

# Gap junction signalling is a stress-regulated component of adrenal neuroendocrine stimulus-secretion coupling in vivo.

Michel Desarménien, Carole Jourdan, Bertrand Toutain, Emilie Vessières,  
Sheriar Hormuzdi, Nathalie Guérineau

## ► To cite this version:

Michel Desarménien, Carole Jourdan, Bertrand Toutain, Emilie Vessières, Sheriar Hormuzdi, et al.. Gap junction signalling is a stress-regulated component of adrenal neuroendocrine stimulus-secretion coupling in vivo.. Nature Communications, Nature Publishing Group, 2013, 4, pp.2938. <10.1038/ncomms3938>. <inserm-00924268>

HAL Id: inserm-00924268

<http://www.hal.inserm.fr/inserm-00924268>

Submitted on 23 Jun 2014

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

**Gap junction signaling is a stress-regulated component of adrenal neuroendocrine stimulus-secretion coupling *in vivo***

Michel G. Desarménien<sup>1,2,3\*</sup>, Carole Jourdan<sup>1,2,3\*</sup>, Bertrand Toutain<sup>4,5,6,7</sup>, Emilie Vessières<sup>4,5,6,7</sup>, Sheriar G. Hormuzdi<sup>8</sup> and Nathalie C. Guérineau<sup>4,5,6,7</sup>,

<sup>1</sup>CNRS, UMR-5203, Institut de Génomique Fonctionnelle, Montpellier, F-34094, France;

<sup>2</sup>INSERM, U661, Montpellier, F-34094, France; <sup>3</sup>Universités de Montpellier 1 & 2, UMR-5203,

Montpellier, F-34094, France; <sup>4</sup>Department of Integrated Neurovascular and Mitochondrial

Biology, Angers, France; <sup>5</sup>CNRS UMR6214, Angers, France; <sup>6</sup>INSERM U1083, Angers, France;

<sup>7</sup>University of Angers, Angers, France; <sup>8</sup>Division of Neuroscience, Ninewells Hospital and

Medical School, University of Dundee, Dundee, DD1 9SY, United Kingdom.

\*M.G.D. and C.J. contributed equally to this study.

Correspondence should be addressed to Nathalie C. Guérineau, Laboratoire de Biologie Neurovasculaire et Mitochondriale Intégrée, Rue Haute de Reculée, 49045 Angers CEDEX 01, France. Tel: 33-2-41-73-58-33; Fax: 33-2-41-73-58-95; E-mail: nathalie.guerineau@univ-angers.fr

Key words: *in vivo* excitation-secretion coupling, gap junction blocker, adrenal catecholamine secretion, cold stress, Cx36-deficient mouse.

## **Abstract**

Elucidating the mechanisms whereby neuroendocrine tissues coordinate their input and output signals to ensure appropriate hormone secretion is currently a topical issue. In particular, whether a direct communication mediated by gap junctions between neurosecretory cells contributes to hormone release *in vivo* still remains unknown. Here we address this issue using a microsurgical approach allowing combined monitoring of adrenal catecholamine secretion and splanchnic nerve stimulation in anesthetized mice. Pharmacological blockade of adrenal gap junctions by the uncoupling agent carbenoxolone reduces nerve stimulation-evoked catecholamine release in control mice and to a larger extent in stressed mice. In parallel, the gap junction-coupled cell network is extended in stressed mice. Altogether, this argues for a significant contribution of adrenomedullary gap junctions to catecholamine secretion *in vivo*. As such, gap junctional signaling appears to be a substantial component for neuroendocrine function in the adrenal medulla, as it may represent an additional lever regulating hormone release.

## **Introduction**

Hormone secretion from neuroendocrine tissues is a highly regulated process that needs a finely tuned coordination between incoming and output signals. In this regard, adrenal chromaffin cells are remarkable and the study of their secretory machinery<sup>1,2</sup> and more recently of their synaptic inputs<sup>3-5</sup> has contributed substantially to the current knowledge on cellular mechanisms underlying neurosecretion function. However, compelling evidence obtained in acute adrenal slices indicate that neurosecretory mechanisms are more intricate than previously envisioned. Indeed, catecholamine release appears to be dualistically controlled by an incoming initial command assisted by local modulation<sup>6,7</sup>, supporting the hypothesis of an amplifying system occurring at the chromaffin cell level<sup>8</sup>. The command triggering catecholamine secretion comes from the sympathetic nervous system synapsing onto chromaffin cells. Under basal condition (low splanchnic nerve frequency discharge), acetylcholine (ACh) is the primary neurotransmitter released at the splanchnic nerve terminal-chromaffin synapse. In response to stress or emergency conditions associated with a high nerve frequency firing, a dual splanchnic-adrenal neurotransmission involving both ACh and the pituitary adenylate cyclase-activating polypeptide PACAP occurs<sup>9,10</sup>, thus enhancing chromaffin cell secretion.

In complement to synaptic transmission, direct communication through gap junctions between chromaffin cells ensures propagation of electrical and ensuing calcium signals between neurosecretory cells and amplifies catecholamine release in response to stimulation of a single cell<sup>11</sup>. The differential regulation of gap junctions and cholinergic synapses cells in physiological and pathological conditions associated with specific catecholamine demand, such as during early postnatal development<sup>4,12</sup> and in response to chronic stress<sup>13,14</sup>, is indicative of a functional interaction between incoming commands and local communication and points towards a putative contribution of gap junctions to stimulus-secretion coupling. Converging

arguments have been accumulated that strongly support this hypothesis but the *ex vivo* acute slice preparation used to obtain them does not allow a convincing demonstration.

We present here a new experimental approach to simultaneously monitor the electrical activity of chromaffin cells and catecholamine secretion in response to splanchnic nerve stimulations in anesthetized adult mice. This *in vivo* technique is combined with a series of histological and molecular analysis of gap junction protein connexins and of dye diffusion through gap junctions in control, cold stressed and connexin-deficient animals. We demonstrate that adrenal gap junctional signaling contributes positively to catecholamine release in response to splanchnic nerve stimulation *in vivo*. Furthermore, this contribution is enhanced in stressed animals, providing an additional level of regulation for stimulation-secretion coupling in the adrenal gland.

## **Results**

**Electrical activity in the mouse adrenal gland *in vivo*.** To determine the contribution of gap junctions to stimulus-secretion coupling in anesthetized mice, blood from the left adrenal gland was collected during electrical stimulation of the splanchnic nerve (Figs. 1A and S1; Supplementary Movie 1). The electrical behaviour of the adrenal medullary tissue was monitored using a microelectrode implanted into the adrenal medulla (location identified by injection of Chicago Sky Blue 6B; Supplementary Fig. S2). A spontaneous electrical activity was observed (Fig. 1B, recording duration: 9-22 min, mean frequency  $0.87 \pm 0.26$  Hz (mean  $\pm$  s.e.m.,  $n = 9$  mice)). It consisted of a continuous firing, on which intermittent bursts were superposed in 7/9 mice (1-8 min duration, intra-burst frequency  $2.7 \pm 1.1$  Hz (mean  $\pm$  s.e.m.)). Tetrodotoxin (TTX, 3  $\mu$ M) applied locally on the splanchnic nerve induced a marked ( $89 \pm 3.5\%$  (mean  $\pm$  s.e.m.,  $n = 9$  mice)) and partially reversible decrease of the firing activity (Fig. 1C), indicating that the *in*

*in vivo* spontaneous firing of the mouse adrenal medulla mainly results from excitatory afferent inputs.

Splanchnic nerve stimulation (10 V, 160 ms) triggered a burst of bipolar action potentials occurring specifically during the stimulation (Fig. 1D, upper traces). In the 7 mice recorded, application of 3  $\mu$ M TTX onto the nerve significantly decreased the evoked activity (Fig. 1D, middle traces), as illustrated by the decreased variance of the signal. This unambiguously shows that evoked electrical activity in the medullary tissue results from action potentials running along the splanchnic nerve, and not from a direct activation of the gland. In addition, this activity was reversibly inhibited (by  $78 \pm 12\%$  (mean  $\pm$  s.e.m.,  $n = 7$  mice)), by a local injection of hexamethonium, an antagonist of  $\alpha$ 3-built nAChRs, through the recording pipette (200  $\mu$ M, Fig. 1E). This demonstrates i) the involvement of cholinergic synaptic terminals, as expected for splanchnic nerve-evoked activity and ii) the dominant contribution of nAChRs rather than muscarinic receptors.

**Basal and nerve stimulation-evoked catecholamine release.** To go further in characterizing excitation-secretion coupling in the adrenal medulla *in vivo*, we measured catecholamine secretion, at rest and in response to splanchnic nerve stimulation (Fig. 1A for a schematic illustration of the protocol, Supplementary Fig. S1 and Supplementary Movie 1 for direct visualization). Basal plasma norepinephrine (NE) and epinephrine (E) were assayed in blood samples collected from the left adrenal vein during 2 or 5 min. No statistical difference was observed between catecholamine concentrations measured using the two sampling durations (NE:  $15.4 \pm 2.9$  nM (mean  $\pm$  s.e.m.,  $n = 11$  mice) for 2 min and  $25.1 \pm 5.2$  nM (mean  $\pm$  s.e.m.,  $n = 17$  mice) for 5 min,  $p > 0.05$ , Mann-Whitney *U* test; E:  $318 \pm 111$  nM (mean  $\pm$  s.e.m.,  $n = 11$  mice) for 2 min and  $384 \pm 177$  nM (mean  $\pm$  s.e.m.,  $n = 17$  mice) for 5 min,  $p > 0.05$ , Mann-Whitney *U* test). We next examined splanchnic nerve stimulation-evoked catecholamine

secretion. Two protocols were used: a low frequency (LF, 0.1 Hz, 5 min) to mimic basal tone firing and a high frequency (HF, 4 Hz, 2 min) to mimic stress firing conditions<sup>15</sup>. Two blood samples were successively collected, a control one first and then a test one during nerve stimulation. In the absence of stimulation, NE and E concentrations in the two successive samples did not significantly differ (NE: C =  $36.6 \pm 8.5$  nM (mean  $\pm$  s.e.m.) and T =  $55.7 \pm 22.7$  nM (mean  $\pm$  s.e.m., n = 12 mice),  $p > 0.05$ , Mann-Whitney *U* test; E: C =  $459 \pm 241$  nM (mean  $\pm$  s.e.m.) and T =  $500 \pm 297$  nM (mean  $\pm$  s.e.m., n = 12 mice),  $p > 0.05$ , Mann-Whitney *U* test). Both LF and HF splanchnic nerve stimulations triggered a frequency-dependent release of catecholamines (>2-fold and >10-fold for NE and E, respectively, Fig. 2). It is noteworthy that not only basal, but also evoked E release is much larger than NE release. This is consistent with the fact that mouse chromaffin cells mainly exhibit an adrenergic phenotype (80% of epinephrine-containing cells<sup>16</sup>).

**Contribution of gap junctions to catecholamine release.** The mouse adrenal medullary tissue was much more immunoreactive to Cx36 than to Cx43 (Fig. 3A). The opposite situation was observed in the cortex. Gap junction involvement in nerve stimulation-evoked catecholamine release was determined by blocking junctional coupling. Mice were injected *i.p.* at the beginning of the surgical procedure with either NaCl (9%), the uncoupling agent carbenoxolone (CBX, 100 mg/kg) or its inactive structural analog glycyrrhizic acid (GZA, 135 mg/kg). CBX injection significantly reduced gap junctional coupling in the adrenal gland, as evidenced by decreased diffusion of the fluorescent dye Lucifer yellow (LY) measured by scrape-loading<sup>13</sup> (Fig. 3B). In mice injected with GZA (n = 11) or NaCl (n = 8), LY widely diffused from the cut edge of the gland into the cortex and medulla, while in mice injected with CBX (n = 5), diffusion was greatly decreased and LY accumulated on the cut edge of the medulla (Fig. 3B). Quantification of LY diffusion into the gland (Figs. 3C and 3D) indicates a reduced (25.8% decrease) diffusion

in the depth of the gland in CBX-treated mice. Similarly, CBX treatment (5 mice) reduced by 30.5% Alexa Fluor 594®-conjugated cadaverine diffusion when compared to GZA (n = 7 mice,  $p < 0.05$ , Mann-Whitney  $U$  test, Fig. 3D and Supplementary Fig. S3). These findings unambiguously show the presence of functional gap junctions in the mouse adrenal medullary tissue and demonstrate the efficiency of CBX to block gap junctional coupling in our experimental conditions.

To go further, we performed similar scrape-loading experiments using mefloquine (MFQ), another gap junction uncoupling agent potently active on Cx36-built gap junction channels<sup>17</sup>. When compared to vehicle (10 glands from 5 mice), MFQ (25 mg/kg) significantly reduced (by 22.7%) LY diffusion throughout the adrenal gland (n = 14 glands from 7 mice,  $p < 0.05$ , Mann-Whitney  $U$  test, Supplementary Fig. S4A-C). It is of note that this percentage is comparable to that obtained in CBX-treated mice (Fig. 3D).

Is catecholamine secretion (at rest and in response to splanchnic nerve stimulation) altered when gap junctional communication is reduced? First, basal catecholamine release is significantly higher in mice treated with GZA, CBX, MFQ or its vehicle than in NaCl-injected mice (Supplementary Fig. S5). However, MFQ by itself did not significantly enhance basal catecholamine concentration when compared to its vehicle. Because GZA and CBX have been previously described as hypertensinogenic compounds when infused intra-cerebroventricularly<sup>18</sup>, we investigated whether GZA, CBX and MFQ were able to modify arterial blood pressure under our experimental conditions. Systolic arterial blood pressure (SBP) was measured by tail-cuff plethysmography in anesthetized mice, either not injected (sham) or injected with NaCl, GZA, CBX, vehicle or MFQ. As illustrated in Supplementary Fig. S6, neither CBX nor GZA increased SBP in our conditions. Conversely, when compared to control values measured before drug injection, CBX, GZA and MFQ decreased SBP, by significant values (Mann-Whitney  $U$  test) for CBX (24.6%) and MFQ (27.4%). This suggests that the increased basal catecholamine secretion



measured in the adrenal-venous blood in response to GZA, CBX and MFQ might be a functionally adaptive response to counteract a hypotension-induced stress. It is also of note that none of the injected compounds altered cardiac frequency during the recording period (data not shown).

The contribution of gap junctions to splanchnic nerve stimulation-evoked catecholamine release was addressed by comparing the T/C ratio in CBX- and GZA-injected mice. Indeed, since GZA and CBX affected basal release to the same extent, the action of CBX on nerve stimulation-evoked release was measured by comparison with that of GZA. As illustrated in Fig. 4, CBX decreased both NE and E release evoked by a LF (Fig. 4A) or HF (Fig. 4B) stimulation, although a statistically significant difference was found only for NE release triggered by HF stimulation (58% decrease,  $n = 9$  GZA- and 8 CBX-treated mice,  $p < 0.05$ , Mann-Whitney  $U$  test). Similarly, MFQ trended to decrease HF stimulation-evoked norepinephrine release (47% decrease,  $n = 5$  vehicle- and 9 MFQ-injected mice,  $p > 0.05$ , Mann-Whitney  $U$  test) without affecting epinephrine release (Supplementary Fig. S4D). This finding demonstrates that gap junction-mediated intercellular communication in the adrenal gland has a notable contribution to hormone release, at least NE, *in vivo*.

**Increased contribution of gap junctions in stressed mice.** We next investigated the effect of CBX in a condition in which gap junctions are increased. The procedure was identical as in our previous work<sup>13,14</sup>, in which we have demonstrated that a chronic systemic stress (cold exposure) induces a persistent increase in gap junctional coupling between chromaffin cells and up-regulates both Cx36 and Cx43 expression in rat. Since the effects of chronic stress may last several hours or days after the end of the stressful stimulus, they can be studied in an animal sacrificed or anaesthetized immediately after return to normal conditions. As illustrated in figure 5A, Cx36 immunostaining in the adrenal medulla was 2-fold higher in cold stressed ( $n = 6$  mice)

than in control (n = 5) mice ( $p < 0.05$ , Mann-Whitney  $U$  test). It is noteworthy that expression of Cx43, dominant in the cortex, was not changed by stress (Supplementary Fig. S7). LY diffusion was significantly reduced in CBX- as compared to GZA-injected mice (Fig. 5B and associated wireframe plots). This reduction (51.2%, Figs. 5C and 5D) is greater than in control mice (Fig. 3D). Altogether, these data are consistent with a robust increase of medullary gap junction-mediated cell-to-cell communication in cold stressed mice. Basal plasma NE and E concentrations were surprisingly lower in cold stressed mice than in unstressed mice (Supplementary Fig. S5). However, since these basal concentrations remained similar for CBX- and GZA-treatments, contribution of gap junctions to stimulus-secretion coupling could be evaluated by comparing nerve-evoked release after these two treatments. As illustrated in figure 6, CBX decreased HF stimulation-evoked NE release to a larger extent in cold stressed than in control mice (reduction by 76% in stressed mice, n = 11 animals, as compared to 58% in control mice (n = 11),  $p < 0.05$ , Mann-Whitney  $U$  test). In addition, while CBX did not significantly affect E release in control mice (21% reduction), it reduced dramatically E secretion in cold stressed mice (75% reduction, n = 11 GZA- and 11 CBX-injected animals,  $p < 0.05$ , Mann-Whitney  $U$  test). All together, data obtained in control and cold stressed mice demonstrate i) the contribution of adrenal medullary gap junctions to splanchnic nerve stimulation-evoked catecholamine secretion *in vivo* and ii) their predominant involvement in stressful situations.

**Impaired nerve stimulation-evoked secretion in Cx36<sup>-/-</sup> mice.** Stimulus-secretion coupling was also investigated in Cx36 deficient (Cx36<sup>-/-</sup>) mice. As expected, the transcript encoding Cx36 was robustly expressed in wild-type mice but not detected in Cx36<sup>-/-</sup> mice, while Cx43 mRNA expression was similar in wild-type and Cx36<sup>-/-</sup> mice (Fig. 7A). Interestingly, the expression of mRNA encoding Cx29, a connexin expressed in S100-positive medullary cells<sup>6</sup> was significantly enhanced in Cx36<sup>-/-</sup> mice. Expression of Cx36 and Cx43 proteins was also

assessed by immunohistofluorescence (Fig. 7B). In glands from Cx36<sup>-/-</sup> mice, Cx36 was detected neither in the cortex nor in the medulla. Cx43 was not detected in the medulla but was expressed in the cortex, at a level comparable to that in control mice ( $6.7 \pm 0.5$  AU (mean  $\pm$  s.e.m.,  $n = 11$  glands from 6 mice) for Cx36<sup>-/-</sup> and  $7.5 \pm 1.1$  AU (mean  $\pm$  s.e.m.,  $n = 11$  glands from 6 mice) for wild type,  $p > 0.05$ , unpaired Student's  $t$  test).

Scrape-loading experiments show an accumulation of LY at the cut-edge of the gland and equal diffusion of the dye within the gland of GZA- and CBX-treated Cx36<sup>-/-</sup> mice (Fig. 7C,  $n = 3$  GZA- and 4 CBX-injected mice). These data are consistent with a lack of LY-permeable gap junctions in the medulla of Cx36<sup>-/-</sup> mice and strengthen a dominant expression of Cx36 in the mouse adrenal medulla.

Basal catecholamine release was comparable between CBX- and GZA-treated Cx36<sup>-/-</sup> mice and similar to the basal release in control mice (Supplementary Fig. S5). However, counter to expectations, HF nerve stimulation did not evoke any significant catecholamine release in Cx36<sup>-/-</sup> mice, whatever the treatment with GZA or CBX (Fig. 8). To try and understand the failure of nerve stimulation to evoke catecholamine release, we tested for integrity of nerve endings and synapses. Nerve terminals were visualized with antibodies against heavy neurofilaments (NF, 200 kDa) to label nerve fibers and against the vesicular acetylcholine transporter (VACHT) to detect synaptic vesicles. As illustrated in Fig. 9A, both NF and VACHT were highly expressed in the control adrenal medulla, decorating the space between lobules and intermingled between individual chromaffin cells within lobules. The staining intensity was significantly attenuated in Cx36<sup>-/-</sup> mice ( $n = 21$  glands from 11 animals,  $p < 0.05$  versus WT,  $n = 22$  glands from 11 mice, unpaired Student's  $t$  test), suggesting an impairment of cholinergic presynaptic terminal function. To further address whether deficits also occurred postsynaptically, expression of mRNA encoding nAChR subunits was evaluated by real-time RT-PCR. None of the 5 tested

subunits ( $\alpha 3$ ,  $\alpha 7$ ,  $\alpha 9$ ,  $\beta 2$  and  $\beta 4$ ) was differentially expressed in Cx36<sup>-/-</sup> mice when compared to wild-type mice (Fig. 9B).

## Discussion

Our study reports *in vivo* basal and splanchnic nerve stimulation-evoked catecholamine secretion from the adrenal gland in anesthetized mouse. In addition, using a combination of *in vivo* monitoring of adrenal catecholamine secretion and splanchnic nerve stimulation, we shed light on the functional relevance of gap junctions in the adrenal medullary tissue. It becomes now unquestionable that gap junctions contribute positively to nerve-evoked catecholamine release.

A striking observation is that the spontaneous electrical activity in the adrenal medullary tissue *in vivo* was sporadic and bursting, contrasting with the high frequency firing observed *ex vivo*<sup>13,19</sup> or in cultured chromaffin cells<sup>20-22</sup>. Although our *in vivo* conditions were designed to minimally disrupt adrenal function, we cannot rule out the possibility that the firing pattern recorded in an anesthetized animal is attenuated. The bursting pattern is likely to be physiologically relevant since catecholamine release from the adrenal is known to be optimized when nerve impulses are delivered in bursts rather than continuously<sup>23,24</sup>. More importantly, we observed that electrical activity was suppressed when action potential propagation along the splanchnic nerve was prevented by TTX. This indicates a predominant nerve-driven activity and a sparse spontaneous electrical firing of chromaffin cells. This finding strengthens the conclusion that chromaffin cell firing discharge is prevalingly paced by synaptic inputs. However, it contrasts with observations made *in vitro*<sup>25-27</sup> and *ex vivo*<sup>13,19</sup>, in which adrenal chromaffin cells were disconnected from their sympathetic inputs and exhibited a sustained autorhythmic activity<sup>20</sup> supporting robust spontaneous action potential firing. The TTX-sensitivity of *in vivo* spontaneous activity is in agreement with Petrovic et al.<sup>28</sup>, who demonstrated that spontaneous catecholamine release in an adrenal medulla slice is dramatically decreased by hexamethonium, indicating that release does

not result from spontaneous chromaffin cells activity but rather from the commonly observed spontaneous activation of synaptic inputs<sup>4,5,12</sup>. The electrical chromaffin cell response to nerve-evoked stimulation was also robustly reduced by hexamethonium, supporting a dominant contribution of nAChRs over muscarinic receptors. This corroborates previous findings in mouse acute slices reporting an inhibition of catecholamine release by hexamethonium<sup>28,29</sup>. However, it contrasts with a study performed in hemisectioned mouse adrenal, in which catecholamine release evoked by acetylcholine mainly depended on muscarinic receptors<sup>19</sup>.

In previous *in vivo* measurements of basal and stimulated adrenal catecholamine release, E and NE were alternatively assayed from adrenal-venous blood or from the general blood circulation. In addition, the specie, collection technique and stimulation protocols varied from one study to the other, thus precluding trivial quantitative comparison. Basal and splanchnic nerve stimulation-evoked release of adrenal catecholamines has been directly measured in adrenal venous blood only in a few cases, in the cat<sup>30,31</sup>, rat<sup>15,23,32,33</sup>, and dog<sup>34,35</sup>, but not in the mouse. Our *in vivo* data indicate a basal E:NE ratio close to 10:1 in mouse. This is consistent with previously reported ratios ranging from 2 to 7<sup>15,31, 33-35</sup>. It is noteworthy that it is the opposite when catecholamines are assayed in the plasma, in anesthetized animals<sup>30,33,35,36</sup>, including mouse<sup>37-40</sup>. In response to electrical splanchnic nerve stimulation (0.1 and 4 Hz), adrenal catecholamine output significantly increased with stimulation frequency. In particular, HF stimulation vigorously enhanced the release of both E and NE, as reported in other species<sup>15,23,24,30,34</sup>. E and NE can be differentially released from the adrenal, depending on splanchnic nerve stimulation frequency. While NE is the main hormone secreted during LF stimulation (1-5 Hz) in cat<sup>31,41</sup>, our results in mouse are in favor of a preferential E secretion at both LF and HF stimulation. The same is observed in rat<sup>15</sup> and dog<sup>35</sup>. The stimulation frequency-dependent increase in catecholamines is a robust event, since it also occurs in acute adrenal slices<sup>42</sup>. In cold stressed mice, basal catecholamine concentration in adrenal-venous blood was

unexpectedly lower than in control mice. This mismatches with previous data reporting a rise in catecholamine concentration in the adrenal vein in response to various stressors<sup>33,36,43</sup>. This also contrasts with the typical *in vivo* increase in catecholamine concentrations measured in plasma, in response to stressful situations<sup>33,36,40,44</sup>. However, the situation in the adrenal-venous blood appears more intricate, such that a robust catecholamine increase detected in plasma is not always associated with equally vigorous catecholamine amounts flowed from the adrenal<sup>44</sup>. The low basal catecholamine concentration we found here was unlikely due to depletion of adrenal catecholamine resources, since the gland further secreted in response to nerve stimulation. Alternatively, the anesthesia and surgical procedure may have uncontrolled consequences on the basal release. Although ketamine does not modify basal catecholamine secretion<sup>45</sup>, a direct measurement of plasma catecholamines, in a freely moving mouse submitted or not to various stress paradigms, will be necessary to definitely answer this question.

Even though a direct cell-cell communication through gap junctions has been described in many endocrine/neuroendocrine tissues<sup>46-49</sup>, including the adrenal medulla<sup>6,7,11</sup>, its contribution to hormone secretion *in vivo* has been unambiguously established so far only for insulin<sup>50,51</sup> and renin<sup>49,52</sup>. Very recent studies performed in the anterior pituitary<sup>53,54</sup> strongly support a crucial role for gap junctions in endocrine tissue output, suggesting that the plasticity of gap junction-coupled endocrine cell network is a key element underlying functional adaptation of hormone release at the population level. More remarkable is the observation that functional adaptation to specific hormonal demand persists within the coupled network<sup>55</sup>. If transposed to the chromaffin cell network, this could be particularly relevant in terms of functional tissue adaptation in response to repeated stressful conditions. Regarding the adrenal gland, contribution of gap junctions to stimulation-secretion coupling was previously inferred from *in situ* experiments in rodent adrenal acute slices<sup>11,56</sup>. Conversely, Petrovic et al.<sup>28</sup> reported that gap junctions do not contribute to catecholamine release evoked by localized electrical stimulation. However, the

mechanisms engaged when a bipolar electrode is positioned on top of a slice are probably hardly comparable to those involved in the response to distant stimulation of the splanchnic nerve and propagation of action potentials along sympathetic axons. Our *in vivo* observation that CBX or MFQ decreases nerve stimulation-evoked catecholamine release in conditions in which they prevent intercellular diffusion of fluorescent dyes within the adrenal gland, is a strong indication for a role of gap junctions in catecholamine release. Furthermore, this finding is reinforced by demonstration of a more robust effect of CBX on catecholamine secretion in cold stressed mice, in which the medullary gap junctional communication is up-regulated. Unfortunately, in the reverse experiment in which Cx36-built gap junctions were reduced by genetic manipulation, nerve-evoked release was reduced so much that the contribution of gap junctions could not be reliably evaluated. It is noticeable that gap junctions contribute chiefly to NE release in control animals, and to both E and NE release in stressed animals. The observation that CBX decreased the release evoked by high, but not low frequency stimulation, suggests a dominant contribution of connexin-dependent signaling in emergency conditions when the hormonal demand is high. As previously reported<sup>6</sup>, our data identified Cx36 as the ascendant connexin expressed in the mouse medullary tissue and documented the expression of Cx29 transcript. At least two findings support a role of gap junctional communication in catecholamine secretion in mouse. First, we show here that Cx36 and Cx29 are up-regulated after an episode of cold stress, as reported for Cx36 and Cx43 in rat<sup>14</sup>. Second, PACAP, a splanchnic-derived peptide transmitter which is released during elevated nerve firing<sup>10</sup>, enhances gap junction-mediated electrical coupling in the mouse adrenal medulla<sup>57</sup>.

The most surprising result concerns the considerable decrease of nerve stimulation-evoked catecholamine release in the adrenal medulla of Cx36<sup>-/-</sup> mice. The underlying default likely occurs presynaptically, as suggested by the substantial reduction of both NF and VAcHT immunolabeling. Although connexin-dependent signaling and chemical synaptic

neurotransmission interact with each other in various neuronal tissues<sup>58</sup>, including in the adrenal medulla<sup>6,7,59</sup>, our finding reveals an unanticipated role for Cx36 at the splanchnic nerve-chromaffin cell synapse. During post-natal development, down-regulation of adrenal gap junctional communication parallels the acquisition of the neurogenic control of catecholamine secretion<sup>4,12</sup>. In adults, gap junctional communication is under a tonic inhibitory control by cholinergic neurotransmission<sup>5,12</sup>. In view of this mutual regulation, one would have expected the lack of gap junctions in Cx36<sup>-/-</sup> mice to help implementation of cholinergic nerve fibres rather than to prevent it. The impairment of nerve stimulation-evoked catecholamine release and the decreased expression of NF and VAcHT are strong and correlative indications of a degraded innervation of chromaffin cells in Cx36<sup>-/-</sup> mice. Defective propagation of signals generated by sympathetic nerve stimulation has been also observed in Cx32 deficient mice<sup>60</sup>. More recently, alterations in physiological properties of neurons *via* early postnatal blockade of gap junctional communication have been reported, thus shoring up the idea that Cx36 deletion may cause changes in presynapses<sup>61</sup>. This opens intriguing questions as to the inter-relationship between gap junctions and synapses, especially during the early phases of neuronal development.

Altogether, the present study ultimately demonstrates that gap junctions are indeed positively contributing to adrenal excitation-secretion coupling *in vivo*. The contribution of connexin-dependent signaling seems particularly decisive when the organism demand in catecholamines increases. Our study substantiates previous data showing that, in the adrenal medulla, synaptic transmission and gap junctional communication act in synergy to ensure a secretory response appropriate to the organism need. In addition to being the first report demonstrating a regulation of gap junction-mediated cell-to-cell communication in the adrenal medullary tissue *in vivo*, our study sheds light on the contributions of gap junctions to neurosecretory function. Importantly, our findings might be extended to other neuroendocrine tissues. Indeed, its ubiquitous distribution merits that gap junction-mediated coupling be



revisited in most neurosecretory tissues since our study suggests that it is a defining characteristic of endocrine/neuroendocrine organ function *in vivo*.

## **Material and Methods**

**Animal preparation and surgery.** All procedures conformed to the animal welfare guidelines of the European Community and were approved by the French Ministry of Agriculture (authorizations No. 34.128 and 49.247). Experiments were performed on 2 month-old male C57BL/6J and Cx36-deficient mice<sup>62</sup>, acclimatised (12h light-dark cycle, 22°C) for a minimum of 7 days before use. For *in vivo* experiments, mice were anesthetized with ketamine (100 mg/kg) and xylazine (10 mg/kg), *i.p.* in 160  $\mu$ l NaCl (9‰). The surgery started with a laparotomy; visceral organs were gently shifted on the right side of the animal to uncover the left adrenal gland. The renal vein and the conjunctive tissue containing the splanchnic nerve were isolated. The splanchnic nerve was placed on a home-made bipolar stimulation electrode. The renal vein was ligated on both the kidney and cava vein sides and then cannulated with a heparinised (20 U/ml) catheter in order to form a reservoir collecting the blood flowing from the left adrenal (Supplementary Fig. S1 and Supplementary Movie 1).

**Blood collection and catecholamine assay.** Twenty minutes after surgery, two blood samples were successively collected during 2 or 5 min each, depending on the stimulation protocol, one control first (C) and then a test one (T) during stimulation. Stimulation-evoked catecholamine release was expressed as the T/C ratio. Samples were centrifugated at 4°C (7000 g, 10 min) and stored at -20°C until use. Catecholamines (epinephrine and norepinephrine) were assayed from 7  $\mu$ l of plasma using a 2-CAT ELISA KIT (Mediane Diagnostics, Plaisir, France), as reported<sup>63</sup>.

**Arterial blood pressure measurement.** Systolic blood pressure (SBP) was measured by tail-cuff plethysmography (BP-2000 blood pressure analysis system, Visitech Systems, Bioseb, Chaville, France) in ketamine/xylazine anesthetized mice. Mice were introduced into a restraint

device with the blood pressure measure cuff placed on the base of the tail. The thermal heating pad underneath was set to 30°C. SBP and cardiac frequency were monitored every minute for 60 min (15 min as a control period before injection and up to 40-45 min after injection as the test period), to reliably compare the results with adrenal-venous blood catecholamine levels assayed 40-50 min after drug injection).

**Electrophysiology.** Square electrical pulses (10 V, 160 ms) were applied to the nerve at low (0.1 Hz, 5 min) or high frequency (4 Hz, 2 min). To block nerve conduction, a tetrodotoxin (TTX)-soaked cotton bud (3  $\mu$ M) was applied on the nerve, as far as possible (2-3 mm) from the gland. Extracellular recordings were performed with a glass microelectrode (in mM: 150 NaCl, 12.5 KCl, 10 HEPES, pH 7.4). The recording site was located by injection of Chicago Sky Blue 6B (2%, Supplementary Fig. S2). In some experiments, the recording electrode contained hexamethonium (200  $\mu$ M) to impair cholinergic neurotransmission.

***In vivo* blockade of gap junctions.** Mice were injected *i.p.* (120  $\mu$ l NaCl 9‰) before the surgical procedure with either carbenoxolone<sup>64</sup> (CBX, 100 mg/kg) or its inactive analog glycyrrhizic acid<sup>65</sup> (GZA, 135 mg/kg). In some experiments, mice were injected with the Cx36 uncoupling agent mefloquine<sup>17</sup> (MFQ, 25 mg/kg in 75  $\mu$ l DMSO + H<sub>2</sub>O, 75/25 v/v) or with the vehicle (DMSO + H<sub>2</sub>O, 75/25 v/v).

**Scrape-loading dye transfer.** *In vivo* surgery and pharmacological treatments were identical to those described above. Excised glands were included in agarose, cooled on ice, hemisectioned and incubated for 45 s either in Lucifer yellow (LY, lithium salt; 0.2% in NaCl 9‰, charge -2/MW 457) or Cadaverine (Alexa Fluor 594®-conjugated, 50  $\mu$ M in NaCl 9‰, charge +1/MW 807)<sup>66</sup>. Hemi-glands were immediately fixed in PFA (4% in 0.1 M phosphate buffer) and stored at 4°C. Hemi-glands were sliced (50  $\mu$ m) and imaged with an epifluorescence microscope and the ImageJ software. Average light intensity was measured in the adrenal medulla, along an axis perpendicular to the cutting edge (distance = 0 at the edge and increasing with penetration into

the medulla), leading to a “fluorescence *versus* distance” curve. Results were further expressed as the area under the curve for statistical analysis.

**Connexin immunolabeling.** To process for connexin immuno-staining, adrenal glands were rapidly removed from an anesthetized mouse, embedded in Tissue-Tek<sup>®</sup> O.C.T<sup>™</sup> compound and frozen by immersion in isopentane. The glands were next cut into 20 µm-thick sections using a cryostat. Gland sections were mounted on coverslips, post-fixed at -20°C for 10 min in acetone, and incubated overnight with a rabbit anti-Cx36 or a rabbit anti-Cx43 (1:250; Zymed Laboratories, CliniSciences, Montrouge, France, Cat N°364600 and 483000) polyclonal antibody. Sections were then rinsed and incubated for 2 h at room temperature with a rabbit secondary antibody conjugated to Cy3 (1:2000; Jackson ImmunoResearch Laboratories, Inc., West Grove, PA, Cat N°711-166-152). Primary and secondary antibodies were diluted in PBS containing 2% BSA and 0.1% Triton X-100. Negative controls were performed by omitting primary antibodies. Stained sections were imaged with an epifluorescence microscope (Axioimager, Zeiss, x20 objective). The fluorescence was quantified with Volocity<sup>®</sup> software (Perkin Elmer, Courtaboeuf, France), the total surface of fluorescent areas positive for connexins (number of fluorescent pixels) was divided by the surface (total number of pixels) of the gland, and expressed as a percentage.

**Vesicular ACh transporter and neurofilament immunolabeling.** Adrenal glands were rapidly removed from anesthetized mice, immediately fixed in PFA (4% in 0.1 M phosphate buffer, 2 h, room temperature) and then stored at 4°C until processing. Glands were then sliced (50 µm thickness) using a vibratome. Slices were first incubated in mouse FAB IgG (1:200, Eurobio-AbCys, Les Ulis, France, Cat N°BI 3013C) for 2 h at room temperature and then in a rabbit anti-VACht (1:500, Synaptic systems, Goettingen, Germany, Cat N°130 103) polyclonal antibody and a mouse anti-neurofilament 200 (clone N52, 1:500, Sigma, Cat N°NO142) monoclonal antibody, for 48 h at 4°C. Slices were then rinsed and incubated for 2 h at room temperature with

a rabbit secondary antibody conjugated to Cy3 and a mouse secondary antibody conjugated to Cy5 (both at 1:2000; Jackson ImmunoResearch Laboratories, Inc., West Grove, PA, Cat N° 711-166-152 and 711-496-152). Primary and secondary antibodies were diluted in PBS containing 2% BSA and 0.1% Triton X-100. Negative controls were performed by omitting primary antibodies. Images acquisition was performed using a LSM 510 Meta confocal microscope (Zeiss, Le Pecq, France) and quantification with the image J software.

**Quantification of mRNA expression by real-time RT-PCR.** Total RNA was extracted from macrodissected mouse adrenal medulla using the RNeasy<sup>®</sup> Micro extraction kit (Qiagen, Courtaboeuf, France). RNA (200 ng) was first reverse transcribed using the QuantiTect<sup>®</sup> Reverse Transcription kit (Qiagen) in a final volume of 10 µl. Real-time PCR analyses of the target genes *Gjc3*, *Gjd2*, *Gja1*, *Chrna3*, *Chrna7*, *Chrna9*, *Chrnb2*, *Chrnb4* and the reference genes *Gusb*, *Hprt* and *Gapdh* were performed using Sybr<sup>®</sup> Green PCR master mix (Applied Biosystems, Foster City, CA) with 1:60 of the reverse-transcription reaction, and were carried out on an ABI 7500 SDS Real-Time PCR system (Applied Biosystems). Primer sequences are given in Supplementary Table S1 and the concentration used was 300 nM for all genes. After an initial denaturation step for 10 min at 95°C, the thermal cycling conditions were 40 cycles at 95°C for 15 s and 60°C for 1 min. Each sample value was determined from duplicate measurements. Expression of the target transcripts was normalized to the mean of the expression level of the three reference genes according to the formula  $E = 2^{-(Ct[\text{Target}] - Ct_{\text{mean}}[\text{Reference}])}$ , where Ct is the threshold cycle.

**Solutions and chemicals.** Xylazine and ketamine chlorhydrate were from Alcyon (Pau, France), Alexa Fluor<sup>®</sup> 594-Cadaverine from Life Technologies (Saint Aubin, France) and mefloquine hydrochloride from BioBlocks, Inc. (San Diego, CA). All other compounds were purchased from Sigma.

**Statistics.** Numerical data are expressed as the mean  $\pm$  s.e.m. Differences between groups were assessed using the non-parametric Mann-Whitney *U* test. The effects of nerve stimulation frequency on catecholamine release was assessed using two-way analysis of variance (followed by Bonferroni's post-hoc test). Unpaired Student's *t*-test was used to compare means. Differences with  $P < 0.05$  were considered significant.

## References

1. Heinemann, C., von Ruden, L., Chow, R.H. & Neher, E. A two-step model of secretion control in neuroendocrine cells. *Pflugers Arch* **424**, 105-112 (1993).
2. Moser, T. & Neher, E. Rapid exocytosis in single chromaffin cells recorded from mouse adrenal slices. *J Neurosci* **17**, 2314-2323 (1997).
3. Barbara, J.G. & Takeda, K. Quantal release at a neuronal nicotinic synapse from rat adrenal gland. *Proc Natl Acad Sci U S A* **93**, 9905-9909 (1996).
4. Martin, A.O., Alonso, G. & Guerineau, N.C. Agrin mediates a rapid switch from electrical coupling to chemical neurotransmission during synaptogenesis. *J Cell Biol* **169**, 503-514 (2005).
5. Colomer, C., *et al.* Functional characterization of alpha9-containing cholinergic nicotinic receptors in the rat adrenal medulla: implication in stress-induced functional plasticity. *J Neurosci* **30**, 6732-6742 (2010).
6. Colomer, C., Desarmenien, M.G. & Guerineau, N.C. Revisiting the stimulus-secretion coupling in the adrenal medulla: role of gap junction-mediated intercellular communication. *Mol Neurobiol* **40**, 87-100 (2009).

7. Colomer, C., Martin, A.O., Desarmenien, M.G. & Guerineau, N.C. Gap junction-mediated intercellular communication in the adrenal medulla: An additional ingredient of stimulus-secretion coupling regulation. *Biochim Biophys Acta* **1818**, 1937-1951 (2012).
8. Cena, V., Nicolas, G.P., Sanchez-Garcia, P., Kirpekar, S.M. & Garcia, A.G. Pharmacological dissection of receptor-associated and voltage-sensitive ionic channels involved in catecholamine release. *Neuroscience* **10**, 1455-1462 (1983).
9. Hamelink, C., *et al.* Pituitary adenylate cyclase-activating polypeptide is a sympathoadrenal neurotransmitter involved in catecholamine regulation and glucohomeostasis. *Proc Natl Acad Sci U S A* **99**, 461-466 (2002).
10. Smith, C.B. & Eiden, L.E. Is PACAP the Major Neurotransmitter for Stress Transduction at the Adrenomedullary Synapse? *J Mol Neurosci* **48**, 403-412 (2012).
11. Martin, A.O., Mathieu, M.N., Chevillard, C. & Guerineau, N.C. Gap junctions mediate electrical signaling and ensuing cytosolic Ca<sup>2+</sup> increases between chromaffin cells in adrenal slices: A role in catecholamine release. *J Neurosci* **21**, 5397-5405 (2001).
12. Martin, A.O., Mathieu, M.N. & Guerineau, N.C. Evidence for long-lasting cholinergic control of gap junctional communication between adrenal chromaffin cells. *J Neurosci* **23**, 3669-3678 (2003).
13. Colomer, C., Lafont, C. & Guerineau, N.C. Stress-induced intercellular communication remodeling in the rat adrenal medulla. *Ann N Y Acad Sci* **1148**, 106-111 (2008).
14. Colomer, C., *et al.* Functional remodeling of gap junction-mediated electrical communication between adrenal chromaffin cells in stressed rats. *J Neurosci* **28**, 6616-6626 (2008).
15. Akiyama, T., Yamazaki, T., Mori, H. & Sunagawa, K. Simultaneous monitoring of acetylcholine and catecholamine release in the in vivo rat adrenal medulla. *Neurochem Int* **44**, 497-503 (2004).

16. Iwasa, K., Oomori, Y. & Tanaka, H. Acetylcholinesterase activity, and neurofilament protein, and catecholamine synthesizing enzymes immunoreactivities in the mouse adrenal gland during postnatal development. *J Vet Med Sci* **61**, 621-629 (1999).
17. Cruikshank, S.J., *et al.* Potent block of Cx36 and Cx50 gap junction channels by mefloquine. *Proc Natl Acad Sci U S A* **101**, 12364-12369 (2004).
18. Gomez-Sanchez, E.P. & Gomez-Sanchez, C.E. Central hypertensinogenic effects of glycyrrhizic acid and carbenoxolone. *Am J Physiol* **263**, E1125-1130 (1992).
19. Nassar-Gentina, V., Pollard, H.B. & Rojas, E. Electrical activity in chromaffin cells of intact mouse adrenal gland. *Am J Physiol* **254**, C675-683 (1988).
20. Marcantoni, A., *et al.* L-type calcium channels in adrenal chromaffin cells: role in pacemaking and secretion. *Cell Calcium* **42**, 397-408 (2007).
21. Marcantoni, A., *et al.* Loss of Cav1.3 channels reveals the critical role of L-type and BK channel coupling in pacemaking mouse adrenal chromaffin cells. *J Neurosci* **30**, 491-504 (2010).
22. Guerineau, N.C., Desarmenien, M.G., Carabelli, V. & Carbone, E. Functional chromaffin cell plasticity in response to stress: focus on nicotinic, gap junction, and voltage-gated  $Ca^{2+}$  channels. *J Mol Neurosci* **48**, 368-386 (2012).
23. Yoshizaki, T. Participation of muscarinic receptors on splanchnic-adrenal transmission in the rat. *Jpn J Pharmacol* **23**, 813-816 (1973).
24. Edwards, A.V. Adrenal catecholamine output in response to stimulation of the splanchnic nerve in bursts in the conscious calf. *J Physiol* **327**, 409-419 (1982).
25. Biales, B., Dichter, M. & Tischler, A. Electrical excitability of cultured adrenal chromaffin cells. *J Physiol* **262**, 743-753 (1976).
26. Kidokoro, Y. & Ritchie, A.K. Chromaffin cell action potentials and their possible role in adrenaline secretion from rat adrenal medulla. *J Physiol* **307**, 199-216 (1980).

27. Gullo, F., *et al.* ERG K<sup>+</sup> channel blockade enhances firing and epinephrine secretion in rat chromaffin cells: the missing link to LQT2-related sudden death? *FASEB J* **17**, 330-332 (2003).
28. Petrovic, J., Walsh, P.L., Thornley, K.T., Miller, C.E. & Wightman, R.M. Real-time monitoring of chemical transmission in slices of the murine adrenal gland. *Endocrinology* **151**, 1773-1783 (2010).
29. Arroyo, G., *et al.* Amperometric study of the kinetics of exocytosis in mouse adrenal slice chromaffin cells: physiological and methodological insights. *J Neurophysiol* **96**, 1196-1202 (2006).
30. Marley, E. & Paton, W.D. The output of sympathetic amines from the cat's adrenal gland in response to splanchnic nerve activity. *J Physiol* **155**, 1-27 (1961).
31. Gaumann, D.M., Yaksh, T.L., Tyce, G.M. & Stoddard, S.L. Adrenal vein catecholamines and neuropeptides during splanchnic nerve stimulation in cats. *Peptides* **10**, 587-592 (1989).
32. Yoshizaki, T. A method for pharmacological studies on adrenaline release from the rat adrenal. *Jpn J Pharmacol* **23**, 689-694 (1973).
33. Kuzmin, A.I., *et al.* The effect of histamine receptor antagonists on stress-induced catecholamine secretion: an adrenomedullary microdialysis study in the rat. *Eur J Pharmacol* **378**, 311-316 (1999).
34. Damase-Michel, C., *et al.* Effects of clonidine, dihydralazine and splanchnic nerve stimulation on the release of neuropeptide Y, MET-enkephalin and catecholamines from dog adrenal medulla. *Naunyn Schmiedeberg's Arch Pharmacol* **348**, 379-384 (1993).
35. Lamouche, S., Martineau, D. & Yamaguchi, N. Modulation of adrenal catecholamine release by PACAP in vivo. *Am J Physiol* **276**, R162-170 (1999).



36. Weinstock, M., Razin, M., Schorer-Apelbaum, D., Men, D. & McCarty, R. Gender differences in sympathoadrenal activity in rats at rest and in response to footshock stress. *Int J Dev Neurosci* **16**, 289-295 (1998).
37. Kobayashi, K., *et al.* Alteration of catecholamine phenotype in transgenic mice influences expression of adrenergic receptor subtypes. *J Neurochem* **65**, 492-501 (1995).
38. Bornstein, S.R., *et al.* Deletion of tyrosine hydroxylase gene reveals functional interdependence of adrenocortical and chromaffin cell system in vivo. *Proc Natl Acad Sci U S A* **97**, 14742-14747 (2000).
39. Tsunoda, M., Takezawa, K., Yanagisawa, T., Kato, M. & Imai, K. Determination of catecholamines and their 3-O-methyl metabolites in mouse plasma. *Biomed Chromatogr* **15**, 41-44 (2001).
40. Grouzmann, E., *et al.* Blood sampling methodology is crucial for precise measurement of plasma catecholamines concentrations in mice. *Pflugers Arch* **447**, 254-258 (2003).
41. Klevans, L.R. & Gebber, G.L. Comparison of differential secretion of adrenal catecholamines by splanchnic nerve stimulation and cholinergic agents. *J Pharmacol Exp Ther* **172**, 69-76 (1970).
42. Kuri, B.A., Chan, S.A. & Smith, C.B. PACAP regulates immediate catecholamine release from adrenal chromaffin cells in an activity-dependent manner through a protein kinase C-dependent pathway. *J Neurochem* **110**, 1214-1225 (2009).
43. Feuerstein, G. & Gutman, Y. Preferential secretion of adrenaline or noradrenaline by the cat adrenal in vivo in response to different stimuli. *Br J Pharmacol* **43**, 764-775 (1971).
44. Vollmer, R.R., Balcita, J.J., Sved, A.F. & Edwards, D.J. Adrenal epinephrine and norepinephrine release to hypoglycemia measured by microdialysis in conscious rats. *Am J Physiol* **273**, R1758-1763 (1997).

45. Ko, Y.Y., Jeong, Y.H. & Lim, D.Y. Influence of ketamine on catecholamine secretion in the perfused rat adrenal medulla. *Korean J Physiol Pharmacol* **12**, 101-109 (2008).
46. Guerineau, N.C., McKinney, R.A., Debanne, D., Mollard, P. & Gahwiler, B.H. Organotypic cultures of the rat anterior pituitary: morphology, physiology and cell-to-cell communication. *J Neurosci Methods* **73**, 169-176 (1997).
47. Guerineau, N.C., Bonnefont, X., Stoeckel, L. & Mollard, P. Synchronized spontaneous Ca<sup>2+</sup> transients in acute anterior pituitary slices. *J Biol Chem* **273**, 10389-10395 (1998).
48. Fauquier, T., Guerineau, N.C., McKinney, R.A., Bauer, K. & Mollard, P. Folliculostellate cell network: a route for long-distance communication in the anterior pituitary. *Proc Natl Acad Sci U S A* **98**, 8891-8896 (2001).
49. Bosco, D., Haefliger, J.A. & Meda, P. Connexins: key mediators of endocrine function. *Physiol Rev* **91**, 1393-1445 (2011).
50. Charollais, A., *et al.* Junctional communication of pancreatic beta cells contributes to the control of insulin secretion and glucose tolerance. *J Clin Invest* **106**, 235-243 (2000).
51. Head, W.S., *et al.* Connexin-36 gap junctions regulate in vivo first- and second-phase insulin secretion dynamics and glucose tolerance in the conscious mouse. *Diabetes* **61**, 1700-1707 (2012).
52. Krattinger, N., *et al.* Connexin40 regulates renin production and blood pressure. *Kidney Int* **72**, 814-822 (2007).
53. Hodson, D.J., *et al.* Coordination of calcium signals by pituitary endocrine cells in situ. *Cell Calcium* **51**, 222-230 (2012).
54. Hodson, D.J. & Mollard, P. Navigating pituitary structure and function - defining a roadmap for hormone secretion. *J Neuroendocrinol* **25**, 674-675 (2013).
55. Hodson, D.J., *et al.* Existence of long-lasting experience-dependent plasticity in endocrine cell networks. *Nat Commun* **3**, 605 (2012).

56. Moser, T. Low-conductance intercellular coupling between mouse chromaffin cells in situ. *J Physiol* **506 (Pt 1)**, 195-205 (1998).
57. Hill, J., Lee, S.K., Samasilp, P. & Smith, C. Pituitary adenylate cyclase-activating peptide enhances electrical coupling in the mouse adrenal medulla. *Am J Physiol Cell Physiol* **303**, C257-266 (2012).
58. Hatton, G.I. Synaptic modulation of neuronal coupling. *Cell Biol Int* **22**, 765-780 (1998).
59. Guerineau, N.C. & Desarmenien, M.G. Developmental and stress-induced remodeling of cell-cell communication in the adrenal medullary tissue. *Cell Mol Neurobiol* **30**, 1425-1431 (2010).
60. Nelles, E., *et al.* Defective propagation of signals generated by sympathetic nerve stimulation in the liver of connexin32-deficient mice. *Proc Natl Acad Sci U S A* **93**, 9565-9570 (1996).
61. Yu, Y.C., *et al.* Preferential electrical coupling regulates neocortical lineage-dependent microcircuit assembly. *Nature* **486**, 113-117 (2012).
62. Hormuzdi, S.G., *et al.* Impaired electrical signaling disrupts gamma frequency oscillations in connexin 36-deficient mice. *Neuron* **31**, 487-495 (2001).
63. Iankova, I., *et al.* Regulator of G protein signaling-4 controls fatty acid and glucose homeostasis. *Endocrinology* **149**, 5706-5712 (2008).
64. Ishimatsu, M. & Williams, J.T. Synchronous activity in locus coeruleus results from dendritic interactions in pericoerulear regions. *J Neurosci* **16**, 5196-5204 (1996).
65. Bani-Yaghoub, M., Underhill, T.M. & Naus, C.C. Gap junction blockage interferes with neuronal and astroglial differentiation of mouse P19 embryonal carcinoma cells. *Dev Genet* **24**, 69-81 (1999).

66. Risley, M.S., Tan, I.P. & Farrell, J. Gap junctions with varied permeability properties establish cell-type specific communication pathways in the rat seminiferous epithelium. *Biol Reprod* **67**, 945-952 (2002).

**Acknowledgements.** The authors thank Drs. G. Guillon and C. Legros for helpful discussions, P. Didier, A. Carrette, E. Galibert, D. Haddou and M. Gien-Asari for technical assistance, Drs. I. Iankova for help with catecholamine assay, A. Vincent, M. Corbani and N. Chauvet for help in imaging and histology. Work supported by grants from Fondation pour la Recherche Médicale, National Biophotonics and Imaging Platform (Ireland), Région Languedoc-Roussillon, Région Pays de la Loire, Conseil Général de Maine et Loire, Angers-Loire Métropole and Wellcome Trust (Grant No. WT078791).

#### **Author contributions**

M.G.D. and N.C.G. conceived and designed the study. M.G.D., C.J., B.T. and E.V. performed the experiments. S.G.H. provided experimental material and offered suggestions. M.G.D., C.J., B.T. and N.C.G. performed analysis. M.G.D. and N.C.G. wrote the paper.

**Competing financial interests:** The authors declare no competing financial interests.

#### **Figure legends**

**Figure 1: *In vivo* recording of spontaneous and splanchnic nerve stimulation-evoked electrical activity.** **A.** Experimental protocol used to simultaneously monitor intra-adrenal electrical activity and collect blood outflow from the adrenal vein. **B.** Spontaneous electrical activity recorded in the medullary tissue comprising both individual spikes (left inset) and

bursting activities (right inset). **C.** Reversible blockade of spontaneous activity by 3  $\mu$ M TTX locally applied on the nerve. \*,  $p < 0.05$  versus control, Mann-Whitney *U* test,  $n = 9$  mice. Values represent mean  $\pm$  s.e.m. **D and E.** Reversible decrease of splanchnic nerve stimulation-evoked electrical activity (10 V, 160 ms, 4 iterations, 1/min), in response to TTX (**D**,  $n = 7$  mice) and to hexamethonium (**E**, 200  $\mu$ M, injection through the recording pipette,  $n = 7$  mice). \*,  $p < 0.05$  versus control, Mann-Whitney *U* test. Values represent mean  $\pm$  s.e.m.

**Figure 2: Stimulus frequency-dependent catecholamine secretion.** Ratio between catecholamine concentrations before (C) and after (T) splanchnic nerve stimulation. In the absence of stimulation (no stim), T/C was slightly but not significantly higher than 100%. LF and HF columns indicate increased NE (**A**) and E (**B**) concentrations in response to graded stimulation. LF, low frequency (0.1 Hz, 160 ms, 10 V, 5 min), HF, high frequency (4 Hz, 1 ms, 10 V, 2 min). \*,  $p < 0.05$ , two-way analysis of variance (followed by Bonferroni's post-hoc test). Values represent mean  $\pm$  s.e.m. The number of recorded mice for each condition is indicated in parentheses.

**Figure 3: Significant reduction of fluorescent dye diffusion in the adrenal gland after *i.p.* injection of carbenoxolone.** **A.** Cx36 and Cx43 immunostainings showing that Cx36 is specifically expressed in the medulla, whereas the cortex displays a robust labeling for Cx43. Scale bar, 50  $\mu$ m. The punctuate fluorescent staining typical of a gap junction marker is evidenced at higher magnification (scale bar, 10  $\mu$ m). **B-D** Intra-peritoneal injection of carbenoxolone (CBX, 100 mg/kg) attenuates LY diffusion when compared to glycyrrhizic acid (GZA, 135 mg/kg) or NaCl (9‰). **B.** Representative snapshots (scale bar, 100  $\mu$ m) and associated wireframe plots illustrating the cutting edge. Quantification of LY diffusion measured as the fluorescence intensity *versus* distance from the edge (**C**) and as the area under this curve

(**D**, \*,  $p < 0.05$  versus GZA, Mann-Whitney  $U$  test,  $n = 5$  CBX- and 11 GZA-injected animals. Values represent mean  $\pm$  s.e.m.). **E**. Quantification measured as the area under the curve of Alexa Fluor® 594-Cadaverine diffusion in GZA- and CBX-injected mice showing that CBX significantly reduced cadaverine diffusion extent within the adrenal gland. \*,  $p < 0.05$  versus GZA, Mann-Whitney  $U$  test,  $n = 5$  CBX- and 7 GZA- injected mice. Values represent mean  $\pm$  s.e.m.

**Figure 4: Effect of carbenoxolone on stimulation-evoked catecholamine secretion. A.** CBX modifies neither NE nor E release triggered by LF stimulation ( $n = 9-10$  mice. Values represent mean  $\pm$  s.e.m.). **B.** Significant CBX-induced decrease in NE secretion evoked by HF stimulation (\*,  $p < 0.05$  versus GZA, Mann-Whitney  $U$  test,  $n = 8-9$  mice. Values represent mean  $\pm$  s.e.m.).

**Figure 5: Carbenoxolone-induced reduction of Lucifer yellow diffusion in the adrenal gland of cold stressed mice. A.** Significant increase of Cx36 immunolabeling in the medulla. Scale bar, 50  $\mu\text{m}$ . Inset is at higher magnification (scale bar, 10  $\mu\text{m}$ ). \*,  $p < 0.05$  versus control, Mann-Whitney  $U$  test,  $n = 5$  control mice and 6 cold stressed mice. Values represent mean  $\pm$  s.e.m. **B-D.** Scrape-loading data illustrating the CBX-induced reduction of LY diffusion. **B.** Representative snapshots (scale bar, 100  $\mu\text{m}$ ) and wireframe plots. **C.** Quantification of LY diffusion measured as the fluorescence intensity *versus* distance from the edge. **D.** Quantification of LY diffusion measured as the area under this curve (**D**, \*,  $p < 0.05$  versus GZA, Mann-Whitney  $U$  test,  $n = 8$  GZA- and 8 CBX-treated mice. Values represent mean  $\pm$  s.e.m.).

**Figure 6: Effect of carbenoxolone on stimulation-evoked catecholamine secretion in cold stressed mice.** Intra-peritoneal CBX injection ( $n = 11$  animals) induced a robust decrease in both

NE and E secretion evoked by a HF stimulation (\*,  $p < 0.05$  versus GZA ( $n = 11$  animals), Mann-Whitney  $U$  test. Values represent mean  $\pm$  s.e.m.).

**Figure 7: Carbenoxolone fails to reduce Lucifer yellow diffusion in the adrenal medulla of Cx36<sup>-/-</sup> mice.** **A.** Quantitative RT-PCR illustrating connexin expression in wild type (WT,  $n = 6$  mice) and Cx36<sup>-/-</sup> mice ( $n = 6$ ) macrodissected medulla. Data confirm invalidation of the Cx36 gene. While expression of Cx43 transcripts is not modified in Cx36<sup>-/-</sup> mice, Cx29 transcript is up-regulated (\*,  $p < 0.05$  versus WT, Mann-Whitney  $U$  test. Values represent mean  $\pm$  s.e.m.). **B.** Immunohistofluorescence images obtained from Cx36<sup>-/-</sup> (large photographs, scale bar, 50  $\mu$ m) and WT (insets, scale bar, 10  $\mu$ m) mice show that i) Cx36 is absent from the medulla of Cx36<sup>-/-</sup> mice and ii) Cx43 is exclusively detected in the cortex of Cx36<sup>-/-</sup> and WT animals. **C.** Scrape-loading data (scale bar, 100  $\mu$ m) and associated analyses show that CBX failed to reduce LY diffusion in adrenal glands of Cx36<sup>-/-</sup> mice (Mann-Whitney  $U$  test,  $n = 3$  GZA- and 4 CBX-treated mice. Values represent mean  $\pm$  s.e.m.).

**Figure 8: Impaired catecholamine secretion in Cx36<sup>-/-</sup> mice.** High frequency splanchnic nerve stimulation does not evoke a significant increase in catecholamine secretion, neither in CBX- nor in GZA-treated mice (Mann-Whitney  $U$  test,  $n = 3$  GZA- and 6 CBX-injected mice. Values represent mean  $\pm$  s.e.m.).

**Figure 9: Deficits occurring at the splanchnic nerve-chromaffin cell synapse in Cx36<sup>-/-</sup> mice.** **A.** Robust decrease of both vesicular acetylcholine transporter (VACHT) and neurofilament (NF) immunostaining in Cx36<sup>-/-</sup> compared with WT mice. Scale bar, 100  $\mu$ m. \*,  $p < 0.05$  versus WT, unpaired Student's  $t$  test,  $n = 22$  glands from 11 WT mice and 21 glands from 11 Cx36<sup>-/-</sup> mice. Values represent mean  $\pm$  s.e.m. **B.** No change in the expression level of

transcripts encoding five nicotinic acetylcholine receptor subunits in Cx36<sup>-/-</sup> mice (n = 6) compared to WT mice (n = 6, Mann-Whitney *U* test). Values represent mean ± s.e.m.