

**Peroxisome Proliferator-Activated Receptor γ activation induces 11 β -Hydroxysteroid
Dehydrogenase type 1 activity in human alternative macrophages**

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Short title: PPAR γ induces 11 β -HSD1 activity in M2 macrophages

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Abstract

Objectives - 11 β -hydroxysteroid dehydrogenase type 1 (11 β -HSD1) catalyses the intracellular reduction of inactive cortisone to active cortisol, the natural ligand activating the glucocorticoid receptor (GR). Peroxisome Proliferator-Activated Receptor gamma (PPAR γ) is a nuclear receptor controlling inflammation, lipid metabolism and the macrophage polarization state. In this study, we investigated the impact of macrophage polarization on the expression and activity of 11 β -HSD1 and the role of PPAR γ therein.

Methods and Results- 11 β -HSD1 gene expression is higher in pro-inflammatory M1 and anti-inflammatory M2 macrophages than in resting macrophages (RM), whereas its activity is highest in M2 macrophages. Interestingly, PPAR γ activation induces 11 β -HSD1 enzyme activity in M2 macrophages, but not in RM or M1 macrophages. Consequently, human M2 macrophages displayed enhanced responsiveness to the 11 β -HSD1 substrate cortisone, an effect amplified by PPAR γ -induction of 11 β -HSD1 activity, as illustrated by an increased expression of GR target genes.

Conclusions - Our data identify a positive cross-talk between PPAR γ and GR in human M2 macrophages via the induction of 11 β -HSD1 expression and activity.

Key words: 11 β -HSD1 - PPAR γ - human alternative macrophages - inflammation - GR.

Condensed abstract

In this study, we report that 11 β -HSD1 has highest expression and activity in human anti-inflammatory M2 macrophages. PPAR γ activation induces 11 β -HSD1 activity specifically in M2 macrophages leading to an acquired responsiveness to cortisone, as illustrated by the induction of GR-target genes.

Introduction

Macrophages are heterogeneous cells^{1,2}. A pro-inflammatory cytokine-rich environment triggers an activation profile yielding classically activated macrophages (M1) which are involved in the Th1 immune response. In the presence of Th2 cytokines, such as IL-4 and IL-13, monocytes orient toward an alternative activation state (M2) and are involved in the Th2 immune response¹. M1 macrophages mainly produce pro-inflammatory mediators, express MHC class II molecules and have a high microbicidal activity, whereas M2 macrophages produce high amounts of anti-inflammatory molecules (IL-10, TGF β and IL-1Ra), promote cell growth and tissue repair and display a high endocytic capacity¹.

11 β -hydroxysteroid dehydrogenase type 1 (11 β -HSD1) catalyses the intracellular reduction of inactive cortisone and 11-dehydrocorticosterone to, respectively, active cortisol and corticosterone which are ligands of the glucocorticoid receptor (GR)³. Glucocorticoid-activated GR regulates numerous metabolic and homeostatic processes, including carbohydrate, protein and lipid metabolism, and exerts anti-inflammatory and immunosuppressive properties⁴. 11 β -HSD1 is expressed in several tissues, including liver, adipose tissue and brain^{5,6} and myeloid cells, such as dendritic cells and macrophages⁷.

11 β -HSD1 is not expressed in human monocytes, but is induced upon differentiation into macrophages, and its expression is further enhanced by the anti-inflammatory Th2 cytokines IL-4 and IL-13⁸.

The inflammatory status of macrophages is controlled by nuclear receptors, such as the Peroxisome Proliferator-Activated Receptors (PPARs) α , β/δ and γ ⁹ and the GR. Indeed, GR activation results in the polarization of monocytes to a specific subtype of M2 macrophages, termed M2c¹⁰. PPAR γ is activated by natural (15-deoxy- Δ 12, 14-prostaglandin J2 (15d-PGJ2))¹¹ and synthetic ligands, including the antidiabetic glitazones (rosiglitazone and pioglitazone) and the GW1929 compound¹². In activated M1 macrophages, the anti-

inflammatory PPAR γ -dependent trans-repression pathway is initiated by sumoylation of the liganded-PPAR γ ligand-binding domain maintaining the co-repressor complex on NF- κ B response elements¹³. PPAR γ also enhances the alternative activation and differentiation of monocytes into macrophages^{14, 15} thus leading to a more pronounced anti-inflammatory M2 phenotype¹⁵. However, it is not known whether PPAR γ and GR cross-talk in the control of macrophage polarization and functions.

Here, we show that 11 β -HSD1 enzyme activity is highest in M2 macrophages and that PPAR γ ligands specifically induce 11 β -HSD1 enzyme activity in anti-inflammatory M2, but not in RM and M1 macrophages. This novel mechanism may contribute to the anti-inflammatory activities of PPAR γ in M2 macrophages and could have consequences in inflammatory diseases, such as atherosclerosis.

Methods

Cell culture

Human peripheral blood mononuclear cells were isolated from healthy donors by density gradient centrifugation¹⁶. Monocyte differentiation to RM occurred after 7 days of culture in RPMI1640 medium (Invitrogen, France) supplemented with gentamicin (40µg/mL), L-glutamine (2mM) (Sigma-Aldrich, France) and 10% human serum (Abcys, France). M2 macrophages were obtained by differentiating freshly isolated monocytes with human IL-4 (15ng/mL, Promocell, Germany) for 7 days. M1 macrophages were obtained by activating RM macrophages with LPS (100ng/ml) during 4h. Agonists for PPAR γ (GW1929, rosiglitazone), PPAR α (GW7647), PPAR β/δ (GW501516), the PPAR γ antagonist (GW9662), RU486, cortisone or dexamethasone were added as indicated.

Bone marrow-derived macrophages (BMDM) prepared from C57BL/6J mice¹⁷, were treated with GW1929 (0.5, 1, 5µM) or rosiglitazone (1, 5, 10µM) for 24h.

RNA extraction and analysis

RNA was isolated from macrophages using Trizol (Invitrogen), reverse transcribed and cDNAs quantified by Q-PCR on a MX4000 apparatus (Agilent Biotechnologies) using the Brilliant II SYBR® Green Q-PCR Master Mix (Agilent Biotechnologies) with specific primers (table 1). TNF α and IL-6 were measured using Brilliant Multiplex® Q-PCR Master Mix kit (Agilent Biotechnologies) with the primers/probes: TNF α : 5'-CCCATGTTGTAGCAAACCCTCA-3' (forward), 5'-ATCTCTCAGCTCCAGGCCATTG-3' (reverse) and 5'Cy5 TTGGCCCGGCGGTTTCAGCCACT 3'DDQ2 (probe); IL-6: 5'CAATAACCACCCCTGACCCAAC3' (forward), 5'-AAGCTGCGCAGAATGAGATGAG-3' (reverse) and 5'FAM TGCCAGCCTGCTGACGAAGCTGCA 3'BHQ1 (probe) and cyclophilin: 5'-TGGTCAACCCACCGTGTTTC-3' (forward), 5'-

TGCAAACAGCTCAAAGGAGACG-3' (reverse) and 5'Yakima Yellow
TTGCCGTCGACGGCGAGCCCTT 3'EDQ (probe).

Electrophoretic Mobility Shift Assay (EMSA)

PPAR γ and RXR α were *in vitro* transcribed from the pSG5-hPPAR γ and pSG5-hRXR α plasmids (Promega, Madison, WI). For the preparation of nuclear extracts, RM, M1 and M2 macrophages were collected in hypotonic buffer (15mM Hepes pH 7.8, 10mM KCl, 2 mM MgCl₂, 0.1mM EDTA, 3mM DTT and protease inhibitors). After 15min incubation at 4°C, NP40 was added before centrifugation. The cell pellet was dissolved in hypertonic buffer (30mM Hepes, 50 mM KCl, 300mM NaCl, 0.1mM EDTA, 3mM DTT, 10% glycerol); after 30min incubation at 4°C, nuclear extracts were collected. Proteins were incubated for 10min in binding buffer (Hepes 10mM pH 7.8, NaCl 100mM, EDTA 0.1mM, 10% glycerol, 1mg/ml BSA) containing poly(dI-dC) (1 μ g) and herring sperm DNA (1 μ g). Double stranded oligonucleotides containing the wild-type DR1-PPAR γ response element (PPRE) present between nucleotides -2406/-2393 of the human 11 β -HSD1 promoter, end-labeled using T4 polynucleotide kinase and γ^{32} P-ATP, was added as probe to the binding reaction. For supershift assays, monoclonal anti-human PPAR γ antibody (Santacruz Biotechnology) was added to the binding reaction. DNA/protein complexes were resolved by non-denaturing polyacrylamide gel electrophoresis in 0.25X Tris-Borate-EDTA. Classical DR1-PPAR γ response element was used as positive control.

Plasmid cloning and transient transfection experiments

The reporter plasmid (11 β -HSD1-PPREwt)₆-TK-pGL3 was generated by inserting 6 copies of the double-strand oligonucleotides (for 5'-CTT GAA GGG TTG AAA GGT CAA AAC TAT -3'; rev 5'-ATA GTT TTG ACC TTT CAA CCC TTC AAG-3) into the pTK-pGL3 plasmid.

Macrophages were transfected overnight in RPMI medium containing 10% human serum with reporter plasmids and expression vectors (pSG5-empty or pSG5-hPPAR γ) using jetPEI (Polyplus transfection, France). Subsequently, cells were incubated for additional 24h with GW1929 (600nM) or DMSO and luciferase and β -galactosidase activities were measured.

Chromatin Immunoprecipitation (ChIP) and ChIP-seq assays

Macrophages were cross-linked with 1% formaldehyde/PBS for 10min¹⁸, collected in PBS and nuclear extracts were prepared. The chromatin was fragmented to ~500 bp in a 10mM Tris buffer pH 7.5 containing 1mM EDTA, 0.5mM EGTA, 100mM NaCl, 0.1% Na-Deoxycholate, 0.5% N-lauroyl sarcosine and 0.4% SDS using a bioruptor sonication bath (Diagenode, Belgium). Half of the chromatin was immunoprecipitated overnight in a 10 mM Tris buffer pH 7.5 containing 1mM EDTA, 0.5mM EGTA, 100mM NaCl, 0.1% Na-Deoxycholate, 0.5% N-lauroyl sarcosine and 0.08% SDS and 1% Triton X100 using either PPAR γ antibodies or control IgG (Santacruz Biotechnology) in combination with Protein A/G Dynabeads (Invitrogen). Precipitates were washed with RIPA buffer. Eluates were obtained with 1% SDS in 50mM Tris pH8.0/1mM EDTA and the cross-linking was reversed at 65°C overnight. Immunoprecipitated DNA was purified by phenol-chloroform-isoamylc alcohol extraction and ethanol precipitation. Then, either a 150 bp region of the 11 β -HSD1 promoter area was amplified by PCR, using the following primers 5'-

GACACAAAGTGGATTAGATGTTTATT and 5'-ACATCCTGTAGCCTCTGGAA, or a 77 bp region of β -actin was amplified using 5'-AGTGTGGTCCTGCGACTTCTAAG and 5'-CCTGGGCTTGAGAGGTAGAGTGT.

Chromatin was used for immunoprecipitation using antibody against H3K9ac (Millipore). ChIP-seq was performed and analysed as described¹⁹, including total tag number equalization before data visualization. H3K9ac ChIP-seq data were visualized using the University of

California at Santa Cruz (UCSC) genome browser. H3K9ac ChIP-QPCR was performed on chromatin from several different donors using the following primers: 11 β -HSD1 promoter 1:

5'-CAGGACCACTTCCAAGCATT-3' and 5'-CCAAAGAGAAGCCAGAGTGG-3';

11 β -HSD1 promoter 2: 5'CTTGGCCATCTGGAAGTCTC-3' and 5'-

TGCTAGCCAATTTCCCTGTC-3'; negative control 5'-

CAGGATATACACCCCGTGA-3' and 5'-CAAAGTGCGTACACCTTGGTA-3'.

Short-interfering (si)RNA assays

Differentiated RM macrophages were transfected with siRNA specific for human PPAR γ and non-silencing control scrambled siRNA (Ambion), using the transfection reagent DharmaFECT4 (Dharmacon). After 16h, cells were incubated with GW1929 (600nM) or vehicle (DMSO) and harvested 24h later.

Adenovirus generation

The recombinant adenovirus (Ad)-GFP (Green Fluorescent Protein) and Ad-PPAR γ were obtained as described²⁰. 1.5×10^6 macrophages were infected at a multiplicity of 100 viral particles/cell by adding virus stocks directly to the culture medium. After 16h, cells were incubated for 24h in the absence or presence of GW1929 (600nM).

Protein extraction and western blot analysis

Cells were harvested in ice-cold lysis buffer (RIPA), proteins collected by centrifugation, separated by 10% SDS-PAGE, transferred to nitrocellulose membranes (Amersham) and revealed using goat polyclonal antibodies against 11 β -HSD1 (Abcam) or β -actin (Santacruz Biotechnology). After incubation with a secondary peroxidase-conjugated antibody

(Santacruz Biotechnology), immunoreactive bands were revealed using ECL detection (Amersham) and band intensity quantified by QuantityOne software.

11 β -HSD1 enzyme activity assay

Cellular 11 β -HSD1 reductase activity was determined as described²¹. Cells were incubated for the indicated time periods (from 90min to 8h) in serum-free medium containing cortisone (200nM) as substrate and [³H]-cortisone (10 nM; 50-60 Ci/mmol; ARC, USA) as tracer. After incubation, medium was collected and steroids extracted with ethyl acetate. The extracts were dried under nitrogen, reconstituted with isopropanol containing a mixture of cortisol (F) and cortisone (E) (1.5mg/ml each) and applied to silica gel TLC plates. Cortisol and cortisone were separated in a chloroform/methanol solvent system (9:1,vol:vol) and steroids visualized under iodine, scraped off the plate and radioactivity counted. The conversion of cortisone to cortisol was calculated as the radioactivity index of (F/E+F) and reflects 11 β -HSD1 enzyme activity. Each analysis was performed in triplicate.

Statistical analysis

Statistically differences between groups were analyzed by ANOVA and Student's t test and considered significant when $p < 0.05$.

Results

11 β -HSD1 reductase expression and activity is highest in M2 macrophages

To determine whether 11 β -HSD1 expression and activity are dependent on the macrophage polarization states, its expression was analyzed and compared between human RM, M1 and M2 macrophages. Expression of the M2 markers MR, AMAC1 (figure 1B,C), IL-1Ra, IL-10 and TGF β (supplemental figure 1), measured as positive control of alternative polarization, was elevated in M2 macrophages. 11 β -HSD1 mRNA levels were higher in M1 and, most pronouncedly, in M2 compared to RM macrophages (figure 1A). By contrast, in mouse BMDM, 11 β -HSD1 expression was not induced by IL-4 during differentiation, indicating that 11 β -HSD1 regulation is species-specific and that is not secondary to cellular differentiation (data not shown).

To investigate whether M1 and M2 macrophages also exhibit increased 11 β -HSD1 reductase activity compared to RM macrophages, cells were incubated with radio-labeled cortisone as substrate and conversion into cortisol was measured at different times. Surprisingly, the percentage of cortisone conversion to active cortisol was approximately 12% after 90min and 37% after 8h incubation of RM macrophages with cortisone, whereas it was 10% after 90min and 25% after 8h in M1 macrophages (figure 2A,B), indicating that the elevated 11 β -HSD1 mRNA in M1 macrophages does not translate in higher enzyme activity. By contrast, cortisone to cortisol conversion was already 50% after 90min of cortisone incubation and reached 80% within 8h in M2 macrophages (figure 2C). Thus, 11 β -HSD1 reductase activity is higher in M2 compared to RM and M1 macrophages. Moreover, expression of hexose-6-phosphate-dehydrogenase (H6PDH), a cofactor for 11 β -HSD1²², was also higher in M2 compared to RM macrophages (data not shown). This increased conversion of exogenous cortisone into active cortisol conferred a higher transactivation response to cortisone of a GR response element (GRE) driven-reporter vector in M2 compared to RM macrophages, despite

lower GR mRNA levels (supplemental figure 2). In addition, cortisone inhibited LPS-induced expression of IL-6, TNF α and IL-1 β in M2 but not RM macrophages, indicating that cortisone to cortisol conversion results in anti-inflammatory effects only in M2 macrophages (figure 2D,E,F). Dexamethasone, a GR-specific agonist which does not require metabolism by 11 β -HSD1 to be active, exerted anti-inflammatory properties both in RM and M2 macrophages (figure 2).

Cortisone induces the expression of GR target genes more pronouncedly in M2 macrophages

To determine the functional consequences of the higher 11 β -HSD1 activity in M2 macrophages, the expression of known GR target genes was measured in RM and M2 macrophages after cortisone activation for different time periods (figure 3). Cortisone induces the expression of glucocorticoid-induced leucine zipper (GILZ) (A), angiopoietin-like 4 (ANGPTL4)²³ (B) and pyruvate dehydrogenase kinase 4 (PDK4)²⁴ (C) more strongly in M2 compared to RM macrophages.

11 β -HSD1 gene expression is induced by PPAR γ activation in RM, M1 and M2 macrophages

Since PPAR γ plays a role in macrophage functions and polarization, we next investigated whether PPAR γ regulates 11 β -HSD1 gene expression. Treatment with the PPAR γ ligand GW1929 significantly increased 11 β -HSD1 gene and protein expression both in RM, M1 and M2 macrophages (figure 4A,B). This regulation occurred in a dose-dependent manner with rosiglitazone (supplemental figure 3A). 11 β -HSD1 gene induction by GW1929 was already detectable within 12h after activation and further increased up to 48h (supplemental figure 3B). By contrast, treatment with PPAR α or PPAR β/δ specific agonists did not modify 11 β -HSD1 mRNA (supplemental figure 3C). Expression of the 11 β -HSD1 gene was significantly

reduced upon PPAR γ activation in mouse BMDM, indicating the existence of species-specific regulatory mechanisms (supplemental figure 3D).

The increase of 11 β -HSD1 gene expression by GW1929 was abolished in the presence of GW9662, a potent and selective PPAR γ antagonist²⁵ (figure 4C). In line, 11 β -HSD1 induction upon PPAR γ activation was completely lost in PPAR γ siRNA- compared to scrambled siRNA-transfected macrophages (figure 4D). Moreover, infection of macrophages with an adenovirus coding for PPAR γ (Ad-PPAR γ) increased 11 β -HSD1 expression by approximately 3.5-fold compared to macrophages infected with an adenovirus coding for GFP (Ad-GFP) and addition of GW1929 resulted in a further 2.5-fold induction of 11 β -HSD1 mRNA levels (figure 4E).

PPAR γ regulates 11 β -HSD1 gene expression at the transcriptional level

To determine whether 11 β -HSD1 is a direct PPAR γ target gene, the human 11 β -HSD1 promoter was analyzed *in silico*. A putative PPRE was identified between nucleotides 2406-2393. EMSA were performed to examine whether the PPAR γ -RXR heterodimer binds to this 11 β -HSD1-PPRE. Incubation of this labeled PPRE oligonucleotide with *in vitro* translated PPAR γ and RXR α (indicated as RXR) resulted in the formation of a retarded complex (figure 5A; indicated by the arrow). Incubation with an anti-PPAR γ antibody prevented complex formation, indicating binding-specificity of PPAR γ to the PPRE site (figure 5A). To test whether PPAR γ activates transcription of the (-2406-2393)11 β -HSD1-PPRE, 6 copies of this element were cloned in front of the herpes simplex virus thymidine kinase promoter yielding the (11 β -HSD1-PPRE)₆-Tk-Luc reporter vector which was transfected into differentiated primary human macrophages. Co-transfection of the pSG5-PPAR γ expression vector led to a significant induction of (11 β -HSD1-PPRE)₆ transcriptional activity, an effect further

enhanced by GW929 (600nM) (figure 5B). The consensus DR1- PPRE site cloned in 6 copies, used as positive control, was also induced (figure 5C).

Incubation of the 11 β -HSD1 labelled PPRE oligonucleotide with nuclear extracts from RM, M1 and M2 macrophage resulted in a weak retarded complex visible only in M2 macrophages. The presence of *in vitro* translated RXR α amplified the formation of the complex, which was again stronger with extracts from M2 macrophages and which was blocked by an anti-PPAR γ antibody (figure 5D). To evaluate whether PPAR γ binds to the 11 β -HSD1 promoter *in cell*, a ChIP assay was performed on RM and M2 macrophages. The genomic DNA region encompassing the PPRE of the 11 β -HSD1 gene was immunoprecipitated with an anti-PPAR γ antibody. Q-PCR analysis revealed that PPAR γ binding was stronger in M2 compared to RM macrophages (figure 5E). Since active promoters are marked with elevated levels of acetylation of histone H3 lysine 9 (H3K9ac) around their transcription start site ²⁶, we monitored the presence of H3K9ac at both known 11 β -HSD1 promoters ²⁷ in RM and M2 macrophages. ChIP followed by high-throughput sequencing (ChIP-seq) indicated that H3K9ac was specifically present at the P2 promoter, which contains the identified PPRE, but not at the P1 promoter, in both RM and M2 macrophages (figure 5F). Moreover, H3K9ac levels at promoter P2 were higher in M2 macrophages than in RM as confirmed using ChIP-qPCR performed on chromatin from 2 different donors (figure 5F). Finally, activity of the 11 β -HSD1-TK-Luc PPRE reporter vector was compared in RM and M2 macrophages in the absence or presence of GW1929 (600nM). Basal transcriptional activity was higher in M2 macrophages (figure 5G), an effect further enhanced by GW1929. Overall, these data suggest that 11 β -HSD1 is expressed from the P2 promoter in human macrophages and that this promoter confers a stronger 11 β -HSD1 expression and response to PPAR γ activation in M2 macrophages.

11 β -HSD1 reductase activity is induced by PPAR γ ligands in M2 macrophages

To assess whether the induction of 11 β -HSD1 expression by PPAR γ agonists results in an increased enzyme activity, differentiated RM, M1 and M2 macrophages were treated for 24h in the presence or absence of GW1929 and subsequently incubated with radio-labeled cortisone. Surprisingly, 11 β -HSD1 activity was not influenced by GW1929 in RM or M1 macrophages (figure 6A,B), despite induction of 11 β -HSD1 mRNA expression (figure 4A). However, GW1929-treated M2 macrophages displayed a 1.6-fold higher cortisone to cortisol conversion within 60min, indicative of increased 11 β -HSD1 activity (figure 6C). Thus, PPAR γ activation induces 11 β -HSD1 enzyme activity specifically in M2 macrophages. To determine the functionality of PPAR γ -induced 11 β -HSD1 enzyme activity in M2 macrophages, the expression of GR target genes was measured. Cortisone treatment more pronouncedly increased GILZ, PDK4 and ANGPTL4 expression in M2 compared to RM macrophages, an effect amplified by pre-treatment with GW1929 (figure 6D,E,F) and inhibited by the GR antagonist RU486²⁸ (supplemental figure 4). These data further indicate that 11 β -HSD1 enzyme activity is higher and induced by PPAR γ only in M2 macrophages.

Discussion

In this paper we report that expression of 11 β -HSD1, an enzyme which amplifies intracellular GC action, is dependent on the macrophage phenotype. M2 macrophages express higher gene and protein levels of 11 β -HSD1 than RM and M1 macrophages. In line with the expression levels, 11 β -HSD1 enzyme activity is higher in M2 than in RM and M1 macrophages, resulting in a more pronounced conversion of cortisone into active cortisol. Moreover, we demonstrate that PPAR γ activation increases the expression and activity of 11 β -HSD1 in alternative human macrophages. *In vitro* experiments show that different PPAR γ ligands, rosiglitazone and GW1929, increase the expression of 11 β -HSD1 mRNA in RM, M1 and M2 macrophages. The induction of 11 β -HSD1 expression by PPAR γ appears species-specific, since PPAR γ inhibits its expression in mouse adipocytes²⁹. Our data indicate that PPAR γ ligands exert their effects via a PPAR γ -dependent mechanism. Indeed, both the PPAR γ antagonist GW9662 and silencing of PPAR γ expression abolished 11 β -HSD1 induction by GW1929. In addition, over-expression of PPAR γ in macrophages induced 11 β -HSD1 gene expression, an effect enhanced by GW1929. PPAR γ activates 11 β -HSD1 transcription *via* a PPRE located in the 11 β -HSD1 P2 promoter. Binding of PPAR γ to this site occurs also *in cell*, as demonstrated by CHIP experiments, and is enhanced in M2 compared to RM macrophages.

The regulation of 11 β -HSD1 enzyme activity by PPAR γ activation is dependent on the macrophage phenotype. Surprisingly, PPAR γ activation induced 11 β -HSD1 enzyme activity selectively in M2 macrophages, but not in RM nor M1 macrophages, although mRNA levels of 11 β -HSD1 were induced, albeit not to the same extent, by PPAR γ ligands in all three cell types. Concomitantly, PPAR γ induced cortisone conversion into cortisol in M2 macrophages,

which was accompanied by an increased expression of the GR-target genes GILZ, PDK4 and ANGPTL4.

We have previously demonstrated that PPAR γ activation promotes M2 macrophage polarization¹⁵. On the basis of our results, 11 β -HSD1 activity appears to be a marker of macrophage M2 activation. More importantly, this high expression level of 11 β -HSD1 could contribute to the functions of M2 macrophages by converting cortisone to cortisol and thus activating GR target genes. Interestingly, it has been reported that ANGPTL4 binds to the extra-cellular matrix in ischemic tissues and reduces endothelial cell adhesion, prevents the organisation of focal adhesion molecules and actin stress fibers and decreases cell migration³⁰. Therefore, M2 macrophage ANGPTL4 can exert paracrine effects on endothelial cells thus participating in the modulation of angiogenesis in a hypoxic environment. M2 macrophages play also a role in tissue remodelling and repair and hence contribute to the resolution of inflammation¹. Similarly, a role of 11 β -HSD1 in the resolution of inflammation has also been reported³¹. Using macrophages isolated from 11 β -HSD1-deficient mice, it has been shown that 11 β -HSD1-mediated intracellular active GC formation enhances phagocytosis of neutrophils undergoing apoptosis during the resolution of inflammation³¹. Also macrophages from 11 β -HSD1-deficient mice overproduce pro-inflammatory cytokines and are hyper-responsive to LPS stimulation, suggesting that such macrophages exhibit an M1 phenotype³².

Our data indicate that the activity 11 β -HSD1 is tightly related to the inflammatory status of human macrophages. Elevated 11 β -HSD1 activity in polarized M2 macrophages might contribute to their function in the anti-inflammatory response and the resolution of inflammation, with potential consequences on inflammatory diseases, such as atherosclerosis. M2 macrophage 11 β -HSD1 could thus contribute to the anti-inflammatory action of these cells within the atherosclerotic plaques. Finally, our findings demonstrate a novel cross-talk between the nuclear receptors PPAR γ and GR.

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References

1. Gordon S. Alternative activation of macrophages. *Nat Rev Immunol*. 2003;3:23-35.
2. Chinetti-Gbaguidi G, Staels B. Macrophage polarization in metabolic disorders: functions and regulation. *Curr Opin Lipidol*. 2011;22:365-372.
3. Chapman KE, Coutinho A, Gray M, Gilmour JS, Savill JS, Seckl JR. Local amplification of glucocorticoids by 11beta-hydroxysteroid dehydrogenase type 1 and its role in the inflammatory response. *Ann N Y Acad Sci*. 2006;1088:265-273.
4. Karin M. New twists in gene regulation by glucocorticoid receptor: is DNA binding dispensable? *Cell*. 1998;93:487-490.
5. Ricketts ML, Verhaeg JM, Bujalska I, Howie AJ, Rainey WE, Stewart PM. Immunohistochemical localization of type 1 11beta-hydroxysteroid dehydrogenase in human tissues. *J Clin Endocrinol Metab*. 1998;83:1325-1335.
6. Pelletier G, Luu-The V, Li S, Bujold G, Labrie F. Localization and glucocorticoid regulation of 11beta-hydroxysteroid dehydrogenase type 1 mRNA in the male mouse forebrain. *Neuroscience*. 2007;145:110-115.
7. Freeman L, Hewison M, Hughes SV, Evans KN, Hardie D, Means TK, *et al*. Expression of 11beta-hydroxysteroid dehydrogenase type 1 permits regulation of glucocorticoid bioavailability by human dendritic cells. *Blood*. 2005;106:2042-2049.
8. Thieringer R, Le Grand CB, Carbin L, Cai TQ, Wong B, Wright SD, *et al*. 11 Beta-hydroxysteroid dehydrogenase type 1 is induced in human monocytes upon differentiation to macrophages. *J Immunol*. 2001;167:30-35.
9. Chinetti G, Fruchart JC, Staels B. Peroxisome proliferator-activated receptors and inflammation: from basic science to clinical applications. *Int J Obes Relat Metab Disord*. 2003;27 Suppl 3:S41-45.
10. Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol*. 2004;25:677-686.
11. Kliewer SA, Lenhard JM, Willson TM, Patel I, Morris DC, Lehmann JM. A prostaglandin J2 metabolite binds peroxisome proliferator-activated receptor γ and promotes adipocyte differentiation. *Cell*. 1995;83:813-819.
12. Willson TM, Brown PJ, Sternbach DD, Henke BR. The PPARs: from orphan receptors to drug discovery. *Journal of Medicinal Chemistry*. 2000;43:527-550.
13. Pascual G, Fong AL, Ogawa S, Gamliel A, Li AC, Perissi V, *et al*. A SUMOylation-dependent pathway mediates transrepression of inflammatory response genes by PPAR-gamma. *Nature*. 2005;437:759-763.
14. Odegaard JI, Ricardo-Gonzalez RR, Goforth MH, Morel CR, Subramanian V, Mukundan L, *et al*. Macrophage-specific PPARgamma controls alternative activation and improves insulin resistance. *Nature*. 2007;447:1116-1120.
15. Bouhrel MA, Derudas B, Rigamonti E, Dievart R, Brozek J, Haulon S, *et al*. PPARg activation primes human monocytes into alternative M2 macrophages with anti-inflammatory properties. *Cell Metabolism*. 2007;6:137-143.
16. Chinetti G, Griglio S, Antonucci M, Pineda Torra I, Delerive P, Majd Z, *et al*. Activation of peroxisome proliferator-activated receptors α and γ induces apoptosis of human monocyte-derived macrophages. *JBC*. 1998;273:25573-25580.
17. Wang N, Lan D, Chen W, Matsuura F, Tall AR. ATP-binding cassette transporters G1 and G4 mediate cellular cholesterol efflux to high-density lipoproteins. *PNAS*. 2004;9774-9779.

18. Odom DT, Dowell RD, Jacobsen ES, Nekludova L, Rolfe PA, Danford TW, *et al.* Core transcriptional regulatory circuitry in human hepatocytes. *Mol Syst Biol.* 2006;2:2006 0017.
19. Nielsen R, Pedersen TA, Hagenbeek D, Moulos P, Siersbaek R, Megens E, *et al.* Genome-wide profiling of PPAR γ :RXR and RNA polymerase II occupancy reveals temporal activation of distinct metabolic pathways and changes in RXR dimer composition during adipogenesis. *Genes Dev.* 2008;22:2953-2967.
20. Rigamonti E, Fontaine C, Lefebvre B, Duhem C, Lefebvre P, Marx N, *et al.* Induction of CXCR2 Receptor by Peroxisome Proliferator-Activated Receptor { γ } in Human Macrophages. *Arterioscler Thromb Vasc Biol.* 2008.
21. Low SC, Chapman KE, Edwards CR, Seckl JR. 'Liver-type' 11 beta-hydroxysteroid dehydrogenase cDNA encodes reductase but not dehydrogenase activity in intact mammalian COS-7 cells. *J Mol Endocrinol.* 1994;13:167-174.
22. Lavery GG, Walker EA, Draper N, Jeyasuria P, Marcos J, Shackleton CH, *et al.* Hexose-6-phosphate dehydrogenase knock-out mice lack 11 beta-hydroxysteroid dehydrogenase type 1-mediated glucocorticoid generation. *J Biol Chem.* 2006;281:6546-6551.
23. Koliwad SK, Kuo T, Shipp LE, Gray NE, Backhed F, So AY, *et al.* Angiopoietin-like 4 (ANGPTL4, fasting-induced adipose factor) is a direct glucocorticoid receptor target and participates in glucocorticoid-regulated triglyceride metabolism. *J Biol Chem.* 2009;284:25593-25601.
24. Connaughton S, Chowdhury F, Attia RR, Song S, Zhang Y, Elam MB, *et al.* Regulation of pyruvate dehydrogenase kinase isoform 4 (PDK4) gene expression by glucocorticoids and insulin. *Mol Cell Endocrinol.* 2010;315:159-167.
25. Leesnitzer LM, Parks DJ, Bledsoe RK, Cobb JE, Collins JL, Consler TG, *et al.* Functional consequences of cysteine modification in the ligand binding sites of peroxisome proliferator activated receptors by GW9662. *Biochemistry.* 2002;41:6640-6650.
26. Ernst J, Kheradpour P, Mikkelsen TS, Shoresh N, Ward LD, Epstein CB, *et al.* Mapping and analysis of chromatin state dynamics in nine human cell types. *Nature.* 2011;473:43-49.
27. Staab CA, Stegk JP, Haenisch S, Neiss E, Kobsch K, Ebert B, *et al.* Analysis of alternative promoter usage in expression of HSD11B1 including the development of a transcript-specific quantitative real-time PCR method. *Chem Biol Interact.* 2011;191:104-112.
28. Beck CA, Estes PA, Bona BJ, Muro-Cacho CA, Nordeen SK, Edwards DP. The steroid antagonist RU486 exerts different effects on the glucocorticoid and progesterone receptors. *Endocrinology.* 1993;133:728-740.
29. Engel T, Lorkowski S, Lueken A, Rust S, Schluter B, Berger G, *et al.* The human ABCG4 gene is regulated by oxysterols and retinoids in monocyte-derived macrophages. *Biochemical Biophysical Research Communications.* 2001;288:483-488.
30. Cazes A, Galaup A, Chomel C, Bignon M, Brechot N, Le Jan S, *et al.* Extracellular matrix-bound angiopoietin-like 4 inhibits endothelial cell adhesion, migration, and sprouting and alters actin cytoskeleton. *Circ Res.* 2006;99:1207-1215.
31. Gilmour JS, Coutinho AE, Cailhier JF, Man TY, Clay M, Thomas G, *et al.* Local amplification of glucocorticoids by 11 beta-hydroxysteroid dehydrogenase type 1 promotes macrophage phagocytosis of apoptotic leukocytes. *J Immunol.* 2006;176:7605-7611.

32. Zhang TY, Daynes RA. Macrophages from 11beta-hydroxysteroid dehydrogenase type 1-deficient mice exhibit an increased sensitivity to lipopolysaccharide stimulation due to TGF-beta-mediated up-regulation of SHIP1 expression. *J Immunol.* 2007;179:6325-6335.

Figure legends

Figure 1. *11 β -HSD1* gene expression is higher in M2 macrophages.

Primary human macrophages were cultured for 7 days in the absence (RM) or in the presence of IL-4 (15 ng/ml) (M2). Pro-inflammatory M1 macrophages were obtained by activating RM macrophages with LPS (100ng/ml) during 4h. 11 β -HSD1 (A), MR (B) and AMAC1 (C) mRNA levels measured by Q-PCR and normalized to those of cyclophilin. Results are expressed as the mean value \pm SD of triplicate determinations, representative of three independent experiments. Statistically significant differences are indicated (**p<0.01, ***p<0.001).

Figure 2. *11 β -HSD1* enzyme activity is highest in M2 macrophages leading to pronounced anti-inflammatory effects of cortisone.

RM (A), M1 (B) and M2 macrophages (C) were incubated with radio-labeled cortisone (E) for the indicated time periods. Production of cortisol (F) was then measured. 11 β -HSD1 reductase activity was determined as the percentage conversion of cortisone to cortisol. Results are representative of two independent experiments. Statistically significant differences are indicated (*p<0.05; **p<0.01, ***p<0.001).

RM and M2 macrophages were treated with cortisone or dexamethasone (Dexa, 1 μ M) for 2h before stimulation with LPS (100ng/ml) for 4h. mRNA levels of IL-6 (D), TNF α (E) and IL-1 β (F) were measured by Q-PCR. Results are representative of three independent experiments. Statistically significant differences between control and treated cells are indicated (*p<0.05, **p<0.01).

Figure 3. *Cortisone* more pronouncedly induces the expression of GR target genes in M2 macrophages.

RM and M2 macrophages were incubated with cortisone (1 μ M) for different time points. GILZ (A), ANGPTL4 (B), and PDK4 (C) mRNA levels were measured by Q-PCR. Statistically significant differences between control and treated cells are indicated (* p <0.05;** p <0.01;*** p <0.001).

Figure 4. *11 β -HSD1* expression is induced by PPAR γ in RM, M1 and M2 macrophages.

(A) RM, M2 and M1 macrophages were treated in the absence or in the presence of GW1929 (600nM); 11 β -HSD1 mRNA level was measured by Q-PCR. Results are representative of three independent experiments. Statistically significant differences between control and treated cells (* p <0.05;** p <0.01;*** p <0.001) and basal RM or M1 and M2 macrophages are indicated ([§] p <0.05; ^{§§} p <0.01). (B) Intracellular 11 β -HSD1 and β -actin protein expression analyzed by western blot and immunoreactive band intensity was quantified. Results are representative of two independent experiments. (C) RM were treated with vehicle or the PPAR γ antagonist GW9662 (1 μ M) in the absence or presence of GW1929 (600nM) for 24h. (D) RM were transfected with scrambled or human PPAR γ siRNA and subsequently treated with GW1929 (600nM) or DMSO during 24h. (E) RM were infected with Ad-GFP or Ad-PPAR γ and subsequently stimulated for 24h with or without GW1929 (600nM). 11 β -HSD1 mRNA levels were measured by Q-PCR and normalized to those of cyclophilin. Results are expressed as the mean value \pm SD of triplicate determinations, representative of three independent experiments. Statistically significant differences are indicated (* p <0.05;** p <0.01;*** p <0.001).

Figure 5. PPAR γ binds more avidly to the 11 β -HSD1 PPPE in M2 macrophages.

(A) EMSA was performed using the end-labeled DR1-consensus-PPPE or 11 β -HSD1-PPPE oligonucleotide in the presence of unprogrammed reticulocyte lysate or *in vitro* translated

hPPAR γ and hRXR α . Supershift assays were performed using an anti-PPAR γ antibody. **(B)** RM were transfected with the indicated reporter constructs (DR1-11 β -HSD1 PPRE)₆ or (DR1-consensus PPRE)₆ **(C)**, in the presence of pSG5-empty or pSG5-PPAR γ vector, treated or not with GW1929 (600nM) and luciferase activity was measured. Statistically significant differences are indicated (*p<0.05;**p<0.01;***p<0.001). **(D)** EMSA was performed using *in vitro* produced RXR and PPAR γ or nuclear extracts from RM, M1 and M2 in the absence or in the presence of exogenous hRXR α and supershift assays performed using an anti-PPAR γ antibody. **(E)** ChIP assays were performed and quantified using chromatin from RM and M2 macrophages, immunoprecipitated with rabbit IgG or PPAR γ -specific antibodies and then subjected to PCR using primer pairs covering either the 11 β -HSD1 gene promoter or the β -actin gene. **(F)** H3K9ac ChIP-seq data for RM and M2 macrophages. The Y-axis shows the number of mapped tags sequenced on ChIP DNA from both RM and M2 macrophages. Promoters P1 and P2 as well as the identified PPRE are indicated. ChIP experiments were performed on H3K9-immunoprecipitated chromatin from two independent donors using primers covering the two identified 11 β -HSD1 PPRE sites on the P1 and P2 promoters. Relative fold enrichments relative to a negative control region (set at 1) are shown. **(G)** RM and M2 macrophages were transfected with the (DR1-11 β -HSD1 PPRE)₆ construct, treated or not with GW1929 (600nM). Statistically significant differences are indicated (*p<0.05;**p<0.01).

Figure 6. PPAR γ activation increases 11 β -HSD1 activity in M2 macrophages leading to the induction of GR-target genes by cortisone.

RM **(A)**, M1 **(B)** and M2 **(C)** macrophages were treated for 24h in the absence or in the presence of GW1929 (600nM) and subsequently incubated with radio-labelled cortisone **(E)** for the indicated time periods. Production of cortisol **(F)** was then measured. 11 β -HSD1

reductase activity was determined as the percentage conversion of cortisone to cortisol.

Results are representative of two independent experiments. Statistically significant differences between control and treated cells are indicated (* $p < 0.05$). RM and M2 macrophages were activated or not with GW1929 (600nM) for 24h and subsequently treated for another 24h with cortisone (1 μ M). PDK4 (**D**), GILZ (**E**) and ANGPTL4 (**F**) mRNA levels were measured by Q-PCR. Statistically significant differences between control and treated cells are indicated (* $p < 0.05$; * $p < 0.01$; *** $p < 0.001$).

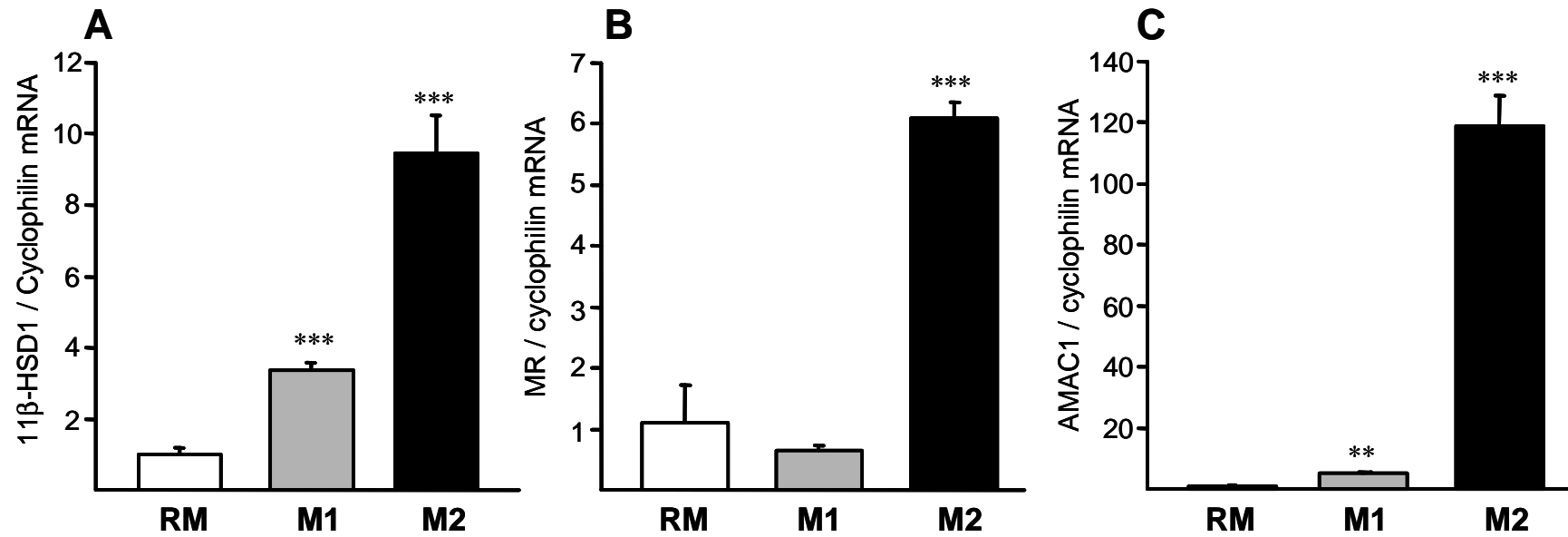


Figure 1

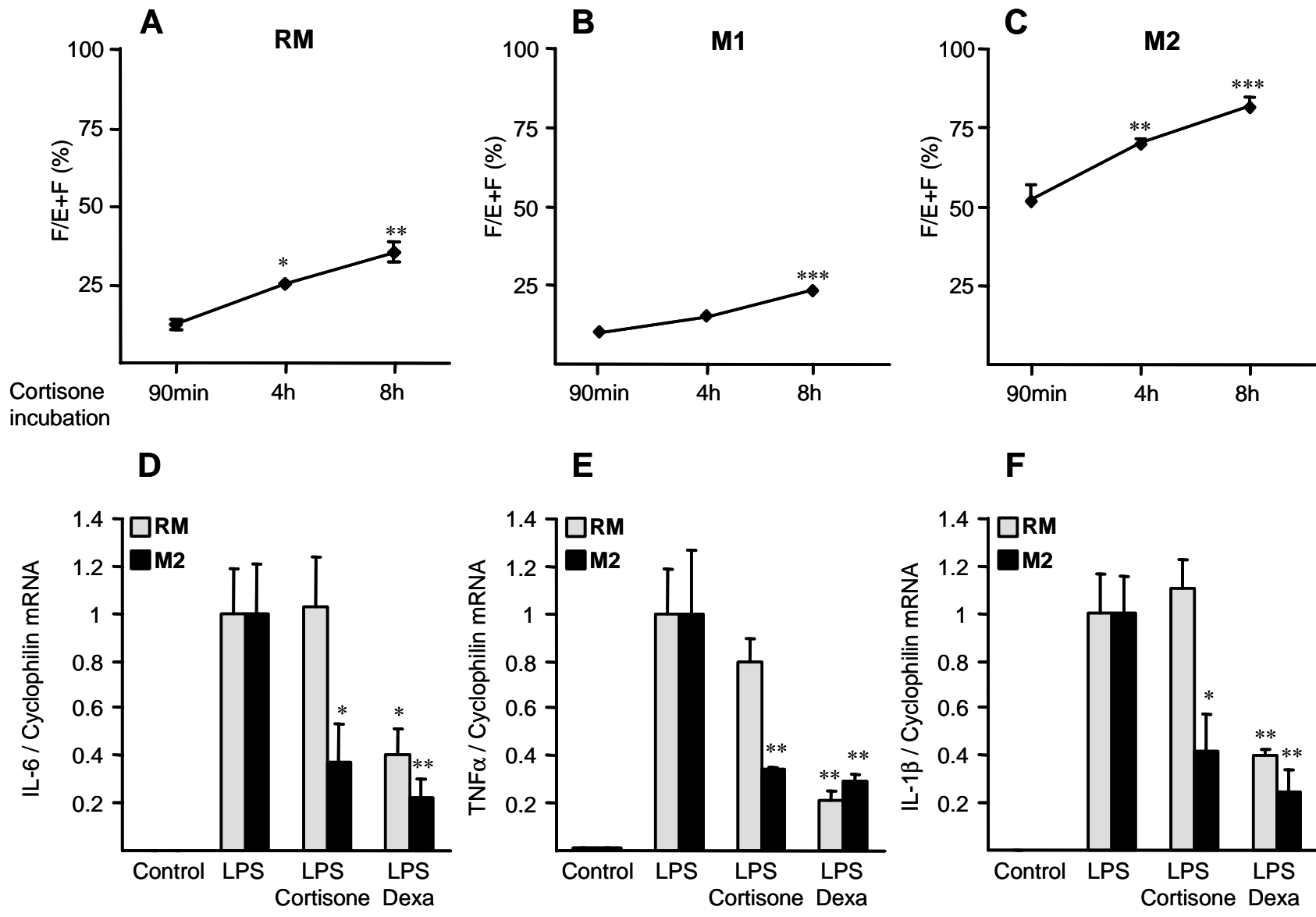


Figure 2

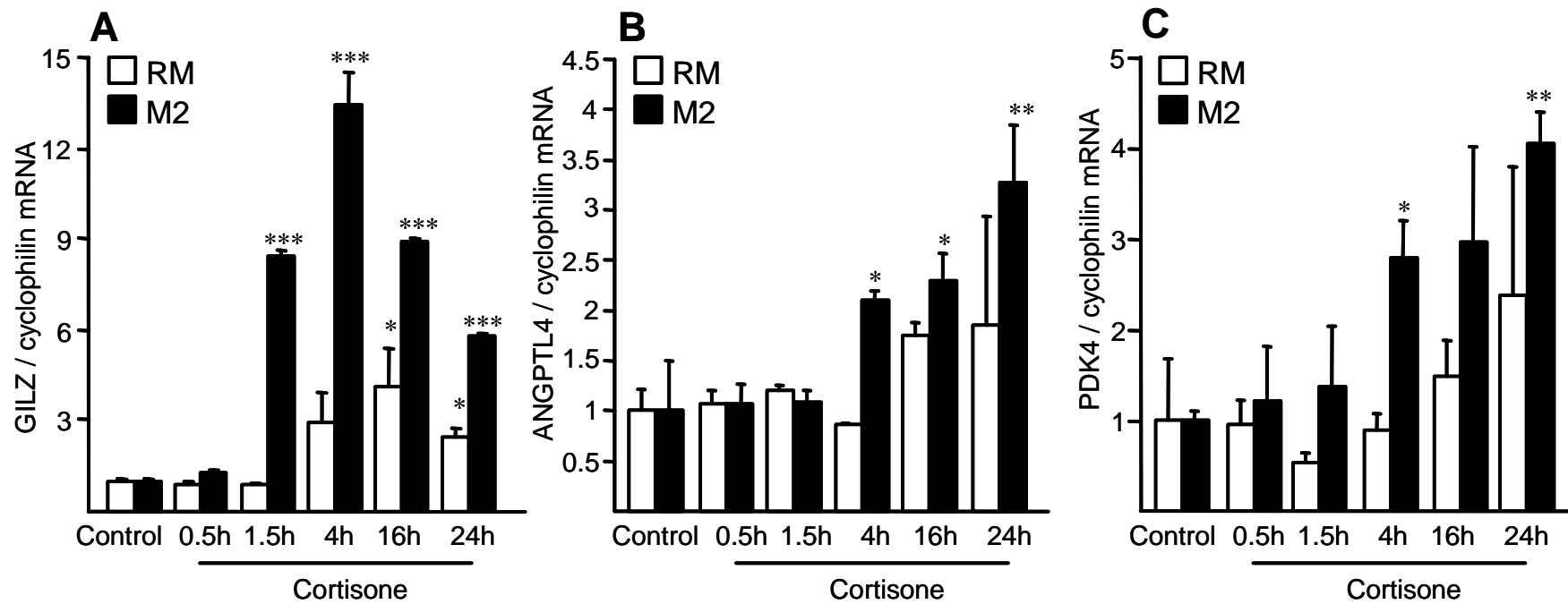


figure 3

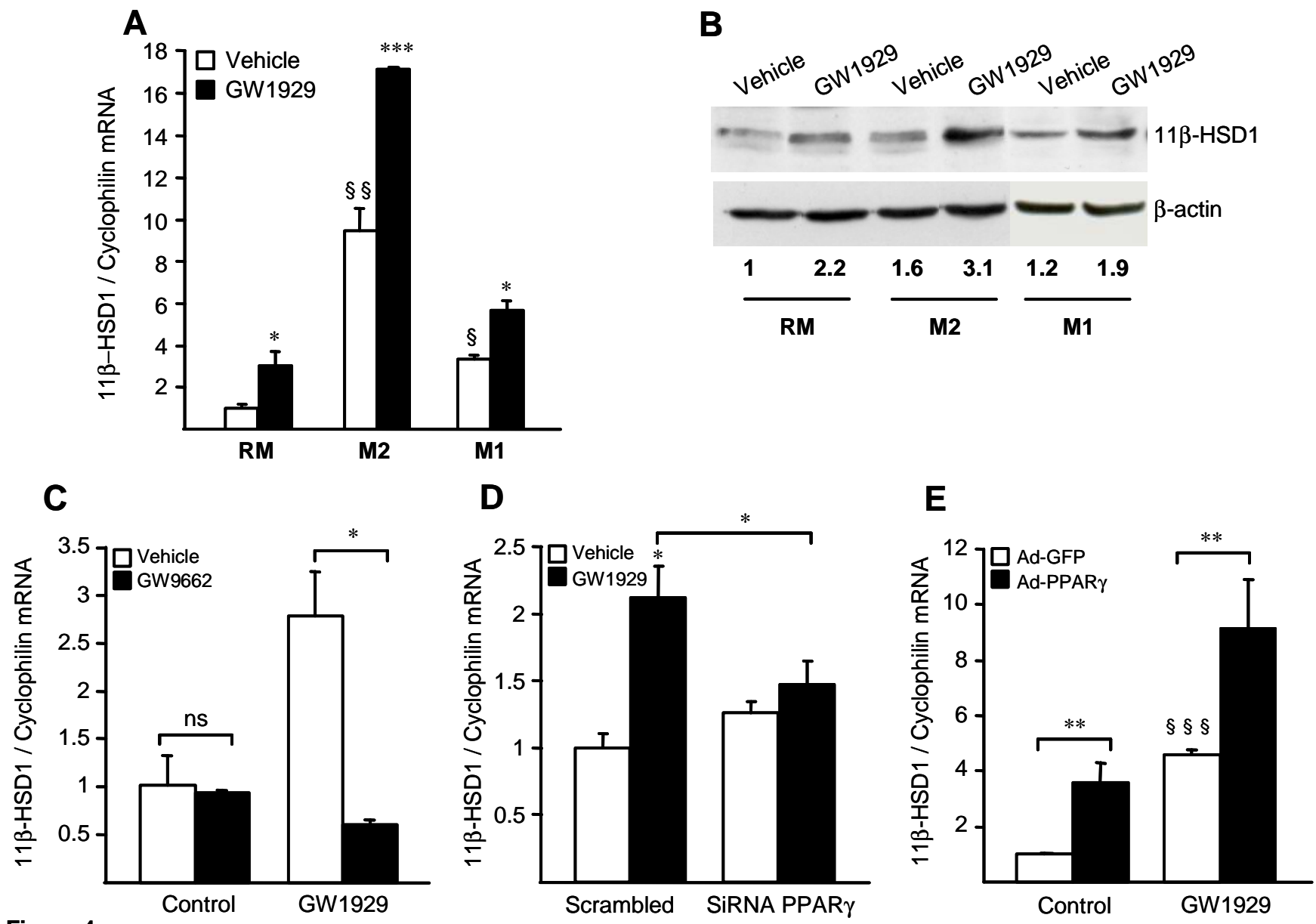


Figure 4

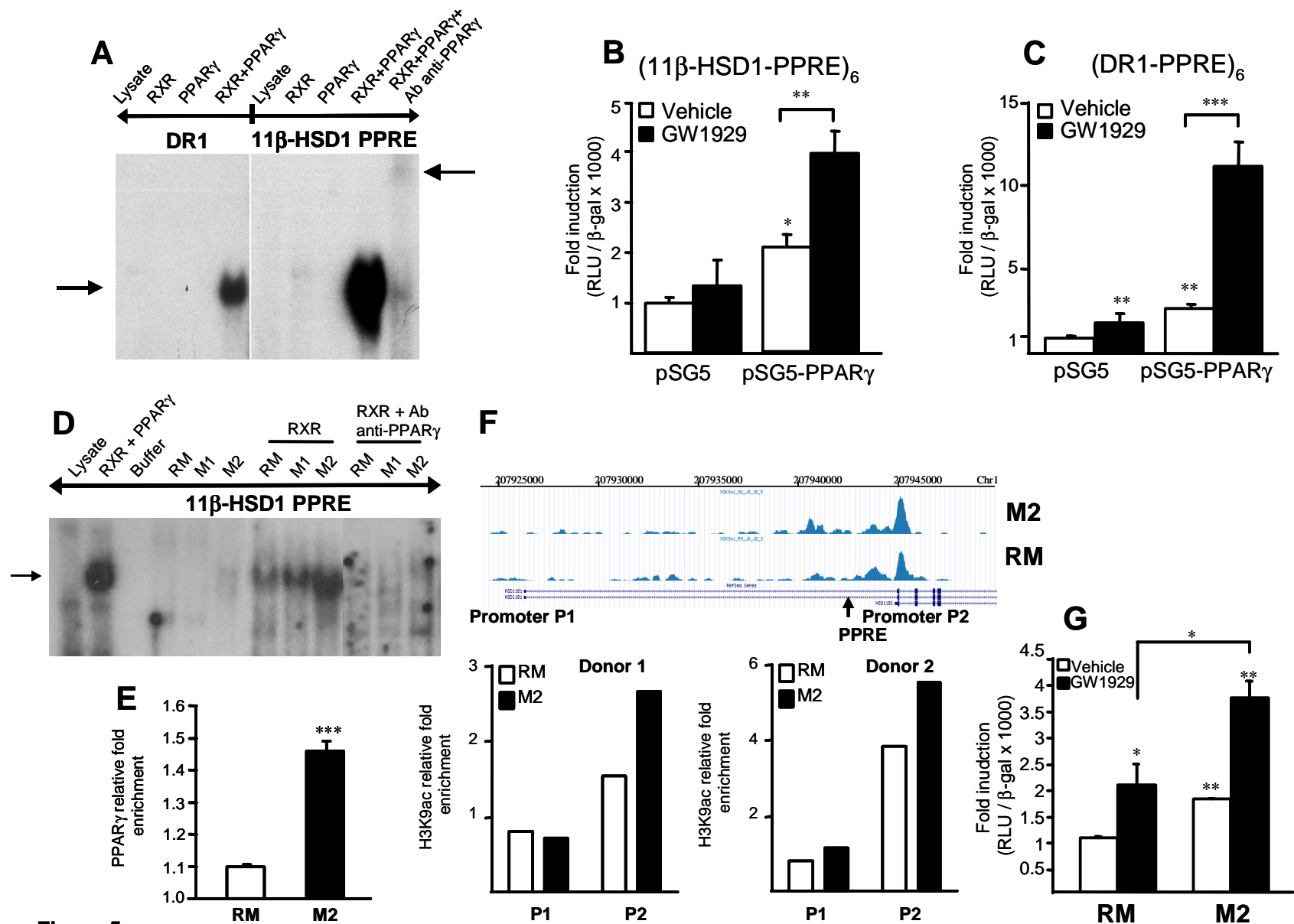


Figure 5

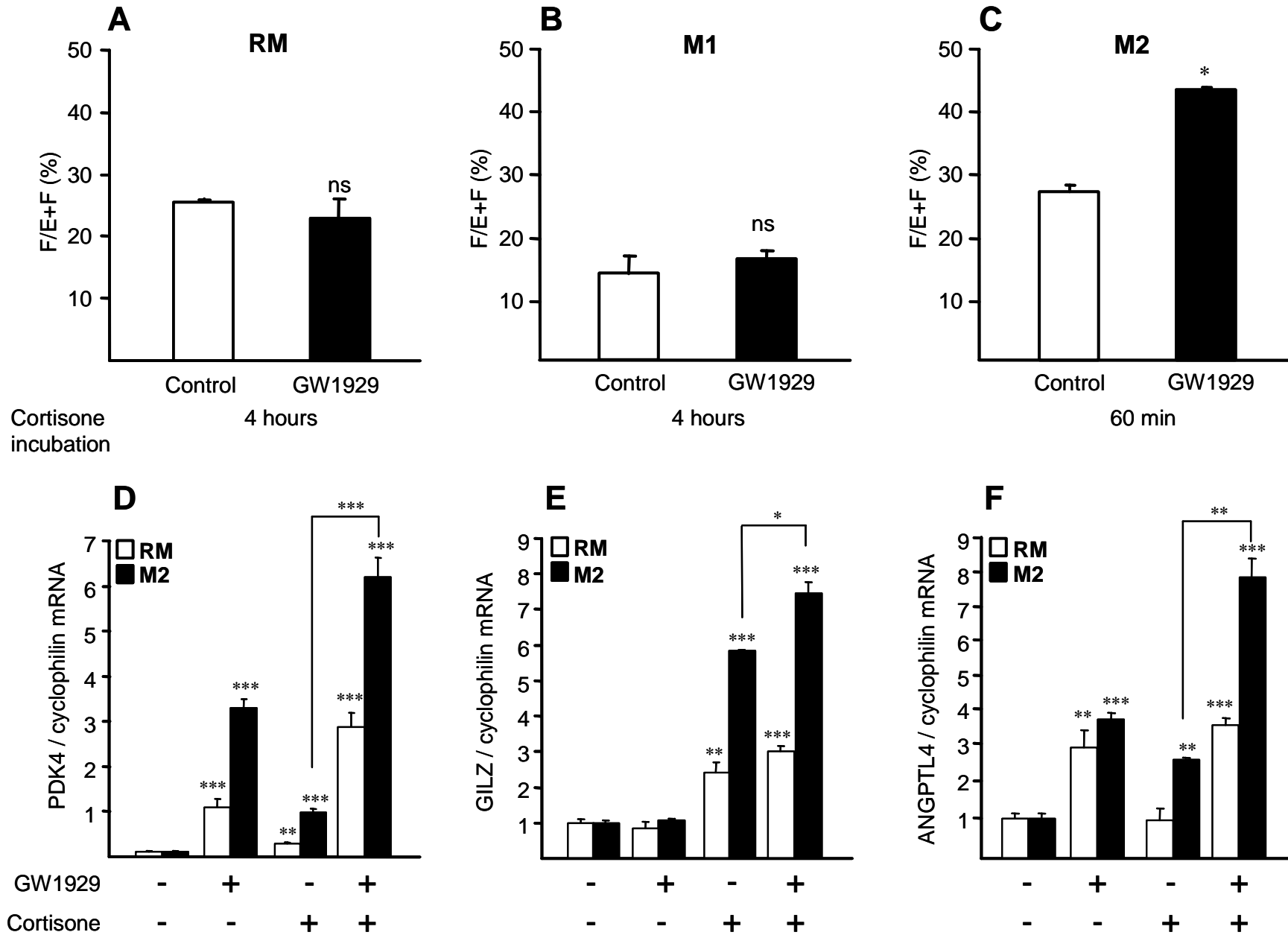


Figure 6