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Oxytocin-induced postinhibitory rebound firing facilitates bursting activity in oxytocin neurons

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ABSTRACT

During parturition and lactation, neurosecretory oxytocin (OT) neurons in the hypothalamus achieve pulsatile hormone secretion by coordinated bursts of firing that occur throughout the neuronal population. This activity is partly controlled by somatodendritic release of OT which facilitates the onset and recurrence of synchronized bursting. To further investigate the cellular mechanisms underlying the control exerted by OT on the activity of its own neurons, we studied the effects of the peptide on membrane potential and synaptic activity in OT neurons in hypothalamic organotypic slice cultures. Bath application of low concentrations of OT (<100 nM) facilitated GABA-A receptor-mediated inhibitory transmission through a presynaptic mechanism without affecting membrane potential and excitatory glutamatergic synaptic activity. The facilitatory action of OT on GABAergic transmission was dose-dependent, starting at 25 nM and disappearing at concentrations above 100 nM. As previously shown, higher concentrations of OT (>500 nM) had the opposite effect, inhibiting GABA-A receptors via a postsynaptic mechanism. Surprisingly, OT-mediated facilitation of GABAergic transmission promoted action potential firing in 40% of the neurons. Each action potential occurred at the end of the repolarizing phase of an inhibitory potential. Pharmacological dissection revealed that this firing involved the activation of low-threshold activated calcium channels. Detailed statistical analysis showed that OT-mediated firing up-regulated bursting activity in OT neurons. It is thus likely to optimize OT secretion and, as a consequence, facilitate delivery and milk ejection in mammals.

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INTRODUCTION

Oxytocin (OT) is a hormone synthesized in magnocellular neurons that are located in the paraventricular (PVN) and supraoptic (SON) nuclei of the hypothalamus. During parturition and lactation in the rat, OT neurons display periodic high frequency bursts of action potentials (AP) that are synchronized in the whole OT neuron population. This triggers a massive and pulsatile release of OT in the blood stream which, in turn, promotes pup delivery and milk ejection (Poulain and Wakerley,1982; Wakerley et al.,1988). OT is also released from the somatodendritic compartment, a process that enables OT neurons to regulate their own activity (Richard et al., 1991). Locally released OT is essential to the onset of the milk ejection reflex (Moos et al., 1989; Neumann et al., 1993) and it enhances the amplitude and frequency of suckling-induced bursts, an effect mimicked by injections of OT in the 3rd ventricle (Freund-Mercier and Richard,1984). Conversely, injection of an OT receptor (OT-R) antagonist directly in the SON or PVN greatly reduces bursting activity (Lambert et al., 1993). Furthermore, OT appears to have the property of priming and inducing its own release, thereby amplifying its local and long distance action (Moos et al., 1984; Ludwig and Leng, 2006).

In vitro recordings have revealed that the periodic high frequency bursting of OT neurons was driven by OT-sensitive glutamatergic inputs (Jourdain et al., 1998; Israel et al., 2003). In addition, OT at fairly high concentrations (1-10 μM) is known to inhibit glutamate (Kombian et al., 1997) and GABA release (De Kock et al., 2003) through presynaptic mechanisms as well as GABA-A receptor-mediated responses via a postsynaptic process (Brussaard et al.,1996). How these different effects come into play to modulate OT neuron excitability, especially during lactation, remains unknown. To address this issue we investigated the effects of OT applied at very low to large concentration (25 to 2000 nM) on identified OT neurons in vitro (Jourdain et al., 1996). Low concentration of

OT (<100 nM) did not affect membrane potential and excitatory postsynaptic activity but triggered or accelerated GABA-A receptor mediated synaptic responses, through a presynaptic action. In about 40% of OT neurons, the enhancement of inhibitory transmission had the unexpected consequence of facilitating AP firing through a postinhibitory rebound (PIR) following individual inhibitory postsynaptic potentials and involving the activation of a low voltage-activated calcium current, as revealed by pharmacological analysis. We found that OT-mediated PIR firing was responsible for the increase in firing rate observed in OT neurons prior to burst occurrence, an increase tightly correlated to burst amplitude, thus providing a further mechanism for optimizing hormone output during pup delivery and lactation in mammals.

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MATERIAL AND METHODS

Slice preparation

Cultured slices were prepared using the roller tube method, as described previously (Jourdain et al., 1996). Briefly, 4- to 6-day-old female Wistar rats were anaesthetized with isoflurane (95% O2 and 5 % isoflurane) for 1 minute and decapitated. Brains were removed, and tissue blocks that included the hypothalamus were quickly dissected and sectioned (400 µm). Frontal slices containing the supraoptic nucleus (SON) were cut into two parts along the third ventricle, and each part was placed on a glass coverslip coated with heparinized chicken plasma. Thrombin was then added to the coverslip to coagulate the plasma and permit adhesion of the slice to the coverslip. The coverslip was inserted into a plastic flat-bottomed tube (Nunc, Roskilde, Denmark) containing 750 µl of medium (pH 7.4; 295 mosmol/kg), composed of 50% Eagle's basal medium (Life Technologies, Gaithersburg, MD), 25% heat-inactivated horse serum (Life Technologies), and 25% HBSS (Life Technologies) enriched with glucose (7.5 mg /ml) and 2 mM L-glutamate (Seromed, Berlin, Germany). No antibiotics were used. The tubes were tightly capped and inserted in a roller drum; the tubes were rotated approximately 15 turns/hr. The medium was replaced twice a week.

Recordings were performed in 2-10 week-old cultures using a temperature-controlled chamber (36.0±0.2 °C) perifused with a solution containing (in mM): NaCl 125; KCl 3; MgSO4 1; KH_2PO_4 1.25; $NaHCO_3$ 5; $CaCl_2$ 2; glucose 5; HEPES 10 (pH 7.25; 290-295 mosmol/kg). Intracellular microelectrodes were filled with 1 M potassium acetate and 1 % biocytin (Sigma). Electrode resistance varied from 150 to 250 Mohms. The patch clamp technique was used in whole cell configuration (current or voltage clamp mode) using electrodes (4-8 Mohms) filled with a solution containing (in mM) 120 K-gluconate, 20 KCl, 10 HEPES, 1 EGTA, 1.3 $MgCl_2$, 0.1 $CaCl_2$, 2 Mg-ATP and 0.3 GTP. For IPSC recording, electrodes were filled with (in mM): 141 CsCl, 10 HEPES, 5 QX-314-Cl and 2 Mg-ATP.

Series resistance (10-25 Mohms) was monitored on line and cells were excluded if more than 20% change occurred during the experiment.

Signals were filtered at 2 kHz, digitized at 5 kHz and analyzed using pClamp 9 (Molecular Device, USA). Firing rate preceding high frequency burst was estimated from frequency histograms calculated over 0.5 s integration periods and plotted versus time using pClamp9. Pre-burst period was defined as the period occurring 20 s before burst incidence. A change in basal firing frequency was considered as significantly different when changes exceeded 10% of control values measured during the 200 s period preceding the pre-burst period (Gouzènes et al, 1998). Detection of synaptic events was achieved off-line using a sliding template whereas action potentials were detected using an amplitude threshold (AxoGraph Scientific, USA). An action potential was considered triggered by an IPSP if occurring within 300 ms of IPSP onset. Values are expressed as means ± SD. Data obtained were compared statistically with the nonparametric Kolmogorov-Smirnov test or the paired Student's test.

Drugs

The following were added to the bath medium when required: synthetic OT (Peninsula), the OT-R agonist, [4-threonine, 7-glycine]-oxytocin ([Thr4, Gly7]-OT ([4-7] OT), the OT-R antagonist, desGly-NH2d(CH2)5[-DTyr2,Thr4]OVT (d-OVT; gifts from Dr. Manning), 6-cyano-7-nitroquinoxaline-2,3-dione (CNQX, RBI), D(-)-2-amino-5-phosphonopentanoic acid (AP5), ZD 7288 (Tocris), bicuculline, CsCl, picrotoxin and tetrodotoxin, mibefradil (Sigma). GABA (Sigma) was dissolved in normal medium at 0.5 mM and was locally delivered through a micropipette (1-2 µm in diameter) positioned at 50-100 µm from the tested cell and connected to a pneumatic ejection system (Picospritzer, Intracel Ltd, UK).

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Identification of recorded neurons

At the end of the recording, neurons were filled with biocytin (1%) using hyperpolarizing current pulses. This was not necessary for patch clamp recording. Slices were then fixed in 4% paraformaldehyde and 0.15% picric acid for 2 h at room temperature and rinsed in 4% paraformaldehyde (2 x 20 min). Biocytin was visualized with streptavidin-conjugated Texas Red fluorescence (Biosys, Compiègne, France) with appropriate filters (Leitz DMR microscope, Leica, Paris). Slices then underwent double immunofluorescence for OT or vasopressin, using a mixture of primary antibodies, one being a monoclonal mouse immunoglobulin (Ig) raised against OT-related neurophysin (OT-NP, provided by Dr. H. Gainer), the other, a polyclonal rabbit serum raised against vasopressin-associated neurophysin (VP-Np, provided by Dr. A. Robinson).

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RESULTS

All the results reported in this study have been obtained in 153 OT magnocellular neurons which were identified according to two criteria: i) their ability to display high frequency bursts of action potentials which are a specific property of OT neurons (Jourdain et al, 1998; Israel et al, 2003), and ii) post-hoc immuno-identification. Intracellular recordings (n=122) obtained from these neurons revealed a mean resting membrane potential of 54.6±5.0 mV (n=50), a mean input resistance of 237.0±40.9 Mohms (n=50) and action potentials (APs) of 71.9±10.9 mV (n=250 from 50 cells).

OT up-regulates GABAergic transmission

Intracellular recordings in current clamp mode in the presence of TTX (1µM), bicuculline (15 µM), CNQX (10 µM) and APV (40 µM) indicated that bath-applications of OT (10 to 1000 nM) did not affect the resting membrane potential nor the input resistance of OT neurons (Fig. 1A1-A3). Bath-application of OT (100 nM) in normal medium did not alter the amplitude (98.4±5.2 % of control; n=8, P>0.05) or frequency (95.4±3.6 % of control, n=8, P>0.05) of excitatory postsynaptic potentials (EPSPs) recorded at -80 mV (data not shown). Under conditions where EPSPs were blocked with CNQX (10µM) and AP5 (40 µM), low concentrations (25 nM to 100 nM) of OT significantly and reversibly increased inhibitory postsynaptic potentials (IPSPs; Fig.1B) and currents (IPSCs; Fig. 2A) in 56 out of 78 neurons (72%). In the remaining neurons (28%), OT did not affect GABAergic activity (Fig. 1B) and these cells were thus considered as non-responsive to the neurohypophysial peptide. In OT-responsive cells, the enhanced IPSP activity was associated with an increase in the frequency (257±41 % of control; P<0.05; n=7) and amplitude (211±52 % of control; P<0.05; n=7) of spontaneous events (Fig. 2B and 2C).

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OT-sensitive IPSPs/IPSCs were blocked by bicuculline (15 µM), showing that synaptic events modulated by OT were mediated by GABA-A receptors (not shown). The effect of OT was mimicked by a specific OT-R agonist, [4-7] OT (100 nM; n=6) which increased the frequency (377±82 % of control; P< 0.05) and amplitude (283±78 % of control; P<0.05) of spontaneous IPSPs (Fig. 2C). Conversely, the effect of 50 nM OT was blocked in the presence of 1 µM d-OVT, a specific OT-R antagonist (79±31 % and 91±9 % of control in frequency and amplitude, respectively; n=4; Fig. 2C).

To identify the locus of action of OT, TTX (1 µM) was added to the external solution to block AP-driven inhibitory synaptic events and thus make sure that monoquantal synaptic responses (miniatures) were recorded. For these experiments we used the whole-cell patch clamp technique in voltage-clamp configuration instead of intracellular sharp electrode recording. Under these conditions, OT (50 nM) significantly increased the frequency (198.3±29.3 % of control, n=4; P<0.05) but not the amplitude (103.5± 8.1 % of control, n=4; P>0.05) of miniature IPSCs (Fig. 2D-F). Although this set of data implies that OT-R are located presynaptically on GABAergic neuron terminals impinging upon OT cells, they do not rule out the possibility that OT-R are also located on GABA neuron somata or on other neurons contacting GABAergic cells, and that these receptors also contribute to facilitate inhibitory activity in OT neurons.

Bimodal dose-dependent action of OT on inhibitory transmission

The stimulatory effect of OT on GABAergic transmission was dose-dependent, with a threshold of 25 nM and a maximal facilitation at 50 nM (Fig. 3A). Higher concentrations of OT progressively inhibited GABAergic synaptic activity. The inhibition was almost total with 300 nM OT, a result in agreement with the postsynaptic inhibitory action of OT on GABA-A receptors previously reported in the SON (Brussaard et al., 1996). The dynamics and dose-dependency of these two opposite effects of OT on GABAergic transmission

were then compared by monitoring simultaneously synaptic currents and responses obtained with local puffs of GABA. Whereas low concentrations of OT (50 nM) triggered IPSP activity without affecting the amplitude of GABA-induced responses, IPSP amplitude and frequency gradually decreased when increasing OT concentrations, with a complete inhibition obtained at 1000 nM (Fig. 3B and 3D). In the same recordings, GABA-induced responses were slightly affected when OT concentration reached 500 nM and were completely inhibited with 2000 nM (Fig. 3C and 3D). These findings demonstrate that OT acts both at pre- and postsynaptic levels, depending on its concentration, to up- or down-regulate GABAergic transmission.

OT-mediated IPSPs facilitate firing

In intracellular current-clamp recordings, low concentrations of OT (50-100 nM) increased the firing activity in 11 out of 27 OT neurons (183±17 % of control; Fig. 4A1-A2). To identify the cellular mechanism responsible for this increase in firing rate, and its possible relation to OT-mediated facilitation of inhibitory transmission, we investigated the action of OT on OT neuron electrical activity in the presence of CNQX to block EPSPs (Fig. 4B). Whereas CNQX inhibited AP firing, as previously reported (Jourdain et al., 1996), OT still triggered spiking activity at resting membrane potential in 15 out of 36 neurons (41%). Examination of recordings at high resolution revealed that during such OT-triggered activity, most APs occurred at the end of the repolarizing phase of individual IPSPs (Fig. 4B3-B4). This was particularly clear during OT washout where IPSP frequency decreased to values < 5 Hz making it easier to reveal the link between IPSP and AP firing (Fig. 4B3d). That APs were exclusively governed by IPSPs was confirmed in a series of experiments where this OT-triggered firing occurring in the presence of CNQX was completely inhibited by the specific GABA-A receptor antagonist picrotoxin (5 µM; Fig. 5A; n=5). It is worth noting that we never observed an increased in AP firing without an

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increase in IPSP activity.

The interaction between OT-mediated IPSP activity and the facilitation of firing activity was confirmed in another set of experiments in which APs were triggered in response to membrane depolarization in the presence of ionotropic glutamate receptors inhibitors (CNQX and AP5). As illustrated in figure 5B1, OT neurons started to fire APs only once the membrane potential reached spike threshold (-38.1±4.3 mV, n=7). In the same recordings, OT triggered spike discharge without depolarizing the membrane potential (Fig. 5B2), an effect that was accompanied by a consistent increase in IPSP activity preceding the appearance of APs. These findings suggest that OT, by triggering or dramatically increasing the occurrence of IPSPs, paradoxically facilitates firing activity in OT neurons. These results prompted us to investigate the cellular mechanism underlying this phenomenon.

Ionic mechanisms underlying OT-induced firing

In presence of CNQX, each OT-triggered spike occurred at the end of the repolarizing phase of an individual IPSP (Fig. 4B4). Such process is reminiscent of postinhibitory rebound (PIR), as described in other structures (Angstadt et al., 2005; Bertrand and Cazalets, 1998 ; Sohal et al., 2006). PIR is defined as the depolarization that occurs at the offset of a hyperpolarizing period. At least, two non-exclusive mechanisms might account for PIR-induced spikes. One involves low voltage-activated (LVA) Ca^{2+} channels that are first de-inactivated by hyperpolarization and then activated upon the repolarization period (Bertrand and Cazalets, 1998, Jahnsen and Llinas, 1984; Fan et al., 2000), thereby generating a depolarization known as a low threshold spike (LTS). This LTS, if of a sufficient amplitude, can generate APs (Huguenard, 1996). A second possibility is the activation of a hyperpolarization-activated inward current (I_H) which underlies rebound responses in many neurons (Matsushima, 1993; Straub et al., 2001;

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Sekirnjak and du Lac, 2002).

Both LVA Ca^{2+} current and I_H have been described in SON and PVN neurons (Fisher and Bourque, 1995; Ghamari-Langroudi and Bourque, 2000; Luther and Tasker, 2000). Thus, we first checked for their presence in our cultured slices before studying their respective contribution to OT-mediated PIR-firing. Since the presence of LVA Ca^{2+} channels is associated with the generation of a LTS, we applied brief (50 ms) hyperpolarizing pulses in the presence of TTX to block Na^+-dependent APs. As illustrated in figure 6A1, such pulses triggered a rebound depolarization typical of a LTS in 17 out of 32 neurons, a process that was compromised when the amplitude of the negative step was reduced, as previously described (Erickson et al., 1993). In the absence of TTX, such hyperpolarizing pulses triggered rebound APs (Fig. 6A2). To study the contribution of LVA Ca^{2+} channels to this process, we bath-applied Ni^{2+} at 100 µM, a concentration that inhibits T-type Ca^{2+} currents (Fisher and Bourque, 1995) and blocks LTS (Erickson et al., 1993) in SON neurons. In agreement with a role for these channels in LTS generation, APs triggered by long (>100 ms; n= 5; Fig 6B1 and 7B1) or brief (25 ms; n=6; Fig 6B2 and 7B2) hyperpolarizing pulses were completely abolished in the presence of this inhibitor. Because Ni^{2+} might also interact with other voltage-gated Ca^{2+} channels, we tested the action of mibefradil, a compound considered to be a specific T-type channel antagonist (Van der Vring et al, 1999). At a concentration of 40 µM, mibefradil also inhibited pulse-triggered APs (n=5; Fig. 6B3 and 7B1). Taken together, these data suggest that the LVA Ca^{2+} channels mediating LTS in these neurons are of the T-type.

We then examined the role of I_H in this process. During 100 ms long hyperpolarizing pulses, we reliably observed the typical depolarizing sag in the voltage response (e.g. Fig. 6C1 and 6D1) that reflects activation of I_H (Ghamari-Langroudi and Bourque, 2000). As previously reported (Ghamari-Langroudi and Bourque, 2001), this sag was inhibited by 3 mM external Cs^+ (n=4; Fig. 6C1) or by 50 µM ZD 7288 (ZD; n=5; Fig. 6D1), two well-

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known blockers of I_H. Conversely, this sag was not affected in the presence of Ni^{2+} or mibefradil (Fig. 6B1 and 6B3). Interestingly, blockade of I_H with Cs or ZD did not prevent pulse-triggered rebound spikes (Fig. 6C1, 6D1 and 7B1) even when hyperpolarization was adjusted to that obtained in control conditions to compensate for changes in membrane resistance (Fig. 6C2 and 6D2). The lack of effect of ZD on pulse-triggered APs was also observed with pulses of shorter duration (25 ms; n=6; Fig 6D3 and 7B2).

Ni^{2+}, mibefradil, Cs^+ and ZD were then used to assess the respective contribution of LVA Ca^{2+} channels and I_H to OT-mediated PIR firing. As illustrated in figure 7A and summarized in figure 7C, Ni^{2+} (n=4) and mibefradil (n=5) inhibited AP firing, but not IPSP activity (Fig. 7D), triggered by OT. On the other hand, neither Cs^+ (n=4) nor ZD (n=4) affected significantly OT-triggered firing activity (Fig. 7A and 7C) or IPSPs (Fig. 7D). These data suggest that IPSPs can trigger rebound firing through the recruitment of LVA Ca^{2+} channels. If this is true, a rebound depolarization should be observed following individual IPSPs. In 4 cells where OT-triggered AP firing and IPSP activity were not intense enough to mask such a phenomenon, rebound excitations were clearly observed following inhibitory synaptic potentials (Fig. 8A1 and 8A2; also see fig 4B3 trace d). These rebounds were not affected by the subsequent application of ZD (Fig. 8B1 and 8B2) whereas they were completely abolished in the presence of Ni^{2+} (Fig. 8C1 and 8C2). As illustrated on the averaged traces in figure 8D and from cumulative histograms in figure 8E, inhibition of the rebound depolarization with Ni^{2+} resulted in an increased IPSP width, an effect that was not observed with ZD. We thus used IPSP duration to assess the effect of the different blockers on IPSP-triggered postinhibitory rebound excitation. Whereas both Ni^{2+} and mibefrendil increased IPSP duration (126±12 % of control, n=4 for Ni^{2+}; 121±9 %, n=4, for Mibefradil), neither ZD nor Cs^+ modified significantly this parameter (Fig 8F). Taken together, these findings reveal the involvement of LVA Ca^{2+} channels, but not of I_H, in IPSP-mediated postinhibitory rebound firing. These data also indicate that IPSPs have to

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be of sufficient amplitude to trigger AP firing, which may not be the case under control conditions.

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Physiological relevance

Throughout this study, OT neurons recorded in the absence of glutamatergic and GABAergic blockers usually displayed a bursting activity, either spontaneously or in response to bath application of 100 nM OT (Fig. 9A; Jourdain et al., 1998, Israel et al., 2003). This activity is characteristic of that recorded in vivo in lactating rats (Lincoln and Wakerley,1975). Careful analysis of this activity in cultured slices revealed an increase in firing rate occurring just before burst onset in 40% of the neurons (Fig. 9A). Such increases in background firing rate immediately preceding the bursts have been already reported in vivo where they are directly and positively correlated to the magnitude of the bursting activity itself (Lincoln and Wakerley,1975). This prompted us to investigate whether a similar type of correlation prevailed in OT neurons recorded from organotypic slice cultures, and to test whether OT-mediated PIR firing was playing a role in this process.

Within bursts, both the mean AP frequency and the peak frequency (over 0.5 s) were increased (175±38 % of control and 183±40 % of control, respectively, n=25) in neurons showing an enhanced background firing activity prior to burst onset (Fig. 9B1-B2). To further analyze burst magnitude, we used the same index as described by Lincoln and Wakerley which corresponds to the number of spikes within the burst multiplied by the peak frequency (Lincoln and Wakerley, 1975). As illustrated in figure 9C, the burst index was positively correlated (r=0.61; n=48 bursts from 15 cells) to the firing frequency measured 20 seconds before the incidence of each burst, a result in complete agreement with previous in vivo data (Lincoln and Wakerley, 1975; Brown et al., 2000).

Although the bursts are driven by glutamatergic inputs (Jourdain et al., 1998; Israel et

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al., 2003), the origin of the increase in background firing rate in OT cells is unknown. One possibility is that this phenomenon is due to OT-mediated PIR firing. To test this hypothesis, we analyzed synaptic activity just before the occurrence of each burst. EPSP activity occurring during this period remained unchanged whether an increase in background firing rate occurred or not (Fig. 10A). On the contrary, in neurons that displayed an increase in firing rate prior to the bursts, a marked increase in both amplitude (155±12 % of control; n=10; $P<0.05$) and frequency (189±39 % of control; n=10; $P<0.05$) of IPSPs occurred (Fig. 10A-B). These findings strongly support a relationship between IPSP activity and increased background firing rate. Because IPSP-mediated PIR firing is related to OT, it is likely that the increased firing observed in these neurons resulted from the dendritic release of endogenous OT. If this is true, then activation or inhibition of OT-R should affect background firing and, consequently, the burst index. In agreement with this hypothesis, OT (50-100 nM) increased the mean firing rate prior to burst onset from 2.3±0.7 Hz to 4.78±1.24 Hz (211±41 % of control, n=5; Fig. 9C, 10C1 and 10D) whereas the OT-R antagonist, d-OVT, decreased it from 2.98±0.20 Hz to 0.98±0.40 (32±14 % of control, n=4; Fig. 9C, 10C2 and 10D). In these neurons, OT and d-OVT respectively augmented (163±26 %, n=5) and diminished (36±19 % of control) the burst index as expected (Fig. 10D). It is noteworthy that in the presence of OT, a concomitant increase in IPSP frequency (190±26 % of control, n=5, $P<0.05$) and amplitude (191±35 % of control, n=5, $P<0.05$) occurred (Fig. 10E) whereas d-OVT by itself, induced an opposite effect (frequency: 73±6 % of control, n=4, $P<0.05$; amplitude: 56±9 % of control, n=4, $P<0.05$; Fig. 10E). This suggests that endogenous ambient OT has a positive action on IPSP activity and, consequently, on burst magnitude.

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DISCUSSION

Action potential firing in neurons is usually obtained when the membrane potential is depolarized above spike threshold. This generally occurs through activation of ion channels or ligand-gated receptors or through the relief of tonic inhibition, a process known as disinhibition. Another mechanism promoting neuronal firing, although less usual, is postinhibitory rebound (PIR). In this phenomenon, one to several APs can be generated during the membrane repolarization that follows the offset of a hyperpolarizing event. PIR may involve hyperpolarization-activated current (I_H), de-inactivation of voltage-gated Ca^{2+} currents or both. Such process is responsible for triggering activity in motoneurons (Bertrand and Cazalets, 1998) in thalamocortical neurons (Sohal et al., 2006) and in rat caudal hypothalamic neurons (Fan et al., 2000) for example. Here we described a process in which a peptide, oxytocin, by facilitating the occurrence of hyperpolarizing GABAergic synaptic potentials, promotes AP discharge through PIR firing. This process potentiates bursting activity of OT neurons which is responsible for the massive and intermittent release of OT in the blood, and thus for pup delivery and milk ejection.

OT modulation of firing activity in OT neurons

The low concentrations of OT that we used here (25-100 nM) is more compatible with physiological concentration, as suggested by microdialysis experiments (Neumann et al., 1993). Interestingly, the facilitatory action of OT on IPSP activity was observed in 72% of OT neurons. That such low concentrations of OT accelerate firing in OT-responsive neurons through the generation of IPSPs is paradoxical since hyperpolarizing synaptic potentials are usually associated with inhibition rather than facilitation of firing activity. The action of OT was receptor-mediated since it was mimicked by an OT-R agonist and inhibited by an OT-R antagonist. Similar up-regulation of GABAergic activity has been reported in CA1 hippocampal neurons (Zaninetti and Raggenbass, 2000) and in putative

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vasopressin hypothalamic neurons (Hermes et al., 2000) in response to OT and VP, respectively. Furthermore, our experiments revealed that OT-mediated GABAergic activation facilitated AP firing in about 40 % of OT neurons. Although moderate, this increase in firing rate was in the range of that reported in lactating rat *in vivo* in response to local OT applications (Brown et al., 2000).

How can a GABAergic inhibitory synapse become excitatory? Several mechanisms may underlie IPSP-mediated PIR firing. One involves an LTS resulting from de-inactivation of LVA Ca^{2+} currents, as previously described in the SON (Fisher and Bourque, 1995, Erickson et al., 1993, Dudek et al., 1989). This is likely to be the case here for several reasons. First, the percentage of neurons exhibiting an LTS is similar to that displaying OT-triggered firing. Second, rebound OT-triggered depolarizations and spikes were entirely blocked by Ni^{2+} at concentrations known to inhibit LTS and LVA Ca^{2+} current in these cells (Fisher and Bourque, 1995; Erickson et al., 1993) and by mibefradil, a more specific T-type channel antagonist (Van der Vring et al, 1999). Although these data suggest that T-type channels are responsible for mediating IPSP-induced PIR firing in OT neurons, the definitive demonstration for the implication of these channels awaits new and more specific pharmacological tools. Similarly, it remains to be determined which Ca_VT subunits among those already detected in hypothalamic neurons (Craig et al., 1999; Talley et al., 1999), are implicated in this process. In SON neurons, activation of I_H could also account for PIR firing (Ghamari-Langroudi and Bourque, 2000): this can be ruled out since Cs^+ and ZD 7288 inhibited I_H without affecting OT-mediated firing.

Heterogeneity of OT actions

It is clear from our observations that OT mediates distinct effects in the SON according to its concentration. At 50 nM, OT stimulated the frequency and amplitude of IPSPs/IPSCs, suggesting that it can act both pre- and post-synaptically. In the presence of TTX, the frequency but not the amplitude of mIPSCs was increased, whereas postsynaptic

responses to applications of GABA were unaffected, indicating a presynaptic site of action of OT. At concentrations higher than 50 nM, OT progressively inhibited both the frequency and amplitude of IPSPs with a complete blockade obtained at 1000 nM, a result in agreement with the inhibitory action of OT on GABA release previously described (De Kock et al., 2003) and which has been related to the release of endocannabinoids acting on presynaptic CB1 receptors (Oliet et al, 2007). At such high concentrations, OT also inhibited GABA-A receptor mediated postsynaptic responses (Brussaard et al., 1996). Taken together, our results reveal, therefore, that depending on its concentration, OT has presynaptic effects, increasing then decreasing the probability of GABA release, and postsynaptic effects, inhibiting GABA-A receptors on OT neurons.

It is obvious from this and our previous studies (Jourdain et al., 1998; Israel et al., 2003) that OT acts differently, according to its concentration and its targets, namely, GABA, glutamate and OT neurons. Such heterogeneity of actions may reflect differences in OT-R mediating these responses. However, although there is evidence supporting the existence of different receptor subtypes, only one type of OT-R has been described so far (Gimpl and Fahrenholz, 2001). Alternatively, if there is only one type of OT-R, there may be a differential expression of this receptor in different cells and/or different OT-R-coupled second messenger pathways (Verbalis, 1999).

Physiological considerations

At parturition and during suckling, local release of OT from the somatodendritic compartment is necessary to trigger and facilitate the periodic activation of OT neurons (Freund-Mercier and Richard, 1984; Moos et al., 1984). We have shown previously that OT neuron bursting is controlled by an intrahypothalamic network in which bursting glutamate neurons govern OT neurons. In turn, OT somatodendritic release is essential to modulate the bursting pattern of glutamatergic neurons (Jourdain et al., 1998; Israel et al., 2003). The modulation of GABA transmission by OT as reported here may provide another

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mean of generating APs during background activity, in addition to those generated by EPSPs. Such a process may also explain previous in vivo data obtained in lactating rats showing that locally applied GABA unexpectedly facilitated bursting activity (Moos, 1995) whereas the same activity was impaired when GABA-A receptors were inhibited (Voisin et al., 1995).

We here showed a strong correlation between background firing activity before each burst and the magnitude of the bursts, a result similar to that reported *in vivo* (Lincoln and Wakerley, 1975; Brown and Moos, 1997). One likely explanation to account for this observation is that such an increase in firing rate facilitates the somatodendritic release of OT, thereby increasing its ambient concentration and range of action in the extracellular space. This, in turn, could positively modulate the intrahypothalamic pacemaker neurons responsible for the bursting activity of OT-secreting cells (Jourdain et al., 1998). In agreement with this hypothesis, we noticed that activation of OT-R with exogenous OT increased background firing rate and, consequently burst magnitude, as reported *in vivo* (Brown et al., 2000) whereas inhibiting OT-R with d-OVT had the opposite effect. In view of these data, it appears that OT-mediated PIR firing is an important process by which OT neurons could not only regulate their own activity but also influence the efficacy of the intrahypothalamic network generating the bursting behavior responsible for pup delivery and milk ejection. These results are reminiscent of those obtained *in vivo* where OT neurons showing an increase in their background firing rate prior to the bursts have been described as “leader” neurons whose task is to recruit “follower” neurons to optimize the activation of the entire OT network, thereby maximizing synchronized bursting activity (Moos et al, 2004).

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LEGENDS

Figure 1: OT facilitates inhibitory transmission.

A1: Example of a recording where OT was applied at two different concentrations (50 nM and 1 µM) in the presence of TTX, bicuculline, CNQX and AP5. The peptide did not affect membrane potential nor membrane resistance measured from negative current pulses of decreasing amplitude (from –150 to 0 pA by successive 15 pA steps). A2: Voltage-current relationship obtained from the experiment shown in A1. The presence of OT did not modify this relationship. A3: Histogram summarizing the lack of action of OT (10 nM to 1 µM) on membrane potential (Em) and membrane resistance (Rm) in OT sensitive-cells. The numbers of experiments are indicated in brackets. B: In the presence of CNQX, application of 50 nM OT reversibly increased IPSP activity in 72 % of OT neurons as illustrated in the example on the left panel (OT responsive cell). In the remaining neurons (28%), GABAergic synaptic transmission was unaffected as shown on the example illustrated in the right panel (OT non-responsive cell).

Figure 2: OT acts at presynaptic sites.

A: IPSC activity was increased in a reversible manner by 50 nM OT. B: Cumulative distributions of IPSC intervals (left) and amplitude (right) indicated that OT increased both the frequency and the amplitude of these events. C: Histogram summarizing the action of OT, the agonist [4-7] OT and OT+ the antagonist d-OVT on IPSP frequency (Hz) and amplitude (q). The number of cells is indicated in brackets. D: The activity of miniature IPSCs recorded in the presence of TTX was also reversibly increased by OT (50 nM). E: Cumulative distributions for miniature IPSC intervals and amplitude indicated that OT increased the frequency without affecting the amplitude of these unitary events. F: Summary histograms illustrating the effect of OT on the frequency and amplitude of

miniature IPSCs (n=4).

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Figure 3: OT has bimodal effects on GABAergic transmission.

A: Histograms summarizing the changes in IPSP amplitude and frequency induced by different concentrations of OT. Insets are example obtained from a cell where OT was successively applied at 50 and 300 nM. OT triggered or facilitated IPSP activity at a threshold concentration of 25 nM, an effect attenuated at higher concentrations (>100 nM). Maximal facilitation for both IPSP amplitude and frequency was obtained at 50-100 nM OT. B-C: Example of an OT neuron where OT, from 25 nM to 1000 nM, had distinct effects on IPSPs (B) and on responses induced by local puffs of GABA (C). At 25 nM, OT slightly increased synaptic activity without modifying the amplitude of the postsynaptic response. With 50 nM OT, IPSP activity was dramatically augmented whereas GABA-induced postsynaptic response was unaffected. At 500 nM, IPSP activity was strongly reduced while the postsynaptic response was only slightly inhibited. Finally, 1000 nM OT inhibited totally IPSPs and almost completely the postsynaptic response. D: Summary histogram illustrating the action of OT at different concentrations on the amplitude of both IPSPs (open bars) and GABA-induced responses (grey bars) obtained from the same neurons. Number of experiments are indicated in brackets.

Figure 4: OT-induced IPSP activity triggers AP firing.

A1: Example of an OT neuron where exogenous application of OT (50 nM) increased firing activity in a reversible manner as illustrated by the change in the sequential frequency histogram. A2: Histogram summarizing the stimulatory effect of OT on firing rate. The number of cells is indicated in brackets. B1: Under conditions where CNQX totally abolished spontaneous firing (b), OT (50 nM) still triggered AP discharge (c). B2:

Summary histogram showing the stimulatory effect of OT in the presence of CNQX. B3: Traces obtained from the recording shown in B1. In control conditions (a), APs were mainly triggered by EPSPs. CNQX totally abolished EPSP activity and APs firing (b). Subsequent addition of OT (50 nM) dramatically increased IPSP activity (arrow heads) and restored AP firing (c). During washout of OT, although IPSP frequency and AP firing decreased, spikes were still occurring at the end of IPSPs (d). B4: Magnification of the trace c shown in B3 revealed that APs (*) occurred at the offset of IPSPs (arrow heads).

Figure 5: OT-triggered APs are exclusively governed by IPSPs.

A: In the presence of CNQX, OT (50 nM) dramatically increased IPSP activity and firing (middle panel), an action that was totally blocked by picrotoxin (5 µM; right panel). B1: In the presence of CNQX and AP5, a positive step current was required to depolarize the cell and trigger firing. B2: OT (50 nM) under the same conditions and in the same neuron, triggered IPSPs (arrowhead) and AP firing without depolarizing the membrane.

Figure 6: Presence of a low threshold spike (LTS) in OT neurons. A1: In the presence of TTX, a negative current (50 ms, -40 pA, thick traces) injected in an OT neuron at -55 mV, triggered a LTS (*) at the offset of the current step. This LTS was abolished when the amplitude of the pulse was decreased (-30 pA, thin traces). A2: When negative current steps (-50 and -70 pA, 40 ms) were applied from -55 mV, only the larger current step triggered a rebound spike (thick trace). B: APs (thin traces) triggered by long (350 ms; B1) **and brief (25 ms; B2)** hyperpolarized pulses were inhibited (thick traces) by 100 µM Ni^{2+} (Ni). 40 µM mibefradil (Mib) also inhibited such pulses-triggered APs (B3). C1: A depolarizing sag (*) typical of I_H activation was observed during voltage responses to

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negative current injections (thin traces). This sag was blocked with Cs^+ (Cs, 3 mM; thick trace) whereas pulse-triggered APs remained unaffected. C2: In Cs^+, adjusting the hyperpolarization to the control value to compensate for the change in membrane resistance associated with I_H blockade did not affect pulse-triggered AP firing. D1: Inhibition of I_H with the specific antagonist ZD 7288 (ZD; 50 µM, thick trace), did not affect 200 ms long pulse-triggered APs. D2: As for Cs^+, pulse-triggered AP firing was not affected when the hyperpolarization was adjusted to the control value in the presence of ZD. D3: APs triggered by brief (25 ms) hyperpolarizing pulses were not affected by ZD.

Figure 7: OT-mediated postinhibitory rebound firing is blocked by low threshold activated calcium current blockers.

A: In CNQX-containing medium (control; left panel), firing activity triggered by 50 nM OT (middle panel) was strongly inhibited by 100 µM Ni^{2+} (Ni) and 40 µM mibefradil (Mib) whereas it was unaffected by 3 mM Cs^+ (Cs) and 50 µM ZD 7288 (ZD). B: Histograms summarizing the action of Ni^{2+} and ZD 7288 on APs triggered by long (>100 ms; B1) and brief (25 ms; B2) hyperpolarizing pulses. C: Histograms summarizing the action of Ni^{2+}, mibefradil, Cs^+ and ZD 7288 on OT-induced PIR firing. D: Ni^{2+}, mibefradil, Cs^+ and ZD 7288 did not affect the frequency (Hz) nor the amplitude (q) of OT-triggered IPSPs.

Figure 8: IPSP-mediated rebound depolarization.

A1: Example of a recording obtained in the presence of CNQX and 50 nM OT showing the presence of rebound depolarizations (*) occurring at the end of IPSPs. A2:

Superimposition of 8 consecutive IPSPs obtained from the recording in A1 clearly shows that a rebound depolarization follows several of these inhibitory potentials. B1 and B2: In the same recording, application of ZD 7288 (ZD) did not affect the occurrence of such

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IPSP-triggered rebound depolarizations. C1 and C2: Subsequent addition of 100 µM Ni^{2+} in the bathing solution completely abolished AP firing and rebound depolarizations. D: Average IPSPs (n=93-105) obtained from the recordings shown in A, B and C. This graph shows that rebound depolarization was unaffected by ZD (grey trace) whereas it was abolished in the presence of Ni^{2+}. Note that IPSP duration was increased with Ni^{2+}. E: Cumulative histograms representing the distribution of IPSP half-width obtained from the recording shown in A, B and C. This distribution was significantly shifted toward higher values in the presence of Ni^{2+}. F: Summary histogram illustrating the percent change in the half-width of OT-triggered IPSPs (n=4 cells). Whereas IPSP duration was significantly increased by Ni^{2+} and mibefradil (Mib), it remained unchanged in the presence of Cs^{+} or ZD.

Figure 9: Pre-burst firing and burst magnitude in OT neurons

A: Frequency histogram (FH) illustrating the bursting activity of an OT neuron recorded in cultured slices. Insets show the raw recording at the time indicated before and during a burst. B1: Superimposition of FHs recorded from 2 OT neurons displaying (grey) or not (black) an increased firing activity prior to the occurrence of a burst. Burst magnitude was larger in the neuron showing such an increase. B2: Summary histogram illustrating the mean intraburst frequency and the mean peak frequency in neurons showing (grey; n=25) or not showing (black; n=25) an increased firing rate prior to burst occurrence. C: Correlation between burst magnitude (index) and firing rate prior to the burst (mean frequency). Data were obtained from control experiments (black dots; n=39), with 50-100 nM OT (grey triangles; n=5) and 100 nM d-OVT (empty squares; n=4). The dotted line represents the linear regression through the data points obtained under control conditions (r=0.61).

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Figure 10: Contribution of OT-mediated rebound firing to OT neuron bursting activity.

A: histograms summarizing the percent change in the amplitude (q) and frequency (Hz) of EPSPs (left panel) and IPSPs (right panel) in OT neurons showing (grey bars) or not showing (black bars) an increased firing rate prior to burst occurrence. B: Sample traces extracted from the recording shown in figure 9A at the time indicated. Detailed analysis revealed a dramatic increase in IPSP activity and AP (*) firing before burst occurrence (b). C1: Superimposition of FHs before (grey) and after (black) application of 100 nM OT. Note that OT induced a marked increase in background activity and burst magnitude. C2: FHs before (grey) and after (black) application of 100 nM d-OVT. Note that the antagonist d-OVT decreased both background activity and burst magnitude. D: Histograms summarizing the changes in background firing (mean Hz) and in burst magnitude (index) caused by OT (grey) and d-OVT (black). The number of experiments are indicated in brackets. E: Under OT treatment (grey bars), IPSPs occurring prior to the burst showed a marked increase in their amplitude (q) and frequency (Hz), whereas d-OVT (black bars) inhibited both parameters.