Pathological lesions in colonic biopsies during Parkinson’s disease.
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CD68 positive cells, were quite evenly distributed in muscle layer of the gallbladder in placebo-treated gallstones patients (A) and were scarcely present in gallbladder of ursodeoxycholic acid (UDCA)-treated patients (B). A similar pattern was exhibited by COX-2 positive cells, in muscle layer of gallbladder in placebo-treated patients (C) and UDCA-treated patients (D). A/C, B/D refer to the same patient. The figure shows corresponding fields for each patient. Original magnification, ×200. High power field, ×400.

Pathological lesions in colonic biopsies during Parkinson’s disease

Parkinson’s disease (PD) is a neurodegenerative condition that affects 1% of the population over 65 years of age. The two pathological hallmarks of PD are a loss of dopaminergic neurons in the substantia nigra (SN) and the presence of cytoplasmic eosinophilic inclusions termed Lewy bodies (LBs), whose main component is phosphorylated α-synuclein. This degeneration of SN neurons leads to a dopamine deficiency

Table 1 Mean number (with SD in parentheses) of positively stained cells in 10 consecutive microscopic fields

<table>
<thead>
<tr>
<th></th>
<th>Controls</th>
<th>Placebo</th>
<th>UDCA</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD68</td>
<td>11.9 (8.1)</td>
<td>36.2 (11)*</td>
<td>19.6 (6)**</td>
</tr>
<tr>
<td>COX-2</td>
<td>10.5 (6.8)</td>
<td>30.6 (12)*</td>
<td>15.2 (5.5)**</td>
</tr>
</tbody>
</table>

*p < 0.001 vs controls; **p < 0.01 vs placebo.

COX-2, cyclooxygenase-2; UDCA, ursodeoxycholic acid.

REFERENCES


(Šanofi-Winthrop, Riells y Viabrea, Spain) for 30 days. Criteria for eligibility, randomisation of patients and blinding were those used in our previous randomised study. Following laparoscopic cholecystectomy, GB specimens were collected for routine histology and immunohistochemistry, the latter performed by the streptavidin–biotin method. Mouse monoclonal antibodies against COX-2 protein (clone CX-294; Dako, Glostrup, Denmark) and mouse monoclonal antibodies against CD68 (clone M7103; Dako) were used. Part of the GB tissue was collected for cell muscle contract to cholecystokinin 8 (CCK-8) studies as well as for measurement of prostaglandin (PG) E2 levels as described elsewhere.

Following randomisation, eight patients received UDCA and 11 received placebo, for 30 days. Staining with haematoxylin & eosin revealed chronic cholecystitis, with eosinophilic inclusions termed Lewy bodies.
During PD, autopsy specimens have shown that the various reports performed on surgical or colonic biopsies and (2) the presence of lesions (1) the ability to study the ENS using routine technique could be a reliable tool to detect to separate the submucosa (containing the internal submucosal plexus) from the mucosa. The submucosa were then fixed in 4% paraformaldehyde. Immunohistochemical studies were then performed on these tissues using a combination of antibodies against rabbit anti-tyrosine hydroxylase (TH) (1:500, Pel-Freez, Rogers, Arkansas, USA), rabbit anti-dopamine-β-hydroxylase (DBH) (1:250, Millipore, Saint-Quentin-en-Yvelines, France), mouse anti-Hu C/D (1:200, Invitrogen, Cergy-Pontoise, France), rabbit anti-phosphorylated α-synuclein (1.5000, WAKO, Osaka, Japan) or rabbit anti-neurofilament 200 kDa (1.250; Millipore) as previously described.3

In control patients, individual biopsies contained 11.2 (SD 7.9) ganglia and each ganglia contained 5.6 (SD 1.9) Hu-immunoreactive (IR) neurons. In PD patients, the number of ganglia per biopsies was similar to controls (15.6 (SD 5.3); p = 0.22). In addition, the number of Hu-IR neurons per ganglion in PD was unchanged as compared to controls (7.0 (SD 1.6); p = 0.25) (fig 1A,C). Constipated controls did not differ from PD patients in the number of ganglia per biopsy (11.3 (SD 1.5); p = 0.57) or in the number of neurons per ganglion (5.5 (SD 0.6); p = 0.19) (fig 1C).

In healthy controls, 11.6 (SD 5.0)% of Hu-IR neurons were TH-IR. In PD patients, the proportion of TH-IR neurons was unchanged as compared to controls (12.3 (SD 3.3)%; p = 0.80) (fig 1D–F). In constipated patients, the proportion of TH-IR neurons was similar to the one of PD patients (8. (SD 2.7)%; p = 0.12) (fig 1F). In all groups no neuronal body was DBH positive, suggesting that all TH-IR neurons in the submucosal plexus were dopaminergic. These results are consistent with a previous report by Singaram et al5 showing the absence of loss of TH-IR neurons in the submucosal and myenteric plexuses of PD patients, suggesting that it is not a marker of choice for detecting PD lesions in the ENS.

However, immunohistochemical staining with an antibody against phosphorylated α-synuclein, revealed that 4 out of 5 PD patients had phospho-α-synuclein-IR neurites (identified with neurofilament (NF) in the submucosa (fig 2A,F). These phospho-α-synuclein-IR neurites were absent in both control and constipated patients. In some cases, large aggregates were observed in dystrophic NF-IR neurites (fig 2E), a pattern reminiscent of Lewy neurites.

Taken together, our pilot study showed that routine colonic biopsies can be used to study the submucosal plexus of the ENS. In addition, we identified for the first time in the gut of living PD patients lesions similar to the ones observed in the brain. This technique could be a reliable tool to detect early lesions in the gut during the course of PD in order to better understand the pathogenesis of the disease and/or to identify novel biomarkers.

We therefore performed routine colonic biopsies in five PD patients complaining of functional constipation (63 (SD 7) years, three men; all had disease duration >5 years). Five healthy age-matched patients (61 (SD 6.5) years, one man) requiring a total colonoscopy for colorectal cancer screening were included as controls. They had no known neurological disease. None suffered from functional digestive symptoms. In order to avoid any specific role for chronic constipation, we included three additional non-age-matched patients (52 (SD 5) years, no men) who underwent total colonoscopy for the assessment of a chronic intractable constipation as additional controls. Written consent was obtained according to the principles of the Declaration of Helsinki.

Four biopsies were taken from the ascending colon during colonoscopy. Biopsies were performed using standard biopsy forceps without needles (FB210K; Olympus, Tokyo, Japan). Samples were immediately immersed in 4˚C saline solution and microdissected in order to separate the submucosa (containing the internal submucosal plexus) from the mucosa. The submucosa were then fixed in 4% paraformaldehyde. Immunohistochemical studies were then performed on these tissues using a combination of antibodies against rabbit anti-tyrosine hydroxylase (TH) (1:5000, Pel-Freez, Rogers, Arkansas, USA), rabbit anti-dopamine-β-hydroxylase (DBH) (1:250, Millipore, Saint-Quentin-en-Yvelines, France), mouse anti-Hu C/D (1:200, Invitrogen, Cergy-Pontoise, France), rabbit anti-phosphorylated α-synuclein (1.5000, WAKO, Osaka, Japan) or rabbit anti-neurofilament 200 kDa (1.250; Millipore) as previously described.3

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**Ethics approval:** The study protocol was approved by the local Committee on Ethics and Human Research on 27 February 2007. TC and TL as well as PD and MN contributed equally to this work.


**REFERENCES**


**CORRECTION**

doi:10.1136/gut.2007.119446corr1

R Spiller, Q Aziz, F Creed, et al. Guidelines on the irritable bowel syndrome: mechanisms and practical management (*Gut* 2007;56:1770–98). In paragraph 4.4.1 the sentence “This in turn acts on the adrenal medulla, resulting in cortisol secretion into the circulation” should read “This in turn acts on the adrenal cortex, resulting in cortisol secretion into the circulation”.

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**Figure 2** Phospho-α-synuclein-positive submucosal neurites differentiate Parkinson’s disease patients from controls. Double labelling with antibodies against neurofilament (NF) (A,B) and phosphorylated α-synuclein (C,D) revealed that some NF-immunoreactive (IR) neuritic structures were also phospho-α-synuclein-IR (merged image in E,F) in the majority of Parkinson’s disease patients, but in none of the controls. Occasionally the inclusion-bearing neurites displayed dystrophic alterations (A,C,E). Scale bar: 30 μm.

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**ANSWER**

From the question on page 1673

The patient had a large inflammatory abdominal aortic aneurysm. The abdominal CT scan shows a large infrarenal aortic aneurysm with a maximum diameter of 7.5 cm extending into the iliac vessels. There is an enhancing soft-tissue cuff surrounding the anterolateral margin of the aneurysm. The aneurysm appears to compress the third part of the duodenum (fig 1 below), which, however, was not detected at endoscopy. These CT findings were suggestive of an inflammatory aneurysm. Inflammatory abdominal aortic aneurysms represent 3–10% of all abdominal aortic aneurysms and occur predominantly in men. They differ from atherosclerotic aneurysms in that patients often present with abdominal symptoms or anorexia, weight loss, and raised inflammatory markers. CT has a specificity of 99.7% for diagnosis of inflammatory aneurysms, usually showing periaortic fibrosis as a cuff of enhancing soft tissue surrounding the anterolateral margin of the aneurysm. If periaortic fibrosis is extensive, adjacent abdominal structures may be compressed and adherent, most commonly the third part of the duodenum. Although rare, inflammatory abdominal aortic aneurysms should be kept in mind as a cause of abdominal pain and/or anorexia, weight loss, and raised inflammatory markers. The natural history of inflammatory abdominal aortic aneurysms remains unknown, with 3.3–14% patients presenting with acute or chronic rupture. As regards to management, the literature supports an operative approach with a 30 day operative mortality rate of up to 9%. Complete regression of fibrosis and inflammatory process occurs in up to one-half of patients at long-term follow-up post-operatively. Clinical symptoms (such as weight loss and gastrointestinal symptoms) reverse in 93% of the patients after an operation. Endovascular therapy is also a potential treatment...

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**Editor’s quiz: GI snapshot**