

AMP-activated protein kinase deficiency reduces ozone-induced lung injury and oxidative stress in mice.

Sébastien Hulo, Hélène Tiesset, Steve Lancel, Jean Louis Edmé, Benoit Viollet, Annie Sobaszek, Rémi Nevière

► **To cite this version:**

Sébastien Hulo, Hélène Tiesset, Steve Lancel, Jean Louis Edmé, Benoit Viollet, et al.. AMP-activated protein kinase deficiency reduces ozone-induced lung injury and oxidative stress in mice.. *Respiratory Research*, BioMed Central, 2011, 12 (1), pp.64. 10.1186/1465-9921-12-64 . inserm-00602564

HAL Id: inserm-00602564

<https://www.hal.inserm.fr/inserm-00602564>

Submitted on 22 Jun 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH

Open Access

AMP-activated protein kinase deficiency reduces ozone-induced lung injury and oxidative stress in mice

Sébastien Hulo^{1,2†}, H  l  ne Tiesset^{1,3†}, Steve Lancel^{1,4†}, Jean J Louis Edm  ^{1,2†}, Benoit Viollet^{5,6,7†}, Annie Sobaszek^{1,2†} and R  mi Nevi  re^{1,4**†}

Abstract

Background: Acute ozone exposure causes lung oxidative stress and inflammation leading to lung injury. At least one mechanism underlying the lung toxicity of ozone involves excessive production of reactive oxygen and nitrogen intermediates such as peroxynitrite. In addition and beyond its major prooxidant properties, peroxynitrite may nitrate tyrosine residues altering phosphorylation of many protein kinases involved in cell signalling. It was recently proposed that peroxynitrite activates 5'-AMP-activated kinase (AMPK), which regulates metabolic pathways and the response to cell stress. AMPK activation as a consequence of ozone exposure has not been previously evaluated. First, we tested whether acute ozone exposure in mice would impair alveolar fluid clearance, increase lung tissue peroxynitrite production and activate AMPK. Second, we tested whether loss of AMP-activated protein kinase alpha1 subunit in mouse would prevent enhanced oxidative stress and lung injury induced by ozone exposure.

Methods: Control and AMPK α 1 deficient mice were exposed to ozone at a concentration of 2.0 ppm for 3 h in glass cages. Evaluation was performed 24 h after ozone exposure. Alveolar fluid clearance (AFC) was evaluated using fluorescein isothiocyanate tagged albumin. Differential cell counts, total protein levels, cytokine concentrations, myeloperoxidase activity and markers of oxidative stress, i.e. malondialdehyde and peroxynitrite, were determined in bronchoalveolar lavage (BAL) and lung homogenates (LH). Levels of AMPK-Thr¹⁷² phosphorylation and basolateral membrane Na(+)-K(+)-ATPase abundance were determined by Western blot.

Results: In control mice, ozone exposure induced lung inflammation as evidence by increased leukocyte count, protein concentration in BAL and myeloperoxidase activity, pro-inflammatory cytokine levels in LH. Increases in peroxynitrite levels (3 vs 4.4 nM, $p = 0.02$) and malondialdehyde concentrations (110 vs 230 μ mole/g wet tissue) were detected in LH obtained from ozone-exposed control mice. Ozone exposure consistently increased phosphorylated AMPK-Thr¹⁷² to total AMPK ratio by 80% in control mice. Ozone exposure causes increases in AFC and basolateral membrane Na(+)-K(+)-ATPase abundance in control mice which did not occur in AMPK α 1 deficient mice.

Conclusions: Our results collectively suggest that AMPK activation participates in ozone-induced increases in AFC, inflammation and oxidative stress. Further studies are needed to understand how the AMPK pathway may provide a novel approach for the prevention of ozone-induced lung injury.

* Correspondence: neviere@univ-lille2.fr

† Contributed equally

¹Univ Lille Nord de France, F-59 000 Lille, France

Full list of author information is available at the end of the article

Background

Ambient ozone is a secondary pollutant formed through a complex series of photochemical reactions involving nitrogen oxides and sunlight. Ozone inhalation causes airway inflammation, direct epithelial injury and decreased lung function in both animal and human studies [1]. Ozone is highly reactive and oxidizes proteins and lipids in the fluid-lining compartment of the lung, which initiates inflammation processes and increased lung permeability. At least one mechanism underlying the lung toxicity of ozone involves excessive production by airway epithelial cells and activated lung immune cells of cytotoxic mediators including pro-inflammatory cytokines, reactive oxygen and nitrogen intermediates such as peroxynitrite [2,3].

Peroxyntirite is an extremely powerful and cytotoxic oxidant, which not only oxidizes lipids, nucleic acids, and protein SH-groups, but also nitrates aromatic amino acid protein residues. Via nitration of tyrosine residues, peroxynitrite alters the function of many proteins, such as superoxide dismutase, prostacyclin synthase PGI₂, Ca²⁺-ATPase, glutathione s-transferase, as well as transcriptional factors [4,5]. Observations that peroxynitrite induces nitration of tyrosine residues and also alters tyrosine phosphorylation have focused attention on phosphorylation cascades, as phosphorylation of mitogen-activated protein kinases modulate many cell signaling processes [4,6]. Moreover, it was recently proposed that peroxynitrite strongly increases phosphorylation of constitutive nitric oxide synthase via 5'-AMP-activated kinase activation, which results in enhanced production of reactive oxygen species contributing to a further oxidative damage [4].

5'-AMP-activated protein kinase (AMPK) is a serine threonine kinase that is highly conserved through evolution. As a cellular metabolic sensor, AMPK acts on a wide variety of substrates and cellular pathways, including the regulation of metabolic pathways (e.g., glycolysis, fatty acid synthesis and oxidation, cellular glucose uptake, and cholesterol synthesis), apoptosis and the cell cycle. It is becoming increasingly recognized that changes in AMPK activation are also implicated in several signaling pathways involved in nitric oxide production, sensing and responding to oxidative stress, and inflammation [7-9]. Downstream effects of AMPK activation may differ depending upon tissue stress intensity and metabolic needs. In peripheral organs such as the heart, activation of AMPK during ischemia reperfusion reduces damage [10]. In the brain, activation of AMPK during low-energy states such as ischemia is detrimental [11]. In the lung, AMPK activation following hypoxic challenge results in pulmonary vasoconstriction and transepithelial Na⁺ transport inhibition [12]. Hence, biological

consequences of stress-mediated AMPK activation are complex and remain highly variable.

Alterations in AMPK activation as a consequence of ozone exposure have not been previously evaluated. It is likely that production of peroxynitrite induced by ozone exposure may activate AMPK, which in turn could modulate transepithelial ion transport, oxidative stress and inflammation. Our objectives were twofold. First, we tested whether acute ozone exposure in mice would impair alveolar fluid clearance, increase lung tissue peroxynitrite production and activate AMPK. Second, we tested whether loss of AMP-activated protein kinase alpha1 subunit in mouse would prevent enhanced oxidative stress and lung injury induced by ozone exposure.

Methods

Animal used

Control and AMPK α 1 KO male mice (129/Sv and C57BL/6 mixed background) were studied at 20-24 wk of age. Animals were housed under controlled temperature and lighting with free access to water and standard mouse chow diet. All procedures were performed in accordance with the principles and guidelines established by the European Convention for the Protection of Laboratory Animals. All animal studies were approved by the Institutional Review Board of the National Institute (Agreement number 59-350146, Direction départementale de Protection des Populations DDPP, Nord, France).

Ozone exposure

Exposure to ozone [2 parts/million (ppm) for 3 h] or filtered room air in conscious mice was conducted in a stainless steel chamber with a Plexiglas door (~2.5 liters in volume). Ozone was generated by passing dry 100% oxygen through ultraviolet light and mixing it with filtered room air in the chamber. Chamber atmosphere was drawn continuously via a sampling port, and ozone concentration was measured by an ozone chemiluminescent analyzer (Model 49I; Thermo-environmental Instruments, Franklin, Massachusetts, USA). Chamber air temperature and relative humidity were maintained constant throughout experiments. Mice were then allowed to recover in ambient air for 24 h. Mice were then euthanized with an i.p. injection of a mixture of ketamine and xylazine (100 mg/kg and 20 mg/kg respectively).

Bronchoalveolar lavage (BAL)

Mice were sacrificed, and the trachea was cannulated with a 20-gauge catheter. BAL was performed twice with 0.8 ml of ice-cold PBS (pH 7.4) each. At least 1 mL of injected volume was recovered in all mice. The BAL fluid was spun at 800 g for 5 min at 4°C, and

supernatant was collected for the measurement of total proteins. Total proteins from cell free supernatant of the BAL fluids were assessed using the Bradford Assay (BioRad, CA). Pelleted cells were harvested, and red cells were lysed, then washed and resuspended in cold PBS. Total cells were enumerated by counting on a hemocytometer. For differential cell counting, cells were spun onto glass slides, air-dried, fixed in ethanol, and stained with Diff-Quick reagents (Baxter Scientific, Miami, FL). The number of macrophages, neutrophils and lymphocytes was counted based on morphology.

Myeloperoxidase activity, malondialdehyde level and peroxynitrite content in lung homogenates

Animals were euthanized and perfused with cold saline via the heart. The left lobes were removed and homogenized in 1 ml of PBS containing complete protease inhibitor cocktail (Sigma, St. Louis, MO). Then, the samples were centrifuged for 10 min at 800 g. Supernatants were filtered and kept in -70°C until used. Myeloperoxidase (MPO) activity, an indicator of polymorphonuclear leukocyte (PMN) accumulation, was determined as previously described [13]. Briefly, following lung homogenate preparation, myeloperoxidase activity was determined by measuring hydrogen peroxide-dependent oxidation of 3,3',5,5'-tetramethylbenzidine. The rate of change in absorbance was measured spectrophotometrically at 650 nm. Myeloperoxidase activity was defined as the quantity of enzyme degrading $1\ \mu\text{mol}$ of peroxide min^{-1} at 37°C and was expressed in units per gram weight of wet tissue. Malondialdehyde (MDA) level was utilized to quantify the lipid peroxidation in tissues as previously described [14]. Briefly, following lung homogenate preparation, MDA content was based on the reaction of MDA with thiobarbituric acid at $90\text{-}100^{\circ}\text{C}$. Samples were then heated for 1 h at 95°C and centrifuged at 3000 g for 10 min. The absorbance of the supernatant was measured by spectrophotometry at 532 nm using 1,1,3,3-tetramethoxypropane as an external standard. MDA was expressed in micromole per g weight of wet tissue. The formation of peroxynitrite was estimated by means of peroxynitrite-dependent oxidation of dihydrorhodamine-123 (DHR123, Molecular Probes, Eugene, Oregon) to rhodamine as previously described. This method is an indirect measurement of peroxynitrite production since other oxidant species can oxidize dihydrorhodamine-123 [15]. Briefly, duplicate aliquots of lung homogenate supernatants incubated with $5\ \mu\text{M}$ DHR123 were taken for rhodamine fluorescence evaluation using a fluorometer at an excitation wavelength of 500 nm and an emission wavelength of 530 nm. A fluorescence standard curve was obtained with authentic rhodamine and results were expressed in nmol/L.

Multiplex for detection of cytokines

Lung homogenate supernatants were assayed for a panel of cytokines and chemokines (IFN- γ , IL-1 α , IL-1 β , TNF- α , IL-6, IL-10, IL-2, IL-4, IL-5, IL-7, IL-9, IL-10, IL-12p40/p70, IL-13, IL-15, IL-17, IP-10, MCP-2, G-CSF, MIP-1 α , GM-CSF, and RANTES) using a multiplex-assay (xMAP Luminex Bioplex 200, Luminex Corp, Biorad, France) according to the manufacturer's instructions. All samples were assayed in duplicate and results were calculated as pg/mL.

Alveolar fluid clearance

After sacrifice, the trachea was cannulated by using a 20-gauge catheter. The lungs were then inflated with 7 cm H₂O continuous airway pressure with 100% oxygen throughout the experiment. An infrared lamp placed above the body maintained body temperature at 37°C . The lungs were instilled with Ringer lactate containing 5% bovine serum albumin and fluorescein isothiocyanate-tagged albumin as an alveolar protein tracer. Osmolarity of the instillate was adjusted to 340 mosM, which we have shown to be isosmolar with mouse plasma. Alveolar fluid clearance (AFC) was measured over 15 min. As in previous studies, AFC was determined by measuring the increase in the final fluorescence intensity of the alveolar protein tracer compared with the initial instilled tracer protein fluorescence intensity [16,17]. AFC was calculated as $\text{AFC} = 1 - (C1/C2)$, where C1 is the fluorescence intensity of the initial instilled tracer protein and C2 is the final total tracer protein fluorescence intensity in the alveolar fluid.

Basolateral cell membrane isolation

Pieces of peripheral lung tissue were collected from each lobe and homogenized to obtain whole cell lysates (each piece approximately represented a cube of 2 millimeters side). Cells were prepared by addition of lysis buffer and centrifugation at 14,000 g to eliminate the insoluble material. Basolateral membranes were prepared using ultracentrifugation as previously described [18]. Peripheral lung tissue was homogenized and centrifuged twice to discard the nuclear and mitochondrial pellet. Supernatant was centrifuged at 48,000 g for 30 min, and the basolateral membrane fraction was recovered after the membrane pellet was centrifuged in a 16% Percoll gradient at 48,000 g for 30 min.

Western blot analysis

Proteins from cell lysates and basolateral membrane fractions were resolved by 10% SDS-PAGE and analyzed by immunoblotting. Mouse monoclonal antibodies anti total AMPK- α (Cell Signaling, France) and rabbit monoclonal antibodies anti-phospho-AMPK- α at Thr172 (Cell Signaling, France) were used for cell

lysate immunoblotting. Mouse monoclonal antibody anti-Na,K-ATPase α_1 (Upstate Biotechnology, Lake Placid, NY) and rabbit polyclonal antibody anti-GLUT1 (Millipore, France) were used for basolateral membrane fractions immunoblotting, respectively. AMPK activation was assessed as the ratio of phospho-AMPK to total AMPK. GLUT1 was measured by immunoblotting as a loading control for plasma membrane protein. The amount of Na,K-ATPase at the plasma membrane was normalized to GLUT1.

Statistical analysis

All statistical analyses were performed using GraphPad Prism 5.0 (GraphPad Software Inc., San Diego, CA). When a significant difference was found, we examined between-group differences using a sequentially rejective Bonferroni procedure. After applying Bonferroni's correction, the level of statistical significance was set at $p < 0.05$ for comparisons with control. Experimental results were expressed as mean \pm S.D.

Results

Ozone exposure induces lung inflammation

In the first part of our study, we observed the impact of acute ozone exposure on lung inflammation. We therefore evaluated cell recruitment and total protein level in the BAL. We found a significant increase in proportion of neutrophils in the ozone-exposed mice BAL (Figure 1). The recruitment of neutrophils observed in mice BAL has been confirmed in lung homogenates by measurement of MPO activity which was significantly higher in ozone-exposed mice than in control mice ($p = 0.037$) (Table 1). The total protein was also higher in ozone-exposed mice BAL than in control mice. We also showed a significant increase in major pro-inflammatory cytokine levels such as IL-6 and IL-1 β in ozone-exposed mice lung homogenates (Figure 1).

Ozone exposure increases lung oxidative stress

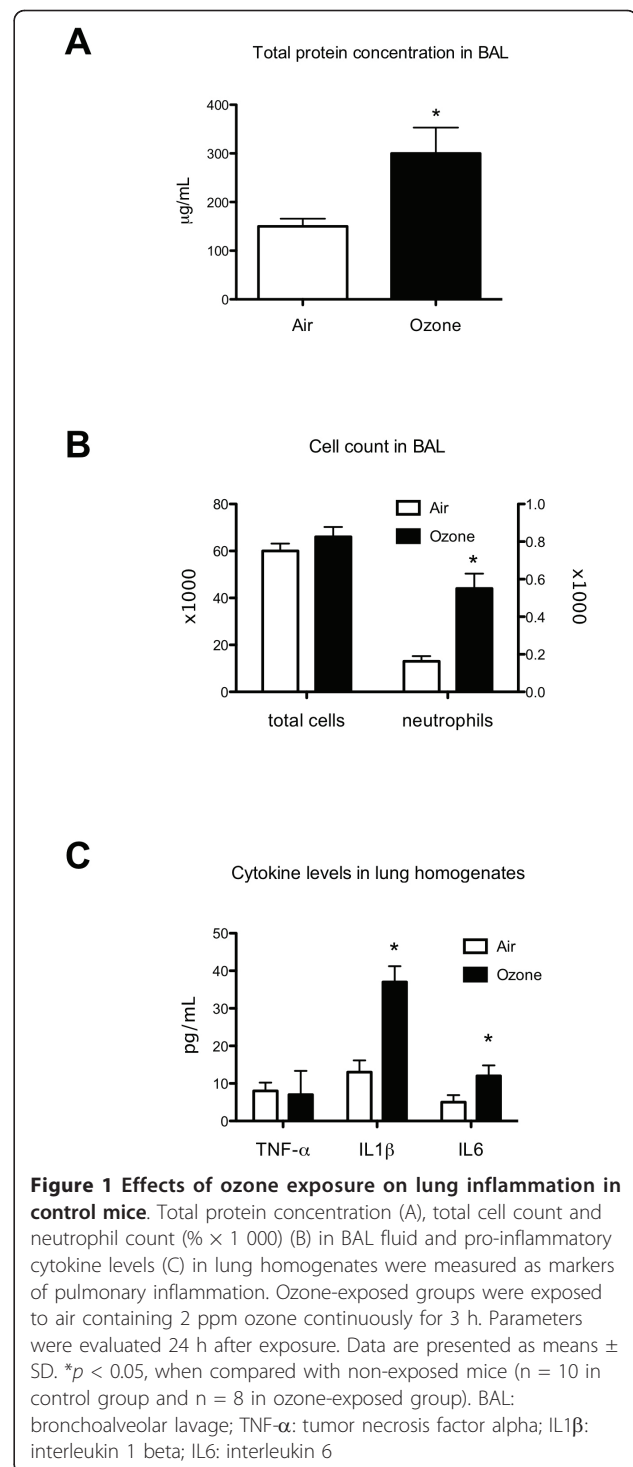
There were significant increases in oxidation of DHR123 ($p = 0.02$) and MDA concentration ($p < 0.0001$) in ozone-exposed mice lung homogenates (Table 1).

Ozone exposure increases AMPK phosphorylation

As we observed a peroxynitrite increase in ozone-exposed mice lung homogenates, we next evaluated AMPK phosphorylation. There was a strong increase of phospho-AMPK to total AMPK ratio in ozone-exposed mice lung homogenates compared with the unexposed group ($p = 0.04$) (Figure 2).

Ozone exposure increases alveolar fluid clearance

As we observed an increase in the total protein in BAL of mice exposed to ozone, which indicated an increase



in crude epithelial permeability, we explored the effects of ozone on the alveolar fluid clearance increase. So, we hypothesized that activation of Na,K-ATPase contributes to the increase in clearance of alveolar fluid. There was a significant increase of alveolar fluid clearance in ozone-exposed control mice ($p = 0.05$). We then wanted

Table 1 MDA levels, MPO activity and peroxynitrite levels in lung homogenates WT or AMPK α 1 deficient mice

	control mice		AMPK α 1 deficient mice		
	Air	Ozone	Air	Ozone	
MDA (μ mol/g wet tissue)	110 +/- 14.1*	230 +/- 10*	160 +/- 13	147 +/- 12.6	*p < 0.0001
MPO (U/g wet tissue)	0.13 +/- 0.03*	0.27 +/- 0.04*	0.13 +/- 0.02	0.16 +/- 0.06	*p = 0.04
Peroxynitrite (nmol/L)	3.0 +/- 0.2*	4.4 +/- 1.0*	2.8 +/- 0.5	2.8 +/- 0.4	*p = 0.03

Mice were exposed to air with or without ozone at 2 ppm for 3 h. Parameters were evaluated 24 h after exposure. Malondialdehyde (MDA) was expressed as micromole per g weight of wet tissue; Myeloperoxidase activity (MPO) was expressed in units per gram weight of wet tissue. Peroxynitrite was expressed as nmol/L. Data are presented as means \pm SD. *: p < 0.05; μ mol/g wet tissue: micromole per g weight of wet tissue; U/g wet tissue: units per gram weight of wet tissue

to know if the Na,K-ATPase was partly responsible for the increased alveolar fluid clearance. Basolateral membrane Na,K-ATPase abundance normalized to GLUT1- was twofold higher in ozone-exposed control mice (Figure 3).

AMPK α 1 deficient mice are protected from ozone lung injury

There were no changes in MPO activity, MDA concentration and peroxynitrite levels in AMPK α 1 deficient mice after ozone exposure (Table 1). Alveolar fluid clearance did not change significantly in AMPK α 1 deficient mice after ozone exposure (Figure 3). The

basolateral membrane Na,K-ATPase abundance was not different between the exposed and unexposed group in AMPK α 1 deficient mice.

Discussion

Because AMPK has been shown to modulate airway transepithelial ion transport, inflammation and oxidative stress, we tested the hypothesis that AMPK deficiency would prevent ozone-induced lung injury in mice. Important findings of this study revealed that acute ozone exposure increase lung fluid clearance, leukocyte infiltration and oxidative stress and that ozone-induced lung injury was largely prevented in AMPK alpha1 subunit deficient mice.

Acute inhalation of ozone was found to cause structural alterations in the lung including disruption of the alveolar epithelial barrier, followed by alveolar epithelial type II cell hypertrophy and hyperplasia. Recruitment of inflammatory cells into the lung following O₃ exposure presents another risk of tissue damage through the release of toxic mediators by activated macrophages and neutrophils inflammatory cells, including cytokines, reactive oxygen and nitrogen species and proteolytic enzymes [1]. Following ozone inhalation, TNF- α levels rapidly peaked, predominantly in alveolar macrophages and has been implicated in lung injury by stimulating the production of cytotoxic inflammatory mediators such as cytokines and chemokines. In rodents, many of the pulmonary manifestations of acute O₃ exposure have been attributed to the effects of these cytokines and chemokines, as interruption of IL-1, IL-6, signaling significantly diminishes O₃-induced neutrophil emigration to the airspaces and/or epithelial cell injury. Twenty-four hours after exposure early TNF- α peak concentration was not detected, whereas concentrations of IL-1 and IL-6 increased in BAL of ozone-exposed mice. This is consistent with previous studies indicating that cytokine secretion profiles are related to compartmentalization and migration of different inflammatory cell-types during the development of ozone injury [1-3]. In addition, characteristics of recruitment of inflammatory cells in mice lung following ozone exposure have been shown varying with ozone concentration, exposure

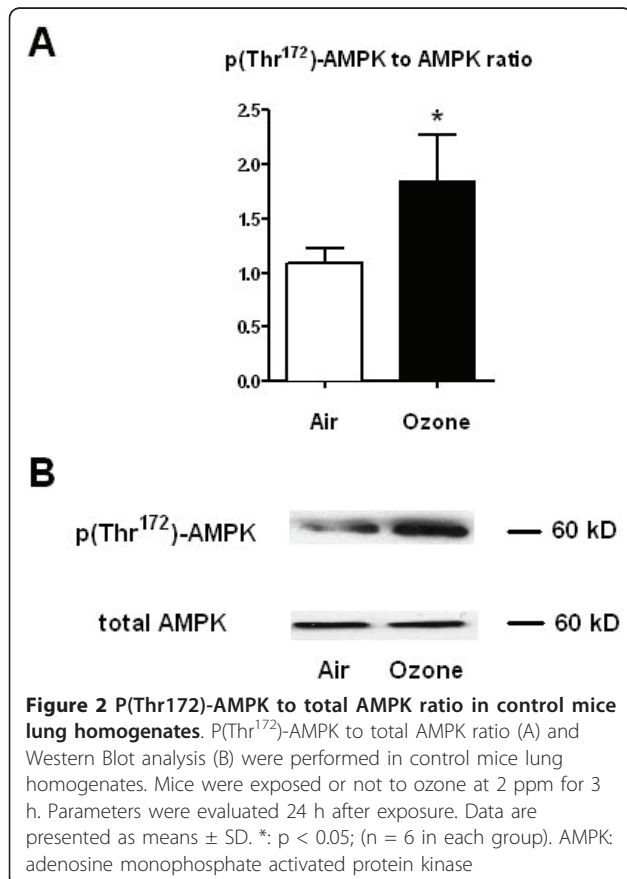
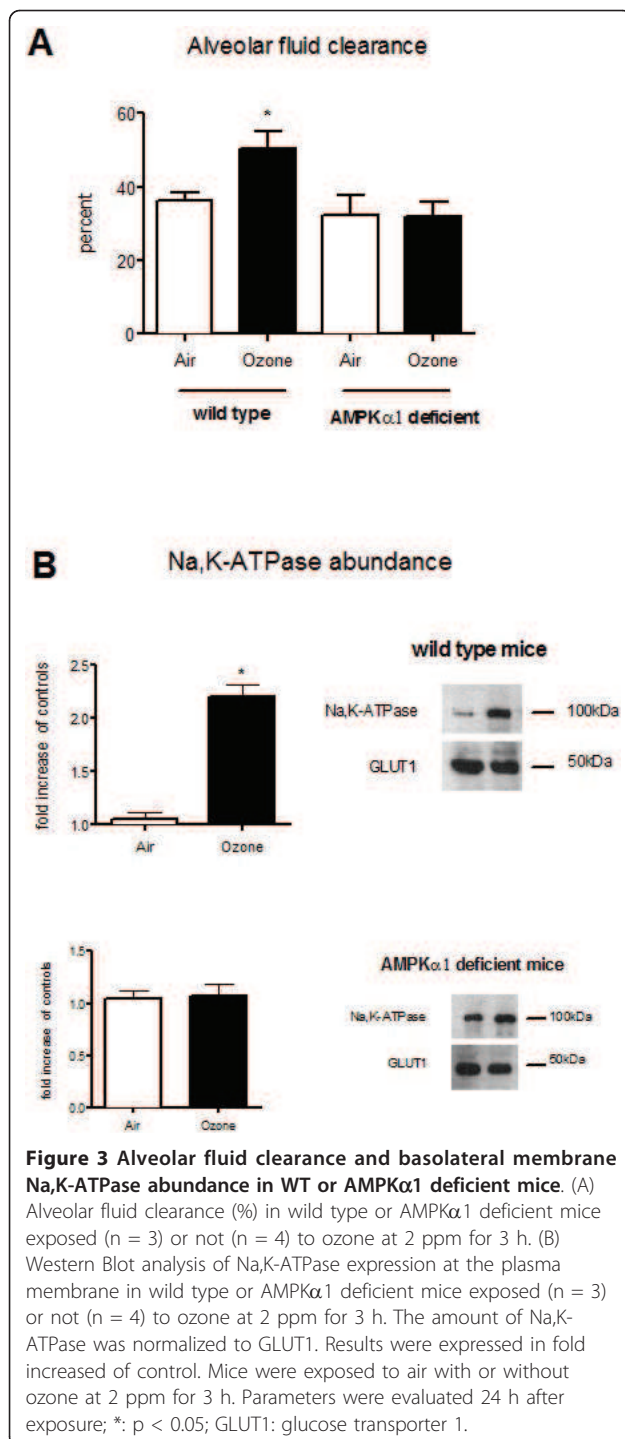


Figure 2 P(Thr172)-AMPK to total AMPK ratio in control mice lung homogenates. P(Thr¹⁷²)-AMPK to total AMPK ratio (A) and Western Blot analysis (B) were performed in control mice lung homogenates. Mice were exposed or not to ozone at 2 ppm for 3 h. Parameters were evaluated 24 h after exposure. Data are presented as means \pm SD. *: p < 0.05; (n = 6 in each group). AMPK: adenosine monophosphate activated protein kinase



duration, age and strains [1]. In these regards, C57bl6/129sv mice used in our model are strong responders for the lung response to O₃ [19] and seem to be little affected by age in term of O₃ response [20]. Consistently, our model of acute ozone exposure (3 h, 2 ppm) elicited alveolar neutrophil recruitment and increases in biomarkers of lung tissue inflammation and oxidative

stress. Expression of concentrations of biomarkers per gram of wet lung in this study would have caused us to underestimate increases in MDA, MPO and peroxynitrite levels, as O₃ exposure likely induced edema formation leading to increases in lung wet weight, which we used as a normalization parameter.

Fluid and solute resorption from the alveolus is critical in clearing fluid from lungs [21]. Active alveolar epithelial Na⁺ ion transport is well established as the primary mechanism of fluid clearance from distal airspaces [22]. In the current model of fluid balance in the distal lung, Na⁺ ions enter alveolar type II epithelial cells at the apical surface primarily through amiloride-sensitive sodium channels and are pumped out on the basolateral surface by Na,K-ATPase. In conditions of lung inflammation, such as ozone exposure, active Na⁺ transport and alveolar fluid clearance may be stimulated leading to increased ability of the lungs to clear edema. Indeed, a large variety of endogenous mediators, which are released by the lung in response to ozone exposure, may be responsible for the stimulation of alveolar liquid clearance. For example, increased TNF-alpha production has been consistently shown to stimulate alveolar fluid transport [23]. Likewise, authentic nitric oxide and nitrogen intermediates have been shown to modulate active Na⁺ transport in alveolar type 2 epithelial cells, and both stimulation [24] and inhibition [25] of transport characteristics have been reported. Hence, it may be hypothesized that alveolar fluid clearance increase observed in our ozone exposure model may be the result of ozone-induced proinflammatory cytokine and peroxynitrite increases.

Concomitant changes in permeability, oxidative stress and recruitment of inflammatory cells in the lung following ozone exposure have been shown varying with ozone concentration, exposure duration, and animal species [1]. The exact mechanism influencing the severity of ozone-induced pulmonary reaction and molecules involved in the modulation of this response are yet to be fully determined. Proposed mechanisms include the regulation of the oxidant anti-oxidant balance through functionally relevant post-translational protein modification. In this context, peroxynitrite can play an important role in this balance through the nitration of tyrosine residues that alters the function of many proteins, such as superoxide dismutase and NADPH oxidase [6]. In addition, via increased c-Src, phosphatidylinositol 3-kinase or protein kinase C activities, peroxynitrite may also activate AMPK, which has been shown to alter cell response to oxidative stress [26-28]. In our study, we confirmed that increased lung peroxynitrite production was associated with AMPK phosphorylation in ozone-exposed mice. Importantly, AMPK alpha1 subunit deficient mice were protected against ozone-induced lung injury, suggesting that AMPK activation was detrimental.

AMPK is a heterotrimeric Ser/Thr kinase composed of a catalytic α subunit and regulatory β and γ subunits. The $\alpha 1$ and $\alpha 2$ subunit isoforms have ~90% homology in their N-terminal catalytic domains and ~60% homology in their C-terminal domains, suggesting that they may have distinct downstream targets. For example, AMPK $\alpha 2$ deletion results in mild insulin-resistance and impaired glucose tolerance, whereas deletion of AMPK $\alpha 1$ has no detectable effects on metabolic phenotype. In addition and beyond the regulation of energy homeostasis, AMPK also exerts non-metabolic functions, such as maintenance of cell polarity and normal cell division, or control of cell growth and survival [29]. In lung cells, the $\alpha 1$ catalytic subunit of AMPK is predominantly expressed and its pharmacological activation leads to inhibition of Na⁺ transport processes across lung epithelium [30]. It has been previously reported that AMPK activation inhibits Na⁺ transport processes across lung epithelium via PKC ζ activation and Na,K-ATPase endocytosis, whereas inhibition of AMPK by compound C enhances Na⁺ transport [31]. In our model, lung AMPK phosphorylation observed in control mice exposed to ozone was not associated with expected inhibition of Na⁺ transport processes across lung epithelium and Na,K-ATPase endocytosis related to AMPK activation. Instead, ozone exposure induced increases in alveolar fluid clearance and the presence of Na,K-ATPase at the plasma membrane. Yet, increases in alveolar fluid clearance and plasma membrane Na,K-ATPase abundance were not observed in AMPK $\alpha 1$ deficient mice exposed to ozone, suggesting that AMPK activation modulates mechanisms involved in Na⁺ transport processes across lung epithelium independent of Na,K-ATPase activity.

Hence, we next reasoned that protection against ozone-induced lung injury in AMPK $\alpha 1$ deficient mice was related to the reduction of oxidative stress and leukocyte lung infiltration. Under conditions of cell stress, the actions of AMPK are mainly dependent on cell type and metabolism [32]. AMPK activation is beneficial in peripheral tissues, such as the ischemic heart [10]. However, AMPK activation may exacerbate brain oxidative stress and injury whereas its pharmacological inhibition provided neuroprotection in the ischemic and reperfused brain [33,34]. It has been proposed that in cells lacking key glycolytic enzymes and/or with limited ability to store nutrients such as neurons, AMPK activation leads to acidosis, enhanced oxidative stress and cell apoptosis [35]. It is, hence, our contention that in lung epithelial cells, which have low nutrient storage capacity, AMPK activation may be detrimental and that defect in AMPK $\alpha 1$ may be protective by limiting ozone-induced lung oxidative stress. Another common feature of the host response to ozone is neutrophilic

inflammation and up-regulation of proinflammatory cytokines. Recent evidence suggests that AMPK participates in the modulation of the inflammatory response. For example, it has been reported that pharmacological activation of AMPK has anti-inflammatory effects [36], whereas AMPK $\alpha 1$ deficient mice display normal immune responses. As neutrophilic recruitment is mainly redox-sensitive in the acute ozone-exposed lung, we speculated that reduced lung leukocyte sequestration was related to the decreased oxidative stress in AMPK $\alpha 1$ deficient mice.

Conclusion

Our results collectively suggest that activation of AMPK is detrimental in ozone-induced lung injury. The use of AMPK $\alpha 1$ knockout mice clearly shown that AMPK activation participates in ozone-induced increases in lung fluid clearance, leukocyte infiltration and oxidative stress. Further studies are needed to delimit the role of AMPK pharmacological inhibition and to demonstrate that manipulation of the AMPK pathway may provide a novel approach for the prevention of ozone-induced lung injury.

Abbreviations

AFC: Alveolar fluid clearance; AMPK: 5'-AMP-activated protein kinase; BAL: bronchoalveolar lavage; DHR123: dihydrorhodamine-123; G-CSF: granulocyte colony-stimulating factor; GLUT: glucose transporter; GM-CSF: granulocyte-macrophage colony stimulating factor; IFN: interferon; IL: interleukin; MCP: monocyte chemotactic protein; MDA: malondialdehyde; MIP: macrophage inflammatory protein; MPO: myeloperoxidase; NADPH oxidase: nicotinamide adenine dinucleotide phosphate oxidase; PKC: protein kinase C; PMN: polymorphonuclear leukocyte; ppm: parts/million; RANTES: Regulated on Activation Normal T Cell Expressed and Secreted; SDS-PAGE: Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis; TNF: tumor necrosis factor

Acknowledgements

This work was supported by the "Agence Nationale de Sécurité Sanitaire de l'alimentation, de l'environnement et du travail", BICEL-campus Lille 2 IMPRT facility for access to instruments and technical services and University of Lille 2 (EA 4483 and 4484). We thank B. Besson-Lescure and N. Brunel in IFR65 for Bioplex analysis, J. Franczak and S. Hassoun for their help in experiments.

Author details

¹Univ Lille Nord de France, F-59 000 Lille, France. ²UDSL, EA 4483, F-59 000 Lille, France. ³INSERM U 1019, F-59 000 Lille, France. ⁴UDSL, EA 4484, F-59 000 Lille, France. ⁵INSERM, U1016, Institut Cochin, Paris, France. ⁶Cnrs, UMR8104, Paris, France. ⁷Univ Paris Descartes, Paris, France.

Authors' contributions

All authors have read and approved the final manuscript. SH was responsible for carrying out the experiments, for data analysis, and for drafted this manuscript; HT was responsible for the alveolar fluid clearance study; SL and RN oversaw the animal experiments, instructed SH in his implementation; RN, SL, JLE and AS assisted in the experimental design and the data analysis and interpretation. BV is expert in AMPK $\alpha 1$ knockout mice and has provided AMPK $\alpha 1$ knockout mice. All authors contributed to the drafting and revisions of the manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 13 January 2011 Accepted: 19 May 2011
Published: 19 May 2011

References

1. Uysal N, Schapira RM: Effects of ozone on lung function and lung diseases. *Curr Opin Pulm Med* 2003, **9**:144-150.
2. Laskin DL, Fakhrzadeh L, Laskin JD: Nitric oxide and peroxyntirite in ozone-induced lung injury. *Adv Exp Med Biol* 2001, **500**:183-190.
3. Roberts RA, Laskin DL, Smith CV, Robertson FM, Allen EMG, Doorn JA, Slikker W: Nitrate and oxidative stress in toxicology and disease. *Toxicol Sci* 2009, **112**:4-16.
4. Liaudet L, Vassalli G, Pacher P: Role of peroxyntirite in the redox regulation of cell signal transduction pathways. *Front Biosci* 2009, **14**:4809-4814.
5. Pacher P, Beckman JS, Liaudet L: Nitric oxide and peroxyntirite in health and disease. *Physiol Rev* 2007, **87**:315-424.
6. Peluffo G, Radi R: Biochemistry of protein tyrosine nitration in cardiovascular pathology. *Cardiovasc Res* 2007, **75**:291-302.
7. Hardie DG: AMP-activated/SNF1 protein kinases: conserved guardians of cellular energy. *Nat Rev Mol Cell Biol* 2007, **8**:774-785.
8. Hardie DG, Hawley SA, Scott JW: AMP-activated protein kinase—development of the energy sensor concept. *J Physiol (Lond)* 2006, **574**:7-15.
9. Kim AS, Miller EJ, Young LH: AMP-activated protein kinase: a core signalling pathway in the heart. *Acta Physiol (Oxf)* 2009, **196**:37-53.
10. Mark Evans A, Ward JPT: Hypoxic pulmonary vasoconstriction—invited article. *Adv Exp Med Biol* 2009, **648**:351-360.
11. Ramamurthy S, Ronnett GV: Developing a head for energy sensing: AMP-activated protein kinase as a multifunctional metabolic sensor in the brain. *J Physiol (Lond)* 2006, **574**:85-93.
12. Gusarova GA, Dada LA, Kelly AM, Brodie C, Witters LA, Chandel NS, Sznajder JI: Alpha1-AMP-activated protein kinase regulates hypoxia-induced Na,K-ATPase endocytosis via direct phosphorylation of protein kinase C zeta. *Mol Cell Biol* 2009, **29**:3455-3464.
13. Lancel S, Tissier S, Mordon S, Marechal X, Depontieu F, Scherpereel A, Chopin C, Neviere R: Peroxyntirite decomposition catalysts prevent myocardial dysfunction and inflammation in endotoxemic rats. *J Am Coll Cardiol* 2004, **43**:2348-2358.
14. Ohkawa H, Ohishi N, Yagi K: Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem* 1979, **95**:351-358.
15. Kooy NW, Royall JA, Ischiropoulos H, Beckman JS: Peroxyntirite-mediated oxidation of dihydrorhodamine 123. *Free Radic Biol Med* 1994, **16**:149-156.
16. Planès C, Leyvraz C, Uchida T, Angelova MA, Vuagniaux G, Hummler E, Matthay M, Clerici C, Rossier B: In vitro and in vivo regulation of transepithelial lung alveolar sodium transport by serine proteases. *Am J Physiol Lung Cell Mol Physiol* 2005, **288**:L1099-1109.
17. Fukuda N, Folkesson HG, Matthay MA: Relationship of interstitial fluid volume to alveolar fluid clearance in mice: ventilated vs. in situ studies. *J Appl Physiol* 2000, **89**:672-679.
18. Litvan J, Briva A, Wilson MS, Budinger GRS, Sznajder JI, Ridge KM: Beta-adrenergic receptor stimulation and adenoviral overexpression of superoxide dismutase prevent the hypoxia-mediated decrease in Na,K-ATPase and alveolar fluid reabsorption. *J Biol Chem* 2006, **281**:19892-19898.
19. Savov JD, Whitehead GS, Wang J, Liao G, Usuka J, Peltz G, Foster WM, Schwartz DA: Ozone-induced acute pulmonary injury in inbred mouse strains. *Am J Respir Cell Mol Biol* 2004, **31**:69-77.
20. Hamade AK, Misra V, Rabold R, Tankersley CG: Age-related changes in cardiac and respiratory adaptation to acute ozone and carbon black exposures: interstrain variation in mice. *Inhal Toxicol* 2010, **22**(Suppl 2):84-94.
21. Vadász I, Raviv S, Sznajder JI: Alveolar epithelium and Na,K-ATPase in acute lung injury. *Intensive Care Med* 2007, **33**:1243-1251.
22. Hummler E, Planès C: Importance of ENaC-mediated sodium transport in alveolar fluid clearance using genetically-engineered mice. *Cell Physiol Biochem* 2010, **25**:63-70.
23. Rezaiguia S, Garat C, Delclaux C, Meignan M, Fleury J, Legrand P, Matthay MA, Jayr C: Acute bacterial pneumonia in rats increases alveolar epithelial fluid clearance by a tumor necrosis factor-alpha-dependent mechanism. *J Clin Invest* 1997, **99**:325-335.
24. Hardiman KM, McNicholas-Bevensee CM, Fortenberry J, Myles CT, Malik B, Eaton DC, Matalon S: Regulation of amiloride-sensitive Na(+) transport by basal nitric oxide. *Am J Respir Cell Mol Biol* 2004, **30**:720-728.
25. Jain L, Chen XJ, Brown LA, Eaton DC: Nitric oxide inhibits lung sodium transport through a cGMP-mediated inhibition of epithelial cation channels. *Am J Physiol* 1998, **274**:L475-484.
26. Han Y, Wang Q, Song P, Zhu Y, Zou MH: Redox regulation of the AMP-activated protein kinase. *PLoS ONE* 2010, **5**:e15420.
27. Choi HC, Song P, Xie Z, Wu Y, Xu J, Zhang M, Dong Y, Wang S, Lau K, Zou MH: Reactive nitrogen species is required for the activation of the AMP-activated protein kinase by statin in vivo. *J Biol Chem* 2008, **283**:20186-20197.
28. Xie Z, Dong Y, Zhang M, Cui MZ, Cohen RA, Riek U, Neumann D, Schlattner U, Zou MH: Activation of protein kinase C zeta by peroxyntirite regulates LKB1-dependent AMP-activated protein kinase in cultured endothelial cells. *J Biol Chem* 2006, **281**:6366-6375.
29. Viollet B, Athes Y, Mounier R, Guigas B, Zarrinpashneh E, Horman S, Lantier L, Hebrard S, Devin-Leclerc J, Beauloye C, Foretz M, Andreelli F, Ventura-Clapier R, Bertrand L: AMPK: Lessons from transgenic and knockout animals. *Front Biosci* 2009, **14**:19-44.
30. Almajra J, Kongsuphol P, Hieke B, Ousingsawat J, Viollet B, Schreiber R, Amaral MD, Kunzelmann K: AMPK controls epithelial Na(+) channels through Nedd4-2 and causes an epithelial phenotype when mutated. *Pflügers Arch* 2009, **458**:713-721.
31. Hallows KR: Emerging role of AMP-activated protein kinase in coupling membrane transport to cellular metabolism. *Curr Opin Nephrol Hypertens* 2005, **14**:464-471.
32. Steinberg GR, Kemp BE: AMPK in Health and Disease. *Physiol Rev* 2009, **89**:1025-1078.
33. Li J, Zeng Z, Viollet B, Ronnett GV, McCullough LD: Neuroprotective effects of adenosine monophosphate-activated protein kinase inhibition and gene deletion in stroke. *Stroke* 2007, **38**:2992-2999.
34. McCullough LD, Zeng Z, Li H, Landree LE, McFadden J, Ronnett GV: Pharmacological inhibition of AMP-activated protein kinase provides neuroprotection in stroke. *J Biol Chem* 2005, **280**:20493-20502.
35. Li J, McCullough LD: Effects of AMP-activated protein kinase in cerebral ischemia. *J Cereb Blood Flow Metab* 2010, **30**:480-492.
36. Zhao X, Zmijewski JW, Lorne E, Liu G, Park YJ, Tsuruta Y, Abraham E: Activation of AMPK attenuates neutrophil proinflammatory activity and decreases the severity of acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 2008, **295**:L497-504.

doi:10.1186/1465-9921-12-64

Cite this article as: Hulo et al.: AMP-activated protein kinase deficiency reduces ozone-induced lung injury and oxidative stress in mice. *Respiratory Research* 2011 **12**:64.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

