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Maternal Oxytocin Triggers a Transient Inhibitory Switch in GABA Signaling in the Fetal Brain During Delivery

Roman Tyzio, *et al.*
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How is the LPFC activity related to the activity in MT+ in the task-irrelevant condition? The correlation coefficient between the task-irrelevance-related activity (28) in MT+ and the LPFC was -0.90 . This is in accord with the view that when the LPFC is activated, it provides direct or indirect inhibitory control on the activity of MT+.

One might think that the low performance at the 5% coherence ratio was obtained because, despite the instructions to focus on RSVP task performance, the participants may have tried to find a coherent motion direction or to detect whether coherent motion was presented. If these motion tasks are difficult, they may leave fewer resources available for the RSVP task. However, this is not likely. If the participants engaged in the search for motion direction, this task should be hardest at 0% coherence and therefore, the lowest RSVP performance should have occurred at 0% and not at 5% coherence. Second, if the participants engaged in motion detection, this task should be hardest at 5% coherence because indecision may be greatest near the coherent motion threshold, and thus in accordance with the observed RSVP performance result. However, the lowest blood oxygen level-dependent (BOLD) activity was observed at 5% coherence ratio in the LPFC and cannot be directly explained by this possibility.

The results of the present study demonstrate two important points. First, a weak task-irrelevant stimulus feature that is below but near the perceptual threshold more strongly activates the visual area (MT+) that is highly related to the stimulus feature and more greatly disrupts task performance. There was a tendency for activity in the posterior occipitotemporal sulcus (pOTS) (29, 30) and the left angular gyrus (31), which are sensitive to letters and words and may be related to the RSVP task, to be lower at the 5% coherence than at the other coherent motion ratios. This contradicts the general view that irrelevant signals that are stronger in stimulus properties have a greater influence on the brain and performance and that the influence of a subthreshold stimulus is smaller than that of a suprathreshold stimulus.

Second, the results may reveal important bidirectional interactions between a cognitive controlling system and the visual system. The LPFC, which has been suggested to provide inhibitory control on task-irrelevant signals (22–26), may have a higher detection threshold for incoming signals than the visual cortex. Task-irrelevant signals around the threshold level may be sufficiently strong to be processed in the visual system but not strong enough for the LPFC to notice and, therefore, to provide effective inhibitory control on the signals (Fig. 4A). In this case, such signals may remain uninhibited, take more resources for a task-irrelevant distractor, leave fewer resources for a given task (32, 33), and disrupt task performance more than suprathreshold signals. On the other

hand, suprathreshold coherent motion may be noticed, may be given successful inhibitory control by the LPFC, and may leave more resources for a task (Fig. 4B) (22–26). This mechanism may underlie the present paradoxical finding that subthreshold task-irrelevant stimuli activate the visual area strongly and disrupt task performance more than some suprathreshold stimuli. It could also be one of the reasons why subthreshold stimuli often lead to relatively robust effects (2, 11, 14).

References and Notes

- S. He, P. Cavanagh, J. Intriligator, *Nature* **383**, 334 (1996).
- T. Watanabe, J. E. Nanez, Y. Sasaki, *Nature* **413**, 844 (2001).
- J. K. O'Regan, R. A. Rensink, J. J. Clark, *Nature* **398**, 34 (1999).
- A. Mack, I. Rock, *Inattentional Blindness* (MIT Press, Cambridge, MA, 1998).
- M. M. Chun, *J. Exp. Psychol. Hum. Percept. Perform.* **23**, 738 (1997).
- V. Stuphorn, J. D. Schall, *Nat. Neurosci.* **9**, 925 (2006).
- R. Blake, R. Fox, *Nature* **249**, 488 (1974).
- S. H. Lee, R. Blake, D. J. Heeger, *Nat. Neurosci.* **8**, 22 (2005).
- N. K. Logothetis, J. D. Schall, *Science* **245**, 761 (1989).
- S. Dehaene *et al.*, *Nature* **395**, 597 (1998).
- M. Bar, I. Biederman, *Psychol. Sci.* **9**, 464 (1998).
- J. D. Haynes, G. Rees, *Nat. Neurosci.* **8**, 686 (2005).
- A. Sahraie, M. Milders, M. Niedeggen, *Vision Res.* **41**, 1613 (2001).
- A. R. Seitz, T. Watanabe, *Nature* **422**, 36 (2003).
- M. Niedeggen, A. Sahraie, G. Hesselmann, M. Milders, C. Blakemore, *Brain Res. Cogn. Brain Res.* **13**, 241 (2002).
- Materials and methods are available as supporting material on Science Online.
- W. T. Newsome, E. B. Pare, *J. Neurosci.* **8**, 2201 (1988).
- G. Rees, K. Friston, C. Koch, *Nat. Neurosci.* **3**, 716 (2000).
- J. Stroop, *J. Exp. Psychol.* **18**, 643 (1935).
- A. M. Treisman, G. Gelade, *Cogn. Psychol.* **12**, 97 (1980).
- S. R. Friedman-Hill, L. C. Robertson, R. Desimone, L. G. Ungerleider, *Proc. Natl. Acad. Sci. U.S.A.* **100**, 4263 (2003).
- R. Dias, T. W. Robbins, A. C. Roberts, *Nature* **380**, 69 (1996).
- J. M. Fuster, *The Prefrontal Cortex: Anatomy, Physiology, and Neuropsychology of the Frontal Lobe* (Lippincott-Raven, New York, ed. 3, 1997).
- R. T. Knight, W. R. Staines, D. Swick, L. L. Chao, *Acta Psychol. (Amst.)* **101**, 159 (1999).
- J. G. Kerns *et al.*, *Science* **303**, 1023 (2004).
- A. W. MacDonald III, J. D. Cohen, V. A. Stenger, C. S. Carter, *Science* **288**, 1835 (2000).
- In the control condition in which motion was task-relevant (Fig. 3B, green), no significant difference was found between any pair of coherence levels.
- Task-irrelevance-related activity is defined as a BOLD signal amount in the task-relevant condition subtracted from that in the task-irrelevant condition, for each motion coherence and for each cortical area.
- M. Ben-Shachar, R. F. Dougherty, G. K. Deutch, B. A. Wandell, *Cereb. Cortex* **10.1093/cercor/bhl071** (2006).
- S. Dehaene, L. Cohen, M. Sigman, F. Vinckier, *Trends Cogn. Sci.* **9**, 335 (2005).
- A. M. Callan, D. E. Callan, S. Masaki, *Neuroimage* **28**, 553 (2005).
- R. Desimone, J. Duncan, *Annu. Rev. Neurosci.* **18**, 193 (1995).
- K. Miller, J. D. Cohen, *Annu. Rev. Neurosci.* **24**, 167 (2001).
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Supporting Online Material

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Materials and Methods

Figs. S1 and S2
References

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Maternal Oxytocin Triggers a Transient Inhibitory Switch in GABA Signaling in the Fetal Brain During Delivery

Roman Tyzio,¹ Rosa Cossart,¹ Ilgam Khalilov,¹ Marat Minlebaev,¹ Christian A. Hübnér,² Alfonso Represa,¹ Yehezkel Ben-Ari,^{1*} Rustem Khazipov¹

We report a signaling mechanism in rats between mother and fetus aimed at preparing fetal neurons for delivery. In immature neurons, γ -aminobutyric acid (GABA) is the primary excitatory neurotransmitter. We found that, shortly before delivery, there is a transient reduction in the intracellular chloride concentration and an excitatory-to-inhibitory switch of GABA actions. These events were triggered by oxytocin, an essential maternal hormone for labor. In vivo administration of an oxytocin receptor antagonist before delivery prevented the switch of GABA actions in fetal neurons and aggravated the severity of anoxic episodes. Thus, maternal oxytocin inhibits fetal neurons and increases their resistance to insults during delivery.

Delivery is a stressful event associated with high risks to the fetal brain (1); however, whether the fetal brain prepares for

delivery remains largely unknown. We addressed this issue by studying γ -aminobutyric acid (GABA)-mediated (GABAergic) signaling in the

rat hippocampus. GABA is the principal inhibitory neurotransmitter in the adult brain. However, during fetal and postnatal periods GABA has a depolarizing action (2–4) and provides the major, and often the only, excitatory synaptic input to immature neurons (5). Excitatory action of GABA is due to elevated intracellular chloride and depolarized value of the GABA type A ($GABA_A$) reversal potential (E_{GABA}) (6–10). Excitatory GABA is pivotal in control of neuronal firing, generation of the primitive patterns of activity, intracellular calcium signaling, and neuronal development (2, 3). GABA signaling is therefore expected to be an essential element in physiological adaptation to stress during delivery.

To characterize the properties of GABA signaling during the perinatal period, we used cell-attached recordings from fetal and neonatal rat hippocampal slices [from embryonic day (E) 18 to postnatal day (P) 5; term is E21] (11). During fetal (E18 and E19) and postnatal periods (P1 to P5), activation of $GABA_A$ receptors increased the firing

of action potentials in the majority of CA3 pyramidal cells [Fig. 1A; also (6, 7, 12)]. However, during a brief period extending from E20 to day of birth (P0), the proportion of cells excited by GABA sharply decreased. The loss of the excitatory effect of GABA peaked at E21 [~1 to 2 hours before delivery (Fig. 1A)].

The action of GABA depends on the direction of the transmembrane current elicited by GABA, its driving force (DF_{GABA}) being the difference between E_{GABA} and resting membrane potential (E_m). Positive values of DF_{GABA} determine excitatory actions of GABA in immature neurons (2, 3, 6, 7). To estimate DF_{GABA} , we used cell-attached recordings of single $GABA_A$ channels (10) (Fig. 1, B to D), which affect neither E_{GABA} nor E_m . With 1 μ M GABA in the pipette solution, $GABA_A$ channels were observed in ~90% of patches ($n = 553$) but not in the presence of the $GABA_A$ receptor antagonist picrotoxin [100 μ M; $n = 25$ (Fig. 1B)]. We found that DF_{GABA} is strongly depolarizing in fetal and postnatal periods (Fig. 1D). However, DF_{GABA} negatively shifted during a brief near-term period (from E20 to P0), switching to a hyperpolarizing value of -8.4 ± 2.5 mV at term [mean \pm SEM, $n = 34$ (Fig. 1D)]. Thus, the disappearance of GABA-mediated excitation at term (Fig. 1A) coincides with a switch in polarity of GABA signals.

The perinatal changes in DF_{GABA} could be due to a negative shift of E_{GABA} or to a depolarizing shift of E_m . By using cell-attached recordings of single *N*-methyl-D-aspartate (NMDA) channels as voltage sensors (12, 13), we found that during early fetal and postnatal periods E_m is -78.7 ± 1.9 mV ($n = 42$; pooled data for E18 and E19 and P1 to P5) [also (13, 14)], but near term there is a small hyperpolarizing shift to -85.4 ± 1.5 mV [$n = 14$ at birth (Fig. 1E)]. Knowing DF_{GABA} and E_m , we calculated E_{GABA} ($E_{GABA} = DF_{GABA} + E_m$) and found that E_{GABA} switches from -40 mV at E18 to -92 mV at E21 and then returns to depolarizing values shortly after birth (Fig. 1F). This corresponds to a decrease in $[Cl^-]_i$ (intracellular chloride concentration) from 18 mM to 4 mM (Fig. 1F).

Because GABA-induced depolarization raises $[Ca^{2+}]_i$ (12, 15–18), we used a multibeam two-photon microscope to monitor $[Ca^{2+}]_i$ changes in hundreds of CA3 neurons in slices loaded with the calcium indicator fura 2-AM (Fig. 2). The $GABA_A$ receptor agonist isoguvacine (10 μ M) elicited a robust increase of $[Ca^{2+}]_i$ in the majority of neurons at E18 and at P2 to P4 [average of $56 \pm 3\%$, n values of 7 movies and 931 cells (Fig. 2, B to D)]. In contrast, at E21 to P0, $[Ca^{2+}]_i$ increased in only $31 \pm 6\%$ (n values of 10 movies and 1546 cells, $P < 0.01$) cells. Isoguvacine also decreased the frequency of spontaneous calcium

¹Institut de Neurobiologie de la Méditerranée, INSERM U29, Université de la Méditerranée, Campus Scientifique de Luminy, Boite Postale 13, 13273 Marseille Cedex 09, France. ²Institut für Humangenetik, Universitätsklinikum Hamburg-Eppendorf, 20251 Hamburg, Germany.

*To whom correspondence should be addressed. E-mail: ben-ari@inmed.univ-mrs.fr

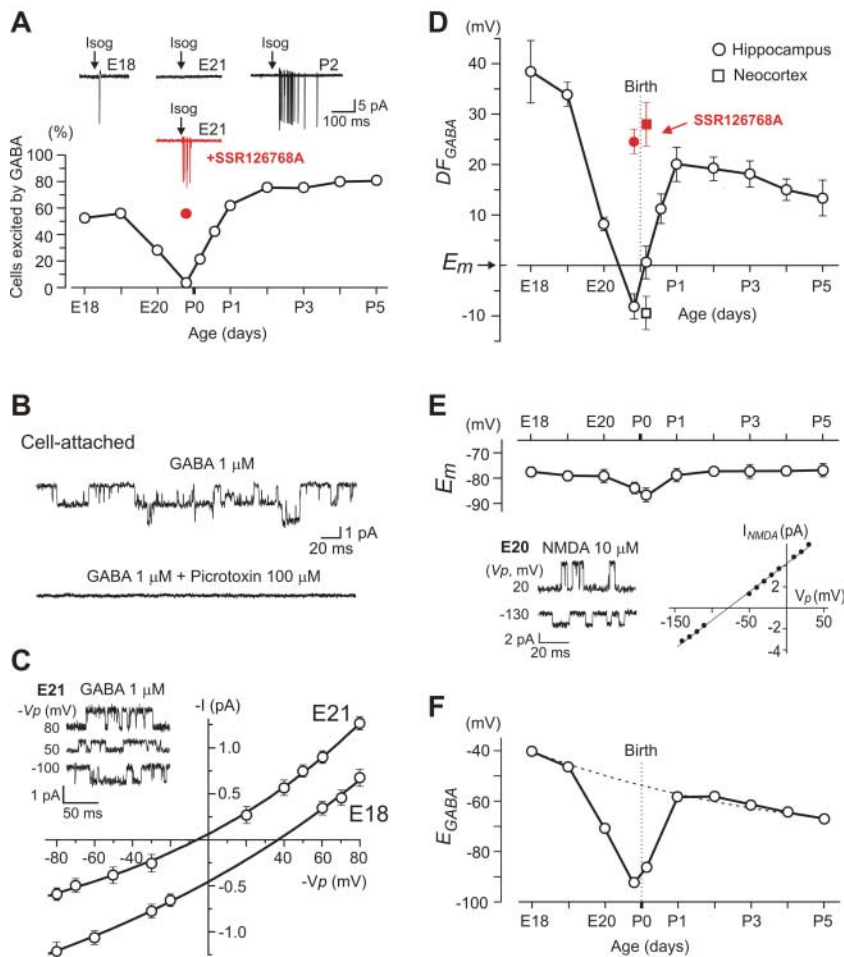


Fig. 1. Transient perinatal loss of the $GABA_A$ -mediated excitation. (A) Responses of CA3 pyramidal cells recorded in cell-attached mode to the $GABA_A$ agonist isoguvacine. Below, summary plot of the proportion of cells excited by isoguvacine during the perinatal period. There is a transient loss of the excitatory effect of isoguvacine near term. Red corresponds to the fetuses whose mothers received SSR126768A. [E21 corresponds to the early phase of delivery (1 to 2 hours before birth); P0 is the day of birth; pooled data from 146 neurons.] (B) Cell-attached recordings of single $GABA_A$ channels with 1 μ M of GABA in patch pipette (top trace); the channels were not observed in the presence of the $GABA_A$ antagonist picrotoxin (100 μ M; bottom trace). (C) *I-V* relationships of the currents through $GABA_A$ channels in two cells at E21 and E18; their reversal potential corresponds to DF_{GABA} . (D) Summary plot of the age dependence of DF_{GABA} inferred from single $GABA_A$ channels recordings [mean \pm SEM; 209 CA3 pyramidal cells (\circ) and 17 neocortical pyramidal cells (\square); 6 to 24 patches for each point]. Red indicates pretreatment with SSR126768A (n values are 25 hippocampal and 9 neocortical patches). (E) Age dependence of the resting membrane potential (E_m) of CA3 pyramidal cells inferred from the reversal of single NMDA channels recorded in cell-attached mode ($n = 84$ cells; 4 to 12 patches for each point). (F) Age dependence of the $GABA_A$ reversal potential ($E_{GABA} = E_m + DF_{GABA}$). There is a transient hyperpolarizing shift of E_{GABA} near birth.

events in many neurons near term [$9 \pm 2\%$ (Fig. 2)]. Thus, during delivery, the ability of GABA to increase $[Ca^{2+}]_i$ is significantly reduced.

What underlies the near-term switch in GABA actions? Because this phenomenon is not observed in fetal neurons grown in culture (8, 9), we hypothesized that it is related to parturition and in particular to maternal hormones released during delivery. Parturition is initiated by a massive release of oxytocin (19). In addition to its pivotal role in parturition, there are also indications that oxytocin exerts multiple effects in the adult central nervous system (19–25). Maternal oxytocin easily crosses the placenta to reach the fetus (26), suggesting that oxytocin may be responsible for the hyperpolarizing switch in action of GABA in fetal neurons during delivery. With selective antibodies, we found a high density of oxytocin receptor immunoreactivity in the hippocampus and neocortex during the perinatal period (Fig. 3A and figs. S1 and S2). During cell-attached recordings from slices at E18 and P2, applications of oxytocin (1 μ M) induced a negative shift in DF_{GABA} (Fig. 3, C and D) and suppressed GABA-mediated excitation (Fig. 3B). The effects of oxytocin were completely prevented by bath application of the selective oxytocin receptor antagonist atosiban (AT, 1 to 5 μ M) (Fig. 3D). At term, application of oxytocin did not cause a significant effect on the hyperpolarizing DF_{GABA} (Fig. 3D), suggesting that the effects of exogenous oxytocin are occluded by the endogenous hormone. In keeping with this hypothesis, we found that atosiban switched DF_{GABA} from hyperpolarizing to depolarizing at term but not in

fetal and postnatal neurons (Fig. 3D). Moreover, intracardial perfusion of E21 fetuses with artificial cerebrospinal fluid (ACSF, to wash out endogenous hormone) produced a similar shift of DF_{GABA} from hyperpolarizing to depolarizing (fig. S3). Addition of 1 μ M oxytocin at the end of the perfusion restored hyperpolarizing values of DF_{GABA} (fig. S3). Therefore, endogenous oxytocin is present in slices at term and provides tonic activation of oxytocin receptors, leading to the shift of DF_{GABA} . We also found that the negative shift in DF_{GABA} from 38.4 ± 6.2 mV ($n = 8$) to -7.2 ± 2.2 mV ($n = 18$) occurred in E18 fetuses obtained from mothers treated with oxytocin (50 μ g/kg) during 2 hours before cesarean delivery.

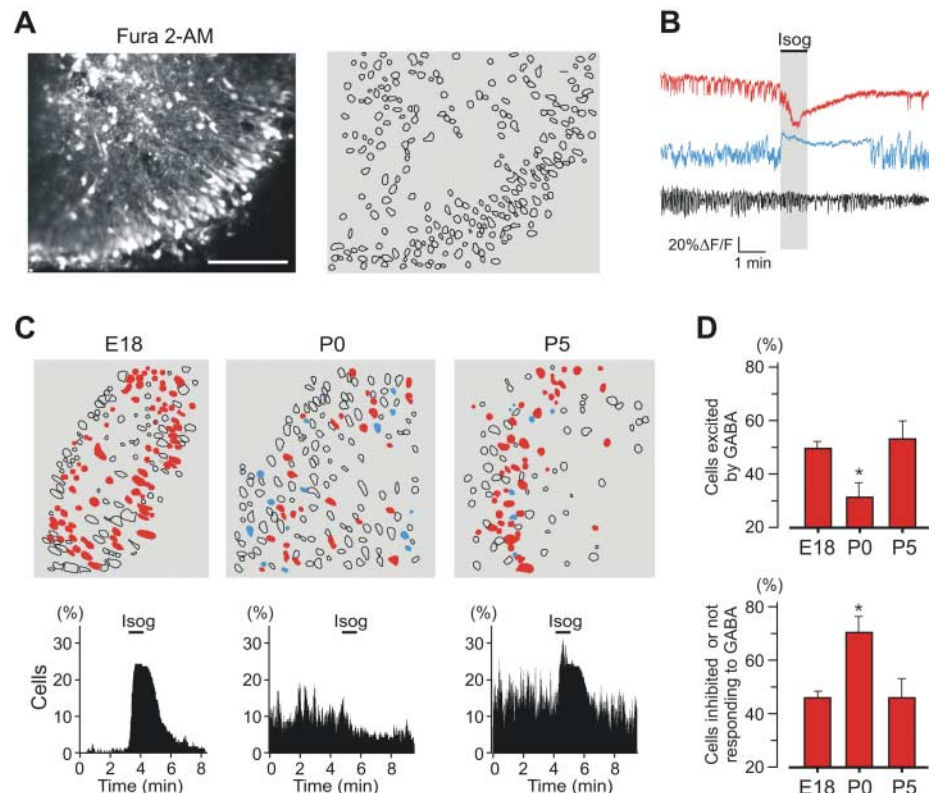
To directly measure the effects of oxytocin on $[Cl^-]_i$, we used two-photon chloride imaging in slices loaded with a chloride-sensitive dye, MQAE (Fig. 3, E and F) (27). In E18 and P4 slices, oxytocin (1 μ M) produced a significant increase in the baseline fluorescence signal in half of the imaged cells, indicating a strong reduction of $[Cl^-]_i$ [average of $43 \pm 4\%$, n values of 10 movies and 1792 cells (Fig. 3F)], and the effect was prevented by atosiban (5 μ M; reduction of $[Cl^-]_i$ in $5 \pm 3\%$ cells, n values of 2 movies and 435 cells). Furthermore, incubation of P4 slices with oxytocin reversed the effect of isoguvacine on spontaneous calcium events [59 $\pm 17\%$ imaged cells not activated by isoguvacine, n values of 3 movies and 228 cells (fig. S4)]. Thus, the effects of oxytocin fully matched the changes in GABA signaling occurring at term. This result indicates that oxytocin is sufficient to trigger changes of $[Cl^-]_i$.

To determine whether endogenous oxytocin is necessary for the near-term switch in the action of GABA, we treated pregnant rats orally with the selective oxytocin receptor antagonist SSR126768A (28) (1 mg/kg, starting from E20) and measured the consequences at term. Cell-attached recordings revealed major differences between treated and age-matched control pups, including strongly depolarizing values of DF_{GABA} (Fig. 1D, red circle) and a high proportion of cells excited by GABA (Fig. 1A, red circle). Similar effects were also observed in neocortical neurons, in which DF_{GABA} was -9.4 ± 3.4 mV in control rat pups ($n = 8$) and 28.3 ± 3.7 mV ($n = 9$) in pups of mothers pretreated with SSR126768A (Fig. 1D). Thus, maternal oxytocin is necessary and sufficient to trigger the near-term switch in GABA action in the hippocampus and neocortex.

What are the mechanisms underlying the oxytocin-mediated reduction of $[Cl^-]_i$ at term? Because the NKCC1 chloride inward cotransporter is responsible for elevated $[Cl^-]_i$ in immature neurons (4, 7, 29, 30), we tested the effects of the selective NKCC1 antagonist bumetanide. We found that bumetanide (10 μ M) produces a robust negative shift of DF_{GABA} and occludes the effects of oxytocin (fig. S5). The short latency of oxytocin actions (Fig. 3F) and the observation that blockade of oxytocin receptors does not alter chloride transporters gene expression (fig. S6) suggest that the hormone down-regulates NKCC1 activity.

Hypoxic-ischemic brain damage is a principal cause of newborn death and neurological impairment (1). We hypothesized that the excitatory-to-

Fig. 2. Perinatal effects of GABA_A receptors activation on $[Ca^{2+}]_i$ in CA3 neurons. (A) Two-photon $[Ca^{2+}]_i$ fluorescence image of the CA3 region from a P0 hippocampal slice loaded with fura 2-AM; scale bar indicates 100 μ m. (Right) Automatically detected contours of the cells. (B) Three types of $[Ca^{2+}]_i$ responses ($\Delta F/F$) to bath application of the GABA_A agonist isoguvacine (10 μ M, 1 min): excited (red), inhibited (blue), or not affected (black) by isoguvacine (Isog). (C) Contour maps representing the distribution of different cellular responses to isoguvacine at E18, P0, and P5: excited (red), inhibited (blue), and not affected (open contours). Bottom histograms show the percentage of cells that are detected as being active on each movie frame corresponding to the above contour plots (140 ms per frame). Isoguvacine application (10 μ M) produces a significant increase in the fraction of active neurons at E18 and P5, whereas it slightly decreased the activity at birth. (D) Histograms of the averaged fraction of imaged neurons excited (top) or inhibited or nonaffected (bottom) by isoguvacine at different ages (for P0, n values of 10 movies and 1546 cells; for E18, n values of 2 movies and 375 cells; and for P5, n values of 3 movies and 328 cells; * $P < 0.01$). Error bars indicate SEM.



inhibitory switch in GABA signaling reduces neuronal activity and metabolic demand, thus helping to protect fetal neurons from hypoxic insults (31). Episodes of anoxia and aglycemia were induced by superfusion with a solution in which oxygen was substituted for nitrogen and glucose was substituted for sucrose. The onset of anoxic depolarization (AD) that is an electrophysiological

marker of neuronal death (31, 32) was measured by using extracellular field potential recordings in the intact hippocampi of E21 rats in vitro after various treatments in vivo to the mother and/or to the fetuses intracardially before the in vitro experiment (Fig 4). In control fetuses, AD occurred after 55.6 ± 1.4 min ($n = 30$) of perfusion with the anoxic-aglycemic solution (Fig. 4). In the hippocampi

prepared from the fetuses intracardially perfused with atosiban ($5 \mu\text{M}$; $n = 15$) or from the fetuses whose mothers received SSR126768A (1 mg/kg ; $n = 19$), AD onset was significantly accelerated to 44.1 ± 1.1 min ($n = 34$; $P < 0.01$). Bumetanide ($10 \mu\text{M}$) applied in the presence of the oxytocin receptor antagonists delayed AD to the control values (53.4 ± 1.7 min; $n = 18$; $P < 0.01$). Bumetanide ($10 \mu\text{M}$)

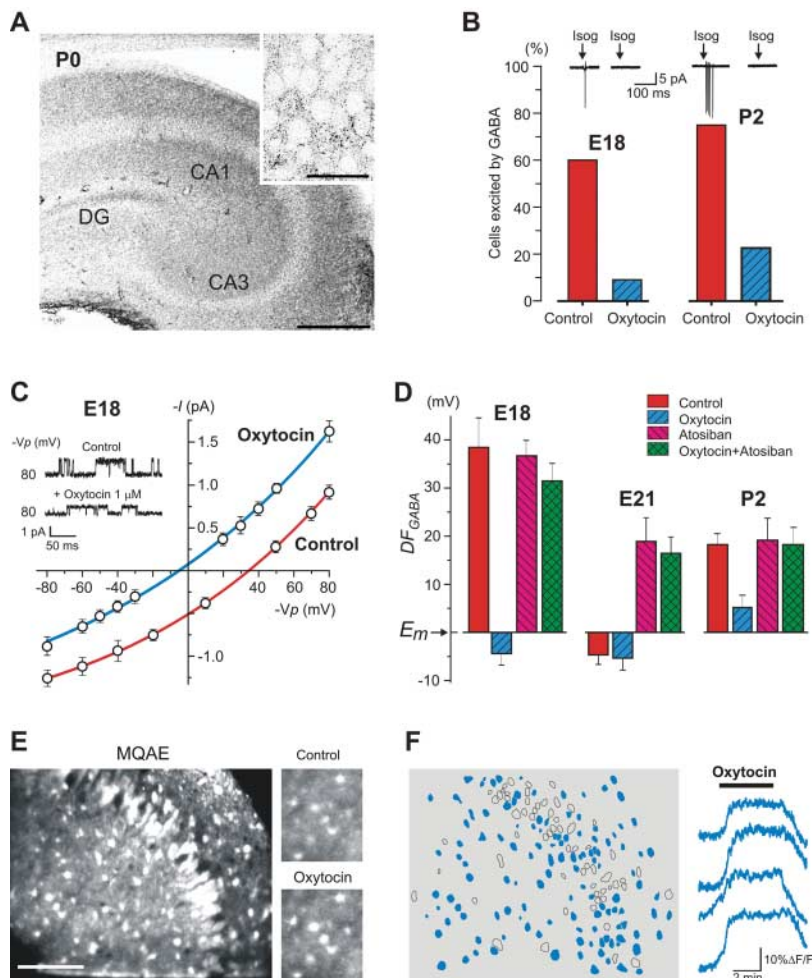
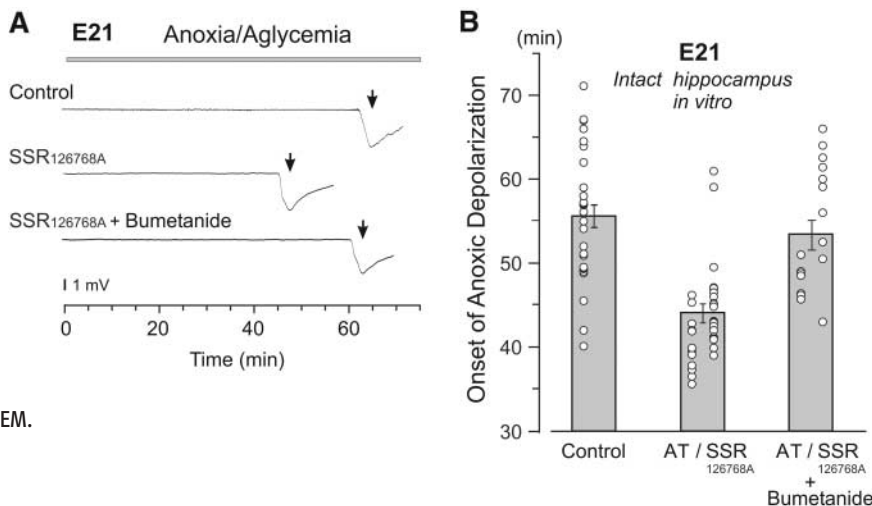


Fig. 3. Oxytocin causes a switch in GABA_A signaling from depolarizing to hyperpolarizing. **(A)** Oxytocin-receptor immunostaining of a P0 rat hippocampus; (inset) CA3 pyramidal cells layer. Bars, 200 μm and (inset) 20 μm . **(B)** Histograms of the proportion of cells excited by brief application of isoguvacine (Isog) in control and in the presence of oxytocin ($1 \mu\text{M}$) at E18 and P2 ($n = 57$ cells). **(C)** *I-V* relationships of the currents through single GABA_A channels recorded from two CA3 pyramidal cells at E18 in control and after addition of oxytocin ($1 \mu\text{M}$). **(D)** Histograms of DF_{GABA} measured at E18, E21, and P2 in control conditions and in the presence of oxytocin ($1 \mu\text{M}$), antagonist of oxytocin receptors atosiban (1 to 5 μM), and oxytocin plus atosiban ($n = 187$ cells). **(E)** Two-photon imaging of $[\text{Cl}^-]_i$ in P4 hippocampal slice loaded with Cl^- indicator MQAE. Scale bar, 100 μm . (Right) $[\text{Cl}^-]_i$ fluorescence change produced by oxytocin application. The intensity of the fluorescence signal increases, and $[\text{Cl}^-]_i$ decreases, in the illustrated region. **(F)** Automatically detected contours of the cells imaged in (E), indicating the distribution of cells in which oxytocin application produced a significant decrease in $[\text{Cl}^-]_i$ (blue filled contours). On the right, chloride fluorescence changes ($\Delta F/F$) in four representative neurons. Time resolution is 100 ms per frame.

Fig. 4. Blockade of oxytocin receptors decreases fetal brain resistance to anoxia-aglycemia at birth. **(A)** Representative traces to illustrate the effects of anoxic-aglycemic solution on extracellular field potential recordings from E21 intact hippocampi. Arrows indicate terminal AD that marks neuronal death. AD occurs earlier in the presence of SSR126768A (middle trace). Addition of bumetanide ($10 \mu\text{M}$) occludes the effects of the antagonist and restores the initial delay (bottom trace). **(B)** Summary plot of the onset of AD in control, in the presence of the oxytocin receptors antagonists atosiban (AT, 5 μM , fetal intracardial perfusion and SSR126768A (1 mg/kg to the mother), and after further addition of bumetanide ($10 \mu\text{M}$). Each circle corresponds to one hippocampus ($n = 82$ intact hippocampi at E21). Error bars indicate SEM.



occluded the effects of the oxytocin receptor antagonists and restored the control delays (53.4 ± 1.7 min, $n = 18$; $P < 0.01$). These data indicate that maternal oxytocin exerts a neuroprotective action on fetal neurons during parturition and that this action is likely due to a reduction of $[Cl^-]_i$.

Our results suggest that oxytocin, in addition to its well-established role in labor and lactation and its multiple effects in the adult central nervous system (19–25), also exerts a powerful action on fetal neurons. This mechanism adds a previously unknown facet to the plasticity of GABA signaling via modulation of $[Cl^-]_i$ (4, 33). The dual action produced by a single messenger in the mother and fetus enables a perfect timing for adaptation of fetal neurons to delivery.

References and Notes

- J. J. Volpe, *Neurology of the Newborn* (Saunders, Philadelphia, ed. 4, 2000).
- Y. Ben Ari, *Nat. Rev. Neurosci.* **3**, 728 (2002).
- D. F. Owens, A. R. Kriegstein, *Nat. Rev. Neurosci.* **3**, 715 (2002).
- J. A. Payne, C. Rivera, J. Voipio, K. Kaila, *Trends Neurosci.* **26**, 199 (2003).
- R. Tyzio *et al.*, *J. Neurosci.* **19**, 10372 (1999).

- Y. Ben-Ari, E. Cherubini, R. Corradetti, J.-L. Gaïarsa, *J. Physiol.* **416**, 303 (1989).
- C. Rivera *et al.*, *Nature* **397**, 251 (1999).
- D. F. Owens, L. H. Boyce, M. B. Davis, A. R. Kriegstein, *J. Neurosci.* **16**, 6414 (1996).
- G. Chen, P. Q. Trombley, A. N. van den Pol, *J. Physiol.* **494**, 451 (1996).
- R. Serafini, A. Y. Valeyev, J. L. Barker, M. O. Poulter, *J. Physiol.* **488**, 371 (1995).
- Materials and methods are available as supporting material on Science Online.
- X. Leinekugel, I. Medina, I. Khalilov, Y. Ben-Ari, R. Khazipov, *Neuron* **18**, 243 (1997).
- R. Tyzio *et al.*, *J. Neurophysiol.* **90**, 2964 (2003).
- D. Maric *et al.*, *Eur. J. Neurosci.* **10**, 2532 (1998).
- J. A. Connor, H. Y. Tseng, P. E. Hockberger, *J. Neurosci.* **7**, 1384 (1987).
- R. Yuste, L. C. Katz, *Neuron* **6**, 333 (1991).
- J. J. LoTurco, D. F. Owens, M. J. Heath, M. B. Davis, A. R. Kriegstein, *Neuron* **15**, 1287 (1995).
- O. Garaschuk, E. Hanse, A. Konnerth, *J. Physiol.* **507**, 219 (1998).
- G. Gimpl, F. Fahrenholz, *Physiol. Rev.* **81**, 629 (2001).
- A. Argiolas, G. L. Gessa, *Neurosci. Biobehav. Rev.* **15**, 217 (1991).
- M. Ragenbass, *Prog. Neurobiol.* **64**, 307 (2001).
- K. Tomizawa *et al.*, *Nat. Neurosci.* **6**, 384 (2003).
- M. Kosfeld, M. Heinrichs, P. J. Zak, U. Fischbacher, E. Fehr, *Nature* **435**, 673 (2005).
- D. T. Theodosis *et al.*, *Mol. Cell. Neurosci.* **31**, 785 (2006).

- D. Huber, P. Veinante, R. Stoop, *Science* **308**, 245 (2005).
- A. Malek, E. Blann, D. R. Mattison, *J. Matern. Fetal Med.* **5**, 245 (1996).
- N. Marandi, A. Konnerth, O. Garaschuk, *Pflugers Arch.* **445**, 357 (2002).
- C. Serradeil-Le Gal *et al.*, *J. Pharmacol. Exp. Ther.* **309**, 414 (2004).
- J. Yamada *et al.*, *J. Physiol.* **557**, 829 (2004).
- V. I. Dzhalala *et al.*, *Nat. Med.* **11**, 1205 (2005).
- V. I. Dzhalala, Y. Ben-Ari, R. Khazipov, *Ann. Neurol.* **48**, 632 (2000).
- P. Lipton, *Physiol. Rev.* **79**, 1431 (1999).
- H. Fiumelli, L. Cancedda, M. M. Poo, *Neuron* **48**, 773 (2005).
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Materials and Methods

Figs. S1 to S6

References

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ATP Release Guides Neutrophil Chemotaxis via P2Y2 and A3 Receptors

Yu Chen,^{1*} Ross Corriden,^{1,2*} Yoshiaki Inoue,¹ Linda Yip,¹ Naoyuki Hashiguchi,¹ Annelies Zinkernagel,⁴ Victor Nizet,⁴ Paul A. Insel,^{2,3} Wolfgang G. Junger^{1†}

Cells must amplify external signals to orient and migrate in chemotactic gradient fields. We find that human neutrophils release adenosine triphosphate (ATP) from the leading edge of the cell surface to amplify chemotactic signals and direct cell orientation by feedback through P2Y2 nucleotide receptors. Neutrophils rapidly hydrolyze released ATP to adenosine that then acts via A3-type adenosine receptors, which are recruited to the leading edge, to promote cell migration. Thus, ATP release and autocrine feedback through P2Y2 and A3 receptors provide signal amplification, controlling gradient sensing and migration of neutrophils.

Neutrophils are primary phagocytic cells with important roles in host defense and tissue repair. However, activated neutrophils damage host tissues and contribute to chronic inflammatory diseases, including rheumatoid arthritis, inflammatory bowel disease, and asthma (1). A key feature of neutrophils is their ability to detect and migrate to compromised tissues by following a concentration gradient of chemotactic substances released from microbial pathogens or injured cells.

Neutrophils can respond to chemoattractant gradients that differ in concentration by as little as 1% across the length of the cell body (2). Chemotaxis must involve signal amplification because a strongly polarized distribution of intracellular signal-transduction components is observed even in shallow gradients. The mechanisms of signal amplification are unclear (3, 4). We identified the polarized release of adenosine triphosphate (ATP), the activation of P2Y2 receptors, and the translocation and activation of A3 adenosine receptors as key mechanisms of signal amplification that control cell orientation and direct the migration of neutrophils.

Membrane deformation caused by mechanical or osmotic stress induces the release of cellular ATP from mammalian cells; however, detailed information on the underlying mechanisms is lacking (5, 6). Because cell migration also involves membrane deformation, we tested whether the

stimulation of human neutrophils with the chemoattractant *N*-formyl-Met-Leu-Phe (FMLP) causes ATP release. Treatment of cells (10^7 cells in 250 μ l of solution) with 100 nM FMLP rapidly tripled extracellular ATP concentrations in bulk media (Fig. 1, A and B) by inducing the release of ~0.5% of their ATP pool. Concentrations of extracellular ATP and its hydrolytic products adenosine monophosphate (AMP) and adenosine peaked 5 min after FMLP stimulation; but while the concentration of ATP returned to basal levels after 15 min, AMP and adenosine concentrations remained >5-fold above baseline (Fig. 1B), which is consistent with the presence of ecto-adenosine triphosphatases (ecto-ATPases) (7, 8). Neutrophils completely hydrolyzed exogenous ATP (5 μ M) within 2 min after ATP addition, which suggests that they have potent ecto-ATPase activity (Fig. 1C).

Because membrane deformation occurs predominantly at the leading edge closest to the chemoattractant source, we hypothesized this region to be the principal site of ATP release. Fluorescent microscopy that was used to visualize ATP release [based on conversion of nicotinamide adenine dinucleotide phosphate (NADP⁺) to its reduced form NADPH] revealed that neutrophils discharge ATP within seconds after FMLP stimulation (Fig. 1D and movies S1 to S3), with ATP release highest near the cell membrane with the greatest degree of protrusion (Fig. 1D, inset).

Extracellular ATP and adenosine modulate neutrophil functions, including chemotaxis (8, 9). We tested the effect of apyrase, which hydrolyzes ATP, on chemotaxis in a trans-well system composed of upper wells with neutrophils and lower wells with 1 nM FMLP separated by a filter with 3- μ m pores. Addition of apyrase to the upper wells reduced chemotaxis by nearly 100% (Fig. 2A).

¹Department of Surgery, University of California San Diego, San Diego, CA 92103, USA. ²Department of Pharmacology, University of California San Diego, La Jolla, CA 92093, USA. ³Department of Medicine, University of California San Diego, La Jolla, CA 92093, USA. ⁴Department of Pediatrics, University of California San Diego, La Jolla, CA 92093, USA.

*These authors contributed equally to this work.

†To whom correspondence should be addressed. E-mail: wjunger@ucsd.edu