relevant to AMD. *PLoS ONE* 4: e7945
- Mahley RW, Rall SC Jr (2000) Apolipoprotein E: far more than a lipid transport protein. *Annu Rev Genomics Hum Genet* 1: 507–537
- Malek G, Johnson LV, Mace BE, Saloupis P, Schmechel DE, Rickman DW, Toth CA, Sullivan PM, Bowes Rickman C (2005) Apolipoprotein E allele-dependent pathogenesis: a model for age-related retinal degeneration. *Proc Natl Acad Sci USA* 102: 11900–11905
- Matsuura F, Wang N, Chen W, Jiang XC, Tall AR (2006) HDL from CETP-deficient subjects shows enhanced ability to promote cholesterol efflux from macrophages in an apoE- and ABCG1-dependent pathway. *J Clin Invest* 116: 1435–1442
- McKay GJ, Patterson CC, Chakravarthy U, Dasari S, Klaver CC, Vingerling JR, Ho L, de Jong PT, Fletcher AE, Young IS et al (2011) Evidence of association of APOE with age-related macular degeneration: a pooled analysis of 15 studies. *Hum Mutat* 32: 1407–1416
- Molday RS, Hicks D, Molday L (1987) Peripherin. A rim-specific membrane protein of rod outer segment discs. *Invest Ophthalmol Vis Sci* 28: 50–61
- Mooijaart SP, Berbee JF, van Heemst D, Havekes LM, de Craen AJ, Slagboom PE, Rensen PC, Westendorp RG (2006) ApoE plasma levels and risk of cardiovascular mortality in old age. *PLoS Med* 3: e176
- Nakai M, Kawamata T, Taniguchi T, Maeda K, Tanaka C (1996) Expression of apolipoprotein E mRNA in rat microglia. *Neurosci Lett* 211: 41–44
- Oh H, Takagi H, Takagi C, Suzuma K, Otani A, Ishida K, Matsumura M, Ogura Y, Honda Y (1999) The potential angiogenic role of macrophages in the formation of choroidal neovascular membranes. *Invest Ophthalmol Vis Sci* 40: 1891–1898
- Ong JM, Zorapapel NC, Rich KA, Wagstaff RE, Lambert RW, Rosenberg SE, Moghaddas F, Pirouzmanesh A, Aoki AM, Kenney MC (2001) Effects of cholesterol and apolipoprotein E on retinal abnormalities in ApoE-deficient mice. *Invest Ophthalmol Vis Sci* 42: 1891–1900
- Park DR, Thomsen AR, Frevert CW, Pham U, Skerrett SJ, Kiener PA, Liles WC (2003) Fas (CD95) induces proinflammatory cytokine responses by human monocytes and monocyte-derived macrophages. *J Immunol* 170: 6209–6216
- Penfold PL, Killingsworth MC, Sarks SH (1985) Senile macular degeneration: the involvement of immunocompetent cells. *Graefes Arch Clin Exp Ophthalmol* 23: 69–76
- Peri F, Nusslein-Volhard C (2008) Live imaging of neuronal degradation by microglia reveals a role for v0-ATPase a1 in phagosomal fusion in vivo. *Cell* 133: 916–927
- Ransohoff RM (2009) Chemokines and chemokine receptors: standing at the crossroads of immunobiology and neurobiology. *Immunity* 31: 711–721
- Raoul W, Keller N, Rodero M, Behar-Cohen F, Sennlaub F, Combadiere C (2008) Role of the chemokine receptor CX3CR1 in the mobilization of phagocytic retinal microglial cells. *J Neuroimmunol* 198: 56–61
- Riddell DR, Zhou H, Atchison K, Warwick HK, Atkinson PJ, Jefferson J, Xu L, Aschmies S, Kirksey Y, Hu Y et al (2008) Impact of apolipoprotein E (ApoE) polymorphism on brain ApoE levels. *J Neurosci* 28: 11445–11453
- Rosenfeld ME, Butler S, Ord VA, Lipton BA, Dyer CA, Curtiss LK, Palinski W, Witztum JL (1993) Abundant expression of apoprotein E by macrophages in human and rabbit atherosclerotic lesions. *Arterioscler Thromb* 13: 1382–1389
- Rutar M, Natoli R, Provis JM (2012) Small interfering RNA-mediated suppression of Ccl2 in Muller cells attenuates microglial recruitment and photoreceptor death following retinal degeneration. *J Neuroinflammation* 9: 221
- Sakurai E, Anand A, Ambati BK, van Rooijen N, Ambati J (2003) Macrophage depletion inhibits experimental choroidal neovascularization. *Invest Ophthalmol Vis Sci* 44: 3578–3585
- Sarks SH (1976) Ageing and degeneration in the macular region: a clinico-pathological study. *Br J Ophthalmol* 60: 324–341
- Sather S, Kenyon KD, Lefkowitz JB, Liang X, Varnum BC, Henson PM, Graham DK (2007) A soluble form of the Mer receptor tyrosine kinase inhibits macrophage clearance of apoptotic cells and platelet aggregation. *Blood* 109: 1026–1033
- Schmitz G, Orso E (2002) CD14 signalling in lipid rafts: new ligands and co-receptors. *Curr Opin Lipidol* 13: 513–521
- Seddon JM, George S, Rosner B, Rifai N (2005) Progression of age-related macular degeneration: prospective assessment of C-reactive protein,

- interleukin 6, and other cardiovascular biomarkers. *Arch Ophthalmol* 123: 774–782
- Sennlaub F, Auvynet C, Calippe B, Lavalette S, Poupel L, Hu SJ, Dominguez E, Camelo S, Levy O, Guyon E et al (2013) CCR2(+) monocytes infiltrate atrophic lesions in age-related macular disease and mediate photoreceptor degeneration in experimental subretinal inflammation in Cx3cr1 deficient mice. *EMBO Mol Med* 5: 1775–1793
- Silverman MD, Zamora DO, Pan Y, Texeira PV, Baek SH, Planck SR, Rosenbaum JT (2003) Constitutive and inflammatory mediator-regulated fractalkine expression in human ocular tissues and cultured cells. *Invest Ophthalmol Vis Sci* 44: 1608–1615
- Smoak KA, Aloor JJ, Madenspacher J, Merrick BA, Collins JB, Zhu X, Cavigiolo G, Oda MN, Parks JS, Fessler MB (2010) Myeloid differentiation primary response protein 88 couples reverse cholesterol transport to inflammation. *Cell Metab* 11: 493–502
- Streilein JW, Ma N, Wenkel H, Ng TF, Zamiri P (2002) Immunobiology and privilege of neuronal retina and pigment epithelium transplants. *Vision Res* 42: 487–495
- Suzuki M, Tsujikawa M, Itabe H, Du ZJ, Xie P, Matsumura N, Fu X, Zhang R, Sonoda KH, Egashira K et al (2012) Chronic photo-oxidative stress and subsequent MCP-1 activation as causative factors for age-related macular degeneration. *J Cell Sci* 125: 2407–2415
- Tsutsumi C, Sonoda KH, Egashira K, Qiao H, Hisatomi T, Nakao S, Ishibashi M, Charo IF, Sakamoto T, Murata T et al (2003) The critical role of ocular-infiltrating macrophages in the development of choroidal neovascularization. *J Leukoc Biol* 74: 25–32
- Um HD, Orenstein JM, Wahl SM (1996) Fas mediates apoptosis in human monocytes by a reactive oxygen intermediate dependent pathway. *J Immunol* 156: 3469–3477
- Wenkel H, Streilein JW (2000) Evidence that retinal pigment epithelium functions as an immune-privileged tissue. *Invest Ophthalmol Vis Sci* 41: 3467–3473
- Zhao L, Lin S, Bales KR, Gelfanova V, Koger D, DeLong C, Hale J, Liu F, Hunter JM, Paul SM (2009) Macrophage-mediated degradation of beta-amyloid via an apolipoprotein E isoform-dependent mechanism. *J Neurosci* 29: 3603–3612
- Zieger M, Ahnelt PK, Uhrin P (2014) CX3CL1 (Fractalkine) protein expression in normal and degenerating Mouse retina: in vivo studies. *PLoS One* 9: e106562



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