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Distinct Changes in cAMP and Extracellular Signal-Regulated Protein Kinase Signalling in L-DOPA-Induced Dyskinesia

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Abstract

Background: In rodents, the development of dyskinesia produced by L-DOPA in the dopamine-depleted striatum occurs in response to increased dopamine D1 receptor-mediated activation of the cAMP - protein kinase A and of the Ras-extracellular signal-regulated kinase (ERK) signalling pathways. However, very little is known, in non-human primates, about the regulation of these signalling cascades and their association with the induction and/or maintenance of dyskinesia.

Methodology/Results: We here studied, in the gold-standard non-human primate model of Parkinson’s disease, the changes in PKA-dependent phosphorylation of DARPP-32 and GluR1 AMPA receptor, as well as in ERK and ribosomal protein S6 (S6) phosphorylation, associated to acute and chronic administration of L-DOPA. Increased phosphorylation of DARPP-32 and GluR1 was observed in both L-DOPA first-ever exposed and chronically-treated dyskinetic parkinsonian monkeys. In contrast, phosphorylation of ERK and S6 was enhanced preferentially after acute L-DOPA administration and decreased during the course of chronic treatment.

Conclusion: Dysregulation of cAMP signalling is maintained during the course of chronic L-DOPA administration, while abnormal ERK signalling peaks during the initial phase of L-DOPA treatment and decreases following prolonged exposure. While cAMP signalling enhancement is associated with dyskinesia, abnormal ERK signalling is associated with priming.

Introduction

In Parkinson’s disease (PD), long-term treatment with L-3,4-dihydroxyphenylalanine (L-DOPA) leads to the emergence of dyskinesia, or involuntary aimless movements [1]. Work in rodents has led to the identification of numerous changes in signalling potentially implicated in L-DOPA-induced dyskinesia (LID) [2]. Prominent among these changes is a marked increase in the dopamine D1 receptor (D1R)-mediated activation of (i) the canonical adenyl cyclase - protein kinase A (PKA) - dopamine and cAMP-regulated phosphoprotein, 32 kDa (DARPP-32) signalling cascade [3,4] and (ii) the non-canonical Ras-extracellular signal-regulated kinase (ERK) signalling cascade [5,6,7,8]. In order to identify potential therapeutic targets, it is essential to understand the role of these signalling cascades in the actual induction, manifestation and/or maintenance of LID. However, the correlation between abnormal PKA/DARPP-32 and Ras-ERK signalling and severity of LID has been established only in rodent models of LID and without the appropriate L-DOPA-treated unlesioned controls. This limits a clear-cut association between specific biochemical events and one or several of the above mentioned key steps in LID elicitation.

In this study, we made use of the gold standard macaque model of LIDs [3,9] to show that dysregulation of PKA/DARPP-32 signalling is mainly associated to the manifestation of dyskinesia, while abnormal Ras-ERK signalling is most evident during the development of this condition.

Materials and Methods

Animals

Thirty-nine F2-bred female rhesus monkeys (Macaca mulatta, Xierxin, Beijing, PR of China; mean weight 5.3±0.8 kg; mean age = 5±1 years) were used. Animals were housed in individual primate cages under controlled conditions of humidity (50±5%), temperature (24±1°C) and light (12 h light/12 h dark cycles, time
lights on 8:00 am) with food and water ad libitum. Experiments were carried out in accordance with European Communities Council Directive of 24 November 1986 (86/609/EEC) for care of laboratory animals in an AAALAC-accredited facility following acceptance of study design by the Institute of Lab Animal Science (Chinese Academy of Science, Beijing, China) IACUC. Veterinarians skilled in the healthcare and maintenance of non-human primates supervised animal care. All reasonable efforts were made to minimize animal suffering. The use of primates was minimized by using an experimental design that permits statistically-significant changes to be demonstrated with the smallest number of animals per group and the smallest number of groups, consistent with scientific rigour. Experiments followed previously published procedures [3,11,12,13,14]. All animals were killed with a sodium pentobarbital overdose (150 mg/kg, i.v.). Dissection of different brain regions were performed on ice with the brain immersed in cold saline (0.9%) in less than 15 min. The dorsolateral striatum was dissected from each hemisphere, immediately frozen at −45°C in isopentane and then stored at −80°C.

Experimental Protocol

The experimental flowchart is described in figure 1. Six animals were kept as untreated controls (control group), five received a single dose of 20 mg/kg of L-DOPA p.o (control acute L-DOPA) and five received 20 mg/kg of L-DOPA twice daily for three months (control chronic L-DOPA). The remaining 32 animals were treated daily (9:00 am) with 1-methyl-4-phenyl 1,2,3,6-tetrahydropyridine (MPTP) hydrochloride (0.2 mg/kg, i.v., Sigma, St Louis, MO) dissolved in saline according to a previously described protocol [10]. Following stabilization of the MPTP-induced syndrome, animals received saline (MPTP; n = 5), a single dose of 20 mg/kg of L-DOPA (MPTP acute L-DOPA; n = 6), or 20 mg/kg of L-DOPA twice daily for three months (MPTP chronic L-DOPA; n = 12). Six animals developed severe and reproducible dyskinesias, presenting choreic–athetoid (characterized by constant writhing and jerking motions), dystonic and sometimes ballistic movements (large-amplitude flinging, flailing movements) (MPTP-intoxicated and chronically L-DOPA-treated, dyskinetic animal group), while 6 other did not (MPTP-intoxicated and chronically L-DOPA-treated, non-dyskinetic animal group). The MPTP intoxication protocol, the chronic L-DOPA treatment, the clinical assessments, the terminal procedure and the characterization of the extent of nigrostriatal denervation were conducted as previously published [3,11,12,13,14]. DAT binding autoradiography using [125I]-(E)-N-(3-iodoprop-2-enyl)-2β-carboxymethyl-3β-(4'-methylphenyl)-nortropane; Chelatec, France) showed a dramatic and similar reduction (>95%) in all MPTP-treated groups in comparison to control animals, as published elsewhere [12].

Behavioural assessment

Animal behaviour was assessed in their home cages. All observers were blinded with regard to the experimental protocol. During each session, two blinded examiners evaluated the level of motor performance of each animal, by coaxing them to perform various tasks by offering appetizing fruits. Animals received supplemental feeding from day 7 onwards to maintain their body weight as constant as possible. The degree of parkinsonism was assessed daily (9 a.m.) for 30 min using a validated parkinsonian macaque clinical scale [3,11,12,13,14] which rates the following symptoms of parkinsonian disability: tremor, variations in the general level of activity, body posture (flexion of spine), vocalization, freezing and frequency of arm movements (reaching for food for each upper limb) and rigidity (for each upper limb). The minimal disability score was 0 and the maximum score was 25. The severity of dyskinesia was rated using the Dyskinesia Disability Scale: 0, dyskinesia absent; 1, mild, fleeting, and rare dyskinetic postures and movements; 2, moderate, more prominent abnormal movements, but not interfering significantly with normal behaviour; 3, marked, frequent and, at times, continuous dyskinesia intruding on the normal repertoire of activity; or, 4, severe, virtually continuous dyskinetic activity, disabling to the

![Figure 1. Experimental flowchart illustrating study design, treatments and group assignments.](https://doi.org/10.1371/journal.pone.0012322.g001)
animal and replacing normal behaviour. Median rating scores and 
SEM were calculated daily for each group.

Western Blotting

Patches collected from 300 µm-thick fresh frozen coronal 
sections containing dorsal motor putamen were sonicated in 1% 
SDS and Western blotting was performed as described [6], using 
antibodies against phospho-Thr34-DARPP-32 [15], phospho-
Ser845-GluR1 (PhosphoSolutions, Aurora, CO, USA), phospho-
Thr202/Tyr204-ERK1/2 and phospho-Ser235/236-S6 (Cell 
Signaling Technology), and against total DARPP-32 [15], S6, 
ERK and GluR1 (Cell Signaling Technology). Detection was 
based on fluorescent secondary antibody binding and quantified 
using a Li-Cor Odyssey infrared fluorescent detection system (Li-
Cor, Lincoln, NE). The levels of each phosphoprotein were 
normalized for the amount of the corresponding total protein 
detected in the sample.

Results

Parkinsonism and dyskinesia status

MPTP-treated animals displayed the classic progression of 
symptoms previously shown with this intoxication regimen [10]. 
Monkeys became increasingly bradykinetic, adopted a stooped 
posture, with increased rigidity of the limbs and decreased 
vocalization. Movement accuracy progressively deteriorated and 
there were occasional episodes of freezing as well as postural 
tremor. Stable parkinsonism was obtained after 19 ± 0.5 MPTP 
injections (parkinsonian score = 9.3 ± 0.34). All MPTP groups 
displayed a similar parkinsonian score (MPTP: 8.5 ± 0.5, MPTP 
acute L-Dopa: 9.5 ± 0.5, MPTP chronic L-Dopa (dyskinetic): 9.33 ± 0.4, MPTP chronic L-Dopa (non dyskinetic): 9.83 ± 0.87). 
Parkinsonian scores of all MPTP groups were significantly 
different from controls (F6,35 = 88.12, p < 0.0001) and identical 
between them. Animals who developed L-Dopa induced dyskine-
sia following chronic L-Dopa treatment had a dyskinetic score of 
2.33 ± 0.21.

cAMP signalling

Administration of L-DOPA, or MPTP-induced lesion, did not 
alter, per se, the phosphorylation of DARPP-32 at Thr34 (Fig. 2A), 
nor did they affect the phosphorylation of another PKA target, the 
GluR1 subunit of the glutamate AMPA receptor (Fig. 2B). In 
contrast, the first ever administration of L-DOPA to MPTP-
lesioned monkeys induced a trend toward increased phosphory-
lization of DARPP-32 at Thr34, and a significant increase in 
phosphorylation of GluR1 at Ser845 (Fig. 2A–B). Following 
chronic treatment with L-DOPA, the phosphorylation of DARPP-
Previous work showed that activated ERK phosphorylates S6 at Ser235/236 and that this regulation promotes 5' cap-dependent initiation of messenger RNA translation [17]. In the mouse, increased S6 phosphorylation has been linked to ERK activation and to the development of LID [18]. In our experiment, the pattern of S6 phosphorylation at Ser235/236 in the various experimental groups exactly matched that of phosphorylated ERK (Fig. 2D). Total levels of ERK and S6 were unaffected by MPTP lesion and/or treatment with L-DOPA (Fig. 3C–D).

Discussion

This study shows that, in a non-human primate model of dyskinesia, the concomitant enhancement of cAMP and ERK signalling in the dorsal striatum is associated to the induction of LIDs. Moreover, once LIDs have been established, there is a preponderant activation of the cAMP/DARPP-32 signalling cascade associated to their acute manifestation.

Acute and chronic administration regimens of L-DOPA to unlesioned monkeys do not affect the state of phosphorylation of DARPP-32 and its target, GluR1. Similarly, L-DOPA per se does not affect ERK and S6 phosphorylation. In contrast, after dopamine denervation, the first ever administration of L-DOPA activates both signalling cascades, as evidenced by the significant

Figure 3. Western blot analysis of levels of (A) DARPP-32 (B) GluR1 (C) ERK1/2 and (D) S6 in the dorsal putamen (mean ± S.E.M.). Groups were prepared as depicted in Figure 1. Representative blots are shown above each panel. doi:10.1371/journal.pone.0012322.g003
The increase in phosphorylation of DARPP-32 and GluR1 produced by L-DOPA following MPTP-lesion is further increased by chronic administration, but only in dyskinetic monkeys. These results suggest that sensitized cAMP signalling plays a pivotal role in the manifestation of dyskinesia. Moreover, they are in agreement with data obtained in rodents, showing that the ability of L-DOPA to increase DARPP-32 phosphorylation is preserved in dyskinetic, but lost in non-dyskinetic animals [6,21].

The ability of L-DOPA to promote ERK signalling is maximal at the first drug administration and declines during chronic treatment. Thus, after three months of daily L-DOPA injections, the phosphorylation of ERK1/2 and S6, albeit still enhanced, is much closer to basal levels. The ability of prolonged administration to attenuate the effects of L-DOPA on ERK signalling has been previously documented in the mouse [6]. However, studies in rodent models show a correlation between LIDs and increased ERK activation [5,6,7]. In contrast, the present data indicate that, following prolonged L-DOPA administration, the levels of phospho-Thr202/Tyr204-ERK and phospho-Ser235/236-S6 found in dyskinetic and non-dyskinetic monkeys are undistinguishable. This discrepancy may be explained by species difference, or by the different length of exposure to chronic L-DOPA employed in the two models (10 to 21 days in rodents vs. three months in monkeys). Furthermore, as protein phosphorylation has here been determined at the single time point of 60 min after administration of L-DOPA, we cannot exclude that the effect of L-DOPA is reduced to a rapid and transient increase in ERK signalling, which cannot be detected under the present experimental conditions. Nevertheless, even this type of change would indicate an extinction of response, as a result of a prolonged administration of L-DOPA.

In conclusion, this study shows that, in non-human primates, dopamine depletion confers to L-DOPA the ability to activate the PKA/DARPP-32 and ERK cascades and that long-term (three months) administration of L-DOPA leads to regression of ERK activation, whereas cAMP/DARPP-32 signalling persists unchanged, or even exacerbated. Given the involvement of ERK and DARPP-32 in LIDs, as indicated by studies performed in rodent models [5,6,7], these results are compatible with the hypothesis that coordinated activation of cAMP/PKA/DARPP-32 and ERK is implicated in the priming processes underlying the emergence of dyskinesia. In this initial phase, activation of the Ras-ERK signalling cascade would lead to transcriptional/translational events necessary to produce and stabilize dyskinetic behaviour. In a later phase, once LIDs have been established, hyper-activation of ERK may become sufficient to maintain their expression independently of ERK, most likely by short-term regulation of neuronal activity through, for instance, regulation of the state of phosphorylation of GluR1 and other ion channels.

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Author Contributions

Conceived and designed the experiments: GF EB. Performed the experiments: ES VF QL MS SD GF EB. Analyzed the data: VF QL MS SD GF EB. Wrote the paper: ES GF EB.

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