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**Fitness and infectivity of drug-resistant and cross-resistant HBV mutants: why and how is it studied?**

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**Summary**

The emergence of HBV drug-resistant (and multidrug-resistant) strains during long-term therapy with nucleos(t)ides analogs is associated with treatment failure and represents therefore a clinical challenge. For clinicians, the close monitoring and management of resistance has become a key issue in clinical practice. For HBV virologists, the understanding of the mechanism of emergence of specific mutant strains in the viral quasispecies during treatment is also an important issue. If a particular viral strain can emerge in the quasispecies within a particular environment, it is likely because its fitness is superior to other strains. The present review focuses on viral fitness as well as viral infectivity, and in particular on technical means that are available to study this viral fitness *in vitro* and in animal models.

## 1 **Chronic hepatitis B: clinical issues and current treatment**

2 Infection by hepatitis B virus (HBV) can be resolved after an acute episode, or leads to persistence  
3 and chronic hepatitis B (CHB). Chronic hepatitis B virus infection is a serious clinical problem and  
4 a major cause of severe liver-related morbidity and premature mortality. Indeed, patients with CHB  
5 have an increased risk of developing decompensated liver disease, cirrhosis and hepatocellular  
6 carcinoma (HCC) [1]. The primary treatment goal is the suppression of HBV replication, ideally  
7 followed by a seroconversion (anti-HBe, and then anti-HBs), and the prevention of active disease in  
8 the long-term [2, 3]. In this respect, the treatment of CHB with either interferons or nucleos(t)ides  
9 analogs (NAs), including lamivudine (LMV), adefovir dipivoxil (ADV), entecavir (ETV),  
10 telbivudine (LdT) and tenofovir (TDF), has resulted in a significant reduction in patient morbidity  
11 and mortality. Yet the efficacy of treatments for CHB can be affected by a number of factors,  
12 including the development of adverse side effects, poor patient compliance, previous treatment with  
13 suboptimal regimes and/or inadequate drug exposure, individual genetic variation, or infection with  
14 drug-resistant virus. As therapy with interferons (“naked” or pegylated) alone remains quite  
15 inefficient, the clinical use of nucleos(t)ide analogs has played a major part in the substantial  
16 advances in CHB treatment that have occurred over the past decade. CHB requires long-term  
17 therapy, and resistance to therapy is a frequent consequence of treatment duration. The emergence  
18 of drug resistance during long-term therapy with NAs is almost inevitable, due to the high  
19 adaptability of viruses and the quasispecies nature of HBV, and represents a clinical challenge [2, 3].  
20 For clinicians, the close monitoring and management of resistance has become a key issue in  
21 clinical practice. For HBV virologists, the understanding of the mechanism of emergence of specific  
22 mutant strains in the viral quasispecies during treatment is also an important issue. If a particular  
23 viral strain can emerge in the quasispecies within a particular environment, it is likely because its  
24 fitness has become superior to other strains. The present review will focus on viral fitness as well as  
25 infectivity, and in particular on technical means that are available to study this viral fitness *in vitro*  
26 or in animal models.

## 27 28 **Treatment failure and HBV resistance**

29 HBV is a DNA virus that replicates its genome via an RNA intermediate, the pregenomic RNA  
30 (pgRNA), that comes from the transcription of cccDNA, i.e. covalently-closed-circular-DNA, the  
31 nuclear form of HBV genome and main template for viral transcription. The pgRNA is reverse-  
32 transcribed by covalently-linked-HBV-polymerase after incorporation in the nucleocapsid [4]. This  
33 step of the viral life cycle is currently the target of NA-based therapy. Long-term therapies with  
34 NAs, which are theoretically necessary to get a chance to clear cccDNA from cells, are confronted  
35 with the emergence of drugs resistant strains in the viral quasispecies. HBV mutants are  
36 spontaneously produced by the low fidelity HBV polymerase, and a drug pressure may select for  
37 viral species that exhibit the best replication capacity in this new treatment environment. Mutations  
38 conferring resistance to NAs are located in the viral polymerase gene. The rapidity of selection of  
39 drug resistant mutants depends on their replication capacity and fitness, their level of resistance, and  
40 free liver space available for infection by these mutants [5, 6]. This may explain, at least in part, the

1 differences in the rate of resistance for the different drugs that are clinically available.  
2 Different mechanisms are involved in drug-resistance under antiviral therapy [3]. First, a complex  
3 mixture of genetically distinct variants develops under selective pressure. A pre-existing or newly  
4 acquired mutation conferring a selective advantage to a variant will give rise to virions, which are  
5 fitter and can spread more rapidly in the liver. This mutant may accumulate and become the  
6 dominant (or at least a well represented) species in the infected liver, under the pressure of the  
7 antiviral drug. The kinetics of replacement of wild type virus in liver cells by a dominant mutant are  
8 generally slow. As resistant mutants mainly infect uninfected cells, the efficient spreading of the  
9 dominant mutant depends on its intrinsic fitness and the availability of free liver space for its  
10 propagation and replication [5, 6]. During antiviral therapy, several months may be needed for the  
11 immune system to clear hepatocytes infected with wild type virus and to generate new cells that are  
12 susceptible to infection by viral drug-resistant mutants. On the other hand, the specific infectivity of  
13 drug resistant mutants may have a major impact on the rapidity of selection of these strains during  
14 therapy. Indeed, some mutations in the viral polymerase gene may result in nucleotide changes in  
15 overlapping surface genes, which in turn may lead to reduced viral fitness, due to impaired  
16 assembly, secretion or infectivity [7, 8]. The level of resistance to a drug conferred by a given  
17 mutation may have profound implication on the fitness of the mutant. This may explain the  
18 difference in drug resistance rates observed with the different antivirals.

19

## 20 **Definition of viral fitness and rationale for studying it**

21 In a simple way, viral fitness can be defined as the sum of parameters that quantify the adaptation of  
22 a viral strain to a given environment. *In vitro*, it can be seen as the growth properties of a given  
23 strain as compared to a reference or wild type viral strain in a defined environment (i.e., in tissue  
24 culture or *in vivo* in the presence of drugs or an intact immune system). The fitness of a given viral  
25 strain is of great importance to understand its emergence or elimination in a particular  
26 microenvironment. Of course in the case of *in vivo* infection other features including the genetic  
27 makeup of host and immune system are also important to explain the emergence of a particular  
28 strain, but this will not be further developed here as this review focuses on virus related parameters.  
29 The therapeutic pressure represents also an important environmental parameter to explain the  
30 emergence of mutant strains in patients. In general, NA-resistant strains do not replicate as well as  
31 wild type counterparts, because mutations in the polymerase gene tend to affect the enzymatic  
32 activity of the protein. But this pattern of replication can be reversed in the presence of the drug(s)  
33 that selected the mutant in the first place [7, 9, 10].

34 The virologic parameters of HBV fitness are the capacity of a strain to synthesize its genome, to  
35 produce infectious particles and (re)-infect cells (i.e. enter and deliver the genome to nucleus). The  
36 synthesis of the genome itself includes the ability of the strain to produce pgRNA (i.e. transcription  
37 via precore promoter, some mutations may alter the level of transcription mediated by host RNA  
38 polymerase II) that is subsequently packaged into the nucleocapsid before being used for reverse  
39 transcription. The study of viral fitness is a complex matter as it relies on the models used to  
40 perform analysis. Cell culture systems or animal models capable of full replication and propagation

1 of the virus are necessary to properly measure the fitness of a viral strain. For HBV such models do  
2 no exist. Therefore to get insight into HBV fitness, several more or less artificial assays that will be  
3 hereafter named replication, assembly and infectivity assays have been developed.

#### 4 5 **Studying viral fitness *in vitro*: critical review of assays**

6 **Replication assays.** Several *in vitro* assays have been developed to determine the replication  
7 capacity of clinically relevant HBV strains, including wild-type and drug-resistant strains. These  
8 assays are also critical to determine the role of a given mutation profile in drug resistance, as well as  
9 to determine the cross-resistance profile of those mutants. Several approaches and assays, also  
10 termed phenotypic assays as they describe the phenotype (i.e. replication capacity + drug resistance  
11 profile) of a given strain *in vitro*, have been described in literature. These assays are mainly based  
12 on the transfection of full-length HBV PCR-amplicons (i.e. vector-free) or plasmids containing 1.1  
13 to 2 HBV genome units into hepatoma cell lines (e.g. Huh7 or HepG2).

14 The vector-free method relies on an original and efficient PCR amplification of full-length HBV  
15 genomes isolated from patients [11]. The linear amplicon is transfected into cells and serves as a  
16 ccc-like DNA template for the initiation of the intracellular HBV replication after re-circularization  
17 by host machinery. This step of circularization is rate limiting and explains the rather low absolute  
18 level of replication obtained with this approach; however the relative replication capacity of two  
19 different strains can be analyzed by this approach. Another advantage of this approach comes from  
20 the fact that the expression of pgRNA is driven by the HBV promoter itself, which is interesting if  
21 precore or core promoter mutants are analyzed.

22 Alternative approaches are based on vectors, that can be plasmid or alternatively recombinant  
23 adenovirus or baculovirus, carrying 1.1 to 3 HBV genome units [12-15]. These vectors contain the  
24 HBV genetic information necessary and sufficient to initiate an HBV replication cycle after  
25 transfection or transduction into cells. The synthesis of HBV pgRNA can be driven either by the  
26 HBV promoter (i.e. 1.3 to 2 genome unit) or a strong mammalian promoter (i.e. 1.1 genome unit).  
27 Until recently, the analysis of the replication capacity of naturally occurring or drug induced HBV  
28 mutants relied either on PCR-mediated transfer of HBV genome cassettes or on site directed  
29 mutagenesis of a well established replication-competent laboratory strain [16, 17]. Despite its  
30 obvious utility to quickly characterize new mutations *in vitro*, the methods based on the exchange of  
31 a cassette or site directed mutagenesis do not take into account the HBV genome variability existing  
32 in other parts of the genome.

33 Several methods based on a standardized cloning strategy of the entire HBV genome isolated from  
34 patient into plasmid vector have been recently described [10, 18] and have proven useful to study  
35 replication capacity and drug susceptibility, as well as cross-susceptibility of clinically relevant  
36 HBV strains [10, 18-23]. These cloning techniques enable the assembly of molecular clones,  
37 containing either 1.1 or 1.3 HBV genome unit, which allows the study of viral replication upon  
38 transfection of one clone or a mixture of clones into eukaryotic cells. By using a vector in which the  
39 expression of HBV pgRNA is driven by heterologous mammalian promoters (e.g. CMV or actin  
40 promoters), the replication level detected post-transfection, as measured by the neo-synthesis of

1 encapsidated relaxed-circular DNA (rcDNA), is artificially elevated. However the replication  
2 capacity of two given HBV strains can be determined in a relative fashion as it measures the  
3 intrinsic ability of HBV polymerase to transform pgRNA into rcDNA [10, 18].

4 Vector-free and vector-based replication assays are complementary approaches to determine the  
5 phenotype (replication capacity + drug susceptibility) of clinical HBV strains. The vector-free  
6 method is interesting as the whole viral population is represented in transfected amplicons, if we  
7 hypothesize that there is no bias during amplification. Vector based assays present the advantage of  
8 strong relative level of DNA synthesis, which render the approach more universal, but the  
9 disadvantage of a cloning step. Altogether, it may be interesting to use both approaches in order to  
10 generate complementary information. According to the approach used several methods can be  
11 utilized to detect HBV replication, including Southern blotting (if the replication level is high, i.e.  
12 when vector based assays are used) or qPCR. The first method is work intensive but gives  
13 qualitatively relevant results (i.e. visualization of all DNA forms of HBV genome), whereas the  
14 second is very sensitive but might be confronted to false positive (i.e. detection of infra-length DNA  
15 sequences). Moreover HBV replication may be monitored by quantifying intracellular or secreted  
16 DNA. If qPCR can be used in both cases, Southern blotting is more adapted for the detection of  
17 intracellular DNA due to the rather low sensibility of the methodology.

18 Beside the molecular approach used, the choice of the cell line for the transfection is also important  
19 as differences in term of replication capacity and drug susceptibility have been observed [9, 24].  
20 Altogether an important effort to standardize assays intra and inter laboratory remains to be made in  
21 order to move from current data that are mainly valid in a relative context (i.e. comparison with the  
22 same approach in the same laboratory) to fully comparable data.

23 It is worth noting that another approach to characterize viral drug resistance in tissue culture is the  
24 use of cell lines permanently expressing the mutants, to allow a more reproducible measure of drug  
25 susceptibility [25-28]. These cell lines are extremely useful to assess the antiviral activity of  
26 approved drugs and those in development on the main resistant mutants. However, one of the  
27 problems of these permanent cell lines, is the need to design and produce new cell lines when new  
28 resistant mutants are identified.

29 ***Virion production, secretion capacity and infectivity - Assembly and infectivity assays.*** Beside the  
30 ability of a strain to replicate its genome in the absence or presence of drugs, the next important step  
31 to ensure a strain a better propagation is the ability to produce particles. Some mutations occurring  
32 in the polymerase gene have a consequence in overlapping envelope genes, in particular in the S  
33 gene. For instance, the rtM204V mutation is associated with the mutation sI195M in the surface  
34 antigen, whereas the rtM204I change is associated with three possible changes: sW196S, sW196L,  
35 or a termination codon. The mutation selected by adefovir, lamivudine or telbivudine at rtA181T  
36 typically results in a stop mutation in the envelope gene (sW172stop), and the ADV-resistance  
37 mutation at rtA181V results in a concomitant change sL173F. Mutations that result in a stop codon  
38 mutation in the envelope gene are present in association with a low percentage of wild type HBV in  
39 order to ensure rescue of viral packaging and release [8].

40 Beside their impact on viral assembly, mutations may also impact the antigenicity and therefore the

1 ability of a strain to be neutralised by circulating or administrated antibodies [29, 30]. This may also  
2 have an impact on viral propagation and therefore represent a component of viral fitness. Several  
3 studies have examined the altered antigenicity of common antiviral drug selected mutations that  
4 also alter HBsAg. In the study by Torresi *et al.*, the mutations rtV173L+rtL180M+rtM204V that  
5 resulted in the mutations sE164D+ sI195M in HBsAg were found to reduce antigen - antibody  
6 binding [31]. Although the reduction was not as great as for the classical vaccine-escape mutant at  
7 sG145R, it was greatly reduced compared with the wild-type virus. These results were recently  
8 confirmed and extended by Sloan and colleagues using cell derived HBVs [30].

9 To study the impact of a given mutation on HBV assembly there are two main methodologies. The  
10 first is based on hepatomas cells replicating HBV after stable or transient transfection. Stably  
11 transformed cell lines are useful to study the assembly and secretion of HBV strains, but are work-  
12 intensive to generate. Moreover, it can be quite difficult to compare two different cell lines as the  
13 sites and numbers of integration into the genome may vary from one cell line to another. By contrast,  
14 the transient transfection of cells with plasmids carrying 1.1 to 2 HBV genome units is more  
15 flexible, although, due to transfection efficiency, a lower number of cells produce HBV particles  
16 compared to stably transformed cell lines. The assembly and secretion of HBV particles is measured  
17 by quantification of secreted HBV DNA by qPCR (or Southern blot) and/or envelope proteins by  
18 Elisa or Western blot. Those measurements are not precise, as they do not distinguish between  
19 enveloped and non-enveloped nucleocapsids for qPCR analysis, and between subviral particles  
20 and Dane particles for Elisa or Western blot analysis. The amount of Dane particle produced can be  
21 more precisely quantified after immunoprecipitation with an anti-PreS1 antibody followed by qPCR  
22 [7]. A more laborious approach is to purify particles by ultracentrifugation in density gradients, and  
23 to analyse the production of each particles (Dane, subviral particles and non-enveloped  
24 nucleocapsids). Electron microscopy can be used to some extent to visualise and distinguish HBV  
25 particles, but is not a quantitative approach.

26 Other approaches to study the impact of mutation in envelope genes on viral assembly are based on  
27 the utilisation of naturally pseudotyped hepatitis delta virus (HDV) particles. Indeed, HDV utilizes  
28 the envelope proteins of HBV for propagation. When introduced into permissive cells, the HDV  
29 RNA genome replicates and associates with multiple copies of the HDV-encoded proteins to  
30 assemble a ribonucleoprotein (RNP) complex. The mechanism necessary to export the RNP from  
31 the cell is provided by the HBV envelope proteins, which have the capacity to assemble lipoprotein  
32 vesicles that bud into the lumen of a pre-Golgi compartment before being secreted [32]. Hence the  
33 co-transfection of HDV genome (contained as trimers in a plasmid) and plasmid either expressing  
34 HBV pgRNA (i.e. like the ones used for replication assays; see above) or coding HBV envelope  
35 proteins leads to the production of either a mixture of HBV/HDV or pure HDV particles. These  
36 models have been successfully used to determine domains and residues of S protein important for  
37 HBV/HDV assembly [33, 34]. They can be used as surrogate model to study the impact of naturally  
38 or drug-selected mutations on HBV assembly. In this case assembly and secretion of HBV/HDV or  
39 HDV particles is measured by quantification of secreted HDV RNA by qRT-PCR (or Northern blot)  
40 and/or envelope proteins by Elisa or Western blot.

1 The infectivity of HBV particles represents another important component of viral fitness to be  
2 analyzed. To perform infectivity assays, HBV particles have to be produced. All the approaches to  
3 produced *in vitro* HBV particles have been already described above and include transiently-  
4 transfected or stably-transformed cells [7], as well as HDV based models [35, 36]. To perform  
5 comparative infectivity analysis, the critical issue is to standardize the inoculum to be used.  
6 Standardization based only on the dosing of HBV (or HDV) genome is not ideal as non-enveloped  
7 nucleocapsid can also be produced together with infectious Dane (or HDV) particles. Therefore a  
8 double standardization based on genome and protein dosing is necessary. An interesting method is  
9 to dose immunoprecipitated (with anti preS1 antibody) HBV (or HDV) genome. Infectivity assays  
10 are performed with cells that can be infected *in vitro*. Currently there are three main models  
11 available: primary human hepatocytes (PHH) [37], primary tupaia hepatocytes [38], or HepaRG  
12 cells [39], the last being the easiest to use because it is an established cell line. After inoculation  
13 with HBV or HDV particles, the intensity of infection is measured by either Northern blot or qRT-  
14 PCR to detect HBV or HDV RNAs. There are differences between HBV and HDV based models.  
15 With HDV, there is a disconnection between entry and replication, as the latter occurs irrespective  
16 of the HBV mutations that are assayed for their impact on entry. With HBV, the measurement of  
17 infectivity includes the ability to enter the cells and to initiate HBV replication, and both aspects are  
18 linked. One main problem with these infectivity models is that there is no propagation of infection  
19 in PHH and HepaRG cells. Thus no competition experiments have been reported with this  
20 experimental setting, although it is one of the main objectives when it comes to compare viral  
21 fitness of two strains.

22 ***Studying viral fitness in vivo: from human to animals models.*** Longitudinal studies in patients  
23 with CHB treated with antivirals are per se very interesting as they provide useful information on  
24 viral fitness. Hence, the emergence of a particular HBV mutant strain in patients treated with NAs  
25 indicates that this strain is fitter than others in this particular microenvironment. Villet *et al.*  
26 managed to obtain *in vitro* data on the viral fitness of a multi-resistant strain that emerged after  
27 treatment with lamivudine, adefovir, and anti-HBV immunoglobulins [7]. The *in vitro* data  
28 confirmed that the finally selected strain was the fittest one amongst the four strains that were yet  
29 present after few weeks of treatment in terms of replication capacity, assembly efficiency and  
30 specific infectivity. This was the first demonstration of a good correlation between clinical and  
31 laboratory findings. The main problem with longitudinal studies on patient cases is that they are  
32 retrospective. It would be interesting to have a small animal model to perform prospective studies.

33 One particularly important point when the viral fitness of a given drug resistant strain is concerned  
34 is to determine whether this strain can be transmitted. The first report of transmission of LMV  
35 resistant HBV was from Thibault *et al.* [40]. In this case report, the transmission of LMV-resistant  
36 HBV (rtL180M+rtM204V) was associated with an incubation period of 2-3 months and resulted in  
37 a typical acute hepatitis. Obtaining evidence for the transmission of HBV mutants in humans  
38 remains difficult and opportunistic. Animal models are necessary to get further information on the  
39 infectivity of HBV mutant strains. Using the chimpanzee infection model, Kamili *et al.* [41] tested  
40 the efficacy of the immunity induced by a commercial hepatitis B vaccine against challenge with a



1 tissue culture-derived, clonal HBV polymerase mutant that contained a combination of three  
2 polymerase mutations (rtV173L, rtL180M, rtM204V), two of which resulted in changes to the  
3 overlapping viral envelope of the HBsAg (sE164D, sI195M). Evidence of HBV replication was  
4 observed in the vaccinated chimpanzees after challenge with the mutant, as well as after rechallenge  
5 with serum-derived wild-type HBV, despite robust humoral and cellular anti-HBV immune  
6 responses after hepatitis B vaccination.

7 Although useful, the chimpanzee model remains difficult to handle. Amongst other animals that  
8 could be used for studying the infectivity of HBV mutant strains, liver-humanized mouse represents  
9 one of the most convenient models [42-44] SCID/Alb-uPA mice can be efficiently infected with *in*  
10 *vitro* produced HBV particles [45], and can be therefore used to study the infectivity of HBV  
11 mutants. Moreover HBV infected SCID/Alb-uPA mice can be treated with LMV and ADV, whereas  
12 infection with LMV-resistant strain was not sensitive to LMV treatment [46, 47]. Altogether this  
13 model could be potentially used to study viral fitness of HBV strains in mono or competitive  
14 challenges in the presence or absence of drugs. However, it is worth noting that quantitative and  
15 qualitative species-dependent differences in the enzymology of nucleos(t)ide metabolism in mouse  
16 tissues might limit the usefulness of this model since the hepatocyte's environment is influenced by  
17 other cells and tissues, that are of mouse origin in this model.

## 18 19 **Concluding remarks**

20 The study of HBV viral fitness *in vitro* is limited as there is currently no really relevant and unique  
21 cell culture model to perform such studies. In the absence of such a model, several assays have been  
22 developed to get insight into various parameters that define viral fitness, including the replication  
23 capacity, assembly efficiency, and specific infectivity of a given HBV strain. These assays have  
24 proven useful to get correlation between *in vitro* and *in vivo* data, and explain the emergence of a  
25 given strain *in vivo* by a better viral fitness of this one, as compared to other wild-type and mutants  
26 strains. Besides *in vitro* testing, animal models could be used to get further information about the  
27 fitness and the infectivity of HBV mutant strains. So far, using chimpanzees, it has been possible to  
28 demonstrate the transmissibility of HBV drug resistant strains, and using HBV vaccinated  
29 chimpanzees it has been possible to demonstrate the transmissibility of HBV vaccine escape  
30 mutants therefore pointing out the potential risk of spreading of such strains. Mice with humanized  
31 liver may also represent an interesting model to get prospective data on the fitness of clinically  
32 relevant and laboratory engineered HBV strains.

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