

Inflammation and impaired endothelium-dependant vasodilatation in non obese women with gestational diabetes mellitus: Preliminary results.

Ines Mrizak, Amel Arfa, Mariem Fekih, Haythem Debbabi, Ali Bouslema, Imen Boumaiza, Monia Zaouali, Naim Akhtar Khan, Zouhair Tabka

► **To cite this version:**

Ines Mrizak, Amel Arfa, Mariem Fekih, Haythem Debbabi, Ali Bouslema, et al.. Inflammation and impaired endothelium-dependant vasodilatation in non obese women with gestational diabetes mellitus: Preliminary results.. *Lipids in Health and Disease*, BioMed Central, 2013, 12 (1), pp.93. 10.1186/1476-511X-12-93 . inserm-00842930

HAL Id: inserm-00842930

<https://www.hal.inserm.fr/inserm-00842930>

Submitted on 9 Jul 2013

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH

Open Access

Inflammation and impaired endothelium-dependant vasodilatation in non obese women with gestational diabetes mellitus: preliminary results

Ines Mrizak^{1*}, Amel Arfa¹, Mariem Fekih², Haythem Debbabi¹, Ali Bouslema³, Imen Boumaiza³, Monia Zaouali¹, Naim A Khan⁴ and Zouhair Tabka¹

Abstract

Background: To evaluate whether abnormal endothelial function, a common finding in gestational diabetes mellitus (GDM) pregnancies, can be explained by inflammatory cytokines.

Methods: Forearm skin blood flow (FSBF), into response to acetylcholine (Ach) (endothelium-dependent vasodilatation), were measured in 24 pregnant control subjects and 28 gestational diabetes mellitus (GDM) women, in the third trimester of gestation. A fasting glycemic and lipidic panel was obtained, and inflammatory cytokines (TNF- α and IL-6) and adiponectin were also determined.

Results: FSBF is significantly reduced in GDM group compared with control subjects (344.59 ± 57.791 vs. 176.38 ± 108.52 , $P < 0.05$). Among all subjects, FSBF showed a strong negative correlation with TNF- α and IL-6 ($r = -0.426$, $P < 0.0001$ and $r = -0.564$, $P < 0.0001$, respectively) and positive correlation with adiponectin ($r = 0.468$, $P < 0.0001$).

Conclusions: Endothelial function, an early marker of macrovascular disease, is present in non-obese pregnancies complicated by GDM. This alteration seems to be directly related to inflammatory status, which may represent a patho-physiological link between GDM and type 2 diabetes and, later on, metabolic syndrome.

Keywords: Forearm skin blood flow, Laser Doppler flowmetry, Gestational diabetes mellitus, Inflammation

Introduction

Gestational diabetes mellitus (GDM), is one of the most commonly observed obstetrical complications affecting from 5% to 8% of all pregnancies [1,2].

GDM is associated with the development of inflammation [3], where adipose tissues play an important role in the regulation of insulin sensitivity by secreting adipokines which are involved in the pathogenesis pregnancy-induced insulin resistance, in the case obese pregnancies [4-6]. However, obesity may be not the only link between inflammation and glucose intolerance during pregnancy, placenta is also an important source of cytokines [7].

Many studies suggest that GDM is characterized by the installation of subclinical inflammation associated with a vascular dysfunction due to the insulin resistance, but to date there has been a dearth of studies, evaluating cytokine levels in women with GDM in order to explain their contribution to endothelium dysfunction [7]. Non-invasive measurement of microcirculatory blood flow in patients has recently emerged as useful tool to investigate the effect of insulin resistance on endothelial function [8,9].

We have conducted the present study to assess whether non-obese GDM women present an impairment of endothelial NO release; we wanted to focus us on the only effect of diabetes on the inflammatory status and its impact on endothelial function. Hence, we have investigated the endothelium-dependent vasodilatation by measuring non-invasively the change of FSBF in

* Correspondence: ines.mrizak@hotmail.com

¹Department of Physiology and Functional Exploration, Farhat Hached University Hospital, Sousse, Tunisia

Full list of author information is available at the end of the article

response to graded infusion of acetylcholine (Ach). We have also assessed whether the inflammatory cytokine levels can be involved in the impairment of endothelial function.

Methods

Subjects

The study was conducted in the Physiology and Functional Explorations Department, University Farhat Hached Hospital, Sousse, Tunisia, and it was approved by Farhat Hached Hospital Ethical Committee for research on humans in Tunisia. An informative written consent was approved and signed by all the women of the study.

A total of 28 GDM patients were selected between June 2010 to June 2011, from High Risk Pregnancy Department, Farhat Hached Maternity. Inclusion criteria were the following: normal glucose tolerance in early pregnancy, diagnosis of GDM in the 2nd or the 3rd trimester of gestation, according to the criteria of National Diabetes Data Group [10], the age range was between 20 and 39 years and gestational age was between 28th and 41th week of gestation.

The gestational age was derived from the last menstrual period, otherwise, gestational age was corrected on the basis of ultrasonographic measurements, consonant with clinical practice.

The selection of control individuals was from among pregnant women with the same characteristics including similar anthropometric parameters and gestational age, but with normal glucose tolerance. Hence, O'Sullivan test was conducted between the 24th and 26th week of gestation [11]. A total control subjects enrolled in the study was 24 pregnant women. Under the same conditions as the diabetic group we collected anthropometric measures of control group. The controls was measured in an earlier gestational week in order to eliminate any eventuality that controls may have glucose intolerance before it was based on monitoring records in High Risk Pregnancy Department, Farhat Hached Maternity. An O'Sullivan test was conducted between the 24th and 26th week of gestation.

Each of the 52 pregnant women who completed the study were first asked to provide details of medical and smoking history and then participated in health screening session and overall medical examination, including anthropometric measurement and resting arterial blood pressure.

In recruiting patients we held that the patients showed no risk to develop a preeclampsia, we followed the screening adopted by the High Risk Pregnancy Department, Farhat Hached Maternity, namely systolic blood pressure 120 mmHg and diastolic blood pressure 90 mmHg, and negative proteinuria because it will in the

risk of influence endothelial function. And all GDM were monitored by insulin treatment.

All pregnancies were conducted under optimal conditions, all deliveries are vaginal, and took place between 36 and 41 weeks, and no special feature of fetal growth (data is not shown).

Biochemical parameters

Blood samples were collected from subjects after 12 hours overnight fast, the blood was maintained at 4°C centrifuged, distributed in aliquots and stored at -80°C until the batched measurement of parameters. Serum total cholesterol (TC) and triglycerides (TG) were determined by standard assays. High-density lipoprotein cholesterol (HDL) was measured by direct assay. Low-density lipoprotein cholesterol (LDL) concentrations were calculated with the Friedwald formula [12]. Fasting glucose was measured by the glucose oxidase method. All biochemical parameters were determined on an automated Synchron CX7 Clinical System (Beckman, Fullerton, CA). Plasma insulin was assayed by IRMA Insulin kit (Immunotech, France). Insulin resistance (IR) was evaluated with the homeostasis model assessment (HOMA) using the following equation: $HOMA-IR = (\text{Fasting insulin } (\mu\text{UI/ml}) * \text{Fasting glucose (mmol/l)}) / 22.5$ as per method of Matthews et al. [13].

Concentrations of maternal cytokines, including TNF- α and IL-6 were measured by ELISA-kits (Immunotech, France). Adiponectin concentrations were measured also by ELISA (R & D System, USA) according to the manufacturer's instructions.

Laser Doppler iontophoresis and hemodynamic measurement of FSBF

Combining Laser Doppler flow measurements with iontophoresis of vasoactive agents is a promising non-invasive tool and an attractive technique for studying the hemodynamic of the skin microcirculation in humans.

The protocol used is well-explained in the study of Miâdi-Messaoud et al. [8]. The equipments used were the following: Laser Doppler flowmeter (Periflux PF5000; Perimed, Stockholm, Sweden), electrode chamber (PF383; Primed Stockholm, Sweden) and acetylcholine chloride (Sigma Aldrich, Switzerland).

The following parameters were determined:

- Basal perfusion index (skin temperature = 33°C, arbitrary unit) during the second 2 min without Ach infusion
- Perfusion index after the third dose of Ach iontophoresis, i.e. the maximal endothelial response (arbitrary unit)
- Maximum perfusion index after heat hyperthermia and without Ach infusion (skin temperature = 44°C, arbitrary unit)

For all groups, cumulative concentration response curves (CCRC) of FSBF to Ach were obtained for each individual. For all groups, FSBF endothelium-dependant was defined as the maximal blood flow in response to Ach iontophoresis divided by the mean baseline blood flow expressed in percent.

Statistical analysis

Statistical analyses are performed using SPSS 17.0. All variables measured are reported as mean (M) ± standard deviation (SD), by applying Kolmogorov-Smirnov normality test; we found that all variables are normally distributed so we applied parametric analyses using the student *t*-test. Spearman correlation was performed on all parameters with FSBF. All correlated variables are introduced in linear regression, for relationship between FSBF as dependant variable and the inflammatory cytokine as independent variables in ascendant model, to assess the magnitude of their individual effects on FSBF. A *p*-value < 0.05 was considered statistically significant for all tests.

Results

Anthropometric parameters of the two groups are shown in Table 1. No significant differences in all anthropometric parameters between two groups were observed as regards age, weight, BMI, systolic blood pressure, diastolic blood pressure, waist circumference and hip circumference. In the two groups gestational age is homogenous, and there is no large variability.

Metabolic parameters are shown in Table 2. As expected, glucose at fasting, HOMA-IR and HbAc (glycated hemoglobin) were significantly higher in GDM group compared to control, whereas fasting insulinemia was insignificantly increased in GDM group.

TC, creatinine were significantly higher in GDM group compared to control whereas triglyceride and LDL levels were higher in the same group but were not significantly different with the control group. The levels of HDL, Apo A1, adiponectin and urea were significantly decreased in GDM group compared to control.

FSBF was significantly decreased in GDM group compared with control. Significant correlations among all groups regarding FSBF with biochemical and inflammatory parameters are regrouped in Table 3. Positive correlation retained, with FSBF, was urea, ApoA1 and adiponectin. Negative correlation was obtained with creatinine, HbAc, HOMA-IR, IL-6 and TNF-α was retained with dependant variable, FSBF.

In order to evaluate the independent contribution of biochemical and inflammatory parameters to predict FSBF, we performed an ascendant regression analysis only of IL-6, TNF-α and adiponectin in the model with a strong linear relationship with FSBF ($r^2 = 0.43$, $p < 0.0001$), $FSBF = 1738.57 - 0.372 * IL-6 - 0.249 * TNF-α + 0.246 * adiponectin$.

Discussion

The results of this study show the following new findings: 1) as compared with control pregnant women, non-obese GDM women show an impaired FSBF; and 2) FSBF is inversely correlated to pro-inflammatory cytokines, IL-6 and TNF-α, and in parallel related to adiponectin.

FSBF, an endothelium-dependant phenomenon, was examined at supine position. Ach induced vasodilatation of skin microcirculation by iontophoretic transdermal administration, involving agonistic endothelium-dependent response [14]. Ach provokes increases in the skin laser Doppler blood flow signal in a dose-dependent manner [15] and with an acceptable reproducibility (coefficients of variation between 10 and 17%) [16,17]. This technique was the method of choice for pregnant women in view of its non-invasiveness and the comfortable conditions of realization.

Although, many studies has explored the endothelium function with GDM women, but, to our knowledge, no previous study has assessed *in vivo* the maternal endothelial function in non-obese pregnancies, complicated by GDM, using combined Laser Doppler blood flow measurement with iontophoresis of vasoactive agent like Ach.

Table 1 Anthropometric parameters

	Control subjects	GDM	p-value
Age (years)	28.3 ± 4.24	31.7 ± 5.5	0.064
Weight (kg)	76.1 ± 3.23	76 ± 2.3	0.239
Height (m)	1.6 ± 0.06	1.63 ± 0.04	0.200
BMI (kg/m ²)	29.7 ± 2.23	28.5 ± 1.8	0.440
Systolic blood pressure (mmHg)	111.13 ± 5.6	113.01 ± 9.28	0.074
Diastolic blood pressure (mmHg)	61.46 ± 7.9	61.97 ± 10	0.298
Waist circumference (cm)	107.7 ± 11.66	115.5 ± 10.58	0.613
Hip circumference (cm)	109.7 ± 6.7	116.1 ± 11.92	0.036

Data are M ± SD. n=24 control subjects; n=28 GDM subjects.

Table 2 Comparison of metabolic and vascular characteristic between control subjects and GDM subjects

	Control subjects	GDM	p-value
Fasting glucose (mmol/l)	5.03 ± 0.46	5.42 ± 1.64*	0.01
Fasting insulinemia (μU/ml)	10.59 ± 11.1	13.19 ± 10.05	0.67
Triglyceride (mmol/l)	1.99 ± 0.23	2.4 ± 0.93	0.38
Total cholesterol (mmol/l)	5.12 ± 1.26	5.43 ± 1.54**	<0.0001
HDL cholesterol (mmol/l)	1.47 ± 0.55	1.31 ± 0.3*	0.018
LDL cholesterol (mmol/l)	2.59 ± 1.39	2.94 ± 1.27	0.47
Apo A1 (g/l)	1.90 ± 0.7	1.50 ± 0.34**	0.001
Apo B (g/l)	1.36 ± 0.5	1.22 ± 0.43	0.64
Urea (mmol/l)	3.83 ± 1.16	2.44 ± 0.89*	0.050
Creatinine (μmol/l)	49.99 ± 0.15	50.74 ± 8.76	0.6
HbAc (%)	5.27 ± 0.5	6.7 ± 1.21*	0.002
HOMA-IR	1.53 ± 0.51	2.38 ± 1.39*	0.02
Adiponectin (μg/ml)	10.02 ± 1.1	8.58 ± 1.7*	0.01
IL-6 (pg/ml)	53.17 ± 1.45	77.84 ± 10.98*	0.002
TNF-α (pg/ml)	61.75 ± 6.5	71.63 ± 12.33	0.091
C-reactive protein (mg/dl)	0.86 ± 1.1	0.80 ± 1.4	0.716
FSBF endothelium-dependent (%)	1176.38 ± 531.67	344.59 ± 305.81*	0.037

Data are M ± SD. n=24 control subjects; n=28 GDM subjects and significant difference is as follow: *p<0.05, **p<0.001.

Our results are consistent with the evidence that women with pregnancies complicated by GDM have impaired endothelial function. This malfunction should be related to deregulation of glucose metabolism according to the literature [18]. There are several investigators who have also reported this impaired resistant in euglycemic women with previous GDM [18]. One of the principal objectives with regard to diabetic patients, at high-risk pregnancies, is to obtain an euglycemic level.

Besides, endothelial function in diabetes has been attributed mainly to hyperglycemia [19,20] but also, and recently to subclinical inflammation [7]. It is interesting to note that among all subjects, FSBF is strongly and positively related to anti-inflammatory adiponectin ($r = 0.468$, $P < 0.0001$), as shown in Figure 1, and negatively related to the levels of pro-inflammatory cytokines, IL-6 and

TNF-α (respectively, $r = -0.564$, $p < 0.0001$; $r = -0.426$, $p < 0.0001$), as shown respectively in Figures 2 and 3.

In our study, insulin resistance, performed with HOMA system, was significantly higher in GDM group compared with control, and it was negatively correlated with FSBF. It has been reported that there is a relationship between insulin resistance and inflammatory status as described by Dandona et al. [21].

According to our results, it seems that the GDM patients might suffer from metabolic syndrome, marked

Table 3 Spearman's bivariate correlation coefficients FSBF with other variables

	r	p-value
Urea	0.334	0.003
Creatinine	0.123	0.207
Apo A1	0.351	0.002
HbAc	-0.514	<0.0001
HOMA-IR	-0.334	0.003
Adiponectin	0.468	<0.0001
IL-6	-0.564	<0.0001
TNF-α	-0.426	<0.0001

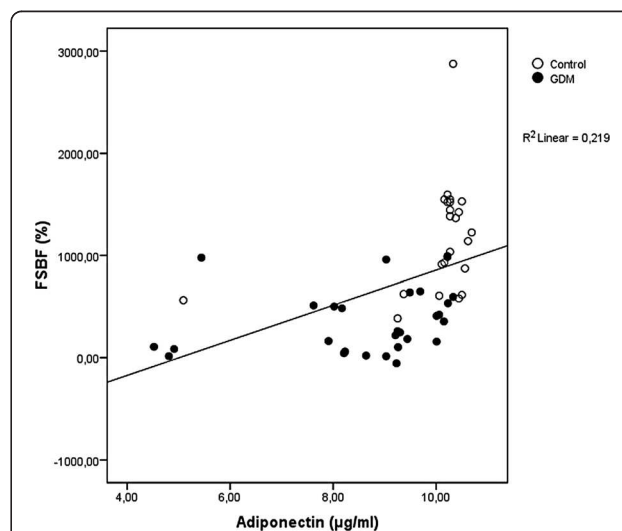
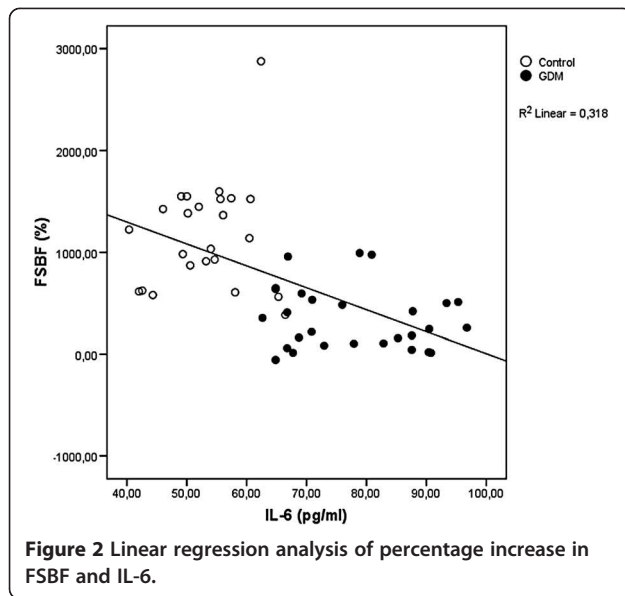
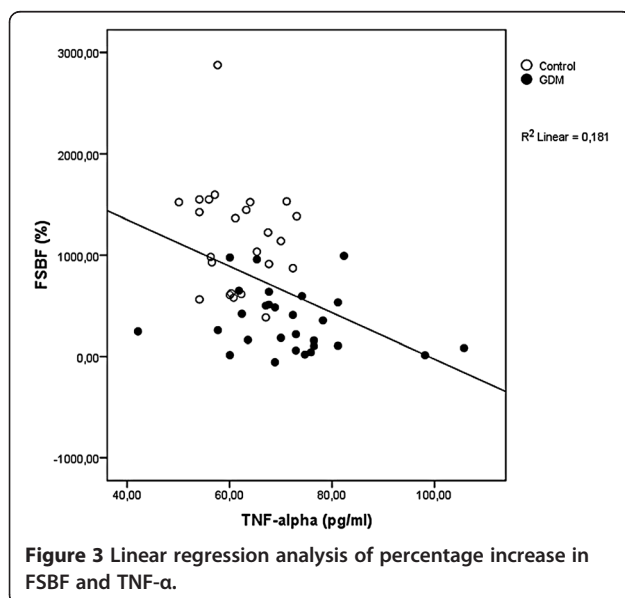


Figure 1 Linear regression analysis of percentage increase in FSBF and adiponectin.



with elevated triglyceride and LDL concentration, low HDL concentrations. This physiological change might be the result of an elevated triglyceride load in the HDL particles by the action of hepatic lipase which hydrolyses the triglycerides [21]. The small HDL particles, which have lost triglyceride, are filtered by the kidney, resulting in a decrease in apolipoprotein (Apo) A and HDL concentrations. Moreover, there are reports that insulin may promote ApoA gene transcription [22,23]. Hence, insulin resistance may be related to diminished ApoA biosynthesis, which seems to be the case in our GDM subjects [24].

In the light of our results, we can propose that there exists a tridimensional relationship between FSBF,



insulin resistance and inflammatory status in GDM group [21,25]. However, the relationship between pro-inflammation and endothelial function is not well-understood. In human endothelial cells, L-arginine is taken up via membrane transport systems grouped a family of proteins known as cationic amino acid transporters (CATs) (hereafter referred as “CATs family”) [26] whose expression and activity, and the mechanisms modulating these phenomena, have been extensively described [27-32], including in the human placenta [33,34].

In another hand, it is now established that GDM are pathological conditions altering hCAT (cationic amino acid transporter-1)-mediated arginine transport and eNOS (endothelial nitric oxide synthase) synthesis of NO in human fetoplacental vasculature, due to abnormal signaling pathway leading to altered vascular reactivity and changes in umbilical vessels blood flow from and to fetus with serious consequence on its growth [35]. Altogether these findings could be crucial for fetal insulin modulation of endothelial-derived NO synthesis in human umbilical vessels from pregnancy diseases associated with hyperinsulinemia, such as GDM, and other states of insulin resistance [27,36-39].

It has been reported that Akt2-null mice develop insulin resistance and mild hyperglycemia with hyperinsulinemia [40] as Akt2 is a key protein involved in signal transduction. Besides, the phosphorylation of Akt2 can be induced by TNF- α and IL-6 which have recently been implicated in inducing SOCS-3 [41-43], a protein known to interfere with tyrosine phosphorylation of the insulin receptor and IRS-1 (insulin receptor substrate 1) [44]. This, in turn, reduces the activation of Akt (protein kinase B), which normally causes the translocation of the insulin-responsive glucose transporter, Glut-4, to the plasma membrane. It also induces the phosphorylation of the enzyme NOS and its activation to generate NO [42].

As discussed above, NO is a key mediator of vascular health, promoting smooth muscle relaxation and exerting anti-inflammatory and antithrombotic effects. Under normal conditions, eNOS transfers electrons to L-arginine to produce citrulline and NO. Decreased NO availability has been observed in clinical studies of patients with insulin resistance [45,46].

What is the origin of this pro-inflammatory state in the GDM women? It is possible that during insulin resistance, increased IL-6 not only diminishes insulin sensitivity but by suppressing insulin signal transduction also interferes with anti-inflammatory effect of insulin, and might favour inflammation during insulin resistance [47].

Indeed, novel non-metabolic actions have been attributed to insulin, like anti-inflammatory effect [21]. Insulin has been shown to suppress several pro-inflammatory transcription factors, such as nuclear factor (NF- κ B),

Egr-1 (early growth response protein 1) and activating protein-1 (AP-1) and the corresponding genes which mediate inflammation [47,48].

The pro-inflammatory state induces insulin resistance, leading to clinical and biochemical manifestations of the metabolic syndrome. This resistance to insulin action promotes inflammation further through an increase in FFA (Free fatty acid) concentration and interferes with the anti-inflammatory effect of insulin [21]. The increase of FFA may be the result of hormonal modifications occurs until gestation like Human placental lactogen (HPL).

Insulin secretion in women with GDM is defective and, therefore, is unable to rise adequately to compensate for the insulin resistance; the result is hyperglycemia. The mechanism by which elevated plasma FFA levels cause insulin resistance in skeletal muscle includes intramyocellular accumulation of diacylglycerol, which activates protein kinase C (the β II and δ isoforms). This results in reduction of tyrosine phosphorylation of the insulin receptor substrate-1 and inhibits activation of phosphoinositol-3 kinase, an enzyme that is essential for normal insulin-stimulated glucose uptake [49,50].

Inflammatory cytokines have a pivotal role in placental function throughout pregnancy. Moreover, the progressive development of insulin resistance during pregnancy is due, in part, to placental cytokines such as TNF- α [51] and leptin [52]. Studies on the expression of inflammatory genes in GDM women have demonstrated major changes in placental gene expression [53].

Conclusion

The particularity of our study is that the studied population was not obese; however, the results showed clearly that the GDM group suffered from dyslipidemia, insulin resistance, pro-inflammatory status and exhibited endothelial dysfunction. Our study is limited by the number of subjects and these results should be supported by researches that extend the number of recruited.

Abbreviations

Akt: Protein kinase B; Apo A1: Lipoprotein 1; Apo B: Lipoprotein B; ELISA: Enzyme linked immunosorbent assay; eNOS: Endothelial nitric oxide synthase; Erg1: Early growth response protein 1; FFA: Free fatty acid; FSBF: Forearm skin blood flow; GDM: Gestational diabetes mellitus; HbAc: Glycated hemoglobin; h-CAT: Cationic amino acid transporter 1; HOMA-IR: Homeostasis model assessment-insulin resistance; IL-6: Interleukin 6; IRMA: Immunoradiometric assay; IRS-1: Insulin receptor substrate 1; NF- κ B: Nuclear factor-kappa B; NO: Nitric oxide; SCOS-3: Suppressor of cytokine signaling 3; T2D: Type 2 diabetes; TC: Total cholesterol; TG: Triglyceride; TNF- α : Tumor necrosis factor alpha.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MI was in charge of the practical work and prepared major parts of the manuscript. AA collected and analyzed data with the involvement of MI. FM participated in interpretation of the gynecology function. DH interpreted

Laser Doppler Data. BA and BI conducted biochemical analyses. MZ conducted hormonal analyses. TZ and KNA planned and organized the study and contributed to the revisions and the final drafts of the manuscripts. All authors read and approved the final manuscript.

Acknowledgements

We would like to thank the Department of Gynecology, Farhat Hached University Hospital Sousse, Tunisia, under the direction of Professor Khairi Hédi, for the kind collaboration. We would also like to express our sincere thanks to Professor Rebai Ahmed Centre of Biotechnology of Sfax, Tunisia.

Writing group

Ines Mrizak, Amel Arfa, Mariem Fekih, Haythem Debbabi, Ali Bouslema, Imen Boumaiza, Monia Zaouali, Naim A Khan and Zouhair Tabka.

Author details

¹Department of Physiology and Functional Exploration, Farhat Hached University Hospital, Sousse, Tunisia. ²Department of Gynecology, Farhat Hached University Hospital Sousse, Sousse, Tunisia. ³Department of Biochemistry, Sahloul University Hospital Sousse, Sousse, Tunisia. ⁴INSERM U866, Physiology of Nutrition & Toxicology, University of Burgundy, Faculty of Life Sciences, Dijon, France.

Received: 8 December 2012 Accepted: 10 June 2013

Published: 27 June 2013

References

1. American Diabetes Association: **Diagnosis and Classification of Diabetes Mellitus.** *Diabetes Care* 2006, **29**(1):43–48.
2. Hanna FW, Peters JR, Harlow J, Jones PW: **Gestational diabetes screening and glycaemic management; national survey on behalf of the Association of British Clinical Diabetologists.** *QJM* 2008, **101**(10):777–784.
3. Wolf M, Sauk J, Shah A, Vossen Smirnakis K, Jimenez-Kimble R, Ecker JL, Thadhani R: **Inflammation and glucose intolerance: a prospective study of gestational diabetes mellitus.** *Diabetes Care* 2004, **27**(1):21–27.
4. Richardson AC, Carpenter MW: **Inflammatory mediators in gestational diabetes mellitus.** *Obstet Gynecol Clin North Am* 2007, **34**(2):213–224.
5. Black PH: **The inflammatory consequences of psychologic stress: relationship to insulin resistance, obesity, atherosclerosis and diabetes mellitus, type II.** *Med Hypotheses* 2006, **67**(4):879–891.
6. Heitritter SM, Solomon CG, Mitchell GF, Skali-Ounis N, Seely EW: **Subclinical inflammation and vascular dysfunction in women with previous gestational diabetes mellitus.** *J Clin Endocrinol Metab* 2005, **90**(7):3983–3988.
7. López-Tinoco C, Roca M, Fernández-Deudero A, García-Valero A, Bugatto F, Aguilar-Diosdado M, Bartha JL: **Cytokine profile, metabolic syndrome and cardiovascular disease risk in women with late-onset gestational diabetes mellitus.** *Cytokine* 2012, **58**(1):14–19.
8. Miadi-Messaoud H, Chouchane A, Abderrazek E, Debbabi H, Zaouali-Ajina M, Tabka Z, Ben-Jebria A: **Obesity-induced impairment of endothelium-dependent vasodilation in Tunisian women.** *Int J Obes* 2010, **34**(2):273–279.
9. Debbabi H, Bonnin P, Ducluzeau PH, Lefthérotis G, Levy BI: **Noninvasive assessment of endothelial function in the skin microcirculation.** *Am J Hypertens* 2010, **23**(5):541–546.
10. National Diabetes Data Group: **Classification and diagnosis of diabetes mellitus and other categories of glucose intolerance.** *Diabetes* 1979, **28**(12):1039–1057.
11. O'Sullivan JB: **Criteria for the glucose tolerance test in pregnancy.** *Diabetes* 1964, **13**:278–285.
12. Friedewald WT, Levy RI, Fredrickson DS: **Estimation of the concentration of low-density lipoprotein cholesterol in plasma without use of preparative ultracentrifuge.** *Clin Chem* 1972, **18**:499–502.
13. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC: **Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man.** *Diabetologia* 1985, **28**:412–419.
14. Morris SJ, Shore AC: **Skin blood flow responses to the iontophoresis of acetylcholine and sodium nitroprusside in man: possible mechanisms.** *J Physiol* 1996, **496**(2):531–542.
15. Christen S, Delachaux A, Dischl B, Golay S, Liaudet L, Feihl F, Waeber B: **Dose-dependent vasodilatory effects of acetylcholine and local warming on skin microcirculation.** *J Cardiovasc Pharmacol* 2004, **44**(6):659–664.

16. Newton DJ, Khan F, Belch JJ: **Assessment of microvascular endothelial function in human skin.** *Clin Sci* 2001, **101**(6):567–572.
17. Kubli S, Waeber B, Dalle-Ave A, Feihl F: **Reproducibility of laser Doppler imaging of skin blood flow as a tool to assess endothelial function.** *J Cardiovasc Pharmacol* 2000, **36**(5):640–648.
18. Paradisi G, Biaggi A, Ferrazzani S, De Carolis S, Caruso A: **Abnormal carbohydrate metabolism during pregnancy : association with endothelial dysfunction.** *Diabetes Care* 2002, **25**(3):560–564.
19. Emery SP, Levine RJ, Qian C, Ewell MG, England LJ, Yu KF, Catalano PM, CPEP study group: **Twenty-four-hour urine insulin as a measure of hyperinsulinaemia/insulin resistance before onset of pre-eclampsia and gestational hypertension.** *BJOG* 2005, **112**(11):1479–1485.
20. Barden A, Singh R, Walters BN, Ritchie J, Roberman B, Beilin LJ: **Factors predisposing to pre-eclampsia in women with gestational diabetes.** *J Hypertens* 2004, **22**(12):2371–2378.
21. Dandona P, Aljada A, Chaudhuri A, Mohanty P, Garg R: **Metabolic syndrome: a comprehensive perspective based on interactions between obesity, diabetes, and inflammation.** *Circulation* 2005, **111**(11):1448–1454.
22. Groenendijk M, Cantor RM, Blom NH, Rotter JJ, de Bruin TW, Dallinga-Thie GM: **Association of plasma lipids and apolipoproteins with the insulin response element in the apoC-III promoter region in familial combined hyperlipidemia.** *J Lipid Res* 1999, **40**(6):1036–1044.
23. Klein BE, Klein R, Lee KE: **Components of the metabolic syndrome and risk of cardiovascular disease and diabetes in Beaver Dam.** *Diabetes Care* 2002, **25**(10):1790–1794.
24. Mooradian AD, Haas MJ, Wong NC: **Transcriptional control of apolipoprotein A-I gene expression in diabetes.** *Diabetes* 2004, **53**(3):513–520.
25. Huda SS, Brodie LE, Sattar N: **Obesity in pregnancy: prevalence and metabolic consequences.** *Semin Fetal Neonatal Med* 2010, **15**(2):70–76.
26. Verrey F, Closs El, Wagner CA, Palacin M, Endou H, Kanai Y: **CATs and HATs: the SLC7 family of amino acid transporters.** *Pflugers Arch* 2004, **447**(5):532–542.
27. Sobrevia L, González M: **A role for insulin on L-arginine transport in fetal endothelial dysfunction in hyperglycaemia.** *Curr Vasc Pharmacol* 2009, **7**(4):467–474.
28. González M, Gallardo V, Rodríguez N, Salomón C, Westermeier F, Guzmán-Gutiérrez E, Abarzúa F, Leiva A, Casanello P, Sobrevia L: **Insulin-stimulated L-arginine transport requires SLC7A1 gene expression and is associated with human umbilical vein relaxation.** *J Cell Physiol* 2011, **226**(11):2916–2924.
29. Tsitsiou E, Sibley CP, D'Souza SW, Catanescu O, Jacobsen DW, Glazier JD: **Homocysteine transport by systems L, A and y+L across the microvillous plasma membrane of human placenta.** *J Physiol* 2009, **587**(Pt 16):4001–4013.
30. Flores C, Rojas S, Aguayo C, Parodi J, Mann G, Pearson JD, Casanello P, Sobrevia L: **Rapid stimulation of L-arginine transport by D-glucose involves p42/44(mapk) and nitric oxide in human umbilical vein endothelium.** *Circ Res* 2003, **92**(1):64–72.
31. Dandona P, Aljada A, Bandyopadhyay A: **Inflammation: the link between insulin resistance, obesity and diabetes.** *Trends in Immunology* 2004, **25**(1):4–7.
32. Athyros VG, Hatzitolios A, Karagiannis A, Didangelos TP, Iliadis F, Dolgyras S, Vosnakidis T, Vasilidiadis P, Malias I, Tziomalos K, Samouilidou M, Mikhailidis DP: **Initiative for a new diabetes therapeutic approach in a Mediterranean country: the INDEED study.** *Curr Med Res Opin* 2009, **25**(8):1931–1940.
33. Dye JF, Vause S, Johnston T, Clark P, Firth JA, D'Souza SW, Sibley CP, Glazier JD: **Characterization of cationic amino acid transporters and expression of endothelial nitric oxide synthase in human placental microvascular endothelial cells.** *FASEB J* 2004, **18**(1):125–127.
34. Arancibia-Garavilla Y, Toledo F, Casanello P, Sobrevia L: **Nitric oxide synthesis requires activity of the cationic and neutral amino acid transport system y+L in human umbilical vein endothelium.** *Exp Physiol* 2003, **88**(6):699–710.
35. Leiva A, Pardo F, Ramírez MA, Fariás M, Casanello P, Sobrevia L: **Fetoplacental vascular endothelial dysfunction as an early phenomenon in the programming of human adult diseases in subjects born from gestational diabetes mellitus or obesity in pregnancy.** *Exp Diabetes Res* 2011, doi:10.1155/2011/349286. Article ID 349286.
36. Westermeier F, Puebla C, Vega JL, Fariás M, Escudero C, Casanello P, Sobrevia L: **Equilibrative nucleoside transporters in fetal endothelial dysfunction in diabetes mellitus and hyperglycaemia.** *Curr Vasc Pharmacol* 2009, **7**(4):435–449.
37. Westermeier F, Salomón C, González M, Puebla C, Guzmán-Gutiérrez E, Cifuentes F, Leiva A, Casanello P, Sobrevia L: **Insulin restores gestational diabetes mellitus-reduced adenosine transport involving differential expression of insulin receptor isoforms in human umbilical vein endothelium.** *Diabetes* 2011, **60**(6):1677–1687.
38. Sobrevia L, Abarzúa F, Nien JK, Salomón C, Westermeier F, Puebla C, Cifuentes F, Guzmán-Gutiérrez E, Leiva A, Casanello P: **Review: Differential placental macrovascular and microvascular endothelial dysfunction in gestational diabetes.** *Placenta* 2011, **32**(2):S159–S164.
39. Ukkola O, Santaniemi M: **Adiponectin: a link between excess adiposity and associated comorbidities?** *J Mol Med* 2002, **80**:696–702.
40. Cho H, Mu J, Kim JK, Thorvaldsen JL, Chu Q, Crenshaw EB 3rd, Kaestner KH, Bartolomei MS, Shulman GI, Birnbaum MJ: **Insulin resistance and a diabetes mellitus-like syndrome in mice lacking the protein kinase Akt2 (PKB beta).** *Science* 2001, **292**(5522):1728–1731.
41. Senn JJ, Klover PJ, Nowak IA, Zimmers TA, Koniaris LG, Furlanetto RW, Mooney RA: **Suppressor of cytokine signaling-3 (SOCS-3), a potential mediator of interleukin-6-dependent insulin resistance in hepatocytes.** *J Biol Chem* 2003, **278**(16):13740–13746.
42. Dimmeler S, Fleming I, Fisslthaler B, Hermann C, Busse R, Zeiher AM: **Activation of nitric oxide synthase in endothelial cells by Akt-dependent phosphorylation.** *Nature* 1999, **399**(6736):601–605.
43. Emanuelli B, Peraldi P, Filloux C, Chavey C, Freidinger K, Hilton DJ, Hotamisligil GS, Van Obberghen E: **SOCS-3 inhibits insulin signaling and is up-regulated in response to tumor necrosis factor-alpha in the adipose tissue of obese mice.** *J Biol Chem* 2001, **276**:47944–47949.
44. Rui L, Yuan M, Frantz D, Shoelson S, White MF: **SOCS-1 and SOCS-3 block insulin signaling by ubiquitin-mediated degradation of IRS1 and IRS2.** *J Biol Chem* 2002, **277**:42394–42398.
45. Segura J, Campo C, Ruilope LM, Rodicio JL: **Do we need to target "prediabetic" hypertensive patients?** *J Hypertens* 2005, **23**(12):2119–2125.
46. Sowers JR, Frohlich ED: **Insulin and insulin resistance: impact on blood pressure and cardiovascular disease.** *Med Clin North Am* 2004, **88**(1):63–82.
47. Dandona P, Aljada A, Bandyopadhyay A: **Inflammation: the link between insulin resistance, obesity and diabetes.** *Trends Immunol* 2004, **25**(1):4–7.
48. Aljada A, Ghanim H, Mohanty P, Kapur N, Dandona P: **Insulin inhibits the pro-inflammatory transcription factor early growth response gene-1 (Egr)-1 expression in mononuclear cells (MNC) and reduces plasma tissue factor (TF) and plasminogen activator inhibitor-1 (PAI-1) concentrations.** *J Clin Endocrinol Metab* 2002, **87**(3):1419–1422.
49. Dandona P, Aljada A, Mohanty P, Ghanim H, Hamouda W, Assian E, Ahmad S: **Insulin inhibits intranuclear nuclear factor kappaB and stimulates I kappaB in mononuclear cells in obese subjects: evidence for an anti-inflammatory effect?** *J Clin Endocrinol Metab* 2001, **86**(7):3257–3265.
50. Sivan E, Boden G: **Free fatty acids, insulin resistance, and pregnancy.** *Curr Diab Rep* 2003, **3**(4):319–322.
51. Kirwan JP, Hauguel-De Mouzon S, Lepercq J, Challier JC, Huston-Presley L, Friedman JE, Kalhan SC, Catalano PM: **TNF-alpha is a predictor of insulin resistance in human pregnancy.** *Diabetes* 2002, **51**(7):2207–2213.
52. Hauguel-de Mouzon S, Guerre-Millo M: **The placenta cytokine network and inflammatory signals.** *Placenta* 2006, **27**(8):794–798.
53. Radaelli T, Lepercq J, Varastehpour A, Basu S, Catalano PM, Hauguel-De Mouzon S: **Differential regulation of genes for fetoplacental lipid pathways in pregnancy with gestational and type 1 diabetes mellitus.** *Am J Obstet Gynecol* 2009, **201**(2):209e1–209e10.

doi:10.1186/1476-511X-12-93

Cite this article as: Mrizak et al.: Inflammation and impaired endothelium-dependant vasodilatation in non obese women with gestational diabetes mellitus: preliminary results. *Lipids in Health and Disease* 2013 **12**:93.