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Abstract

Objective

Multiple myeloma (MM) is a plasma cell neoplasm characterized by the accumulation of malignant plasma cells within the bone marrow. This disease still remains incurable, despite major treatment improvements. However, gene expression profiling of multiple myeloma cells (MMC) may lead to the identification of new therapeutic targets.

Methods

Using Affymetrix microarrays, we identified the overexpression of the MYEOV gene in MMC of 171 patients with newly-diagnosed multiple myeloma, compared to normal plasma cells.

Results

The MYEOV gene was present (Affymetrix call) in 79% of MMC and in 15% of normal plasma cells. MYEOV gene is not expressed in cells of the patients’ bone marrow (BM) environment. The down-regulation of MYEOV gene reduced the growth of a MYEOV\textsuperscript{present} myeloma cell line, unlike a MYEOV\textsuperscript{absent} one. Patients with MYEOV\textsuperscript{absent} MMC have an increased event free survival compared to patients with MYEOV\textsuperscript{present} MMC, after high-dose therapy and stem cell transplantation and a trend for increased overall survival. In a Cox-proportional-hazard model, MYEOV expression in MMC is predictive for EFS for patients independently of ISS stage, t(4;14) translocation, albumin or B2M serum levels. A knock out of MYEOV significantly reduced the growth of MMC.

Conclusion

Thus, MYEOV expression is a prognostic factor for patients with MM, in part through a role of MYEOV in the control of MMC proliferation.
Keywords

Multiple Myeloma, prognostic factor, therapeutic target, tumor growth, gene expression profiling.
Introduction

Multiple myeloma (MM) is a plasma cell neoplasm characterized by the accumulation of malignant plasma cells within the bone marrow (BM). Several autocrine or paracrine factors can promote multiple myeloma cell (MMC) survival and proliferation [1-4] and inhibition of MMC growth factors may have clinical applications in combination with other drugs[5-7].

In order to identify new therapeutic targets in MM, we compared gene expression profiles (GEP) of MMC with those of normal plasma cells, normal plasmablasts and normal peripheral blood B cells. We identified that MYEOV gene (for Myeloma overexpressed gene) was expressed in malignant plasma cells in 79% of newly-diagnosed patients with MM. The MYEOV gene was originally isolated by the application of the NIH/3T3 tumorigenicity assay with DNA from a gastric carcinoma. The chromosomal region 11q13 is frequently associated with genetic rearrangements in a large number of human malignancies, including B cell malignancies[8-10] and overexpression of MYEOV is frequently observed in breast tumors and oral and esophagel squamous cell carcinomas[11]. Although MYEOV is expressed in a subset of human myeloma cell lines (HMCLs) with t(11;14)(q13;q32) translocation[12], MYEOV expression is rarely related with t(11;14) in MM[13]. Recently, Janssen et al. demonstrated MYEOV gene expression is transcriptionally silenced by a DNA-methylation mechanism in esophageal squamous cell carcinomas[14]. The presence of functional domains such as RNP-1 (motif typical of RNA binding protein) and the studies of the short hydrophobic regions and of the C-terminal leucine/isoleucine tail showed that MYEOV might be directed to the membrane[12]. Nevertheless, the biological role of MYEOV remains unclear. Recent studies showed
that MYEOV siRNA decreased proliferation of gastric cancer cells and colon cancer cell lines in vitro [15, 16].

In this study, we demonstrate that MMC of 79% of the patients with newly-diagnosed MM express MYEOV gene. For patients treated with high dose chemotherapy (HDC) and autologous hematopoietic stem cell transplantation (ASCT), MYEOV gene expression is a prognostic factor for EFS independent of ISS stage, HRS, t(4;14) translocation, albumin or β2M serum levels.

Materials and methods

Cell samples
XG human myeloma cell lines were obtained as described [17-20]. SKMM, OPM2, LP1 and RPMI8226 HMCLs were purchased from ATTC (LGC Promochem, France). MMC were purified from 171 patients with newly-diagnosed MM after written informed consent was given in accordance with the Declaration of Helsinki and IRB approval of the University hospitals of Heidelberg (Germany) or Montpellier (France). These 171 patients were treated with high dose therapy (HDC) and autologous stem cell transplantation (ASCT) and were termed in the following Heidelberg-Montpellier (HM) series. Patients’ characteristics are indicated in Supplementary Table S1. The obtaining and purification of MMC, normal bone marrow (BM) plasma cells (BMPC), memory B cells, polyclonal plasmablasts, osteoclasts, BM stromal cell lines, BM CD34 cells, BM CD3 T cells, BM monocytes and BM polymorphonuclear neutrophils were performed as previously described [21]. We also used Affymetrix data of a cohort of 208 purified MMCs from previously untreated patients from the University of Arkansas for Medical Sciences (UAMS, Little Rock, AR). These patients were treated with total therapy 3[22] and termed in the following LR-TT3 series. These data are
publicly available through online Gene Expression Omnibus (Gene Expression Profile of Multiple Myeloma, accession number GSE2658. http://www.ncbi.nlm.nih.gov/geo/. Accessed June 1, 2006). Publicly available data from LR-TT2 cohort were not used because these patients were treated with thalidomide[23] and this information is not publicly available.

**Preparation of complementary RNA (cRNA) and microarray hybridization**

RNA was extracted using the RNeasy Kit (Qiagen, Hilden, Germany). Biotinylated cRNA was amplified with a double *in vitro* transcription reaction and hybridized to the Affymetrix HG U133 set of Gene Chips, according to the manufacturer’s instructions (Affymetrix, Santa Clara, CA). Microarray data have been deposited in the ArrayExpress public database, under accession numbers E-MEXP-2360 for BMPC [24] and E-TABM-937 for B cells, PPC, MMC and BM environment samples.

**Real-time RT-PCR**

Total RNA was converted to cDNA using the Superscript II reverse transcriptase (Invitrogen, Cergy Pontoise, France). The assays-on-demand primers and probes and the TaqMan Universal Master Mix were used according to the manufacturer’s instructions (Applied Biosystems, Courtaboeuf, France). The measurement of gene expression was performed using the ABI Prism 7000 Sequence Detection System and analyzed using the ABI PRISM 7000 SDS Software. For each primer, serial dilutions of a standard cDNA were amplified to create a standard curve, and values of unknown samples were estimated relative to this standard curve in order to assess the PCR efficiency. Ct values were obtained for GAPDH and the respective genes of interest during log phase of the cycle. Gene of interest levels were normalized to GAPDH for each sample ($\delta$Ct = Ct gene of interest – Ct GAPDH) and compared with
the values obtained for a known positive control using the following formula 100/2^{-Ct} where ddCt = dCt unknown – dCt positive control.

5-Aza-2’-deoxycytidine (5-azadC) treatment
The human MM cell lines XG-6, L363, and LP1 were cultured in RPMI 1640 supplemented with 10% fetal bovine serum. Cells (2 x 10^5/mL) were treated either with 0.5 µmol/L Aza-dC or with no drug (control) for 7 days.

Study of apoptosis
After 4 days of culture, cells were washed twice in PBS and apoptosis was assayed with FITC-conjugated annexin V labeling (Boehringer). Fluorescence was analyzed on a FACScan flow cytometer (Becton Dickinson).

Cell cycle analysis
DNA was stained with propidium iodide (PI). Cells were washed in PBS, suspended in 1 mL of 75% ethanol/25% water at room temperature for 2 minutes and washed again. 500 µl of PBS containing PI (40 µg/mL) and RNase (100 µg/mL) (both from Sigma, St Louis, MO, USA) were added to each sample. Cells were incubated for 30 minutes at 37 °C and stored at 4 °C in the dark before analysis with a FACScan flow cytometer using Cell Quest software. The cell cycle was analyzed with the ModFit LT software (Verity Software House, Topsham, ME, USA).

Western blot analysis
Cells were lysed in 10 mM Tris-HCl (pH 7.05), 50 mM NaCl, 50 mM NaF, 30 mM sodium pyrophosphate (NaPPi), 1% triton X-100, 5 µM ZnCl₂, 100 µM Na₃VO4, 1 mM DTT, 20 mM β-glycerophosphate, 20 mM p-nitrophenolphosphate (PNPP), 20 µg/ml aprotinin, 2.5 µg/ml leupeptin, 0.5 mM PMSF, 0.5 mM benzamidine, 5 µg/ml pepstatin, and 50 nM okadaic acid. Lysates were resolved on 12% sodium dodecyl sulfate-polyacrylamide by gel electrophoresis (SDS-PAGE) and transferred to a
nitrocellulose membrane (Schleicher and Schuell, Kassel, Germany). Membranes were blocked for 2 hours at room temperature in 140 mM NaCl, 3 mM KCl, 25 mM Tris-HCl (pH 7.4), 0.1% Tween 20 (TBS-T), 5% non-fat milk and human Ig (1 mg/ml), and then immunoblotted with a rabbit anti-MYEOV (Proteintech Group, Chivago, IL). As a control for protein loading, we used a mouse monoclonal anti-β-actin antibody (Sigma, St Louis, MO). The primary antibodies were visualized with goat anti-rabbit (Sigma) or goat anti-mouse (Bio-Rad, Hercules, CA) peroxidase-conjugated antibodies by an enhanced chemiluminescence detection system. Blots were quantified by densitometry using acquisition into Adobe Photo Shop (Adobe Systems, San Jose, CA), and analyzing with the NIH Image software (National Institutes of Health, Bethesda, MD, USA).

**siRNA transduction**

The MYEOV siRNA smart pool (M-016553-00) (Dharmacon Inc, IL, USA) was transduced by electroporation (Amaxa, Köln, Germany) using nucleofaction. We also used Dharmacon’s negative control siRNA (ON-TARGETplus siCONTROL Non-Targeting siRNA) as a control. RPMI8226, XG-7 and LP1 HMCLs were electroporated using respectively the T solution and programs T-001 or A-023 according to manufacturer’s instructions[25]. After electroporation, HMCLs were cultured for 4 days in 96-well flat-bottom microtiter plates in serum-free culture medium Syn H (ABCell-Bio, Montpellier, France) without cytokine as previously described[3].

**Statistical analysis**

Gene expression data were normalized with the MAS5 algorithm and analyzed with our bioinformatics platforms (RAGE, http://rage.montp.inserm.fr/ and Amazonia,
Survival curves were plotted using the Kaplan-Meier method. The statistical significance of differences in event free or overall survivals between groups of patients was estimated by the log-rank test. Univariate analyses were done to screen for prognostic variables by using Cox proportional hazards regression. The Cox model was also used for multivariate analysis to identify the most significant variables related to survival. A P value ≤ .05 was considered significant in all statistical analyses. Statistical comparisons were done with R (http://www.r-project.org/) or SPSS11 (SPSS Chicago, IL) software. The biological functions and pathways encoded by a gene list was analyzed with Ingenuity software (Ingenuity Systems, www.ingenuity.com).
Results and Discussion

*MYEOV* gene expression was investigated in purified MMC from 171 patients and 20 HMCLs using Affymetrix microarrays. Of the 5 *MYEOV* probe sets, the 227342_s_at probe set was the most strongly correlated with real time RT-PCR data ($r = .86, P < .001$) on the 20 HMCLs (Figure 1A). The use of present or absent Affymetrix call determined by the MAS5 algorithm was also validated for this probe set since only the 9 real-time RT-PCR $^+$ HMCLs had an Affymetrix present call (Figure 1A). In the following, the present and absent Affymetrix call of 27342_s_at probe set was used to combine microarray data from different patients’ cohort performed with A+B or U133 plus 2.0 microarrays. *MYEOV* gene was expressed (present call) in 9/20 HMCLs and in 131 of 171 purified MMCs of newly-diagnosed patients. *MYEOV* gene expression was also confirmed at protein level in 9 HMCLs and primary MMCs from 4 patients using western blot (Figure 1 B & C). Normalized MYEOV protein expression was significantly correlated with *MYEOV* Affymetrix expression ($r = .9, P = .001$) (Figure 1 B). The western blot analysis of MYEOV expression was not easy since only 2 antibodies are commercially available and they were not validated in previous peer-reviewed publications. The antibody we used provided a specific but weak signal after incubation with the anti-MYEOV antibody. *MYEOV* gene expression in primary MMCs was compared to that in normal BM plasma cells, normal plasmablasts or normal memory B cells. Whereas *MYEOV* 27342_s_at probe set had a present call (*MYEOV*$_{\text{present}}$) in 9/20 HMCLs and 131/171 primary MMCs, it had rarely a present call in BMPC (1/7) and was absent (*MYEOV*$_{\text{absent}}$) in normal plasmablasts (0/7) or memory B cells (0/7) (Figure 2). Of note, *MYEOV* probe set had a present call in purified plasma cells from 6/7 patients with MGUS. The expression of *MYEOV* gene, associated with poor prognosis, in
plasma cells from individuals with the premalignant phase of MM is appealing. Such observation was already done for CD200 that is not expressed in normal plasma cells, aberrantly expressed in MMCs in association with a poor prognosis and also expressed in MGUS plasma cells[29]. A likely explanation is that cancer disease is a multi hit disease. MYEOV overexpression could be one oncogenic hit that requires additional hits, occurring in myeloma cells, to be able to promote tumor formation and/or drug resistance.

MYEOV gene expression was investigated in the BM environment from patients with MM. MYEOV gene was not expressed in CD14 monocytes, CD15 polynuclear cells and CD3 T cells purified from the BM of 5 newly-diagnosed patients. It is also not expressed in 7 osteoclast samples, in BM stromal cells and in CD34+ hematopoietic stem cells from 5 patients with MM (Figure 2 and Supplementary Table S3). These data were validated by real-time RT-PCR (Supplementary data, Figure S1).

MYEOV gene expression in MMCs delineates a subset of patients with specific clinical characteristics. The frequencies of patients with lambda-light chain MM, with C-reactive protein level ≥ 5 mg/liter, LDH ≥ 240 IU/liter, or chromosome 13 deletion are increased in patients with MYEOVpresent MMCs (Supplementary data Table S2). Patients with MYEOVabsent MMC had a better event free survival (55 months) compared to patients with MYEOVpresent MMC (26 months) (Figure 3 A) and a trend for better overall survival \( (P = .06) \) (Figure 3 B). In a Cox-proportional-hazard model, the absence or presence of MYEOV \( (P = 0.01, \text{hazard ratio} = 1.9) \) and ISS-stage \( (P = 0.001, \text{hazard ratio} = 1.6) \) are independently predictive for EFS \( (P = 0.04 \text{ and } P = 0.002 \text{ respectively}) \) (Table 1). If MYEOV expression is tested together with classical prognostic factors, \( i.e. \) serum albumin and serum β2M, MYEOV expression \( (P = 0.04) \), β2M \( (P = 0.006) \) and albumin \( (P = 0.02) \) remain independent prognostic
factors. **MYEOV** expression ($P = .05$) is an independent prognostic factor of spiked **MMSET** expression, that is an indicator of t(4;14) translocation ($P = .0001$) [3] and of a spiked c-**MAF** expression, that is an indicator of t(14;16) translocation ($P = .004$) [30] (Table 1). We also looked for the prognostic value of **MYEOV** expression in MMCs in publicly available data from LR-TT3 series. **MYEOV** had a “present” call in MMCs of 73% of these patients. Patients with **MYEOV**\textsuperscript{absent} MMCs had a significant better overall survival in the LR-TT3 cohort ($P = .04$) (Figure 3 C). EFS data for the LR-TT3 cohort were not publicly available. A comparison of **MYEOV** prognostic value with those of other prognostic factors (ISS, genetic abnormalities) could not be done because these data are not publicly available.

**MYEOV** expression was previously described in a subset of HMCLs with t(11;14)(q13;q32) translocation[12]. No correlation between **MYEOV** expression and t(11;14)(q13;q32) translocation was found in primary MMCs (Supplementary data Table S2). Specht, et al also reported that **MYEOV** expression is rarely related with t(11;14) in MM [13]. Since **MYEOV** has been shown to be transcriptionally silenced by a DNA-methylation mechanism in esophageal squamous cell carcinomas, [14], we investigated such an epigenetic regulation of **MYEOV** gene in MMCs. A treatment with 5-aza-2’–deoxycytidine of 2 **MYEOV**\textsuperscript{absent} HMCLs (XG-6 and LP1) induced **MYEOV** expression without affecting that in the **MYEOV**\textsuperscript{present} L363 HMCL (Figure 4). Recently, a knock out of **MYEOV** RNA (siRNA) has been shown to decrease proliferation of gastric cancer cells and colon cancer cell lines in vitro [15, 16] suggesting a role of **MYEOV** in cancer proliferation and invasion. The shorter EFS and OAS in patients with **MYEOV**\textsuperscript{present} MMC could be explained by a role of **MYEOV** in MMC proliferation. **MYEOV** siRNA downregulated **MYEOV** gene expression by 63% and 67% in RPMI 8226 and XG-7 HMCLs respectively and significantly reduced
the growth of these 2 HMCLs by 40% and 65% respectively \((P = .006 \text{ and } P = .001, \ n=5)\) (Figure 5A&C). The \textit{MYEOV} siRNA could also knock down \textit{MYEOV} protein (Figure 5B). The growth of the \textit{MYEOV}\textsuperscript{absent} HMCL LP1 was not affected by the \textit{MYEOV} siRNA (Figure 5C). \textit{MYEOV} siRNA did not significantly induce apoptosis in RPMI 8226 and XG-7 HMCLs, but it blocked the cell cycle entry into the S phase (Figure 5D&E and Table 3).

In order to identify genes that are co-regulated with \textit{MYEOV} gene, gene expression of \textit{MYEOV}\textsuperscript{present} MMCS and \textit{MYEOV}\textsuperscript{absent} MMCS of newly-diagnosed patients were compared using SAM supervised analysis. Probe sets with a present call in less than 3 out of all patients and a variation coefficient \(\leq 100\) were excluded from the analysis, yielding to 7073 probe sets. 25 unique genes were differentially expressed between \textit{MYEOV}\textsuperscript{present} and \textit{MYEOV}\textsuperscript{absent} MMCS (2 fold ratio, 1000 permutations and false discovery rate (FDR) < 5 \%) (Table 2). \textit{MYEOV}\textsuperscript{present} MMCS overexpressed \textit{MAGE-A6} cancer testis antigen. In agreement with the bad prognostic factor of \textit{MYEOV} expression and its control of MMC proliferation, \textit{MAGE-A6} expression is associated with a shorter event free survival in patients with MM[31] and \textit{MAGE-A3/6} protein is associated with elevated proliferation in MMCS[32].

\textit{MYEOV}\textsuperscript{present} MMCS expressed more weakly the CD81 tetraspanin. The \textit{CD81} gene downregulation in \textit{MYEOV}\textsuperscript{present} MMCS was validated by quantitative RT-PCR (Supplementary Figure S2). A low CD81 expression may be also involved in treatment resistance since CD81 can inhibit the adhesion, migration, invasion, and viability of MMCS [33]. The anti-myeloma effect of CD81/CD82 involves a down-regulation of Akt, activation of FoxO transcription factors and a decrease in active mTOR and mTOR/rictor[34]. \textit{MYEOV}\textsuperscript{present} MMCS overexpressed \textit{Homer1} gene. Homer1 may regulate the apoptotic response of MMCS to TRAIL activation, since
TRAIL activation kills Homer1-positive cells unlike Homer1-negative cells[35]. Homer1 is generally more expressed in cancer cells than their normal counterpart suggesting it may confer on tumor cells the possibility to be killed by TRAIL activation[35]. A comprehensive analysis of the biological functions or pathways of the proteins encoded by these 25 genes using Ingenuity did not reveal more information.

In conclusion, we have identified that MYEOV expression by MMC is a new factor of poor prognosis in patients with MM, in part through a role of MYEOV in the control of MMC proliferation.
### EFS

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### EFS

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Table 1. Multivariate proportional hazards analysis.
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<td>---</td>
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</table>

Table 2: 25 unique genes differentially expressed between $MYEOV^{\text{present}}$ and $MYEOV^{\text{absent}}$ MMCs of newly diagnosed patients (SAM analysis, 2 fold ratio, 1000 permutations and false discovery rate (FDR) < 5 %).
### MYEOV<sup>present</sup> HMCL

<table>
<thead>
<tr>
<th></th>
<th>G0-G1</th>
<th>S-Phase</th>
<th>G2-M</th>
<th>P value</th>
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<tbody>
<tr>
<td>XG-7 CT &lt;br&gt; siRNA</td>
<td>40,67</td>
<td>35,97</td>
<td>23,36</td>
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<td>XG-7 &lt;br&gt; MYEOV &lt;br&gt; siRNA</td>
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<td>25,33</td>
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<td>RPMI8226 CT &lt;br&gt; siRNA</td>
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<td>RPMI8226 MYEOV siRNA</td>
<td>63,17</td>
<td>13,48</td>
<td>23,35</td>
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</table>

### MYEOV<sup>absent</sup> HMCL

<table>
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<tr>
<th></th>
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<th>S-Phase</th>
<th>G2-M</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>U266 CT &lt;br&gt; siRNA</td>
<td>50,85</td>
<td>34,75</td>
<td>14,39</td>
<td>NS</td>
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<tr>
<td>U266 MYEOV siRNA</td>
<td>50,95</td>
<td>34,45</td>
<td>14,6</td>
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**Table 3**: After electroporation with MYEOV siRNA or siRNA CT, RPMI8226, XG-7 and U266 myeloma cells were cultured at $10^5$ cells/ml. Cells were recovered after 3 days of culture, DNA was labelled with PI and cells were analyzed on a FACS<sup>a</sup> apparatus. The percentage of cells in the G0-G1, S and G2-M phases of the cell cycle is indicated and was determined using the ModFit LT software. Results are representative of three experiments.
Figure legends

Figure 1. Validation of MYEOV expression.

(A) Gene expression of MYEOV in 20 HMCLs was assayed with real time RT-PCR and normalized with GAPDH expression. The arbitrary value of 100 was assigned to XG-7 positive control. The coefficient of correlation between Affymetrix and real-time RT-PCR values was determined.

(B) Expression level of MYEOV protein in HMCLs using western blot and correlation with MYEOV gene expression Affymetrix signal value. For each cell line, the ratios of MYEOV and beta actin proteins were determined in order to compare MYEOV protein expression between cell lines.

(C) MYEOV protein expression in purified primary myeloma cells using western blot analysis.

Figure 2. MYEOV gene expression in myeloma cells from patients with MM and normal cells.

Affymetrix MYEOV gene expression in normal memory B cells, normal polyclonal plasmablasts, normal BM plasma cells (BMPC), purified myeloma cells from patients with multiple myeloma (MM), human myeloma cell lines, bone marrow (BM) CD34 cells, BM stromal cells, purified BM CD15, CD14 and CD3 cells and osteoclasts.

Figure 3. Association of MYEOV gene expression and survival of newly-diagnosed patients with MM.

(A) Kaplan-Meier plot of the event-free survival in patients with MYEOV$^{\text{present}}$ and MYEOV$^{\text{absent}}$ MMCs in the HM cohort of 171 patients.

(B) Kaplan-Meier plot of the overall survival in patients with MYEOV$^{\text{present}}$ and MYEOV$^{\text{absent}}$ MMCs in our cohort of 171 patients.
(C) Kaplan-Meier plot of the overall survival in patients with $MYEOV^{\text{present}}$ and $MYEOV^{\text{absent}}$ MMCs in the LR-TT3 cohort of 208 patients from the University of Arkansas School of Medical Sciences.

Figure 4. Epigenetic regulation of MYEOV gene in MMCs.

Gene expression of $MYEOV$ in XG-6, LP1 and L363 HMCLs was assayed with real time RT-PCR. Cells $(2 \times 10^5$/$\mu$L) were treated either with $0.5 \mu$mol/L Aza-dC or with no drug (control) for 7 days. For each experiment, the expression of MYEOV in myeloma cells was compared to that of untreated myeloma cells which was assigned an arbitrary value. Data are mean values of five independent experiments.

Figure 5. Inhibition of $MYEOV$ gene expression and myeloma cell growth by a $MYEOV$ siRNA.

(A) Inhibition of $MYEOV$ gene expression by a $MYEOV$ siRNA. For each experiment, the expression of $MYEOV$ in myeloma cells treated with $MYEOV$ siRNA was compared to that of myeloma cells treated with control siRNA, which was assigned the arbitrary value of 100. Results are the mean $MYEOV$ gene expression in five independent experiments. * The mean value is statistically significantly different from that obtained with a non-targeting control siRNA (siRNA CT) using Student’s $t$-test for pairs ($P \leq .05$).

(B) Inhibition of MYEOV protein expression in MMC using $MYEOV$ siRNA. RPMI8226 myeloma cells were treated for 4 days with $MYEOV$ or control siRNA and MYEOV protein was assayed by western blot.

(C) After electroporation with $MYEOV$ siRNA or siRNA CT, LP1, RPMI8226 and XG-7 HMCLs were cultured in serum-free culture medium for 4 days. Results are the mean ± SD values of the RLU fluorescence determined on sextuplet culture wells. Results shown are representative of five independent experiments. * The mean value
is statistically significantly different from that obtained with siRNA CT using Student’s \( t \)-test (\( P \leq .05 \)).

(D) After electroporation with MYEOV siRNA or siRNA CT, RPMI8226 and XG-7 myeloma cells were cultured at \( 10^5 \) cells/ml. Cells were recovered after 3 days of culture and apoptotic cells were detected by annexin V staining. Results are those of one experiment representative of three.

(E) DNA was labelled with PI and cells were analyzed on a FACScan apparatus. The percentage of cells in the S phase of the cell cycle is indicated and was determined using the ModFit LT software. Results are from one experiment representative of three.

**Acknowledgments**

This work was supported by grants from the Ligue Nationale Contre le Cancer (Equipe Labellisée 2009)(Paris, France) and Institut National du Cancer (R07001FN), the Hopp-Foundation, Germany, the University of Heidelberg, Heidelberg, Germany, the National Center for Tumor Diseases, Heidelberg, Germany, and the Tumorzentrum Heidelberg/Mannheim, Heidelberg, Mannheim, Germany.
References


Figure 1A
Figure 1 B

[Image of gel electrophoresis showing bands at 60 kD and 43 kD for MYEOV and Beta actin]

[Graph showing comparison of Affymetrix expression and Protein expression with a linear regression line for MYEOV, r = .9, P = .001]
Figure 1C
Figure 3 A

Event free survival

$P = 0.01$

MYEOV\textsuperscript{present}

MYEOV\textsuperscript{absent}

Days

EFS HM cohort

Figure 3 B

Overall survival

$P = 0.06$

MYEOV\textsuperscript{present}

MYEOV\textsuperscript{absent}

Days

OAS HM cohort
Figure 3 C

Overall survival

\[ P = 0.04 \]

- \( MYEOV^{\text{present}} \)
- \( MYEOV^{\text{absent}} \)

OAS LR-TT3 cohort
Figure 4

MYEOV expression (Real time RT-PCR) in arbitrary units:
- L363
- XG-6
- LP1

Comparison of MYEOV expression:
- XG-6 CT vs. XG-6 5aza: P = 0.02
- LP1 CT vs. LP1 5aza: P = 0.04
- L363 CT vs. L363 5aza: NS
Figure 5
XG-7 CT siRNA
S phase = 36%

XG-7 MYEOV siRNA
S phase = 25%

Figure 5E