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RESEARCH

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# Population genetics analysis during the elimination process of *Plasmodium falciparum* in Djibouti

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## Abstract

**Background:** Case management of imported malaria within the context of malaria pre-elimination is increasingly considered to be relevant because of the risk of resurgence. The assessment of malaria importation would provide key data i) to select countries with propitious conditions for pre-elimination phase and ii) to predict its feasibility. Recently, a sero-prevalence study in Djibouti indicated low malaria prevalence, which is propitious for the implementation of pre-elimination, but data on the extent of malaria importation remain unknown.

**Methods:** Djiboutian plasmodial populations were analysed over an eleven-year period (1998, 1999, 2002 and 2009). The risk of malaria importation was indirectly assessed by using plasmodial population parameters. Based on 5 microsatellite markers, expected heterozygosity (H.e.), multiplicity of infection, pairwise  $F_{st}$  index, multiple correspondence analysis and individual genetic relationship were determined. The prevalence of single nucleotide polymorphisms associated with pyrimethamine resistance was also determined.

**Results:** Data indicated a significant decline in genetic diversity (0.51, 0.59, 0.51 and 0 in 1998, 1999, 2002 and 2009, respectively) over the study period, which is inconsistent with the level of malaria importation described in a previous study. This suggested that Djiboutian malaria situation may have benefited from the decline of malaria prevalence that occurred in neighbouring countries, in particular in Ethiopia. The high  $F_{st}$  indices derived from plasmodial populations from one study period to another (0.12 between 1999 and 2002, and 0.43 between 2002 and 2009) suggested a random sampling of parasites, probably imported from neighbouring countries, leading to oligo-clonal expansion of few different strains during each transmission season. Nevertheless, similar genotypes observed during the study period suggested recurrent migrations and imported malaria.

**Conclusion:** In the present study, the extent of genetic diversity was used to assess the risk of malaria importation in the low malaria transmission setting of Djibouti. The molecular approach highlights i) the evolution of Djiboutian plasmodial population profiles that are consistent and compatible with Djiboutian pre-elimination goals and ii) the necessity to implement the monitoring of plasmodial populations and interventions at the regional scale in the Horn of Africa to ensure higher efficiency of malaria control and elimination.

**Keywords:** Microsatellites, Molecular epidemiology, Genetic diversity, Drug resistance, Malaria elimination, Pyrimethamine, Horn of Africa

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## Background

According to the World Malaria Report 2012 [1], there were about 219 million cases of malaria (with an uncertainty range [10 to 90 percentile] of 154 million to 289 million) and an estimated 660,000 deaths in 2010 (with an uncertainty range of 610,000 to 971,000). Malaria mortality rates have fallen by more than 25% globally since 2000 and by 33% in the World Health Organization (WHO) African region [2]. Today, the WHO is considering that in some regions characterized by favourable conditions, malaria elimination has become a realistic goal [3-5]. As a first step, these areas are eligible for pre-elimination stage. Local malaria transmission can subsequently be interrupted in these areas through incremental stages. A progressive decline in malaria transmission is expected to reduce the surface area of malaria-endemic region, country by country, from hypo-endemic to hyper-endemic areas [6].

However, malaria control and the pre-elimination process may be hampered by multiple insecticide and drug resistance of mosquito vectors and malaria parasites, respectively. In order to limit the impact of drug-resistant malaria and prevent its spread, monitoring of drug resistance is strongly recommended [5]. Moreover, imported malaria may hamper malaria control and cause malaria resurgence in countries that have reached the elimination phase [7,8]. The higher the numbers of imported cases in a country in the pre-elimination phase, the greater will be the risk that malaria returns and even rebounds, and more resources for combating malaria will be required to attain elimination [9]. Therefore, according to the incremental elimination strategy, areas with limited cases of imported malaria should be enlisted first in the pre-elimination phase. When elimination is attained in those areas, malaria elimination in neighbouring areas with higher malaria transmission level may be considered.

Since the level of local malaria endemicity and the number of imported cases are two key factors for the assessment of feasibility and estimation of resources required for the pre-elimination step, national surveillance system in a country aiming for pre-elimination should focus on these two determinants. To that end, classical sero-epidemiological methods as described by Corran *et al.* [10] and Drakeley *et al.* [11] may be completed by a population genetics approach which could take advantage of the population plasticity driven by evolutionary forces. For example, selection pressure or migrations alter parasite populations, and these modifications in plasmoidal population would lead to genetic changes, which can be monitored. Analysis of allelic frequencies in plasmoidal populations is an efficient way to describe such micro-evolution [12-20].

As allelic frequencies within parasite populations are summarized by genetic diversity defined by expected heterozygosity (He),  $[H_e = n/(n-1)][1 - \sum_{i=1}^n p_i^2]$  [21], the

determination of He is an efficient way to monitor changes occurring at the population level. Interestingly, He is positively correlated with local malaria transmission level [22-26]. Low local transmission levels are associated with low He [27,28]. Conversely, high transmission levels are associated with an increase in He [24,29]. He values may also be associated with human migrations which appear to be the main parasite 'transporter and provider' and therefore a 'promoter' of genetic diversity [22,30,31], even over long distances [32]. This is consistent with the correlation observed between human migration rate and genetic similarities among malaria parasite populations in an archipelago model [33]. Local levels of malaria transmission and the number of imported malaria cases are therefore positively correlated with genetic diversity. Since pre-elimination programme aims to decrease local transmission level and control parasite importations, both of these factors together should be associated with a decrease in plasmoidal genetic diversity (He) during the process of pre-elimination.

Additional information on the evolution of allelic frequencies may be obtained from Fst index [34]. This index may be considered as a measure of the variance in allelic frequencies between populations [35]. Pairwise comparison between years over a study period highlights the global evolution and allows better understanding of the parasite turnover rate (i.e. high pairwise Fst between years is consistent with high turnover). Analysis of genotype frequencies and their similarity is also informative to assess and determine the origin of imported malaria cases. It may be possible to identify genetic lineage or recurrent genotypes. In order to derive valid data from genetic analysis, an establishment of genotype database from eligible areas is necessary before the pre-elimination process. In this way, it will be possible to monitor the impact of human actions on *Plasmodium falciparum* populations.

The Republic of Djibouti exhibits an interesting malaria situation to illustrate the usefulness of population genetics analysis within the context of malaria elimination. This country should be eligible for pre-elimination [6,36,37]. Djibouti (geographic coordinates of Djibouti city, the capital of the country, 11°36'N 43°10'E) [38] was formerly known to be a meso- to hypo-endemic country with unstable malaria [39-41]. *P. falciparum* and *P. vivax* co-exist in the country, but the former is the predominant (> 80%) species. The major vector is *Anopheles arabiensis* [1,42,43]. Its climate is semi-arid, with a mean annual rainfall of 147 mm [38]. Over the past 12 years, malaria incidence, as well as recurrence of epidemics, has decreased [44]. The epidemiologic strata have been illustrated by few micro-epidemics [32], with the exception of a major malaria outbreak that occurred in 1999 [45]. Sero-prevalence data from 2002 [43] confirmed the

existence of residual malaria foci. In the city of Djibouti, hotspots occurred on both sides of the main wadi (i.e., Ambouli wadi) and in the quarters of Arhiba and Balbala 4 (Additional file 1 from Khairah *et al.* [43]) where large populations of migrants who travel regularly to and/or from Ethiopia reside [46]. Across the Republic, *P. falciparum* sero-prevalence rates were higher in the southern regions (Dikhil and Ali-Sabieh), particularly along the land routes to Ethiopia, i.e., Tammiro/As-Eyla and Ali-Sabieh (Additional file 2 from Khairah *et al.* [43]). These roads are used frequently by professional truckers, private users, and migrants because they are the only two roads connecting Djibouti and Ethiopia [47].

Recently, a national sero-prevalence investigation including 7,151 individuals (from November 2008 to January 2009) throughout the country (150 clusters of 75 households from the capital [40%] and districts [60%]) confirmed the very low malaria prevalence (0.58%) [48]. National sero-prevalence data are consistent with the decrease in malaria incidence monitored by three main medical services in the city of Djibouti (Additional file 3) where almost two-thirds of the population reside (61.7%, according to the official Djiboutian publication of census [49]). Considering this high Djiboutian urbanization and movement of people between the capital and the rest of the country [49], this correspondence between inside and outside the capital was expected. After the outbreak in 1999, the first decrease in malaria incidence occurred in 2001 (4-fold decrease) and the second decrease in 2006 (11-fold decrease) (Additional file 3). Although the decrease in malaria burden was in favour of the Djiboutian eligibility for pre-elimination step, there are no field data on the risk of malaria importation that could hamper the elimination process. In order to adapt malaria control programme in this country, it would be useful to characterize the circulating *P. falciparum* isolates in Djibouti and importations from neighbouring countries.

In the late 1980s, a study showed multiple events of malaria importation from neighbouring countries [41]. Moreover, previous molecular data suggested the existence of parasite exchange sufficient to provide a moderate genetic diversity [32,45], despite seasonal interruption of local malaria transmission [39,45]. However, the actual extent of imported malaria remains unknown. What is the genetic diversity of imported malaria? What is the parasite turnover rate? The success of the pre-elimination programme will depend, in part, on the responses to these questions.

To complete the understanding of Djiboutian malaria epidemiologic strata within the context of malaria pre-elimination, an indirect genetic approach was adopted. Analyses of genetic diversity, as well as those of relationships among parasites collected over a period of 11 years, shed light on the above-mentioned questions.

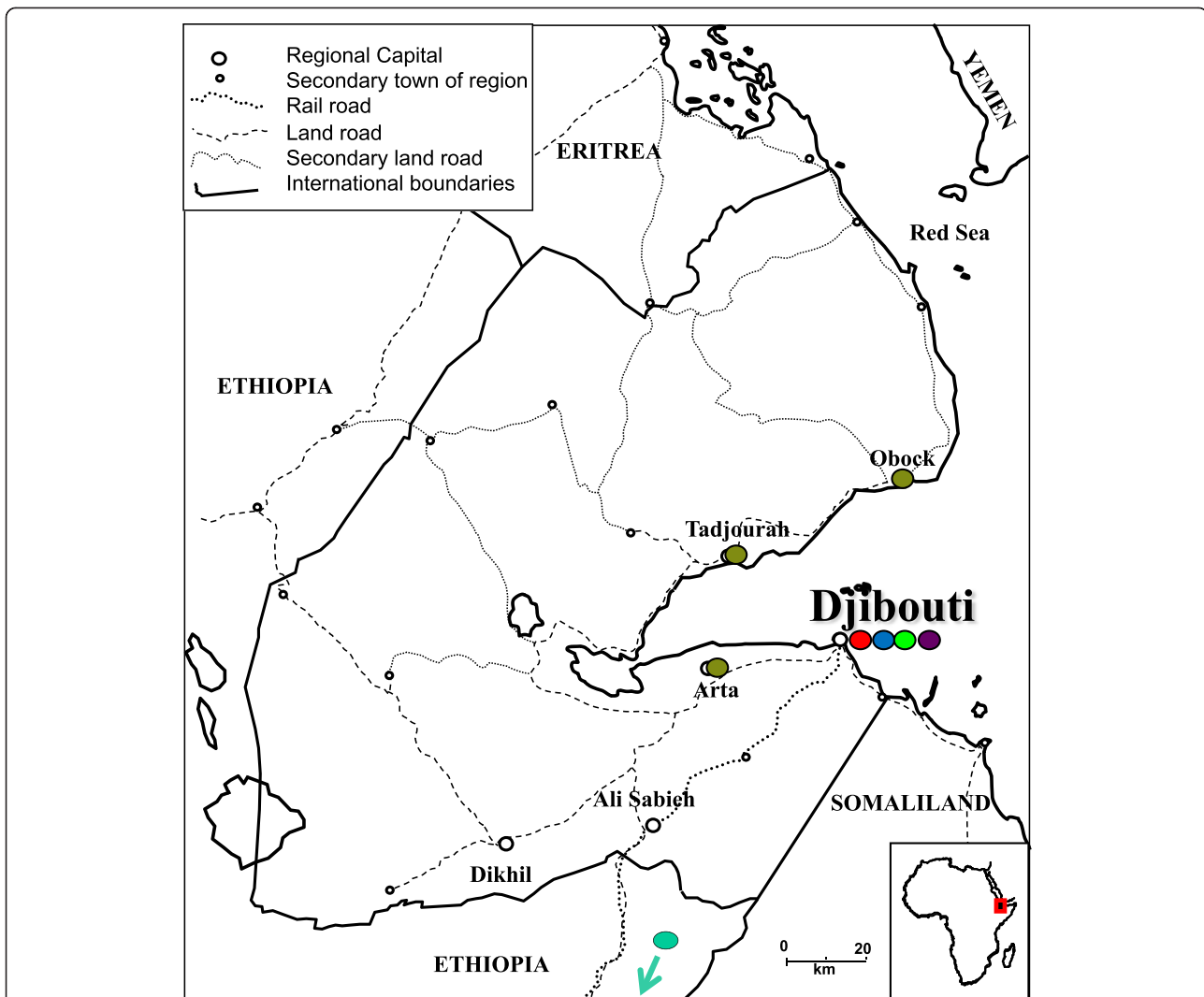
Multilocus microsatellite genotyping was performed in *P. falciparum* isolates collected over 11 years (1998, 1999, 2002, and 2009). Based on WHO recommendations on drug resistance monitoring [3,5] and the fact that sulphadoxine-pyrimethamine is the current partner molecule of artesunate (artemisinin-based combination therapy [ACT]) for the first-line treatment employed in Djibouti since 2008 [50], polymorphisms associated with resistance to pyrimethamine were also genotyped.

## Methods

### *Plasmodium falciparum* isolates

Isolates were collected four times over an 11-year period (1998, 1999, 2002, and 2009). The first three investigations were conducted at the Centre Hospitalier des Armées Bouffard, a French military hospital in Djibouti city serving Djiboutian military and native civilians residing in the city, and other public health facilities in Djibouti city. Blood samples were collected from symptomatic patients with *P. falciparum* who had not travelled outside the city of Djibouti during the preceding month and who denied self-medication with an anti-malarial drug before consultation. Forty-six blood samples were collected between September and December 1998, 61 in April 1999, and 32 between March and May 2002. Venous blood (5 mL) was collected in ethylene diamine tetra acetic acid (EDTA)-coated Vacutainer® tubes (Becton Dickinson, Rutherford, NJ, USA). Aliquots of freshly collected blood were kept at -20°C until DNA extraction. In 2009, during the fourth investigation, 42 samples were provided by the Djiboutian Malaria Control Unit (Ministry of Health). These samples, collected during an investigation of malaria seroprevalence detailed elsewhere, were obtained from three sites outside Djibouti city: Arta, Tadjourah, and Obock [48] (Figure 1). One blood sample was obtained from a Djiboutian military recruit who presented with clinical malaria in January 2009 after staying at the military training camp in Debrezeit (Ethiopia) for 10 months prior to blood collection. Three additional isolates from Ethiopia sampled in 2008 were provided by the Department of Parasitology and Vector Borne Diseases (Ethiopian Health and Nutrition Research Institute).

Blood samples were spotted onto Whatman® 3 MM filter paper, and all samples were frozen and kept at -20°C in the national laboratory at Hôpital General Peltier in Djibouti city. The study was cleared and approved by the Djiboutian Ministry of Health (authorization no. 48/INSP/MS/2012). Informed consent was obtained from all patients before blood collection. Sampling details on geographic location and sampling period are described elsewhere [48]. The characteristics of laboratory-confirmed *P. falciparum* malaria at four different time points are summarized in Figure 1.



**Figure 1** Locations of *Plasmodium falciparum*-positive cases sampled in the Republic of Djibouti (1998, 1999, 2002, and 2009)

Sampling sites where *Plasmodium falciparum* cases were diagnosed with rapid diagnostic tests and confirmed by PCR. Forty-six blood samples were collected from September to December 1998 (red dot), 61 in April 1999 (blue dot), and 32 from March to May 2002 (light green dot) from symptomatic patients with *P. falciparum* who had not travelled outside the city of Djibouti during the preceding month and declared not to have taken any antimalarial drug before blood sampling. In 2009, 42 Djiboutian samples were collected from November 2008 to January 2009 by the Djiboutian Malaria Control Unit (Ministry of Health) (khaki green) [48]. One blood sample was obtained from a Djiboutian military recruit who presented with clinical malaria (purple dot) in January 2009 and who stayed at the training camp in Debrezeit (Ethiopia) for 10 months prior to blood collection. Three blood samples provided by the Department of Parasitology and Vector Borne Diseases (Ethiopian Health and Nutrition Research Institute) were collected in Southeast Ethiopia (dark green dot) in 2008. According to the official census of Djiboutian population, Djibouti exhibits a high urbanization rate. Djibouti city, Ali Sabieh, Dikhil, Tadjourah, Arta, and Obock account for 62%, 11%, 10%, 9%, 5%, and 4% of the national population, respectively [49].

### Molecular markers

DNA was extracted from filter papers and frozen whole blood using EZNA blood DNA kit (Biofidal, Vaulx-en-Velin, France) according to the manufacturer's recommendations. Microsatellite genotyping was performed by amplification fragment length polymorphism (AFLP) analysis. Five complex and putatively neutral microsatellite loci [32] (*Pf*2689, *7A11*, *C4M79*, *Pf*2802, and *TRAP*) were amplified by nested PCR with fluorescent end-labelled

primers, as described previously [32]. Thermocycling was performed using a Biometra (Goettingen, Germany) 96-well T1 thermocycler. The accession numbers and primer sequences are summarized in Table A (Additional file 4).

Single nucleotide polymorphisms (SNPs) of *P. falciparum* dihydrofolate reductase (*Pf**dhfr*) gene at codons 16, 51, 59, 108, and 164 associated with resistance to pyrimethamine and cycloguanil (i.e., the biologically active metabolite of proguanil [51]) were determined using



a primer extension method (SNaPshot®), as described in an earlier study [52]. The accession numbers are presented in Table A (Additional file 4).

AFLP and SNaPshot® products were analysed by capillary electrophoresis on polyacrylamide gels using ABI 3130XL® sequencer (Applied Biosystems®, Warrington, UK). Electrophoregram was interpreted using Genemapper® 4.0 software (Applied Biosystems®, Carlsbad, CA, USA).

### Statistical analysis

The multiplicity of plasmodial infection (MOI, defined as the number of genetically distinguishable parasites per isolate) was estimated for each isolate from the microsatellite locus that exhibited the highest number of alleles. The mean MOI for each collection year of *P. falciparum* population (1998, 1999, 2002, and 2009) was calculated.

The evolution of genetic diversity among Djiboutian *P. falciparum* populations is dependent on local transmission level and parasite flow, a source of genetic diversity. Genetic diversity was assessed by the Nei unbiased expected heterozygosity index [21],  $H_e = [n / (n - 1)] [1 - \sum_{i=1}^n p_i^2]$  (where  $n$  is the number of isolates sampled and  $p_i$  is the frequency of the  $i$ th allele) and calculated from allelic frequencies of five microsatellite loci using GENETIX software version 4.05 [53]. Pairwise comparisons of  $H_e$  values among the four collection periods were performed using FSTAT software version 2.9.4, with a 10,000 permutations bilateral comparison test [54].

Genetic similarity of plasmodial populations was investigated using Wright F statistic ( $F_{ST}$ ) [34]. Pairwise comparisons among the collection years (1998, 1999, 2002, and 2009) were performed based on microsatellite genotype frequencies using FSTAT software version 2.9.4 [35,54].  $F_{ST}$  is a comparison of the sum of genetic variability within and between populations based on the differences in allelic frequencies.  $F_{ST}$  values were interpreted as no differentiation (0), low genetic differentiation ( $>0 - 0.05$ ), moderate differentiation (0.05-0.15), and high differentiation (0.15-0.25).

Multiple correspondence analyses (MCA), also known as factorial correspondence analysis (FCA) [55] according to multilocus genotypes, were conducted to illustrate the genetic similarity of plasmodial populations during the study period. FCA was performed by considering population centroids as active points, using GENETIX software, as described in the help menu [53]. The graphical representation with 95% data concentration ellipse (i.e., including 95% of the projected genotypes on the FCA plan) and centroids was obtained using R software, version R 2.15.1 [56].

The relationships between parasite genotypes were assessed using eBurst algorithm. Based on microsatellite allelic profiles, the algorithm selects the most parsimonious

patterns of genotype evolution and predicts founder(s). The assignment of founders is tested by a bootstrap procedure [57]. The global optimization of the diagram based on goeBurst algorithm [58] was performed with Phyloziz software [59]. The eBurst algorithm implements a simple model of clonal expansion and diversification which is generally used to represent a population of clonal prokaryote [57]. Despite its obligatory sexual stage, *P. falciparum* may evolve as a clonal organism due to inbreeding [30,60-65]. A high self-fertilization rate (i.e. syngamy between genetically identical gametes) may be favoured in low malaria transmission settings [24,25,30,66]. Outbreaks are an extreme situation where the oligo-clonal spreading of parasites (i.e., only few plasmodial populations propagate during epidemics) may occur [67-69]. As malaria epidemics had occurred in Djibouti with oligo-clonal expansion of plasmodial populations [32,45], eBurst diagram is particularly well adapted for the description of Djiboutian malaria situation.

Comparison between the global unstratified eBurst diagram and eBurst diagram stratified by the sampling year was performed to assess the robustness of eBurst algorithm.

The index of discriminatory power (D) was assessed in order to estimate the discriminatory power of the genotyping based on four microsatellite loci. This index is calculated from the number of genotypes and their relative frequencies. The index D is the probability that two unrelated parasites randomly sampled from studied population display different genotypes. By analogy with Nei unbiased expected heterozygosity index ( $H_e$ ), D is an indirect measure of genotypic diversity. According to Hunter and Gaston's formula [70],  $D = 1 - 1 / (N(N - 1) \sum_{j=1}^S n_j(n_j - 1))$  where  $N$  is the total number of parasites in the sample population,  $s$  is the total number of genotypes observed, and  $n_j$  is the number of strains with the  $j$ th genotype.

## Results

### Genetic diversity

When the combination of the profiles at five microsatellite loci was considered, 57, 66 and 43 genotypes were successfully obtained from 46, 61, and 32 samples collected in 1998, 1999, and 2002, respectively. Among 42 samples collected in 2009, the Pf2802 locus did not show enough profiles to perform analysis. The low success rate of Pf2802 genotyping was probably due to: i) the longer amplified fragment by the first PCR that is more sensitive to the quality of DNA template, or ii) the presence of null allele in oligo-clonal plasmodial populations. Therefore, the combination of the profiles at four microsatellite loci was considered, and 36 genotypes were observed in 2009.

Based on the microsatellite loci, a moderate genetic diversity ( $H_e$ ) was observed during the four-year period from 1998 ( $H_e = 0.51 \pm 0.20$ ) to 2002 ( $H_e = 0.51 \pm 0.14$ ). At the

end of the study period in 2009, He decreased considerably and there was no observable genetic diversity (He = 0).

The MOI showed a similar pattern, starting from the baseline mean ( $\pm$  SD) MOI of  $1.86 \pm 0.81$  in 1998 to the MOI value of 1.0 in 2009. This result is particularly noteworthy, considering that the isolates were sampled in 2009 from three areas outside Djibouti city (Arta, Tadjourah, and Obock) during an interval of only a few weeks.

The numbers of alleles by locus that estimate the number of distinct genotypes were 7.8, 8.4, 6.0, and 1.0 in 1998, 1999, 2002, and 2009, respectively. These results suggested a similar tendency as MOI and He values. The results on genetic diversity, MOI, and numbers of alleles are summarized in Table 1.

#### Parasite genetic similarity: population and individual analysis

The pairwise comparisons of plasmodial populations among different years of sample collection displayed statistically significant differences in Fst indices (Fst > 0.12), except between 1998 and 1999 (Fst = 0). The highest Fst index (>0.39) was observed between the isolates in 2009 and those collected during the preceding years. Contiguous centroids and superimposed ellipses of plasmodial populations in 1998 and 1999 were obtained (Figure 2). Plasmodial population in 2002 appeared dissimilar from the other three populations (Fst > 0.12).

Genotype relationships provide additional information about individual relationships and yearly genotype turnover (Figure 3). Based on the analysis of four microsatellite loci, a total of 62 *P. falciparum* genotypes were detected in the present study. Clustering among the isolates collected during the same year was consistent with significant Fst indices between different years. During the 11-year period, the present data exhibited 10, 18, 10, and 0 private genotypes observed only in 1998, 1999, 2002 and 2009, respectively (2 genotypes observed only in 2008 were sampled in southeast Ethiopia). However, 16 genotypes were present across several study periods. Among these, 12 genotypes observed in 1998 were still present in 1999. Major genotypes were even present for

three or four years. Genotype 32 was present in 1998, 1999, and 2002 and also occurred among imported malaria parasites from Ethiopia in 2008 (Figures 3 and 4). Genotype 38 also occurred in 1998, 1999, and 2002. Moreover, genotypes were closely-related from year to year. Most of the new genotypes differed at only one locus from genotypes that were observed in earlier years (i.e., single-locus variant, SLV). The global, unstratified eBurst diagram, without considering the year of sampling, involved only five double-loci variants (DLVs).

In 2009, all Djiboutian isolates exhibited identical genotypes at four microsatellite loci (PF2689, C4M79, TRAP, and 7A11). This genotype, i.e., genotype 36, was observed earlier in 1998. Genotype 27 observed in *P. falciparum* isolated from a Djiboutian military recruit who stayed in a military training camp in Debrezeit (Ethiopia) (8° 45' 00" N 38° 59' 00" E) for 10 months in 2009 was observed earlier in 1999. Ethiopian genotypes found in the present study were either similar to Djiboutian genotypes (genotype 32) or differed by only one locus from Djiboutian genotypes (genotypes 39 and 46).

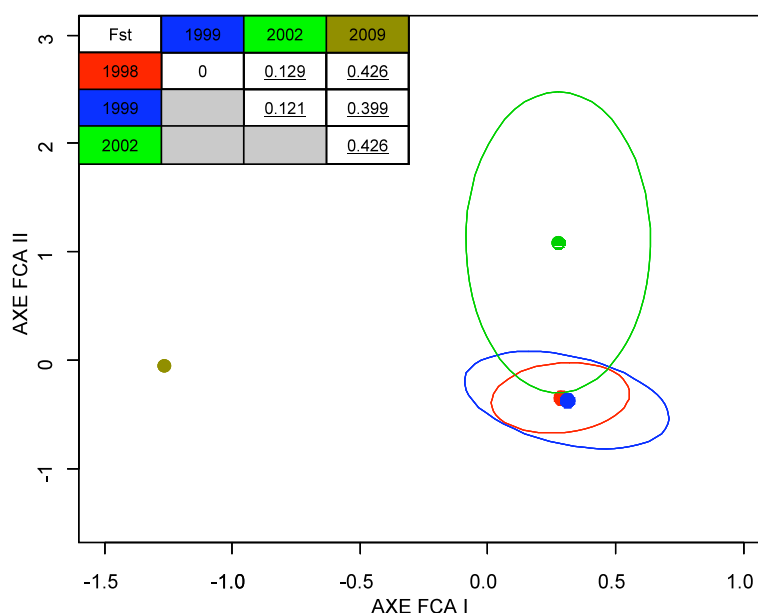
Eburst diagram stratified by the year of detection (Figure 4) showed a similar pattern, with two major genotypes (genotypes 38 and 32) linked to numerous SLVs. This analysis avoided few links between years that had probably not occurred (13 links were modified) and pointed out more clearly the diversification of genotypes over time. Based on the eBurst diagrams of parasites collected in 1998 and 1999, six (genotypes 2, 17, 27, 37, 48, and 54) and five (genotypes 5, 22, 24, 59, and 62) new SLVs emerged in 1999 from genotypes 32 and 38, respectively. Despite this diversification, genetic lineage (56, 38, 32, 49, 51, and 61) was maintained from year to year, and major genotypes were similar (genotypes 32 and 38). To a lesser extent, in 2002, diversification occurred from an earlier major genotype (32).

Based on the frequencies of 62 genotypes observed among 225 parasites, the index of discriminatory power (D) [70] was 91%. The index D by year was 0.87, 0.89, and 0.89 for 1998, 1999, and 2002, respectively. Despite the closely-related genotypes and even recurrent genotypes

**Table 1 Genetic diversity of *Plasmodium falciparum* and evolution of multiplicity of infection (MOI) in the Republic of Djibouti (1998–2009)**

Populations	Microsatellites, individual He						Mean He [ $\pm$ SD]	MOI [ $\pm$ SD]	Mean number of alleles/ locus
	n	7A11	C4M79	Pf 2689	Pf 2802	TRAP			
<b>1998</b>	46	0.5427(72)	0.2838(57)	0.3163(62)	0.6944(66)	0.7060(66)	0.5086[ $\pm$ 0.20]	1.36[ $\pm$ 0.81]	7.8
<b>1999</b>	61	0.5835(76)	0.4585(66)	0.4381(75)	0.7336(80)	0.7245(81)	0.5876[ $\pm$ 0.14]	1.42[ $\pm$ 0.79]	8.4
<b>2002</b>	32	0.4493(47)	0.1762(43)	0.4615(46)	0.7281(46)	0.7467(47)	0.5124[ $\pm$ 0.23]	1.12[ $\pm$ 0.38]	6.0
<b>2009</b>	42	0(42)	0(41)	0(42)	NA	0(36)	0	1.00	1.0

He, Nei unbiased expected heterozygosity index; SD standard deviation, MOI multiplicity of infection mean number of alleles/locus, n, number of samples, NA, no results detected. The number of genotypes successfully obtained is in parentheses.



**Figure 2** Analysis of genetic similarity between Djiboutian *Plasmodium falciparum* populations (1998, 1999, 2002, and 2009) *Plasmodium falciparum* populations in Djibouti (1998 red dot, 1999 blue dot, 2002 light green dot, and 2009 khaki dot; sampling details are presented in the text and Figure 1) were compared based on the genotyping of four microsatellites. Pairwise  $F_{st}$  indices between the years studied are presented in the table at the top left. Statistically significant  $F_{st}$  values ( $P < 0.05$ ) are underlined. Multiple correspondence analysis (also called factorial correspondence analysis) was performed with Genetix software [53], using populations' centroid as the active point. Axes FCA I and FCA II are the first two order factors representing 51.7% and 42.4% of the total inter-centroid variance, respectively. Ellipses around the centroids enclose 95% of the projected genotypes on the FCA plan and were calculated and graphically represented using R software [56]. Genotype frequencies in 2009 did not allow ellipse representation.

observed, these genetic analyses allowed monitoring of parasite populations on a fine scale.

#### Evolution of drug resistance: mutations associated with pyrimethamine resistance

Genotyping of all five *Pfdhfr* codons (16, 51, 59, 108 and 164) associated with pyrimethamine resistance was successful in 36 of 46 (78%), 50 of 61 (82%), and four of 32 (12.5%; only five samples were available after microsatellite study [32]) samples collected in 1998, 1999, and 2002, respectively. Thirteen of 42 (31%) samples obtained in 2009 were successfully genotyped, with the exception of codon 51. The prevalence of mutations in relation to the study periods is summarized in Table 2.

In 1998, there was no triple mutant (defined as N51I, C59R, and S108N) and only one isolate (2.8%) carried C59R and S108N amino acid substitutions. In 1999, two (4%) isolates were triple mutants. In 2002, the prevalence of triple mutants dropped back to 0%. On the contrary, 13 (100%) isolates were double mutants (C59R and S108N) in 2009.

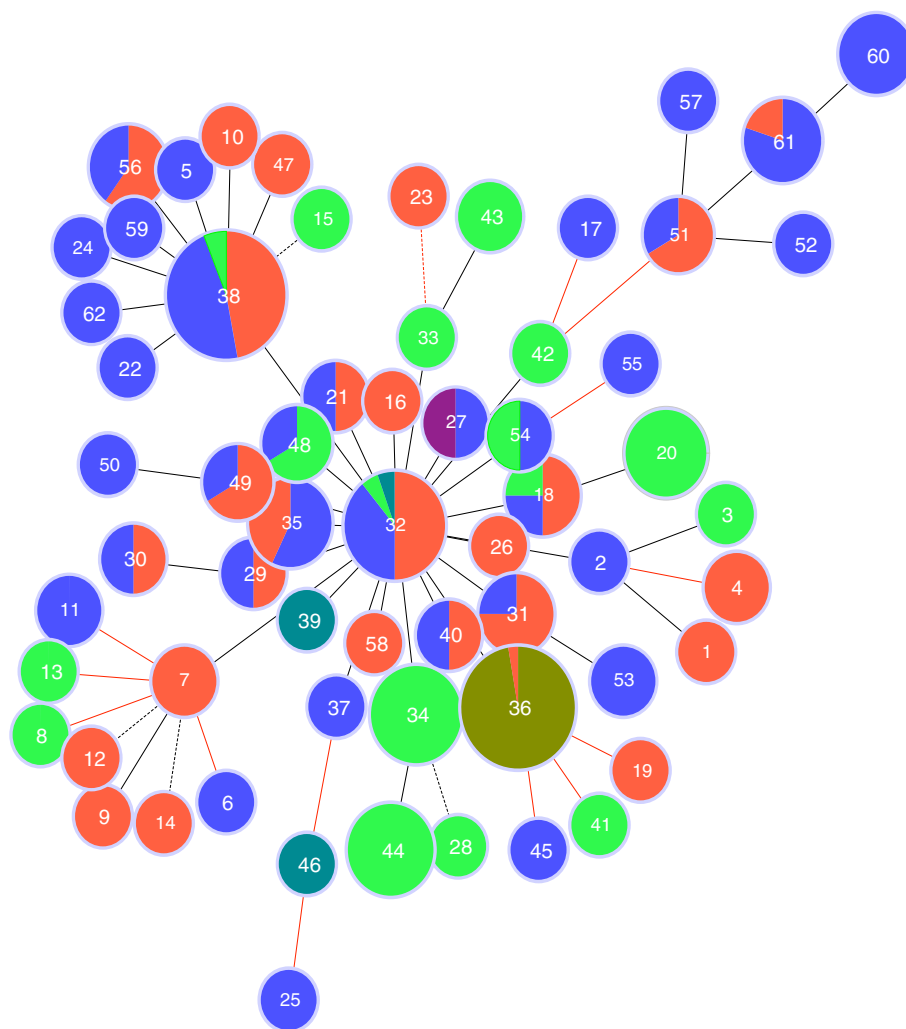
#### Discussion

In this present study, genetic diversity was analysed and applied in innovative ways to assess the risk of malaria importation into low malaria transmission settings. Based

on the results of microsatellite allele frequencies, the present study highlights an important modification in Djiboutian *P. falciparum* population. First, a moderate genetic diversity ( $He = 0.51$ ) was observed from 1998 to 2002, with a slight increase in 1999 ( $He = 0.59$ ). These results are in agreement with antigenic data and previous microsatellite analysis, which showed a similar level of genetic diversity ( $He = 0.53$ ) [32]. Secondly, at the end of the study period, a significant decline in genetic diversity was recorded ( $He_{2009} = 0$ ). This diminution is all the more important because, in 2009, mass screening for malaria was conducted over a period of three months in 150 sites throughout the country, and *P. falciparum*-positive cases were found in only three sites. These three sites are separated by hundreds of kilometres (by road), as compared with samples collected before 2009, which originated from Djibouti city only. In this context, the sampling bias in 2009 should have resulted in an opposite trend. The evolution of genetic diversity is confirmed by a similar reduction in the numbers of alleles (7.8 to 1) and the decrease in MOI at the microsatellite loci analysed during the study period.

Concerning the association between the local level of malaria transmission and plasmodial population parameters, such as genetic diversity and MOI [24,27,29,71-73], the observed significant decrease in malaria transmission

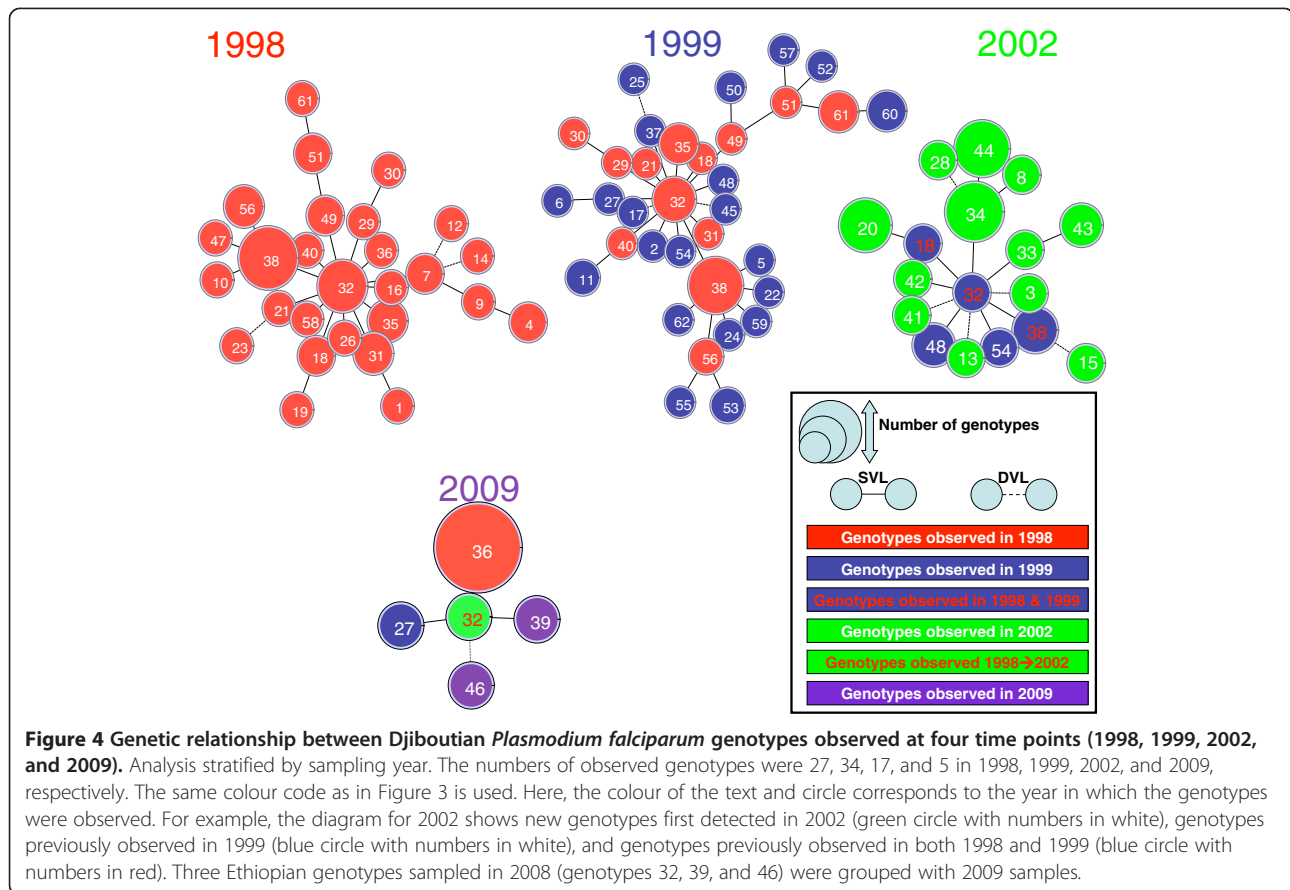




**Figure 3 Genetic relationship between Djiboutian *Plasmodium falciparum* genotypes observed at four time points (1998, 1999, 2002, and 2009). Unstratified analysis that does not take into account the year of sampling.** This graphical representation of relationships between closely-related genotypes of *Plasmodium falciparum* is based on eBurst algorithm [57]. A global optimization selects the more likely evolutionary pattern and highlights potential ancestral (or founding) genotype(s) of cluster of closely-related genotypes [58,59]. A total of 62 genotypes were found (numbers 1 to 62). A separation level between two distinct genotypes is represented by i) a solid line, a different profile in only one locus of microsatellite marker, i.e., single-locus variant (SLV) and ii) a dotted line, a double-loci variant (DLV). The distance between genotypes in the diagram does not have any relation with genetic distance between genotypes. The area of circles depends on the number of individuals with the same genotype. The colour code is as follows: 1998 (red dot), 1999 (blue dot), 2002 (light green dot), and 2009 (khaki dot). In 2009, three malaria cases originated from Southeast Ethiopia (dark green dot) and one from Debrezeit, Ethiopia (purple dot). The proportion of each colour inside one circle depends on the number of individuals with the same genotype for the year considered. Red lines indicate relationships that are modified in stratified analysis.

in Djibouti is in agreement with the results of the recent sero-epidemiological study [48]. Interestingly, such a decrease in genetic diversity is not consistent with the level of malaria importation reported previously [32,41]. So far, even with an interruption of local malaria transmission for several months during the dry season, a long-term carriage of *P. falciparum* [71,74] and, more probably, imported malaria cases were able to maintain an intermediate diversity level [ $0.51 < He_{1998-1999-2002} < 0.58$ ] [32]. Therefore, the absence of genetic diversity i) confirms the decline in the

level of local malaria transmission, and ii) strongly suggests a reduction of imported malaria cases in Djibouti. These observations highlight the likely benefits of a coordinated malaria control programme with the neighbouring countries [75-77]. Since Ethiopia is the main source of imported malaria cases in Djibouti due to geographic proximity and socio-epidemiologic context [39,41,78], a successful Roll Back Malaria Programme in Ethiopia is expected to lower the number of imported malaria cases from Ethiopia to Djibouti.



Based on the analysis of genotype frequencies, a high  $F_{st}$  index ( $>0.15$ ) and distant centroids (Figure 2) between sampling years strongly support a considerable plasmodial population turnover during the study period. These clusters at different time points may be attributable to sub-sampling of plasmodial populations from neighbouring countries (in particular Ethiopia) through human migration. As this random sampling occurred each year, the source of different strains involved in epidemics may vary over time. As fewer parasite strains are imported into Djibouti, the allelic frequencies in plasmodial populations will be less stable due to genetic drift. This phenomenon is

consistent with an increase in  $F_{st}$  recorded from 1999 to 2009 ( $F_{st(1999 \text{ vs } 2002)} = 0.129$  and  $F_{st(2002 \text{ vs } 2009)} = 0.426$ ).

Conversely, a statistically non-significant  $F_{st}$  and contiguous centroids between 1998 and 1999 observed during an earlier malaria situation in Djibouti are consistent with a more stable population. This particular result in 1998–1999 is probably due to i) residual malaria foci in Djibouti, leading to genetically similar populations as the source of yearly epidemics, and/or ii) higher numbers of imported malaria cases from neighbouring countries, leading to an increased genetic exchange in plasmodial populations from different countries. These observations

**Table 2 Evolution of the prevalence of *Pfdhfr* point mutations in codons 16, 51, 59, 108, and 164**

Year	N	51		59		108	
		N (Asn) (W)	I (Ile) (M)	C (Cys) (W)	R (Arg) (M)	S (Ser) (W)	N (Asn) (M)
		AAT	ATT	TGT	CGT	AGC	AAC
1998	36	100	0	97.2	2.8	97.2	2.8
1999	50	96.0	4.0	94.0	6.0	92.0	8.0
2002	4	100	0	100	0	100	0
2009	13	NA	NA	0	100	0	100

Proportions (%) of single nucleotide polymorphism are presented: *Pfdhfr Plasmodium falciparum* dihydrofolate reductase, N, number of samples with at least a result at one codon; W wild-type, M mutant. NA; no results detected. Codons 16 and 164 showed only wild-type codons in all years studied.

lead to the hypothesis that Djiboutian plasmodial populations were more stable in 1998–1999. The analysis of genetic relationship showed closely related genotypes, and even similar genetic lineages, between these two years. The two most recurrent genotypes occurred nine and seven times for genotype 32 and 23 and 23 times for genotype 38 in 1998 and 1999, respectively (Figure 3 detailed in Additional file 5, Table B). To a lesser extent, similar genotypes were observed between 1999 and 2002, which suggested a common source of Djiboutian strains despite the random annual turnover due to sampling and genetic diversification (Figure 4). This data interpretation is in agreement with recurring human migratory flows across Djiboutian borders, including regular visits to Ethiopia [78] and transport of goods and persons via the international Djiboutian port [79].

The decline in genetic diversity in 2009 is due to unexpected genotype frequencies. Only one genotype was found in Djibouti in that year. In fact, after sampling 150 sites throughout the country, *P. falciparum* had been found in three distant sites. Taking into account both the time interval (three months) between isolate collection and distance separating the sites, identical genotypes should not be considered as the same strains but rather that they probably represent similar plasmodial populations. Moreover, genotyping analyses were performed with only five or even four microsatellite loci. Additional loci could have detected more genotypes. Nevertheless, i) the molecular system described in the present paper already attained a discriminatory index of 91% [70] and ii) as the same microsatellite markers were used throughout the 11-year study period, the present data strongly indicate a significant decrease in genetic diversity in Djibouti.

It is important to note that malaria epidemics characterized by a random clonal expansion of few plasmodial populations may lead to a rapid spread of drug resistance throughout the country. Indeed, all isolates collected in 2009 carried single nucleotide polymorphism (SNPs) associated with pyrimethamine resistance at codons 59 and 108. As the Djiboutian national anti-malarial drug policy relies on artesunate-sulphadoxine-pyrimethamine (AS-SP) combination for the first-line treatment of uncomplicated malaria, the field data presented in this paper raise the question of the necessity for a change in drug policy if molecular data are confirmed by poor clinical and parasitological response to AS-SP. However, it should be noted that i) these high percentages of *dhfr* double mutants are based on the analysis of a limited number of isolates and triple mutants have not been observed for the past 10 years in Djibouti, and ii) the observed mutants most likely reflect random fluctuations due to imported strains, as opposed to a sustained increase in the prevalence of *dhfr* mutants which would have been expected if local Djiboutian *dhfr* mutants had been selected and maintained under constant

drug pressure. Molecular studies in southern Ethiopia have shown that double N511/S108N (46%) and triple N511/C59R/S108N *dhfr* mutants (54%) predominate in Jimma, while a large majority (>90%) of isolates are triple *dhfr* mutants in Dilla and Gambo [80–82]. Further molecular studies in Ethiopia closer to the Djiboutian border are required for the evaluation of imported malaria and spread of drug resistance in Djibouti. The results of the present study should be interpreted in the light of the prevalence of *dhfr* mutants in neighbouring countries, which may be informative to assess the risk of the spread of sulphadoxine-pyrimethamine resistance during the next epidemics in Djibouti.

Recent studies have used molecular tools to monitor plasmodial populations in regions of declining endemicity following malaria control effort or in areas of naturally low level malaria transmission ([28,69,83–86]). The most striking finding of the present study, the “clonal expansion” observed after a decline in malaria transmission, is consistent with these previous studies. Clonal expansion may occur in low transmission settings [28,61,66,69,83,87] and may be favoured by control efforts [69,84,85]. Considering the global decline in malaria prevalence over the last decade [1], more situations exhibiting epidemic or clonal expansion of parasites can be expected in the future.

The relations a between decline in malaria transmission and variations in parasite population parameters showed contradictory results, and more field data are required to establish a solid relationship between these indices and malaria situation. An expected decline in genetic diversity ( $H_e$ ) shown in the present study (Additional file 6) was not observed in another recent study conducted in Thailand even after a considerable reduction in malaria transmission [88]. In western Kenya, a decrease in transmission has led to an increase in genetic diversity [86]. Epidemiological context and migration of human population might explain some of these discordant results. However, a comparison of results of the present study with those of other recent studies which used different molecular tools (SNP vs microsatellites) and different indices to monitor parasites should be interpreted with caution.

According to Nkhoma *et al.* [88], index based on MOI (i.e., percentage of polyclonal infection) seems to be more robust or less sensible to genetic diversity resulting from migration into a study area. These authors have found a high correlation (coefficient of determination,  $r^2 = 0.7967$ ) between malaria incidence and proportion of polyclonal infections based on 96xSNP genotyping [88]. A high coefficient of determination ( $r^2 = 0.99$ ) has also been observed using SNP data [66]. The data presented in this paper (four time points, including one outbreak in 2009) did not allow a reliable estimation of correlation ( $r^2 = 0.59$ , Additional file 7). Moreover, indices based on microsatellite loci and comparability with SNP have not been established.

Nevertheless, the present data and those of other studies confirm a decrease in the percentage of polyclonal infections after a decline in malaria transmission [85].

Indices which take into account the genetic linkage disequilibrium (standardized index of association, genotype richness, and genotypic diversity) also show correlation with a decline in malaria transmission [85,88]. However, the self-fertilization rate of *P. falciparum* is largely unknown, which hampers the full understanding of these complex relations. Despite these limitations, analysis of relationship between closely-related genotypes in low malaria transmission settings may highlight various evolutionary patterns, such as clonal expansion, diversification, selection, migration, and residual foci, that could be modified by an intervention of malaria control programmes. Therefore, analysis of relationship between genotypes can be a powerful complementary tool to monitor *Plasmodium* populations, especially during pre-elimination and elimination phases.

## Conclusions

Molecular tools for population genetics analysis play an increasingly important role in monitoring the characteristics of *P. falciparum* populations. On the basis of a positive correlation between local malaria transmission, imported malaria cases and plasmodial genetic diversity, the latter was used in innovative ways to assess the risk of malaria importation into Djibouti, which is a low malaria transmission setting. The significant decrease in He, in unexpected proportions, suggests a significant decrease in local malaria transmission as well as in imported malaria. The present results indicate an evolution of Djiboutian plasmodial populations towards a malaria situation that is favourable for malaria elimination due to the presence of a few genetically distinct strains originating from the same source and spreading throughout the country. In this context, an efficient surveillance system and case management will play a major role to attain malaria elimination. Moreover, the knowledge on the geographic origin of strains would be helpful to target malaria control at the regional scale. To that end, additional molecular data from neighbouring countries, in particular from Ethiopia, are necessary. To limit additional cost required for molecular investigations, such molecular studies can use biological materials that have already been sampled (e.g. RDTs for malaria and thick smear) as part of the regular activities of malaria control units in the region, provided that prior consent from patients had been obtained. Moreover, standardization of protocols (DNA extraction, genotyping and microsatellite panel) is necessary to allow direct comparison among studies and strengthen national monitoring systems in the region for a coordinated malaria control effort in the Horn of Africa.

## Additional files

**Additional file 1: Classes of *P. falciparum* seroprevalence (%).**

**Additional file 2: Clusters localisation in the districts of the country.**

**Additional file 3: Numbers of malaria attacks within Djibouti city from 1998 to 2009 based on three largest Djiboutian surveillance systems: i) Peltier General Hospital (dotted squares), ii) Djiboutian National Healthcare Insurance Program (solid squares), and iii) Bouffard French Military Hospital (hatched squares) (adapted from Ollivier et al. 2011). Colours (red, blue, green, and purple) highlight the years with samples genotyped in the present study.**

**Additional file 4: Table A.** Primer sequences and amplification conditions of the 5 microsatellite loci and *Pfdhfr*: Primer sequences (5' → 3') are given for reactions no. 1 (first round) and no. 2 (second round) of the nested PCRs with fluorescent label (VIC, NED, 6-FAM or PET) and annealing temperature (Ta, °C). Thermal cycling was performed using Biometra® (Goettingen, Germany) 96-well T3 thermocycler. Size in basepairs for 3D7 reference clone. *Pfdhfr*, *P. falciparum* dihydrofolate reductase; Chr, chromosome.

**Additional file 5: Table B.** Number of distinct genotypes of *P. falciparum* and years based on genotyping with 4 microsatellites. MIS: Samples from Malaria Indicator Survey (described in Figure 1), D: samples from a clinical case of Djiboutian military recruit (described in Figure 1), E: South-East Ethiopian samples (described in Figure 3). Genotypes 32, 34, 36, and 38 constituted the largest number of Djiboutian isolates.

**Additional file 6: Malaria incidence and genetic diversity in Djiboutian *P. falciparum* population.** The estimation of malaria incidence was based on the three largest Djiboutian surveillance systems (details in the text and Additional file 3). The calculation of genetic diversity (He) based on four microsatellite genotyping is described in material and methods section. The coefficient of determination  $r^2 = 0.9527y = 0.1557\ln(x) - 0.4437$  suggests a positive non-linear relation between genetic diversity (He) and malaria incidence in the Republic of Djibouti. The linear coefficient of determination was lower ( $r^2 = 0.5047$ ).

**Additional file 7: Malaria incidence and percentage of polyclonal infections in Djiboutian population of *Plasmodium falciparum* (1998, 1999, 2002, and 2009).** The estimation of malaria incidence was based on three largest Djiboutian surveillance systems (details in the text and Additional file 3). The percentage of polyclonal infections (i.e. infection with multiple alleles at one or more of the four microsatellite loci) was based on the analysis of four microsatellites.

## Abbreviations

ACT: Artemisinin-based combination therapy; AFLP: Amplification fragment length polymorphism; DLV: Double-loci variant; FCA: Factorial correspondence analysis; He: Heterozygosity expected; MCA: Multiple correspondence analysis; MOI: Multiplicity of infection; mAll: Mean number of alleles/locus; *Pfdhfr*: *Plasmodium falciparum* dihydrofolate reductase; SD: Standard deviation; SNP: Single nucleotide polymorphism; SLV: Single-locus variant; SP: Sulphadoxine pyrimethamine; WHO: World Health Organization.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

BAK, AA, IHF, HMA, SMB, HHG, SNA, ZA, and HYD collected the data. BAK and AP performed the genotyping analysis. BAK, HB, and MAK analysed the data. BAK wrote the first draft of the manuscript. BAK, LKB, AA, AAA, IHF, HMA, SMB, SB, SNA, HYD, BP, and CR helped to draft the manuscript. BAK, LKB, SB, BP, CR and HB participated in the interpretation of data and revised the paper. HB, BAK and CR designed the study. HB directed the research and contributed to the writing and editing of the manuscript. All authors read and approved the final manuscript.

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#### References

- WHO: *World malaria report 2012*. Geneva: World Health Organization; 2012. [www.who.int/malaria/publications/world\\_malaria\\_report\\_2012/report/en/index.html](http://www.who.int/malaria/publications/world_malaria_report_2012/report/en/index.html).
- WHO: *Malaria Fact sheet*. Geneva: World Health Organization; 2012. <http://www.who.int/mediacentre/factsheets/fs094/en/index.html>.
- RBM: *Global Malaria Action Plan for a Malaria Free World*. 2008. <http://www.rbm.who.int/gmap/2-4a.html> (accessed 2012.05.24).
- WHO: *Global malaria control and elimination: report of a technical review 2008*. Geneva: World Health Organization; 2012. <http://www.who.int/malaria/publications/atoz/9789241596756/en/index.html>. Accessed 24.05.2012.
- RBM: *Eliminating Malaria: Learning from the Past, Looking Ahead*; 2012. <http://www.rbm.who.int/ProgressImpactSeries/docs/report9-en.pdf> (accessed 2012 08 22).
- Feachem RGA, Phillips AA, Hwang J, Cotter C, Wielgosz B, Greenwood BM, Sabot O, Rodriguez MH, Abeyasinghe RR, Ghebreyesus TA, Snow RW: **Shrinking the malaria map: progress and prospects**. *Lancet* 2010, **376**:1566–1578.
- Martens P, Hall L: **Malaria on the move: human population movement and malaria transmission**. *Emerg Infect Dis* 2000, **6**:103–109.
- Cohen JM, Smith DL, Cotter C, Ward A, Yamey G, Sabot OJ, Moonen B: **Malaria resurgence: a systematic review and assessment of its causes**. *Malar J* 2012, **11**:122.
- Crowell V, Hardy D, Briët O, Chitnis N, Maire N, Smith T: **Can we depend on case management to prevent re-establishment of *P. falciparum* malaria, after local interruption of transmission?** *Epidemics* 2012, **4**:1–8.
- Corran P, Coleman P, Riley E, Drakeley C: **Serology: a robust indicator of malaria transmission intensity?** *Trends Parasitol* 2007, **23**:575–582.
- Drakeley C, Cook J: **Potential contribution of sero-epidemiological analysis for monitoring malaria control and elimination: historical and current perspectives**. *Adv Parasitol* 2009, **69**:299–352.
- Djimé A, Doumbo OK, Cortese JF, Kayentao K, Doumbo S, Diouré Y, Coulibaly D, Dicko A, Su XZ, Nomura T, Fidock DA, Wellem TE, Plowe CV: **A molecular marker for chloroquine-resistant falciparum malaria**. *N Engl J Med* 2001, **344**:257–263.
- Noranate N, Durand R, Tall A, Marrama L, Spiegel A, Sokhna C, Pradines B, Cojean S, Guillotte M, Bischoff E, Ekala MT, Bouchier C, Fandeur T, Ariey F, Patarapotikul J, Le Bras J, Trape JF, Rogier C, Mercereau-Puijalon O: **Rapid dissemination of *Plasmodium falciparum* drug resistance despite strictly controlled antimalarial use**. *PLoS One* 2007, **2**:e139.
- Wootton JC, Feng X, Ferdig MT, Cooper RA, Mu J, Baruch DI, Magill AJ, Su XZ: **Genetic diversity and chloroquine selective sweeps in *Plasmodium falciparum***. *Nature* 2002, **418**:320–323.
- Chan CW, Spathis R, Reiff DM, McGrath SE, Garruto RM, Lum JK: **Diversity of *Plasmodium falciparum* chloroquine resistance transporter (pfcrt) exon 2 haplotypes in the Pacific from 1959 to 1979**. *PLoS One* 2012, **7**:e30213.
- Vinayak S, Alam MT, Sem R, Shah NK, Susanti AI, Lim P, Muth S, Maguire JD, Rogers WO, Fandeur T, Barnwell JW, Escalante AA, Wongsrichanalai C, Ariey F, Meshnick SR, Udhayakumar V: **Multiple genetic backgrounds of the amplified *Plasmodium falciparum* multidrug resistance (pfmdr1) gene and selective sweep of 184F mutation in Cambodia**. *J Infect Dis* 2010, **201**:1551–1560.
- McCollum AM, Basco LK, Tahar R, Udhayakumar V, Escalante AA: **Hitchhiking and selective sweeps of *Plasmodium falciparum* sulfadoxine and pyrimethamine resistance alleles in a population from central Africa**. *Antimicrob Agents Chemother* 2008, **52**:4089–4097.
- Nair S, Williams JT, Brockman A, Paiphan J, Mayxay M, Newton PN, Guthmann JP, Smithuis FM, Hien TT, White NJ, Nosten F, Anderson TJ: **A selective sweep driven by pyrimethamine treatment in southeast asian malaria parasites**. *Mol Biol Evol* 2003, **20**:1526–1536.
- Genton B, Betuela I, Felger I, Al-Yaman F, Anders RF, Saul A, Rare L, Baisor M, Lorry K, Brown GV, Pye D, Irving DO, Smith TA, Beck HP, Alpers MP: **A recombinant blood-stage malaria vaccine reduces *Plasmodium falciparum* density and exerts selective pressure on parasite populations in a phase 1-2b trial in Papua New Guinea**. *J Infect Dis* 2002, **185**:820–827.
- Flück C, Schoepflin S, Smith T, Genton B, Alpers MP, Beck HP, Felger I: **Effect of the malaria vaccine Combination B on merozoite surface antigen 2 diversity**. *Infect Genet Evol* 2007, **7**:44–51.
- Nei M: **Estimation of average heterozygosity and genetic distance from a small number of individuals**. *Genetics* 1978, **89**:583–590.
- Paul RE, Hackford I, Brockman A, Muller-Graf C, Price R, Luxemburger C, White NJ, Nosten F, Day KP: **Transmission intensity and *Plasmodium falciparum* diversity on the northwestern border of Thailand**. *Am J Trop Med Hyg* 1998, **58**:195–203.
- Paul RE, Brockman A, Price RN, Luxemburger C, White NJ, Looareesuwan S, Nosten F, Day KP: **Genetic analysis of *Plasmodium falciparum* infections on the north-western border of Thailand**. *Trans R Soc Trop Med Hyg* 1999, **93**:587–593.
- Anderson TJ, Haubold B, Williams JT, Estrada-Franco JG, Richardson L, Mollinedo R, Bockarie M, Mokili J, Mharakurwa S, French N, Whitworth J, Velez ID, Brockman AH, Nosten F, Ferreira MU, Day KP: **Microsatellite markers reveal a spectrum of population structures in the malaria parasite *Plasmodium falciparum***. *Mol Biol Evol* 2000, **17**:1467–1482.
- Iwagami M, Rivera PT, Villacorte EA, Escueta AD, Hatabu T, Kawazu S, Hayakawa T, Tanabe K, Kano S: **Genetic diversity and population structure of *Plasmodium falciparum* in the Philippines**. *Malar J* 2009, **8**:96.
- Campino S, Auburn S, Kivinen K, Zongo I, Ouedraogo JB, Mangano V, Djimde A, Doumbo OK, Kiara SM, Nzila A, Borrmann S, Marsh K, Michon P, Mueller I, Siba P, Jiang H, Su XZ, Amaratunga C, Socheat D, Fairhurst RM, Imwong M, Anderson T, Nosten F, White NJ, Gwilliam R, Deloukas P, MacLinnis B, Newbold CI, Rockett K, Clark TG, Kwiatkowski DP: **Population genetic analysis of *Plasmodium falciparum* parasites using a customized Illumina GoldenGate genotyping assay**. *PLoS One* 2011, **6**:e20251.
- Schoepflin S, Valsangiacomo F, Lin E, Kiniboro B, Mueller I, Felger I: **Comparison of *Plasmodium falciparum* allelic frequency distribution in different endemic settings by high-resolution genotyping**. *Malar J* 2009, **8**:250.
- Pumpaibool T, Amathau C, Durand P, Kanchanakhan N, Siripoon N, Suegorn A, Sitthi-Amorn C, Renaud F, Harnyuttanakorn P: **Genetic diversity and population structure of *Plasmodium falciparum* in Thailand, a low transmission country**. *Malar J* 2009, **8**:155.
- Mobegi VA, Loua KM, Ahouidi AD, Satoguina J, Nwakanma DC, Amambua-Ngwa A, Conway DJ: **Population genetic structure of *Plasmodium falciparum* across a region of diverse endemicity in West Africa**. *Malar J* 2012, **11**:223.
- Paul RE, Packer MJ, Walmsley M, Lagog M, Ranford-Cartwright LC, Paru R, Day KP: **Mating patterns in malaria parasite populations of Papua New Guinea**. *Science* 1995, **269**:1709–1711.
- Paul RE, Day KP: **Mating Patterns of *Plasmodium falciparum***. *Parasitol Today* 1998, **14**:197–202.
- Bogreau H, Renaud F, Bouchiba H, Durand P, Assi SB, Henry MC, Garnotel E, Pradines B, Fusai T, Wade B, Adehossi E, Parola P, Kamil MA, Puijalon O, Rogier C: **Genetic diversity and structure of African *Plasmodium falciparum* populations in urban and rural areas**. *Am J Trop Med Hyg* 2006, **74**:953–959.



33. Rebaudet S, Bogreau H, Silai R, Lepere JF, Bertaux L, Pradines B, Delmont J, Gautret P, Parola P, Rogier C: **Genetic structure of *Plasmodium falciparum* and elimination of malaria, Comoros archipelago.** *Emerg Infect Dis* 2010, **16**:1686–1694.
34. Wright S: **The interpretation of population structure by F-statistics with special regard to systems of mating.** *Evolution* 1965, **19**:395–420.
35. Weir B, Cockerham C: **Estimating F-statistics for the analysis of population structure.** *Evolution* 1984, **38**:1358–1370.
36. Tatem AJ, Smith DL, Gething PW, Kabaria CW, Snow RW, Hay SI: **Ranking of elimination feasibility between malaria-endemic countries.** *Lancet* 2010, **376**:1579–1591.
37. Cohen JM, Moonen B, Snow RW, Smith DL: **How absolute is zero? An evaluation of historical and current definitions of malaria elimination.** *Malar J* 2010, **9**:213.
38. CERD: **Pluviométrie annuel.** Centre d'étude et de recherche de djibouti. 2008.
39. Carteron B, Morvan D, Rodhain F: **The question of endemic malaria in Republic of Djibouti (in French).** *Med Trop (Mars)* 1978, **38**:299–304.
40. Fox E, Abbate EA, Leef M, Mikhail E, Said-Salah Y, Hassan A: **Malaria in the Djibouti Republic. Results of a serologic survey in Ambouli (in French).** *Med Trop (Mars)* 1989, **49**:159–160.
41. Fox E, Bouloumie J, Olson JG, Tible D, Lluberas M, Shakib SO, Parra JP, Rodier G: **[*Plasmodium falciparum* travels by train from Ethiopia to Djibouti] (in French).** *Med Trop (Mars)* 1991, **51**(2):185–189.
42. Manguin S, Carnevale P, Mouchet J, Coosemans M, Julvez J, Richard-Lenoble D, Sircoulo J: **Biodiversity of Malaria in the World.** ISBN : 978-2-7420-0616-8. *John Libbey Eurotext* 2008:140.
43. Khairah BA, Briolant S, Pascual A, Mokrane M, Machault V, Travaille C, Khairah MA, Farah IH, Ali HM, Abdi AIA, Ayeh SN, Darar HY, Ollivier L, Waiss MK, Bogreau H, Rogier C, Pradines B: ***Plasmodium vivax* and *Plasmodium falciparum* infections in the Republic of Djibouti: evaluation of their prevalence and potential determinants.** *Malar J* 2012, **11**:395.
44. Ollivier L, Nevin RL, Darar HY, Bougâre J, Saleh M, Gidenne S, Maslin J, Anders D, Decam C, Todesco A, Khairah BA, Ahmed AA: **Malaria in the Republic of Djibouti, 1998–2009.** *Am J Trop Med Hyg* 2011, **85**:554–559.
45. Rogier C, Pradines B, Bogreau H, Koeck JL, Kamil MA, Mercereau-Pujalon O: **Malaria epidemic and drug resistance, Djibouti.** *Emerg Infect Dis* 2005, **11**:317–321.
46. Ministère de l'intérieur: *Bilan 1999–2005.* 45–46. République de Djibouti.
47. Ministère du Transport: *Rapport d'activité semestriel.* Tech. Rep. 2. Imprimerie Nationale: Ministère du Transport; 2008.
48. Noor AM, Mohamed MB, Mugenyi CK, Osman MA, Guessod HH, Kabaria CW, Ahmed IA, Nyonda M, Cook J, Drakeley CJ, Mackinnon MJ, Snow RW: **Establishing the extent of malaria transmission and challenges facing pre-elimination in the Republic of Djibouti.** *BMC Infect Dis* 2011, **11**:121.
49. Journal Officiel de la République de Djibout: **Décret n°2010-0038/PR/MEFPCP portant publication des résultats du 2ème Recensement Général de la Population et de l'Habitat de 2009.** Tech Rep 2010.
50. PNL: **Programme National de Lutte Contre le Paludisme: Directives nationales pour le diagnostic et le traitement du paludisme.** 2007:11.
51. Kublin JG, Dzinjalama FK, Kamwendo DD, Malkin EM, Cortese JF, Martino LM, Mukadam RAG, Rogerson SJ, Lescano AG, Molyneux ME, Winstanley PA, Chimpeni P, Taylor TE, Plowe CV: **Molecular markers for failure of sulfadoxine-pyrimethamine and chlorproguanil-dapsone treatment of *Plasmodium falciparum* malaria.** *J Infect Dis* 2002, **185**:380–388.
52. Nair S, Brockman A, Paiphun L, Nosten F, Anderson TJC: **Rapid genotyping of loci involved in antifolate drug resistance in *Plasmodium falciparum* by primer extension.** *Int J Parasitol* 2002, **32**:852–858.
53. Belkhir K, Borsari P, Chikhi L, Raufaste L, Bonhomme F: **[GENETIX 4.05, Windows software for population genetics] (in French);** 2012. <http://www.genetix.univ-montp2.fr/genetix/genetix.htm> Accessed 2012 05.24.
54. Goudet J: **FSTAT (version 2.9.3.2), a program to estimate and test population genetics parameters;** 2012. <http://www2.unil.ch/popgen/softwares/fstat.htm>. Accessed 2012 05.24.
55. Benzécri J: *L'analyse des données / leçons sur l'analyse factorielle et la reconnaissance des formes et travaux.* France: Dunod; 1982.
56. Team RC: *R: A Language and Environment for Statistical Computing.* Vienna, Austria: R Foundation for Statistical Computing; 2012. <http://www.R-project.org>. Accessed 2012 05.24
57. Feil EJ, Li BC, Aanensen DM, Hanage WP, Spratt BG: **eBURST: inferring patterns of evolutionary descent among clusters of related bacterial genotypes from multilocus sequence typing data.** *J Bacteriol* 2004, **186**:1518–1530.
58. Francisco AP, Bugalho M, Ramirez M, Carriço JA: **Global optimal eBURST analysis of multilocus typing data using a graphic matroid approach.** *BMC Bioinformatics* 2009, **10**:152.
59. Francisco AP, Vaz C, Monteiro PT, Melo-Cristino J, Ramirez M, Carriço JA: **PHYLOVIZ: Phylogenetic Inference and Data Visualization for Sequence Based Typing Methods.** *BMC Bioinformatics* 2012, **13**:87.
60. Rich SM, Hudson RR, Ayala FJ: ***Plasmodium falciparum* antigenic diversity: evidence of clonal population structure.** *Proc Natl Acad Sci USA* 1997, **94**:13040–13045.
61. Urdaneta L, Lal A, Barnabe C, Oury B, Goldman I, Ayala FJ, Tibayrenc M: **Evidence for clonal propagation in natural isolates of *Plasmodium falciparum* from Venezuela.** *Proc Natl Acad Sci U S A* 2001, **98**:6725–6729.
62. Tibayrenc M, Ayala FJ: **The clonal theory of parasitic protozoa: 12 years on.** *Trends Parasitol* 2002, **18**:405–410.
63. Gauthier C, Tibayrenc M: **Population structure of malaria parasites: the driving epidemiological forces.** *Acta Trop* 2005, **94**:241–250.
64. Razakandrainibe FG, Durand P, Koella JC, De Meeus T, Rousset F, Ayala FJ, Renaud F: **"Clonal" population structure of the malaria agent *Plasmodium falciparum* in high-infection regions.** *Proc Natl Acad Sci USA* 2005, **102**:17388–17393.
65. Annan Z, Durand P, Ayala FJ, Amathau C, Awono-Ambene P, Simard F, Razakandrainibe FG, Koella JC, Fontenille D, Renaud F: **Population genetic structure of *Plasmodium falciparum* in the two main African vectors, *Anopheles gambiae* and *Anopheles funestus*.** *Proc Natl Acad Sci USA* 2007, **104**:7987–7992.
66. Echeverry DF, Nair S, Osorio L, Menon S, Murillo C, Anderson TJC: **Long term persistence of clonal malaria parasite *Plasmodium falciparum* lineages in the Colombian Pacific region.** *BMC Genet* 2013, **14**:2.
67. Arez AP, Snounou G, Pinto J, Sousa CA, Modiano D, Ribeiro H, Franco AS, Alves J, Do Rosario VE: **A clonal *Plasmodium falciparum* population in an isolated outbreak of malaria in the Republic of Cabo Verde.** *Parasitology* 1999, **118**:347–355.
68. Laserson KF, Petralanda I, Almera R, Barker R Jr, Spielman A, Maguire JH, Wirth DF: **Genetic characterization of an epidemic of *Plasmodium falciparum* malaria among Yanomami Amerindians.** *J Infect Dis* 1999, **180**:2081–2085.
69. Griffing SM, Mixson-Hayden T, Sridaran S, Alam MT, McCollum AM, Cabezas C, Marquino Quezada W, Barnwell JW, De Oliveira AM, Lucas C, Arrospe N, Escalante AA, Bacon DJ, Udhayakumar V: **South American *Plasmodium falciparum* after the malaria eradication era: clonal population expansion and survival of the fittest hybrids.** *PLoS One* 2011, **6**:e23486.
70. Hunter PR, Gaston MA: **Numerical index of the discriminatory ability of typing systems: an application of Simpson's index of diversity.** *J Clin Microbiol* 1988, **26**:2465–2466.
71. Manjurano A, Okell L, Lukindo T, Reyburn H, Olomi R, Roper C, Clark TG, Joseph S, Riley EM, Drakeley C: **Association of sub-microscopic malaria parasite carriage with transmission intensity in north-eastern Tanzania.** *Malar J* 2011, **10**:370.
72. Bruce MC, Macheso A, McConnachie A, Molyneux ME: **Comparative population structure of *Plasmodium malariae* and *Plasmodium falciparum* under different transmission settings in Malawi.** *Malar J* 2011, **10**:38.
73. Vafa M, Troye-Blomberg M, Anchang J, Garcia A, Migot-Nabias F: **Multiplicity of *Plasmodium falciparum* infection in asymptomatic children in Senegal: relation to transmission, age and erythrocyte variants.** *Malar J* 2008, **7**:17.
74. Daubersies P, Sallenave-Sales S, Magne S, Trape JF, Contamin H, Fandeur T, Rogier C, Mercereau-Pujalon O, Druilhe P: **Rapid turnover of *Plasmodium falciparum* populations in asymptomatic individuals living in a high transmission area.** *Am J Trop Med Hyg* 1996, **54**:18–26.
75. Hopkins H, Talisuna A, Whitty CJ, Staedke SG: **Impact of home-based management of malaria on health outcomes in Africa: a systematic review of the evidence.** *Malar J* 2007, **6**:134.
76. Yeshiwondim AK, Gopal S, Hailemariam AT, Dengela DO, Patel HP: **Spatial analysis of malaria incidence at the village level in areas with unstable transmission in Ethiopia.** *Int J Health Geogr* 2009, **8**:5.
77. Shargie EB, Ngondi J, Graves PM, Getachew A, Hwang J, Gebre T, Mosher AW, Ceccato P, Endeshaw T, Jima D, Tadesse Z, Tenaw E, Reithinger R, Emerson PM, Richards FO, Ghebreyesus TA: **Rapid increase in ownership and use of long-lasting insecticidal nets and decrease in prevalence of malaria in three regional States of Ethiopia (2006–2007).** *J Trop Med* 2010, **2010**:12. Article ID 750978.

78. Ministère de l'intérieur: *Secrétariat Général: Rapport Semestriel*. République de Djibouti: Tech. rep; 2009.
79. Ministère du Transport: *Rapport d'activité semestriel*, Tech. Rep. République de djibouti; 2008:22.
80. Gebru-Woldearegai T, Hailu A, Grobusch MP, Kun JF: **Molecular surveillance of mutations in dihydrofolate reductase and dihydropteroate synthase genes of *Plasmodium falciparum* in Ethiopia.** *Am J Trop Med Hyg* 2005, **73**:1131–1134.
81. Schunk M, Kumma WP, Miranda IB, Osman ME, Roewer S, Alano A, Löscher T, Bienzle U, Mockenhaupt FP: **High prevalence of drug-resistance mutations in *Plasmodium falciparum* and *Plasmodium vivax* in southern Ethiopia.** *Malar J* 2006, **5**:54.
82. Mula P, Fernández-Martinez A, De Lucio A, Ramos JM, Reyes F, González V, Benito A, Berzosa P: **Detection of high levels of mutations involved in anti-malarial drug resistance in *Plasmodium falciparum* and *Plasmodium vivax* at a rural hospital in southern Ethiopia.** *Malar J* 2011, **10**:214.
83. Chenet SM, Schneider KA, Villegas L, Escalante AA: **Local population structure of *Plasmodium*: impact on malaria control and elimination.** *Malar J* 2012, **11**:412.
84. Griffing SM, Viana GMR, Mixson-Hayden T, Sridaran S, Alam MT, De Oliveira AM, Barnwell JW, Escalante AA, Povoas MM, Udhayakumar V: **Historical Shifts in Brazilian *P. falciparum* Population Structure and Drug Resistance Alleles.** *PLoS One* 2013, **8**:e58984.
85. Daniels R, Chang HH, Séne PD, Park DC, Neafsey DE, Schaffner SF, Hamilton EJ, Lukens AK, Van Tynne D, Mboup S, Sabeti PC, Ndiaye D, Wirth DF, Hartl DL, Volkman SK: **Genetic Surveillance Detects Both Clonal and Epidemic Transmission of Malaria following Enhanced Intervention in Senegal.** *PLoS One* 2013, **8**:e60780.
86. Vardo-Zalick AM, Zhou G, Zhong D, Afrane YA, Githeko AK, Yan G: **Alterations in *Plasmodium falciparum* genetic structure two years after increased malaria control efforts in western Kenya.** *Am J Trop Med Hyg* 2013, **88**:29–36.
87. Anthony TG, Conway DJ, Cox-Singh J, Matusop A, Ratnam S, Shamsul S, Singh B: **Fragmented population structure of *Plasmodium falciparum* in a region of declining endemicity.** *J Infect Dis* 2005, **191**:1558–1564.
88. Nkhoma SC, Nair S, Al-Saai S, Ashley E, McGready R, Phyto AP, Nosten F, Anderson TJC: **Population genetic correlates of declining transmission in a human pathogen.** *Mol Ecol* 2013, **22**:273–285.

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