MMSET is overexpressed in cancers: link with tumor aggressiveness

Alboukadel Kassambara[‡], Bernard Klein^{*†}, Jérôme Moreaux^{*‡}

* CHU Montpellier, Institute of Research in Biotherapy, Montpellier, FRANCE;

[‡] INSERM, U847, Montpellier, F-34197 France;

[†] Université MONTPELLIER1, UFR Médecine, Montpellier, France;

Corresponding author: Pr. Bernard KLEIN
Director of INSERM U847
Institute of Research in Biotherapy CHU Montpellier,
Av Augustin Fliche
34285 Montpellier cedex, FRANCE
Tel 33-(0)467337888
Fax 33-(0)467337905
Mail: bernard.klein@montp.inserm.fr
<http://irb.chu-montpellier.fr/index.htm>

Abstract:

MMSET is expressed ubiquitously in early development and its deletion is associated with the malformation syndrome called Wolf-hirschhorn syndrome. It is involved in the t(4;14)(p16;q32) chromosomal translocation, which is the second most common translocation in multiple myeloma (MM) and is associated with the worst prognosis. MMSET expression has been shown to promote cellular adhesion, clonogenic growth and tumorigenicity in multiple myeloma. MMSET expression has been recently shown to increase with ascending tumor proliferation activity in glioblastoma multiforme. These data demonstrate that MMSET could be implicated in tumor emergence and/or progression. Therefore we compared the expression of MMSET in 40 human tumor types – brain, epithelial, lymphoid – to that of their normal tissue counterparts using publicly available gene expression data, including the Oncomine Cancer Microarray database. We found significant overexpression of *MMSET* in 15 cancers compared to their normal counterparts. Furthermore MMSET is associated with tumor aggressiveness or prognosis in many types of these aforementioned cancers.

Taken together, these data suggest that MMSET potentially acts as a pathogenic agent in many cancers. The identification of the targets of MMSET and their role in cell growth and survival will be key to understand how MMSET is associated with tumor development.

Introduction

MMSET gene is expressed ubiquitously in early development. The Wolf-hirschhorn syndrome is a malformation syndrome associated with a homizygous deletion of the distal short arm of chromosome 4 containing MMSET gene[1].

The *MMSET* gene is involved in the t(4;14)(p16;q32) chromosomal translocation, which is the most common translocation in multiple myeloma (MM) – occurring in 15% of newly-diagnosed patients – and is associated with poor prognosis[1; 2; 3]. The t(4;14) translocation dysregulates both *MMSET* and *FGFR3* genes that are closely located on chromosome 4p16. FGFR3 has oncogenic activity in vitro and in vivo, but approximately one-third of MM tumors harboring t(4;14) translocation lacks *FGFR3* expression, whereas an overexpression of *MMSET* is a universal characteristic of t(4;14) cases[3; 4; 5]. Furthermore, the poor prognosis of t(4;14) is independent of FGFR3 expression. These data suggest that activation of MMSET, not FGFR3, may be the critical transforming event of this recurrent translocation[1; 3; 5].

MMSET gene encodes for three isoforms including MMSET type I (647 aa) and type II (1365 aa) as a result of alternative splicing, and a 584 aa protein, known as RE-IIB (response element II-binding protein) translated from a second transcription initiation site. MMSET isoforms share various conserved protein domains suggesting a role in DNA-binding and chromatin modification, including PWWP domains, an HMG domain, PHD zinc fingers, and a C-terminal SET domain characteristic of histone methyltransferases[1].

The biological role of MMSET in MM disease is still poorly understood. A direct evidence shows that MMSET expression contributes to cellular adhesion, clonogenic

growth and tumorigenicity in multiple myeloma [1]. MMSET contains a SET domain that is found in many histone methyltransferases and determines their enzymatic activity. Histone lysine methylation promotes or prevents binding of proteins and protein complexes that drive particular regions of the genome into active transcription or repression[6]. MMSET has a transcriptional repressor activity, with increased H4K20 methylation gene and loss of histone acetylation. Consistent with this repressive activity, MMSET could form a complex with HDAC1 and HDAC2, mSin3a and the histone demethylase LSD1 suggesting that it is a component of transcription co-repressor complexes[7]. Moreover, RE-IIB (IL5 response element II-binding protein) has been recently demonstrated to repress IL-5 expression, through the histone H3-K27 hyper-methylation around promoter region and H3 hypo-acetylation[8].

Collectively, these data suggest that by acting directly as a modifier of chromatin as well as through binding of other chromatin modifying enzymes, MMSET may influence gene expression and potentially acts as a pathogenic agent in MM. The identification of the targets of MMSET and their role in cell growth and survival will be key to understand how MMSET is associated with tumor development[7].

Recent study has identified an aberrant expression of *MMSET* mRNA and protein in glioblastoma multiform compared to normal brain cortex samples using proteomic approach[9]. They demonstrated that MMSET expression increased with ascending tumor proliferation activity. Furthermore, RNA interference blocking *MMSET* expression suppresses glioma cells growth[9].

These data demonstrate that MMSET could be implicated in tumor emergence and/or progression. Therefore, we focussed on *MMSET* expression in various cancers compared to their normal counterparts and in association with staging.

Methods

Databases

We used oncomine cancer microarray database (<http://www.oncomine.org>)[10] to study gene expression of *MMSET* in 40 human tumor types and their normal tissue counterpart as indicated in table 1. To compare the gene expression of a tumor type to its normal counterpart, we used gene expression data from a same study with the same methodology. All data were log transformed, median centered per array, and the standard deviation was normalized to one per array[10] .

Statistical analysis

Statistical comparisons were done with Mann Whitney or student t-test.

Results and discussion

We investigated the expression of *MMSET* in cancer using publicly available gene expression data. The 40 tumor types investigated corresponded to 4 hematological malignancies and 36 solid tumors (Table 1). We found *MMSET* overexpression in all hematological cancers and in 12/36 solid tumors (Table 1 and Supplementary figure S1). Overexpression of *MMSET* was found in glioblastoma compared to normal brain ($P = 1.7E-14$, $P = .009$, $P = .002$)[11; 12; 13]; in hepatocellular carcinoma compared to normal liver ($P = 2.9E-7$, $1.3E-6$) [14; 15]; in head and neck cancer compared to the normal ($P = .008$, $P = 2.7E-5$)[16; 17]; in bladder carcinoma compared to normal bladder ($P = 4.1E-11$, $P = 1.8E-7$)[18; 19]; in primary colon cancer compared to normal adjacent mucosa ($P = 9.5E-6$)[20]; in esophagus adenocarcinoma compared to normal esophagus ($P = .004$)[21]; in breast carcinoma compared to normal breast ($P = 3.8E-$

8)[22]; in T-cell acute lymphoblastic leukemia compared to normal bone marrow ($P = 5.1E-7$)[23]; in B-cell acute lymphoblastic leukaemia compared to normal bone marrow ($P = .001$)[23]; in lung adenocarcinoma compared to normal lung ($P = .009$, $P = 8.4E-4$, $1.7E-6$)[24; 25; 26]; in lymphoma compared to normal B-cell ($P = 3.5E-5$, $7.3E-5$)[27]; in cutaneous melanoma compared to normal melanocyte ($P = 6.46E-13$, $P = .01$, $P = .009$)[28; 29; 30]; in smoldering multiple myeloma compared to normal bone marrow ($P = 7.3E-4$)[31]; in prostate cancer compared to normal prostate ($P = 1.8E-6$, $P = .039$, $P = 5.1E-8$, $P = .002$, $P = .009$, $P = .006$, $P = .009$)[32; 33; 34; 35; 36; 37; 38]; in yolk sac tumor compared to normal testis ($P = .003$)[39]; in ovarian carcinoma compared to normal ovary ($P = .002$, $P = 1.1E-4$)[40; 41] and in clear cell carcinoma compared to normal kidney tissue ($P = .006$)[42].

MMSET has been shown to be highly overexpressed in MM with t(4;14), in association with poor prognosis. Thus, we looked for whether *MMSET* expression could be associated with tumor progression and prognosis. *MMSET* was significantly overexpressed in oligodendroglioma grade III compared to grade II ($P = .009$)[11] (Figure 1) in agreement with previous data[9]; in patients presenting advanced bladder carcinoma (grade II and III) compared to grade I in four independent studies ($P = 4E-6$, $P = .0001$, $P = .004$, $P = .008$)[19; 43; 44; 45]; in breast carcinoma grades II and III compared to grade I in four independent studies ($P = 2.2E-6$, $P = 9.1E-4$, $P = .014$, $P = .017$)[46; 47; 48; 49]; in advanced prostate carcinoma grades VII, VIII and IX compared to grade VI ($P = 1.4E-4$)[50]; in no differentiated hepatocellular carcinoma compared to differentiated hepatocellular carcinoma ($P = 7.2E-5$)[15]; in undifferentiated compared to differentiated head and neck cancer ($P = .002$)[17]; in undifferentiated compared to

differentiated cervical carcinoma[51]; in advanced papillary renal cell carcinoma stages III and IV compared to stages I and II ($P = .0039$)[52](Figure 1).

Furthermore, as it has been demonstrated in MM, *MMSET* expression is associated with bad prognostic in others cancers (Figure 2). *MMSET* is overexpressed in patients with head and neck squamous cell carcinoma dead at 5 years compared to patients alive at 5 years ($P = 1.2E-4$)[53]; in 1 year relapsing MM patient compared to patients with no relapse ($P = 3.2E-4$)[54]; in MM patients alive compared to dead MM patient at one year ($P = .001$)[54]; in prostate carcinoma patients presenting recurrence compared to prostate carcinoma patients with no recurrence ($P = 1.5E-4$) at 5 years [50] and in dead glioma patients compared to alive glioma patients at 3 years ($P = .009$) [55].

Correlations between *MMSET* and HDAC expression in tumor cells

In patients with MM, it was recently demonstrated that *MMSET* could form a complex with HDAC1 and HDAC2, mSin3a and the histone demethylase LSD1 suggesting that *MMSET* is a component of transcription co-repressor complexes. Interestingly, the genes of at least one of these four *MMSET* partners are overexpressed in cancers presenting a *MMSET* overexpression compared to their normal counterpart (Supplementary Table 1). Furthermore, *HDAC1*, *HDAC2* and *LSD1* overexpression are associated with tumor aggressiveness, as *MMSET*, in brain and liver cancers (Supplementary Table 2). Furthermore a high expression of the *MMSET* partners is associated with prognosis in brain cancer (Supplementary Table 3). *HDAC2* and *LSD1* overexpression is also associated with breast and lung progression; and they are associated with prognosis in breast cancer (Supplementary Table 3).

Conclusions:

The analysis reported here demonstrates that *MMSET* mRNA is overexpressed in at least in 15 cancers compared to their normal counterparts, and within a given tumor categories is associated with tumor progression or bad prognosis. The properties of transcriptional co-factor of MMSET appear to be associated with tumorigenesis. The identification of the targets of MMSET and their role in cell growth and survival will be key to understanding how MMSET is associated with tumor development.

Acknowledgements

This work was supported by grants from the Ligue Nationale Contre le Cancer (équipe labellisée), Paris, France, from INCA (n°R07001FN) and from MSCNET European strep (N°E06005FF).

Figure legends

Figure 1: Association between MMSET expression and progression in various cancers

MMSET gene expression in oligodendroglioma [9], bladder[19; 43; 44; 45], breast carcinoma [46; 47; 48; 49], prostate carcinoma[50], in hepatocellular carcinoma [15], head and neck cancer[17], cervical carcinoma[51], papillary renal cell carcinoma[52].

Figure 2: Association of MMSET expression with prognosis

MMSET expression in alive patients with head and neck carcinoma, in dead patients with head and neck carcinoma [53], in MM patients with no relapse, in MM patients with relapse[54], in alive and dead patients with MM [54], in prostate carcinoma patients with no recurrence, prostate carcinoma patients with recurrence [50], in alive and dead patients with glioma [55].

References

- [1] J. Luring, A.M. Abukhdeir, H. Konishi, J.P. Garay, J.P. Gustin, Q. Wang, R.J. Arceci, W. Matsui, and B.H. Park, The multiple myeloma associated MMSET gene contributes to cellular adhesion, clonogenic growth, and tumorigenicity. *Blood* 111 (2008) 856-64.
- [2] P.L. Bergsagel, and W.M. Kuehl, Molecular pathogenesis and a consequent classification of multiple myeloma. *J Clin Oncol* 23 (2005) 6333-8.
- [3] J.J. Keats, T. Reiman, C.A. Maxwell, B.J. Taylor, L.M. Larratt, M.J. Mant, A.R. Belch, and L.M. Pilarski, In multiple myeloma, t(4;14)(p16;q32) is an adverse prognostic factor irrespective of FGFR3 expression. *Blood* 101 (2003) 1520-9.
- [4] J.J. Keats, C.A. Maxwell, B.J. Taylor, M.J. Hendzel, M. Chesi, P.L. Bergsagel, L.M. Larratt, M.J. Mant, T. Reiman, A.R. Belch, and L.M. Pilarski, Overexpression of transcripts originating from the MMSET locus characterizes all t(4;14)(p16;q32)-positive multiple myeloma patients. *Blood* 105 (2005) 4060-9.
- [5] M. Santra, F. Zhan, E. Tian, B. Barlogie, and J. Shaughnessy, Jr., A subset of multiple myeloma harboring the t(4;14)(p16;q32) translocation lacks FGFR3 expression but maintains an IGH/MMSET fusion transcript. *Blood* 101 (2003) 2374-6.
- [6] C.B. Yoo, and P.A. Jones, Epigenetic therapy of cancer: past, present and future. *Nat Rev Drug Discov* 5 (2006) 37-50.
- [7] J. Marango, M. Shimoyama, H. Nishio, J.A. Meyer, D.J. Min, A. Sirulnik, Y. Martinez-Martinez, M. Chesi, P.L. Bergsagel, M.M. Zhou, S. Waxman, B.A. Leibovitch, M.J. Walsh, and J.D. Licht, The MMSET protein is a histone methyltransferase with characteristics of a transcriptional corepressor. *Blood* 111 (2008) 3145-54.
- [8] J.Y. Kim, H.J. Kee, N.W. Choe, S.M. Kim, G.H. Eom, H.J. Baek, H. Kook, and S.B. Seo, Multiple-myeloma-related WHSC1/MMSET isoform RE-IIBP is a histone methyltransferase with transcriptional repression activity. *Mol Cell Biol* 28 (2008) 2023-34.
- [9] J. Li, C. Yin, H. Okamoto, H. Mushlin, B.M. Balgley, C.S. Lee, K. Yuan, B. Ikejiri, S. Glasker, A.O. Vortmeyer, E.H. Oldfield, R.J. Weil, and Z. Zhuang, Identification of a novel proliferation-related protein, WHSC1 4a, in human gliomas. *Neuro Oncol* 10 (2008) 45-51.
- [10] D.R. Rhodes, J. Yu, K. Shanker, N. Deshpande, R. Varambally, D. Ghosh, T. Barrette, A. Pandey, and A.M. Chinnaiyan, ONCOMINE: a cancer microarray database and integrated data-mining platform. *Neoplasia* 6 (2004) 1-6.
- [11] L. Sun, A.M. Hui, Q. Su, A. Vortmeyer, Y. Kotliarov, S. Pastorino, A. Passaniti, J. Menon, J. Walling, R. Bailey, M. Rosenblum, T. Mikkelsen, and H.A. Fine, Neuronal and glioma-derived stem cell factor induces angiogenesis within the brain. *Cancer Cell* 9 (2006) 287-300.
- [12] P.J. French, S.M. Swagemakers, J.H. Nagel, M.C. Kouwenhoven, E. Brouwer, P. van der Spek, T.M. Luider, J.M. Kros, M.J. van den Bent, and P.A. Sillevius Smitt,

- Gene expression profiles associated with treatment response in oligodendrogliomas. *Cancer Res* 65 (2005) 11335-44.
- [13] R. Shai, T. Shi, T.J. Kremen, S. Horvath, L.M. Liao, T.F. Cloughesy, P.S. Mischel, and S.F. Nelson, Gene expression profiling identifies molecular subtypes of gliomas. *Oncogene* 22 (2003) 4918-23.
 - [14] X. Chen, S.T. Cheung, S. So, S.T. Fan, C. Barry, J. Higgins, K.M. Lai, J. Ji, S. Dudoit, I.O. Ng, M. Van De Rijn, D. Botstein, and P.O. Brown, Gene expression patterns in human liver cancers. *Mol Biol Cell* 13 (2002) 1929-39.
 - [15] E. Wurmbach, Y.B. Chen, G. Khitrov, W. Zhang, S. Roayaie, M. Schwartz, I. Fiel, S. Thung, V. Mazzaferro, J. Bruix, E. Bottinger, S. Friedman, S. Waxman, and J.M. Llovet, Genome-wide molecular profiles of HCV-induced dysplasia and hepatocellular carcinoma. *Hepatology* 45 (2007) 938-47.
 - [16] A. Cromer, A. Carles, R. Millon, G. Ganguli, F. Chalmel, F. Lemaire, J. Young, D. Dembele, C. Thibault, D. Muller, O. Poch, J. Abecassis, and B. Wasylyk, Identification of genes associated with tumorigenesis and metastatic potential of hypopharyngeal cancer by microarray analysis. *Oncogene* 23 (2004) 2484-98.
 - [17] D. Pyeon, M.A. Newton, P.F. Lambert, J.A. den Boon, S. Sengupta, C.J. Marsit, C.D. Woodworth, J.P. Connor, T.H. Haugen, E.M. Smith, K.T. Kelsey, L.P. Turek, and P. Ahlquist, Fundamental differences in cell cycle deregulation in human papillomavirus-positive and human papillomavirus-negative head/neck and cervical cancers. *Cancer Res* 67 (2007) 4605-19.
 - [18] M. Sanchez-Carbayo, N.D. Socci, J. Lozano, F. Saint, and C. Cordon-Cardo, Defining molecular profiles of poor outcome in patients with invasive bladder cancer using oligonucleotide microarrays. *J Clin Oncol* 24 (2006) 778-89.
 - [19] L. Dyrskjot, M. Kruhoffer, T. Thykjaer, N. Marcussen, J.L. Jensen, K. Moller, and T.F. Orntoft, Gene expression in the urinary bladder: a common carcinoma in situ gene expression signature exists disregarding histopathological classification. *Cancer Res* 64 (2004) 4040-8.
 - [20] D.H. Ki, H.C. Jeung, C.H. Park, S.H. Kang, G.Y. Lee, W.S. Lee, N.K. Kim, H.C. Chung, and S.Y. Rha, Whole genome analysis for liver metastasis gene signatures in colorectal cancer. *Int J Cancer* 121 (2007) 2005-12.
 - [21] E.T. Kimchi, M.C. Posner, J.O. Park, T.E. Darga, M. Kocherginsky, T. Karrison, J. Hart, K.D. Smith, J.J. Mezhir, R.R. Weichselbaum, and N.N. Khodarev, Progression of Barrett's metaplasia to adenocarcinoma is associated with the suppression of the transcriptional programs of epidermal differentiation. *Cancer Res* 65 (2005) 3146-54.
 - [22] A.L. Richardson, Z.C. Wang, A. De Nicolo, X. Lu, M. Brown, A. Miron, X. Liao, J.D. Iglehart, D.M. Livingston, and S. Ganesan, X chromosomal abnormalities in basal-like human breast cancer. *Cancer Cell* 9 (2006) 121-32.
 - [23] A. Andersson, C. Ritz, D. Lindgren, P. Eden, C. Lassen, J. Heldrup, T. Olofsson, J. Rade, M. Fontes, A. Porwit-Macdonald, M. Behrendtz, M. Hoglund, B. Johansson, and T. Fioretos, Microarray-based classification of a consecutive series of 121 childhood acute leukemias: prediction of leukemic and genetic subtype as well as of minimal residual disease status. *Leukemia* 21 (2007) 1198-203.
 - [24] A. Bhattacharjee, W.G. Richards, J. Staunton, C. Li, S. Monti, P. Vasa, C. Ladd, J. Beheshti, R. Bueno, M. Gillette, M. Loda, G. Weber, E.J. Mark, E.S. Lander, W.

- Wong, B.E. Johnson, T.R. Golub, D.J. Sugarbaker, and M. Meyerson, Classification of human lung carcinomas by mRNA expression profiling reveals distinct adenocarcinoma subclasses. *Proc Natl Acad Sci U S A* 98 (2001) 13790-5.
- [25] S. Wachi, K. Yoneda, and R. Wu, Interactome-transcriptome analysis reveals the high centrality of genes differentially expressed in lung cancer tissues. *Bioinformatics* 21 (2005) 4205-8.
- [26] L.J. Su, C.W. Chang, Y.C. Wu, K.C. Chen, C.J. Lin, S.C. Liang, C.H. Lin, J. Whang-Peng, S.L. Hsu, C.H. Chen, and C.Y. Huang, Selection of DDX5 as a novel internal control for Q-RT-PCR from microarray data using a block bootstrap re-sampling scheme. *BMC Genomics* 8 (2007) 140.
- [27] K. Basso, A.A. Margolin, G. Stolovitzky, U. Klein, R. Dalla-Favera, and A. Califano, Reverse engineering of regulatory networks in human B cells. *Nat Genet* 37 (2005) 382-90.
- [28] K.S. Hoek, N.C. Schlegel, P. Brafford, A. Sucker, S. Ugurel, R. Kumar, B.L. Weber, K.L. Nathanson, D.J. Phillips, M. Herlyn, D. Schadendorf, and R. Dummer, Metastatic potential of melanomas defined by specific gene expression profiles with no BRAF signature. *Pigment Cell Res* 19 (2006) 290-302.
- [29] D. Talantov, A. Mazumder, J.X. Yu, T. Briggs, Y. Jiang, J. Backus, D. Atkins, and Y. Wang, Novel genes associated with malignant melanoma but not benign melanocytic lesions. *Clin Cancer Res* 11 (2005) 7234-42.
- [30] C. Haqq, M. Nosrati, D. Sudilovsky, J. Crothers, D. Khodabakhsh, B.L. Pulliam, S. Federman, J.R. Miller, 3rd, R.E. Allen, M.I. Singer, S.P. Leong, B.M. Ljung, R.W. Sagebiel, and M. Kashani-Sabet, The gene expression signatures of melanoma progression. *Proc Natl Acad Sci U S A* 102 (2005) 6092-7.
- [31] F. Zhan, B. Barlogie, V. Arzoumanian, Y. Huang, D.R. Williams, K. Hollmig, M. Pineda-Roman, G. Tricot, F. van Rhee, M. Zangari, M. Dhodapkar, and J.D. Shaughnessy, Jr., Gene-expression signature of benign monoclonal gammopathy evident in multiple myeloma is linked to good prognosis. *Blood* 109 (2007) 1692-700.
- [32] J. Lapointe, C. Li, J.P. Higgins, M. van de Rijn, E. Bair, K. Montgomery, M. Ferrari, L. Egevad, W. Rayford, U. Bergerheim, P. Ekman, A.M. DeMarzo, R. Tibshirani, D. Botstein, P.O. Brown, J.D. Brooks, and J.R. Pollack, Gene expression profiling identifies clinically relevant subtypes of prostate cancer. *Proc Natl Acad Sci U S A* 101 (2004) 811-6.
- [33] S. Nanni, C. Priolo, A. Grasselli, M. D'Eletto, R. Merola, F. Moretti, M. Gallucci, P. De Carli, S. Sentinelli, A.M. Cianciulli, M. Mottolose, P. Carlini, D. Arcelli, M. Helmer-Citterich, C. Gaetano, M. Loda, A. Pontecorvi, S. Bacchetti, A. Sacchi, and A. Farsetti, Epithelial-restricted gene profile of primary cultures from human prostate tumors: a molecular approach to predict clinical behavior of prostate cancer. *Mol Cancer Res* 4 (2006) 79-92.
- [34] S.M. Dhanasekaran, A. Dash, J. Yu, I.P. Maine, B. Laxman, S.A. Tomlins, C.J. Creighton, A. Menon, M.A. Rubin, and A.M. Chinnaiyan, Molecular profiling of human prostate tissues: insights into gene expression patterns of prostate development during puberty. *FASEB J* 19 (2005) 243-5.

- [35] D.K. Vanaja, J.C. Cheville, S.J. Iturria, and C.Y. Young, Transcriptional silencing of zinc finger protein 185 identified by expression profiling is associated with prostate cancer progression. *Cancer Res* 63 (2003) 3877-82.
- [36] S.A. Tomlins, R. Mehra, D.R. Rhodes, X. Cao, L. Wang, S.M. Dhanasekaran, S. Kalyana-Sundaram, J.T. Wei, M.A. Rubin, K.J. Pienta, R.B. Shah, and A.M. Chinnaiyan, Integrative molecular concept modeling of prostate cancer progression. *Nat Genet* 39 (2007) 41-51.
- [37] Y.P. Yu, D. Landsittel, L. Jing, J. Nelson, B. Ren, L. Liu, C. McDonald, R. Thomas, R. Dhir, S. Finkelstein, G. Michalopoulos, M. Becich, and J.H. Luo, Gene expression alterations in prostate cancer predicting tumor aggression and preceding development of malignancy. *J Clin Oncol* 22 (2004) 2790-9.
- [38] P. Liu, S. Ramachandran, M. Ali Seyed, C.D. Scharer, N. Laycock, W.B. Dalton, H. Williams, S. Karanam, M.W. Datta, D.L. Jaye, and C.S. Moreno, Sex-determining region Y box 4 is a transforming oncogene in human prostate cancer cells. *Cancer Res* 66 (2006) 4011-9.
- [39] R.I. Skotheim, G.E. Lind, O. Monni, J.M. Nesland, V.M. Abeler, S.D. Fossa, N. Duale, G. Brunborg, O. Kallioniemi, P.W. Andrews, and R.A. Lothe, Differentiation of human embryonal carcinomas in vitro and in vivo reveals expression profiles relevant to normal development. *Cancer Res* 65 (2005) 5588-98.
- [40] N.D. Hendrix, R. Wu, R. Kuick, D.R. Schwartz, E.R. Fearon, and K.R. Cho, Fibroblast growth factor 9 has oncogenic activity and is a downstream target of Wnt signaling in ovarian endometrioid adenocarcinomas. *Cancer Res* 66 (2006) 1354-62.
- [41] K.H. Lu, A.P. Patterson, L. Wang, R.T. Marquez, E.N. Atkinson, K.A. Baggerly, L.R. Ramoth, D.G. Rosen, J. Liu, I. Hellstrom, D. Smith, L. Hartmann, D. Fishman, A. Berchuck, R. Schmandt, R. Whitaker, D.M. Gershenson, G.B. Mills, and R.C. Bast, Jr., Selection of potential markers for epithelial ovarian cancer with gene expression arrays and recursive descent partition analysis. *Clin Cancer Res* 10 (2004) 3291-300.
- [42] M.L. Gumz, H. Zou, P.A. Kreinest, A.C. Childs, L.S. Belmonte, S.N. LeGrand, K.J. Wu, B.A. Luxon, M. Sinha, A.S. Parker, L.Z. Sun, D.A. Ahlquist, C.G. Wood, and J.A. Copland, Secreted frizzled-related protein 1 loss contributes to tumor phenotype of clear cell renal cell carcinoma. *Clin Cancer Res* 13 (2007) 4740-9.
- [43] D. Lindgren, F. Liedberg, A. Andersson, G. Chebil, S. Gudjonsson, A. Borg, W. Mansson, T. Fioretos, and M. Hoglund, Molecular characterization of early-stage bladder carcinomas by expression profiles, FGFR3 mutation status, and loss of 9q. *Oncogene* 25 (2006) 2685-96.
- [44] N. Stransky, C. Vallot, F. Rey, I. Bernard-Pierrot, S.G. de Medina, R. Segraves, Y. de Rycke, P. Elvin, A. Cassidy, C. Spraggon, A. Graham, J. Southgate, B. Asselain, Y. Allory, C.C. Abbou, D.G. Albertson, J.P. Thiery, D.K. Chopin, D. Pinkel, and F. Radvanyi, Regional copy number-independent deregulation of transcription in cancer. *Nat Genet* 38 (2006) 1386-96.
- [45] L. Dyrskjot, K. Zieger, M. Kruhoffer, T. Thykjaer, J.L. Jensen, H. Primdahl, N. Aziz, N. Marcussen, K. Moller, and T.F. Orntoft, A molecular signature in superficial bladder carcinoma predicts clinical outcome. *Clin Cancer Res* 11 (2005) 4029-36.
- [46] A.V. Ivshina, J. George, O. Senko, B. Mow, T.C. Putti, J. Smeds, T. Lindahl, Y. Pawitan, P. Hall, H. Nordgren, J.E. Wong, E.T. Liu, J. Bergh, V.A. Kuznetsov, and

- L.D. Miller, Genetic reclassification of histologic grade delineates new clinical subtypes of breast cancer. *Cancer Res* 66 (2006) 10292-301.
- [47] L.D. Miller, J. Smeds, J. George, V.B. Vega, L. Vergara, A. Ploner, Y. Pawitan, P. Hall, S. Klaar, E.T. Liu, and J. Bergh, An expression signature for p53 status in human breast cancer predicts mutation status, transcriptional effects, and patient survival. *Proc Natl Acad Sci U S A* 102 (2005) 13550-5.
- [48] C. Ginestier, N. Cervera, P. Finetti, S. Esteyries, B. Esterni, J. Adelaide, L. Xerri, P. Viens, J. Jacquemier, E. Charafe-Jauffret, M. Chaffanet, D. Birnbaum, and F. Bertucci, Prognosis and gene expression profiling of 20q13-amplified breast cancers. *Clin Cancer Res* 12 (2006) 4533-44.
- [49] H. Zhao, A. Langerod, Y. Ji, K.W. Nowels, J.M. Nesland, R. Tibshirani, I.K. Bukholm, R. Karesen, D. Botstein, A.L. Borresen-Dale, and S.S. Jeffrey, Different gene expression patterns in invasive lobular and ductal carcinomas of the breast. *Mol Biol Cell* 15 (2004) 2523-36.
- [50] G.V. Glinsky, A.B. Glinskii, A.J. Stephenson, R.M. Hoffman, and W.L. Gerald, Gene expression profiling predicts clinical outcome of prostate cancer. *J Clin Invest* 113 (2004) 913-23.
- [51] B. Bachtary, P.C. Boutros, M. Pintilie, W. Shi, C. Bastianutto, J.H. Li, J. Schwock, W. Zhang, L.Z. Penn, I. Jurisica, A. Fyles, and F.F. Liu, Gene expression profiling in cervical cancer: an exploration of intratumor heterogeneity. *Clin Cancer Res* 12 (2006) 5632-40.
- [52] X.J. Yang, M.H. Tan, H.L. Kim, J.A. Ditlev, M.W. Betten, C.E. Png, E.J. Kort, K. Futami, K.A. Furge, M. Takahashi, H.O. Kanayama, P.H. Tan, B.S. Teh, C. Luan, K. Wang, M. Pins, M. Tretiakova, J. Anema, R. Kahnoski, T. Nicol, W. Stadler, N.G. Vogelzang, R. Amato, D. Seligson, R. Figlin, A. Belldegrun, C.G. Rogers, and B.T. Teh, A molecular classification of papillary renal cell carcinoma. *Cancer Res* 65 (2005) 5628-37.
- [53] C.H. Chung, J.S. Parker, G. Karaca, J. Wu, W.K. Funkhouser, D. Moore, D. Butterfoss, D. Xiang, A. Zanation, X. Yin, W.W. Shockley, M.C. Weissler, L.G. Dressler, C.G. Shores, W.G. Yarbrough, and C.M. Perou, Molecular classification of head and neck squamous cell carcinomas using patterns of gene expression. *Cancer Cell* 5 (2004) 489-500.
- [54] D.R. Carrasco, G. Tonon, Y. Huang, Y. Zhang, R. Sinha, B. Feng, J.P. Stewart, F. Zhan, D. Khatry, M. Protopopova, A. Protopopov, K. Sukhdeo, I. Hanamura, O. Stephens, B. Barlogie, K.C. Anderson, L. Chin, J.D. Shaughnessy, Jr., C. Brennan, and R.A. Depinho, High-resolution genomic profiles define distinct clinico-pathogenetic subgroups of multiple myeloma patients. *Cancer Cell* 9 (2006) 313-25.
- [55] W.A. Freije, F.E. Castro-Vargas, Z. Fang, S. Horvath, T. Cloughesy, L.M. Liao, P.S. Mischel, and S.F. Nelson, Gene expression profiling of gliomas strongly predicts survival. *Cancer Res* 64 (2004) 6503-10.

Table 1

		Gene overexpression compared to normal tissue counterpart				
Tissue	Datasets	<i>MMSET</i>	<i>HDAC1</i>	<i>HDAC2</i>	<i>LSD1</i>	<i>SIN3A</i>
Myeloma	8	<u>Yes</u>	No	<u>Yes</u>	No	Yes
Leukemia	33	<u>Yes</u>	Yes	Yes	Yes	No
Lymphoma	16	<u>Yes</u>	<u>Yes</u>	Yes	Yes	No
Bladder	8	<u>Yes</u>	<u>Yes</u>	Yes	Yes	No
Blood	2	No	No	No	No	No
Brain	23	<u>Yes</u>	<u>Yes</u>	Yes	Yes	No
Breast	44	<u>Yes</u>	No	Yes	No	No
Cervix	1	No	No	No	No	No
Chondrosarcoma	1	No	No	No	No	No
Colon	12	<u>Yes</u>	No	Yes	No	No
Endocrine	1	No	No	No	No	No
Endometrium	4	No	Yes	No	No	No
Esophagus	4	<u>Yes</u>	<u>No</u>	Yes	No	No
Gastric	5	No	No	Yes	No	No
Head & Neck	5	<u>Yes</u>	<u>Yes</u>	Yes	Yes	Yes
Liver	4	<u>Yes</u>	Yes	Yes	Yes	No
Lung	16	<u>Yes</u>	Yes	Yes	Yes	No
Melanoma	10	<u>Yes</u>	Yes	Yes	Yes	No
Mesothelioma	3	No	No	No	No	No
Muscle	2	No	No	No	No	No
Neuroblastoma	2	No	No	No	No	No
Oral	1	No	<u>No</u>	Yes	No	No
Ovarian	14	<u>Yes</u>	Yes	No	No	No
Pancreas	6	No	<u>Yes</u>	No	No	No
Parathyroid	1	No	No	No	No	No
Prostate	20	Yes	Yes	Yes	Yes	No
Rectum	2	No	No	No	No	No
Renal	11	<u>Yes</u>	No	No	Yes	No
Salivary Gland	1	No	No	Yes	No	No
Sarcoma	11	No	No	No	No	No
Seminoma	1	No	yes	No	No	No
Skin	1	No	No	No	No	No
Testis	1	<u>Yes</u>	<u>No</u>	No	No	Yes
Thyroid	1	No	No	<u>No</u>	No	No
Uterus	1	No	No	No	No	No

Table 1

Expression of *MMSET*, *HDAC1*, *HDAC2*, *LSD1*, *Sin3A* in human tumor types to that of their normal tissue counterparts using publicly available gene expression data, including the Oncomine Cancer Microarray database.

Figure 1

Figure 1 : Association of MMSET expression with tumor grade

Cervical Carcinoma - Differentiation

Bachtiary et al. Clin Cancer Res 2006

Papillary Renal Cell Carcinoma - Stage

Yang et al. Cancer Res 2005

Figure 2

Figure 2 : Association of MMSET expression with prognosis

Head and Neck Squamous Cell Carcinoma Overall Survival - 5 Years

Chung et al. Cancer Cell 2004

Multiple Myeloma - Relapse Status - 1 Year

Carasco et al. Cancer Cell 2006

Multiple Myeloma - Survival - 1 Year

Carasco et al. Cancer Cell 2006

Prostate Carcinoma - Biochemical Recurrence - 5 Years

Glinsky et al. J Clin Invest 2004

Glioma - Survival - 3 Years

Freije et al. Cancer Res 2006