



HAL
open science

Exposure-effect population model of inolimomab, a monoclonal antibody administered in first-line treatment for acute graft-versus-host disease.

Céline Dartois, Gilles Freyer, Mauricette Michallet, Emilie Hénin, Benoît You, Isabelle Darlavoix, Claudine Vermot-Desroches, Brigitte Tranchand, Pascal Girard

► To cite this version:

Céline Dartois, Gilles Freyer, Mauricette Michallet, Emilie Hénin, Benoît You, et al.. Exposure-effect population model of inolimomab, a monoclonal antibody administered in first-line treatment for acute graft-versus-host disease.: Exposure-Effect population model of inolimomab. *Clinical Pharmacokinetics*, 2007, 46 (5), pp.417-32. inserm-00172432

HAL Id: inserm-00172432

<https://inserm.hal.science/inserm-00172432>

Submitted on 14 Sep 2009

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

TITLE Exposure-Effect population model of inolimomab, a monoclonal antibody administered in first-line treatment for acute graft-versus-host disease.

Céline Dartois¹, Gilles Freyer^{1,2}, Mauricette Michallet³, Emilie Hénin¹, Benoît You², Isabelle Darlavoix⁴, Claudine Vermot-Desroches⁴, Brigitte Tranchand^{1,5}, and Pascal Girard¹.

1 Université de Lyon, Lyon, F-69003, France ; université Lyon 1, EA3738, CTO, faculté de médecine Lyon Sud, Oullins, F-69600, France.

2 Service d'Oncologie Médicale, Centre Hospitalier Lyon Sud, Hospices Civils de Lyon, F-69310, Pierre-Bénite, France.

3 Hôpital Edouard Herriot, Lyon, France

4 OPi, Limonest, France

5 Centre anticancéreux Léon Bérard, Lyon, F-69008, France.

Runing title : Exposure-Effect population model of inolimomab

Corresponding author:

Pascal Girard

EA 3738, Université de Médecine de Lyon sud BP 12

69 921 Oullins cedex, FRANCE

tél : +33 (0)4 26 23 59 76 / fax: +33 (0)4 26 23 59 76

mail: pascal.girard@adm.univ-lyon1.fr

Word count : 3831

ACKNOWLEDGEMENTS

Authors wish to thank OPi for providing the pharmacokinetic and pharmacodynamic samples, for reviewing and approving the manuscript.

The funding of this study was provided by a support from OPi and Medicine Faculty of Lyon Sud. I. Darlavoix and C Vermot-Desroches are employees of OPi. C. Dartois's PhD is supported by Institut de Recherches Internationales Servier. Pascal Girard is supported by INSERM, France. Other authors have no conflict of interests.

NAME AND ADDRESS FOR CORRESPONDENCE

Pascal Girard

EA 3738, Université de Médecine de Lyon sud BP 12

69 921 Oullins cedex, FRANCE

tél : +33 (0)4 26 23 59 76 / fax: +33 (0)4 26 23 59 76

mail: pascal.girard@adm.univ-lyon1.fr

TABLE OF CONTENTS

BACKGROUND	7
METHODS	9
PATIENTS AND TREATMENT	9
PK SAMPLING.....	10
BIOANALYSIS	10
PD ASSESSMENTS.....	11
PK ANALYSIS	11
PK-PD ANALYSIS.....	11
MODEL QUALIFICATION.....	12
RESULTS	14
PK ANALYSIS	14
PK-PD ANALYSIS.....	14
DISCUSSION	17
CONCLUSION.....	20
REFERENCE LIST	21

ABSTRACT

Background: Inolimomab, a monoclonal antibody against IL-2R α (CD25) has shown promising results in the treatment of steroid-resistant acute graft-versus-host disease (aGvHD).

Objective: Our purpose was to characterize its pharmacokinetic (PK) and pharmacodynamic (PD) properties in first line treatment.

Methods: Data arose from 21 patients with aGvHD (8 with IBMTR at score B, 11 at score C and 2 at score D) following Hematopoietic Stem Cell Transplantation after a median delay of 26 days (10 – 127 days). Inolimomab was administrated at 0.1, 0.2, 0.3 or 0.4 mg/kg daily associated with methylprednisolone (2 mg/kg) for 8 or 16 days depending of status at day 9. Then, for responder patients, administrations were continued three times a week until day 28. Inolimomab concentrations and PD data (aGvHD scores) were collected along the study. PD data were assessed in 4 grades according to IBMTR and Glucksberg classification in parallel with Karnofsky scores. Population analysis was developed using NONMEM to define the pharmacokinetic model, to test covariates, and when apparent, to model the exposure-effect relationship by a proportional odds model. Modelling was finally qualified by predictive check.

Results: The best pharmacokinetic model was bi-compartmental. For each score, the most demonstrative exposure-effect graphics linked cumulative AUC to cumulated probabilities to observe a specific score. This relationship was identified as an E_{\max} model for skin (with 2 patient subpopulations: sensitive/less sensitive) and a linear model for intestinal tract and liver. No covariate was identified as influent on any of these parameters.

Conclusion: Inolimomab exposure-effect relationships in first-line treatment of aGvHD have been identified and modeled. The discovered dose effect relationship allows to confirm the

treatment response, then to establish the first step towards optimizing the doses of future trials.

BACKGROUND

Allogeneic Haematopoietic Stem Cell Transplantation (HSCT) is an effective and curative treatment for many hematological malignancies [1] related to the existence of a graft-versus-malignancy effect. However, it is frequently associated with graft-versus-host disease (GvHD) which is still responsible for a high rate of treatment-related mortality.[2] Acute GvHD usually involves skin, liver and the intestinal tract, but lymphoid and haematopoietic tissues can also be affected. It is induced by alloreactive T-cells from the donor, which react against the recipient's tissues and organs. Standard GvHD prophylaxis consists in administering a combination of immunosuppressive drugs with the classical association of cyclosporine A (CsA) and methotrexate.[3] Other therapeutic approaches have been tested including T-cell depletion, that can be performed *in vivo* or *ex vivo*. The incidence of GvHD decreases, but this is at the cost of a high risk of rejection and relapse associated to a delayed immune reconstitution.[4] The first line treatment of aGvHD is based on steroids with usually methylprednisolone (MP) at the dose of 2 to 2.5 mg/kg/day.[5] However, steroid resistance is observed in approximately 40 % of the patients and therefore requires alternative treatment.[6,7] No standard therapy really exists in steroid-refractory aGvHD. Therapy could be based on high dose of steroids (10 to 15 mg/kg/day) either alone or in combination with antithymocyte globulins or monoclonal anti-T-cell preparations.[8-10] In some cases, although it cures aGvHD, this treatment is responsible for a strong immunosuppression leading to an increased incidence of severe bacterial, viral infections, and an increased risk of Epstein-Barr Virus (EBV)-related lymphoproliferative disorders.[11,12] Inolimomab (Leukotac®, Opi, Limonest, France) because of its inhibitory effects on activated T-cells, could be useful for the treatment of aGvHD. This murin monoclonal antibody (mAb) specifically targets the α chain (CD25) of the interleukine-2 (IL2) receptor. Activated T-cells express the inducible IL-2R α chain whereas resting cells and their precursors do not. Consequently, fewer adverse experiences are expected, due to a lower and more targeted

immunosuppression activity. Some clinical trials have already been performed in steroid-resistant aGvHD and have shown some promising results in terms of response and survival.[13-17]

A clinical trial of inolimomab given in association with steroids, was conducted as initial therapy of aGvHD. As the compound was well tolerated [15], this clinical trial expected to show a better, longer and less heterogeneous overall response than in steroid-resistant patients. This study presents an original population PK-PD modelling of these clinical data, showing for the first time and in this indication, an exposure-effect relationship of a mAb. The specific aims were: (i) to model the PK of inolimomab given as a repeated dose; (ii) to identify inolimomab exposure-effect relationships on different efficacy markers in aGvHD and (iii) to propose a model in order to help future dose optimization of this treatment.

METHODS

Patients and Treatment

Data were collected from an open label, dose-escalating, non-randomized phase I-II study of inolimomab in association with steroids (MP at 2 mg/kg) as first line therapy for grade II to IV aGvHD following allogeneic HSCT. The main objective of this trial was to establish pharmacokinetics of four dosages of inolimomab. Six French institutions participated in the study (Lyon-Hôpital Edouard Herriot, Clermont-Ferrand-Centre Jean Perrin, Marseille-Institut Paoli Calmettes, Lille-Hôpital Claude Hurriez, Nantes-Hôtel Dieu, Créteil-Hôpital Henri Mondor). The study protocol was approved by the Independent Ethics Committee of Lyon-Centre Léon Bérard. Inclusion criteria were as follows: grade II to IV aGvHD following HSCT using either allogeneic bone marrow or allogeneic peripheral blood progenitor cells, and patients had to be 18 years old with written informed consent to participate in the study. Patients were not included if steroids were part of prophylaxis of aGvHD or in case of aGvHD occurring after the donor lymphocyte infusions. Diagnosis and classification of aGvHD was done according to the Seattle criteria (Glucksberg classification) [18,19] and the IBMTR [20] classification.

After eligibility was confirmed, 21 patients were registered and assigned to 1 of 4 cohorts to receive 30-minute intravenous (IV) inolimomab infusion (0.10, 0.20, 0.30, and 0.40 mg/kg), with 5 patients in each dose group except 0.30 mg/kg group which had 6 patients. The treatment was divided into the induction and maintenance regimen phases. The induction regimen was given from day 1 to day 8 and consisted in a once daily IV infusion of inolimomab at the patient's assigned dose level. The clinical response assessed at day 9 determined subsequent treatment. Patients with complete response were assigned to receive the maintenance regimen. Patients with Partial Response, Mixed Response, No Response or Progression disease were reassigned to the induction regimen for one week. The maintenance regimen consisted in the administration of IV inolimomab three times a week at the patient's induction dose level. During

both the induction and maintenance regimens, all patients received concomitant IV infusion of MP. Patients received from 6 to 22 administrations of inolimomab depending on their induction and maintenance phase durations. The entire treatment period lasted a maximum of four weeks.

A total of 21 patients (12 men and 9 women; age range, 29-61 years; weight range, 49-93 kg) were enrolled in the trial (Table I). According to their initial disease, they could be classified in 3 groups: 6 patients with good prognosis transplantation of success (for CML in chronic phase, AML, or ALL in first complete remission), 9 with intermediate prognosis (for AML, ALL in second complete remission or above, NHL, myeloma, or hodgkin disease in partial response), and 6 with a poor prognosis (disease refractory or in relapse).

PK sampling

The median number of PK samples per patient was 12 (range 7-23) and the total number was 318. Blood samples for PK (including peak and trough levels of inolimomab) were collected on day 1 prior to the infusion then 30 minutes, 2, 8 and 16 after completion of the infusion; prior to the infusion and 30 minutes after completion of the administration of study medication on days 2, 3 and 8; and prior to the infusion and 30 minutes after completion of the administration of study medication from day 9 to day 28 for the first three infusions. After collection, they were centrifuged and serums were stored at -20 °C until analysis.

Bioanalysis

Quantification was then carried out in the OPi Research department by a validated inolimomab Enzyme-Linked ImmunoSorbent Assay (ELISA) according to GLP. To trap inolimomab, serum samples were put on a coated plate with goat polyclonal anti-mouse Ig antibodies (GAM). Next, sheep polyclonal anti-mouse IgG1 antibodies used as tracer antibodies were added to the mixture. After incubation with TMB-substrate, the reaction was stopped by the addition of sulfuric acid and absorption was read photometrically to quantify the samples. The range of the immunoassay was from 0.15 to 10 µg/ml, with a sensitivity > 100 ng/ml, a sample intra-assay variation at 8 % and a sample inter-assay variation at 11%.

PD assessments

Acute GvHD grades and performance status were evaluated daily until day 9 then at each administration, from day 10 to day 28, as well as at follow-up (day 60 and day 100) using the Glucksberg–Seattle, IBMTR and Karnofsky classifications [21] (defined in the study as composite scores). Glucksberg criteria determines aGvHD severity (from grades 0, corresponding to no GvHD, to 4, maximum severity) by a combination of different organ scores (skin, intestinal tract and liver) and a decrease in clinical performance. These organs are graded independently from 0 to 4 corresponding respectively to extent of a skin rash, a diarrhea volume, and total bilirubin concentrations. The detailed definition of IBMTR score is given in annex 1. Briefly, it involves the same organs but assigns the score based on maximum involvement in an individual organ system. Therefore, it tends to assign a higher overall grade for aGvHD severity than the Glucksberg.[20] The Karnofsky scale defines overall performance status of a patient from 100% for a normal person with no complaint and no sign of disease to 0 % for a moribund. Independent organs as well as composite scores were considered for pharmacodynamics analysis.

PK analysis

PK and PD analysis were carried out with mixed-effect modeling using NONMEM software version V.[22] Different PK models were tested, including one, two and three compartment models, coupled with linear or nonlinear processes, such as saturable elimination. Inter-individual PK parameter variability was assumed to follow log-normal distribution with non-zero correlations. Residual unexplained variability was modeled as multiplicative. FOCE INTERACTION method was used to fit all PK models. Models were evaluated through goodness of fit plots [23-25] and the parameter precision estimated by asymptotic covariance matrix. Nested models were compared according to likelihood ratio tests (decrease of NONMEM objective function between reduced and full model by $\Delta=3.84$, corresponding to nominal p-value 0.05, for one additional parameter).

PK-PD analysis

For each pharmacodynamic time point assessment, inolimomab exposures were estimated from individual PK profiles predicted from the previously described model. They were defined either as maximal serum concentration (C_{max}), Area Under the serum concentration-time Curve (AUC) over last dosing interval, cumulated AUC, or AUC intensity (cumulated AUC/duration) from the first to the last dosing before PD assessment. Cumulated AUC corresponded to the cumulated sum of all AUCs computed for each dosing interval before PD assessment; duration used in AUC intensity calculation corresponded to the treatment duration before PD assessment. The graphical exploration of the exposure-effect relationships was performed with all PD assessments by plotting estimated cumulative probabilities of ordered scores (composite and organ scores) vs. distribution quartiles (25, 50, 75 and 100%) of the above defined drug exposures. Apparent relationships were then quantified by proportional odds models.[26] Cumulative probabilities of the observed score were linked to PK exposure through logit transformation. The nature of this link was tested with different PD models, like E_{max} , log-linear or linear models. Inter-patient variability of some key parameters was assumed to follow either a normal or a log normal distribution. Parameter estimation was performed using the Laplacian estimation method in NONMEM. The adequacy of the different developed models and selection of the basic model was evaluated by comparing predicted and observed probabilities.

Model Qualification

The model qualification for PK and PK-PD model was conducted in two steps. In regards to the PK model, after inspection of the basic graphics (predictions of a typical patient versus observations, individual predictions versus observations, weighted residuals versus observations, individual predictions and observations versus time), a visual predictive check was conducted.[27] It consisted in simulating (with NONMEM) 200 new datasets with identical patients, dosage regimens, sampling times, and then comparing graphically the simulated

concentrations with the observed ones. The qualification of the PK-PD model was also based on a visual predictive check. Here, the purpose was to test the model performance in order to predict probabilities to observe the different grades. Therefore, we compared graphically simulated grades function of exposure, with the observed ones. Then, in order to qualify the PK-PD model for its clinical purpose, a predictive check was more specifically conducted.[28,29] It consisted in simulating (using NONMEM) 1000 new datasets with identical patients, score record time, and then comparing a test statistic deduced from these simulations with observed ones. This test statistic, which is a test quantity that depends only on data was chosen in order to highlight treatment effect.

RESULTS

PK analysis

Amongst all PK models tested, the best results were obtained with a two-compartment model. The goodness of fit plot for this model showed that the mean population and individual predicted concentrations were in good agreement with the observed ones (close to identity) except for a few concentrations over 20 μ g/ml which were under-predicted. Other tested models included a third compartment or a Michaelis-Menten elimination. The three-compartment model was not identifiable and a non-linear elimination did not show a significant improvement fit, therefore the two-compartment model was eventually retained. The correlation between all PK parameters was then introduced. Parameter estimates are presented on Table II. The half-life of the compound for a typical patient was equal to 44.5h.

The most important goodness-of-fit plots of the final PK model are presented in Figure 1. Although it shows a slight under prediction for high concentrations, a visual predictive check (Figure 2) revealed that this has no impact on predicted concentrations: the proportion of concentration points outside the 80 % confidence interval band was in agreement with the expected one. The PK model was consequently considered to be qualified.

PK-PD analysis

PD assessments included composite scores, with a median number per subject of IBMTR scores at 13, ranging (3-23), a median number per subject of Glucksberg scores at 12, ranging (3–23), and a median number per subject of Karnofsky scores at 18, ranging (7-25). Organ scores (skin, intestinal tract or liver) were recorded with a median number per subject at 18, ranging (7-25). The PK-PD exploratory graphical analysis revealed that composite scores (IBMTR, Glucksberg and Karnofsky) were apparently not related to drug exposure (see Figure 3 with cumulated AUC, other graphics concerning C_{max} , AUC and AUC intensity are not shown). On the contrary, organ scores (skin, intestinal tract and liver) revealed patient improvements (i.e.

lowest grade probability increase) when plotted versus drug exposure expressed as cumulated AUC or AUC intensity (see cumulated AUC in Figure 4). When AUC or C_{max} measured over last dosing interval were used, this relationship was less clear. Consequently, it was chosen to develop the PK-PD model for the 3-organ score cumulated probabilities, conditional on cumulated AUC, and AUC intensity. Based on goodness of fit and parameter uncertainty, only the relationship between cumulated AUC and observed scores was finally considered. In regards to skin, after merging grade 3 and 4 (since only four grades 4 were observed), an E_{max} model with inter-individual variability on logit and EA_{50} (cumulated AUC producing 50 % of maximum effect) gave the best results according to equation:

$$\text{logit}[P(Y \leq j)] = \alpha_j + \left(\frac{E_{max} * \text{cumulatedAUC}^\gamma}{EA_{50}^\gamma + \text{cumulatedAUC}^\gamma} \right) \quad (1)$$

where $P(Y \leq j)$ represents the probability to obtain a score Y for skin inferior or equal to the grade j , α_j represents intercept of the logit for the grade j and γ the coefficient of sigmoidicity. Individual EA_{50i} distribution across the population revealed two groups of patients ($i=1$ or 2), one with high EA_{50} , another with low EA_{50} . A mixture probability was then added to EA_{50i} random effect in order to estimate the proportion of those two sub-populations and their respective EA_{50} . [30] For intestinal tract and liver score, the best results were obtained with a linear model (see equation 2) and an inter-individual variability on the logit. All parameter estimates are presented in Table III.

$$\text{logit}[P(Y \leq j)] = \alpha_j + \text{slope} * \text{cumulatedAUC} \quad (2)$$

where E_{max} model of Equation 1 is replaced by a linear model defined by a slope.

A visual predictive check of the PK-PD models for the three organs (Figure 5) revealed an overall good agreement – between 80 % confidence interval band and observed grade probabilities. The next step consisted in global qualification of the PK-PD analysis. For this purpose we evaluated the model by predictive check. We evaluated whether the combination of the PK model and the 3-organ score models (for skin, intestinal tract and liver), correctly

predicted global effect therapy expressed by the IBMTR score. In this way, our model could be used to verify a treatment effect. The chosen test statistics was the number of the observed grade at a given time, either calculated from the observed data or predicted conditional on the model. Simulation was performed for the overall treatment duration, but test statistic is only presented at treatment start (top of the Figure 6) and day 28 (bottom of the Figure 6), for which 11 patients remained at treatment end. This graph revealed a good agreement of the 90 % confidence interval band with the observed IBMTR score. The combination of the three models to obtain the IBMTR score is qualified and can be used to define the effect of treatment without modelling the IBMTR score itself. The comparison of the two rows revealed that IBMTR scores decreased with treatment time: IBMTR at day 1 had more probability to be observed at a grade 2 or 3 and eventually 4, whereas at day 28 the highest probability observed and predicted is at grade 0 and then 1 or 2. This observation confirmed the global therapy effect as it was already shown in PKPD profiles of each organ.

DISCUSSION

Following promising results observed in 32 steroid-resistant patients who presented aGvHD [31], and more recently, in a retrospective analysis concerning 85 steroid-resistant patients grade II-IV aGvHD [32], inolimomab was proposed as an up-front therapy. As in these clinical trials authors have already observed heterogeneity within organ response with a better and prolonged response for cutaneous aGvHD [32], it therefore clearly appeared that the PD of inolimomab needed to be investigated. Concerning inolimomab and concomitant treatments, patients have received very different exposures in terms of duration as well as dosage. Some authors carried out a multivariate analysis suggesting that a higher total dose of inolimomab might be predictive of a better response.[32] In this context, it also appeared important to identify the PK of this drug [32] in order to take into account in analyses the real inolimomab treatment exposure, and to understand more precisely the PD and its relation to the PK. This type of approach in aGvHD is not very widespread and has not previously involved any type of monoclonal antibody. Only a few authors have already tried to link the PK to the PD for prophylactic treatment. For instance, some noticed a correlation between cyclosporine trough blood concentrations in the early post transplantation period and the probability to observe an aGvHD.[33] By splitting the population into four groups, from no aGvHD, mild aGvHD, moderate aGvHD to severe aGvHD, they observed a decrease in the mean of cyclosporine trough blood concentrations whatever the time periods. Other authors defined binary criteria as the probability to observe at least grade II aGvHD and linked this criteria to busulfan AUC at steady state by a logistic function.[34] Finally, other authors used a threshold value of AUC of unbound mycophenolate in week 1 after transplant to define 2 groups of patients and observed different cumulative proportions of observing a grade II-IV in function of time.[35]

Our study modeled the PK of inolimomab and succeeded to model its exposure-effect relationship. For PK modeling, the population approach taking into account design heterogeneity

and individual treatment history, allowed to identify a two-compartment model. Despite some under predictions in the higher concentrations, observations were on the whole well predicted and the model was qualified for calculating individual treatment exposure, which could not be directly computed from the observed data.

To highlight PK-PD relationships, we initially considered aGvHD according to Glucksberg and IBMTR reference scores, and the Karnofsky classification. Since those combined scores are not arranged in order (i.e. grade 0 < grade I < grade II...), the PK-PD relationship is not easy to reveal. For instance, a grade B in IBMTR classification can correspond to grade 2-0-0 or a grade 0-1-2 for skin, liver and intestinal tract, respectively. We found out that the calculation of IBMTR and Glucksberg grades, as well as of the global performance provided by the Karnofsky score were not adapted to highlight an exposure-effect relationship of inolimomab. Actually, some authors have already identified a better efficacy of this compound on a cutaneous form of aGvHD. [32] In the case of targeted efficacy on one organ, one can easily understand that a composite measure of the effect is not relevant for a PK-PD analysis.

Our PK-PD analysis logically focused on each of the three organs and on treatment exposure. It clearly appeared that a relationship was significant in whatever treatment exposure measurement was chosen. The treatment effect was found saturable for skin and was modelled with an E_{\max} model. A mixture model revealed two populations of patients, sensitive and non sensitive.[36-38] For liver as well as for intestinal tract, effect appeared for high cumulated AUC. It is illustrated by a large decrease of patients with severe symptoms (grade 4) and an important increase of patients without symptoms (grade 0). This means that a larger exposure than for skin is required to reach same efficacy for those organs. It also explains why clinically, skin is the first organ to be cured whereas it seems to be more difficult to treat the liver and the intestinal tract. It is illustrated by PK-PD graphs at day 8: from 15 to 20 % of patients still present highest grade for these two organs. Some authors have explained this phenomenon by the difference of bioavailability or pathophysiology of aGvHD depending on organ involvement.[32]

Although the modelling of the IBMTR score could not be performed, this approach allowed us to easily simulate this score by the combination of the three organs on which it is based. It was also possible to use it to define the overall response at the end of treatment and therefore to verify a treatment effect. However, the benefit of this type of approach is that it takes into account the time dependency of the data. In this way, our models can predict the effect for a given patient and a treatment schedule, in function of time throughout the study (see Figure 7) and also give a more precise idea of the disease evolution under treatment. It is also possible to know if one organ presents a rapid or a slow remission. However, these predictions for the time being must be used with caution. As in our models, the effect is related to cumulated AUC and we modelled the PK-PD relationship by positive monotonous functions, predicted effects automatically increase with time. Therefore, no aGvHD relapse could be predicted by the model. Moreover, based on cumulated AUC, we will obtain the same response for a concentration y during x hours or a concentration x during y hours, regardless of the values of x and y . [39] In these conditions, the dosage or regimen optimization based on modelling is not possible. In fact, we used cumulated AUC because we assumed there would be a delay between concentrations and effect. In our case, the nature of the PD data did not allow us to model this delay in another way. However, perspectives of this work, with future data, will be to link scores to serum concentrations using a more physiologic model taking into account drug and effect accumulation. In this case, dosage and regimen optimization of future trial as well as individual therapeutic monitoring of patients could be performed.

CONCLUSION

Finally, with this analysis, we highlighted and modelled a PK-PD relationship between cumulated AUC of inolimomab and skin, liver and intestinal tract scores. The modelling of the data allowed us to describe observations as well as to predict an overall response at the end of treatment for this population through scores of IBMTR. This approach, validated for its objective, allowed us to understand better the treatment effect over time and represents the first step towards optimizing the doses of future patients. However, it does still present some limitations, due in particular to the limited number of available patients. They were clearly identified, and future trials are needed to improve clinical use of these models.

REFERENCE LIST

1. Storb R. Allogeneic hematopoietic stem cell transplantation--yesterday, today, and tomorrow. *Exp Hematol* 2003 Jan; 31 (1): 1-10
2. Ferrara JL, Deeg HJ. Graft-versus-host disease. *N Engl J Med* 1991 Mar 7; 324 (10): 667-74
3. Storb R, Pepe M, Deeg HJ, et al. Long-term follow-up of a controlled trial comparing a combination of methotrexate plus cyclosporine with cyclosporine alone for prophylaxis of graft-versus-host disease in patients administered HLA-identical marrow grafts for leukemia. *Blood* 1992 Jul 15; 80 (2): 560-1
4. Keever CA, Small TN, Flomenberg N, et al. Immune reconstitution following bone marrow transplantation: comparison of recipients of T-cell depleted marrow with recipients of conventional marrow grafts. *Blood* 1989 Apr; 73 (5): 1340-50
5. Ruutu T, Niederwieser D, Gratwohl A, et al. A survey of the prophylaxis and treatment of acute GVHD in Europe: a report of the European Group for Blood and Marrow, Transplantation (EBMT). Chronic Leukaemia Working Party of the EBMT. *Bone Marrow Transplant* 1997 Apr; 19 (8): 759-64
6. Martin PJ, Schoch G, Fisher L, et al. A retrospective analysis of therapy for acute graft-versus-host disease: secondary treatment. *Blood* 1991 Apr 15; 77 (8): 1821-8
7. Weisdorf D, Haake R, Blazar B, et al. Treatment of moderate/severe acute graft-versus-host disease after allogeneic bone marrow transplantation: an analysis of clinical risk features and outcome. *Blood* 1990 Feb 15; 75 (4): 1024-30
8. Ross WA. Treatment of Gastrointestinal Acute Graft-Versus-Host Disease. *Curr Treat Options Gastroenterol* 2005 Jun; 8 (3): 249-58
9. Akpek G, Lee SM, Anders V, et al. A high-dose pulse steroid regimen for controlling active chronic graft-versus-host disease. *Biol Blood Marrow Transplant* 2001; 7 (9): 495-502
10. Mollee P, Morton AJ, Irving I, et al. Combination therapy with tacrolimus and anti-thymocyte globulin for the treatment of steroid-resistant acute graft-versus-host disease developing during cyclosporine prophylaxis. *Br J Haematol* 2001 Apr; 113 (1): 217-23
11. Curtis RE, Travis LB, Rowlings PA, et al. Risk of lymphoproliferative disorders after bone marrow transplantation: a multi-institutional study. *Blood* 1999 Oct 1; 94 (7): 2208-16
12. Micallef IN, Chhanabhai M, Gascoyne RD, et al. Lymphoproliferative disorders following allogeneic bone marrow transplantation: the Vancouver experience. *Bone Marrow Transplant* 1998 Nov; 22 (10): 981-7

13. Hertenstein B, Stefanic M, Sandherr M, et al. Treatment of steroid-resistant acute graft-vs-host disease after allogeneic marrow transplantation with anti-interleukin-2 receptor antibody (BT563). *Transplant Proc* 1994 Dec; 26 (6): 3114-6
14. Herbelin C, Stephan JL, Donadieu J, et al. Treatment of steroid-resistant acute graft-versus-host disease with an anti-IL-2-receptor monoclonal antibody (BT 563) in children who received T cell-depleted, partially matched, related bone marrow transplants. *Bone Marrow Transplant* 1994 May; 13 (5): 563-9
15. Cuthbert RJ, Phillips GL, Barnett MJ, et al. Anti-interleukin-2 receptor monoclonal antibody (BT 563) in the treatment of severe acute GVHD refractory to systemic corticosteroid therapy. *Bone Marrow Transplant* 1992 Nov; 10 (5): 451-5
16. Herve P, Bordigoni P, Cahn JY, et al. Use of monoclonal antibodies in vivo as a therapeutic strategy for acute GvHD in matched and mismatched bone marrow transplantation. *Transplant Proc* 1991 Feb; 23 (1 Pt 2): 1692-4
17. Herve P, Wijdenes J, Bergerat JP, et al. Treatment of corticosteroid resistant acute graft-versus-host disease by in vivo administration of anti-interleukin-2 receptor monoclonal antibody (B-B10). *Blood* 1990 Feb 15; 75 (4): 1017-23
18. Glucksberg H, Storb R, Fefer A, et al. Clinical manifestations of graft-versus-host disease in human recipients of marrow from HL-A-matched sibling donors. *Transplantation* 1974 Oct; 18 (4): 295-304
19. Thomas ED, Storb R, Clift RA, et al. Bone-marrow transplantation (second of two parts). *N Engl J Med* 1975 Apr 24; 292 (17): 895-902
20. Rowlings PA, Przepiorka D, Klein JP, et al. IBMTR Severity Index for grading acute graft-versus-host disease: retrospective comparison with Glucksberg grade. *Br J Haematol* 1997 Jun; 97 (4): 855-64
21. Karnofsky DA, Burchenal JH. The clinical evaluation of chemotherapeutic agents in cancer. In: Macleod CM, editor. *Evaluation of Chemotherapeutic Agents*. New York: Columbia University Press, 1949: 199-205.
22. Boeckmann AJ, Sheiner L, Beal SL. *NONMEM User's Guides*. NONMEM project group. San Francisco: University of California, 1998
23. Ette EI, Ludden TM. Population pharmacokinetic modeling: the importance of informative graphics. *Pharm Res* 1995 Dec; 12 (12): 1845-55
24. Food and Drug Administration. Guidance for industry - Population pharmacokinetics. U.S. Department of Health and Human Services. Food and Drug Administration. Center for Drug Evaluation and Research (CDER). Center for Biologics Evaluation and Research (CBER).; 1999 Feb.

25. Wade JR, Edholm M, Salmonson T. A guide for reporting the results of population pharmacokinetic analyses: a swedish perspective. *AAPS PharmSci* 2005 Oct; 7 (2): 45
26. Agresti A. Modelling ordered categorical data: recent advances and future challenges. *Stat Med* 1999 Sep 15; 18 (17-18): 2191-207
27. Duffull SB, Chabaud S, Nony P, et al. A pharmacokinetic simulation model for ivabradine in healthy volunteers. *Eur J Pharm Sci* 2000; 10 (4): 285-94
28. Yano Y, Beal SL, Sheiner LB. Evaluating pharmacokinetic/pharmacodynamic models using the posterior predictive check. *J Pharmacokinet Pharmacodyn* 2001 Apr; 28 (2): 171-92
29. Rubin DB. Bayesianly justifiable and relevant frequency calculations for the applied statistician. *Ann Stat* 1984; 12: 1151-72
30. Shiiki T, Hashimoto Y, Inui K. Simulation for population pharmacodynamic analysis of dose-ranging trials: usefulness of the mixture model analysis for detecting nonresponders. *Pharm Res* 2002 Jun; 19 (6): 909-13
31. Cahn JY, Bordigoni P, Tiberghien P, et al. Treatment of acute graft-versus-host disease with methylprednisolone and cyclosporine with or without an anti-interleukin-2 receptor monoclonal antibody. A multicenter phase III study. *Transplantation* 1995 Nov 15; 60 (9): 939-42
32. Bay JO, Dhedin N, Goerner M, et al. Inolimomab in steroid-refractory acute graft-versus-host disease following allogeneic hematopoietic stem cell transplantation: retrospective analysis and comparison with other interleukin-2 receptor antibodies. *Transplantation* 2005 Sep 27; 80 (6): 782-8
33. Martin P, Bleyzac N, Souillet G, et al. Relationship between CsA trough blood concentration and severity of acute graft-versus-host disease after paediatric stem cell transplantation from matched-sibling or unrelated donors. *Bone Marrow Transplant* 2003 Oct; 32 (8): 777-84
34. Andersson BS, Thall PF, Madden T, et al. Busulfan systemic exposure relative to regimen-related toxicity and acute graft-versus-host disease: defining a therapeutic window for i.v. BuCy2 in chronic myelogenous leukemia. *Biol Blood Marrow Transplant* 2002; 8 (9): 477-85
35. Jacobson P, Rogosheske J, Barker JN, et al. Relationship of mycophenolic acid exposure to clinical outcome after hematopoietic cell transplantation. *Clin Pharmacol Ther* 2005 Nov; 78 (5): 486-500
36. Beal SL, Sheiner LB. *NONMEM User's Guide - Part VII. Conditional Estimation Methods*. San Francisco: University of California, 1998
37. Frey N, Laveille C, Paraire M, et al. Population PKPD modelling of the long-term hypoglycaemic effect of gliclazide given as a once-a-day modified release (MR) formulation. *Br J Clin Pharmacol* 2003 Feb; 55 (2): 147-57

38. Zingmark PH, Ekblom M, Odergren T, et al. Population pharmacokinetics of clomethiazole and its effect on the natural course of sedation in acute stroke patients. *Br J Clin Pharmacol* 2003 Aug; 56 (2): 173-83

39. Karlsson MO, Molnar V, Bergh J, et al. A general model for time-dissociated pharmacokinetic-pharmacodynamic relationship exemplified by paclitaxel myelosuppression. *Clin Pharmacol Ther* 1998 Jan; 63 (1): 11-25

TABLES

Table I. Patient's disease, pretreatment, and donor characteristics.

Characteristics	Number
Initial disease	
acute myeloblastic leukemia (AML)	5
chronic lymphoid leukemia (CLL)	2
non hodgkin lymphoma (NHL)	2
hodgkin disease	1
myelodisplasia	2
chronic myeloid leukemia (CML)	1
acute lymphoblastic leukemia (ALL)	1
multiple myeloma	1
myeloid splenomegaly	1
solid tumors	5
Transplantation	
first	17
second	4
Type	
peripheral blood stem cell	15
bone marrow	6
Status after	
complete remission	8
partial response	6
stable disease	0
relapse	1
progressive disease	5
chronic phase	1
Conditioning regimen	
non myeloablative	13
myeloablative	8
Acute GVHD prophylaxis	
Cyclosporine	
yes	20
no	1
Methotrexate	
yes	8
no	13
Steroids	
yes	0
no	21
Other drug	
yes	10
no	11
Donor and recipient compatibility	
Gender	
yes	10
no	11
ABO	
yes	9
minor incompatibility	7
major incompatibility	5
Histocompatibility (HLA)	
matched sibling donor	17
matched unrelated donor	2
mismatched unrelated donor	2

Table II. Population pharmacokinetic parameters obtained from the final model.

Parameters	Estimate	SE (%)
Fixed effects		
Clearance (l/hr)	0.077	19
Volume of compartment 1 (Volume 1) (l)	2.76	26
Inter-compartmental clearance (l/hr)	2.22	52
Volume of compartment 2 (Volume 2) (l)	2.25	18
Random effects		
IIV Clearance ^a	43%	87
IIV Volume 1	68%	90
IIV Inter-compartmental clearance	43%	425
ρ IIV Clearance, Volume1 ^b	-0.05	
ρ IIV Clearance, Inter-compartmental clearance	-0.76	
ρ IIV Volume 1, Inter-compartmental clearance	-0.62	
Residual error	0.0913	7

^a **IIV** = inter-individual variability on a fixed effect; ^b **ρ** = correlation coefficient between all individual estimations of two parameters.

Table III. Population pharmacodynamic parameters obtained from the final models.

Parameters	Estimate	SE (%)
Skin model		
Fixed effects		
logit of baseline probability, grade 0 (alpha 0)	-4.79	21
logit of baseline probability, grade 1 (alpha 1)	3.14	17
logit of baseline probability, grade 2 (alpha 2)	2.64	25
E _{max} (maximum effect)	13.4	23
Sigmoidicity factor	1	^a
EA ₅₀ population 1	15900	35
EA ₅₀ population 2	609	36
Population 1 repartition	0.35	31
Random effects		
IIV EA ₅₀	0.1	45
IIV logit	7.2	57
Gut model		
Fixed effects		
logit of baseline probability, grade 0 (alpha 0)	-0.3	^a
logit of baseline probability, grade 1 (alpha 1)	1.72	13
logit of baseline probability, grade 2 (alpha 2)	1.55	25
logit of baseline probability, grade 3 (alpha 3)	1.75	33
slope	0.000339	60
Random effects		
IIV logit	16.5	36
Liver model		
Fixed effects		
logit of baseline probability, grade 0 (alpha 0)	2	^a
logit of baseline probability, grade 1 (alpha 1)	2.42	14
logit of baseline probability, grade 2 (alpha 2)	3.19	19
slope	0.000429	87
Random effects		
IIV logit	50.2	51

^a fixed parameter.

ANNEXE 1. Criteria for IBMTR severity index for acute GvHD.

Index ^a	Skin involvement		Liver involvement		Gastrointestinal involvement	
	Stage (max)	Extent of rash	Stage (max)	Total bilirubin (µmol/l)	Stage (max)	Volume of diarrhoea (ml/d)
A	1	< 25%	or 0	< 34	or 0	< 500
B	2	25-50%	or 1-2	34-102	or 1-2	550-1500
C	3	> 50%	or 3	103-255	or 3	> 1500
D	4	Bullae	or 4	> 255	or 4	Severe pain and ileus

^a assign index based on maximum involvement in an individual organ system.

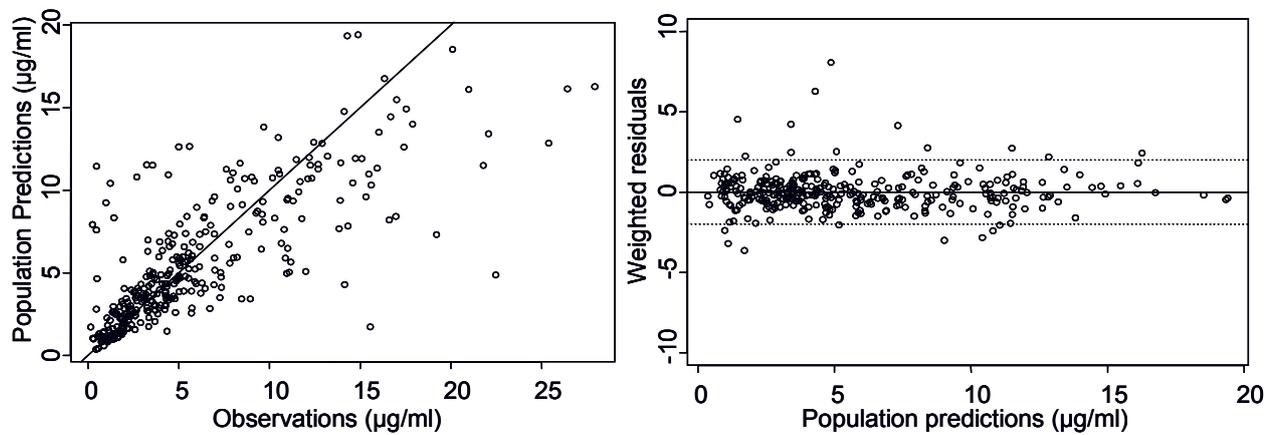


Figure 1. Goodness of fit plots of final PK model. Right plot: solid line represents identity line of population predictions and observation. On weighted residual plots, solid line define zero and dotted lines values for -1.96 and +1.96, the 2.5 and 97.5% quantiles of normal distribution.

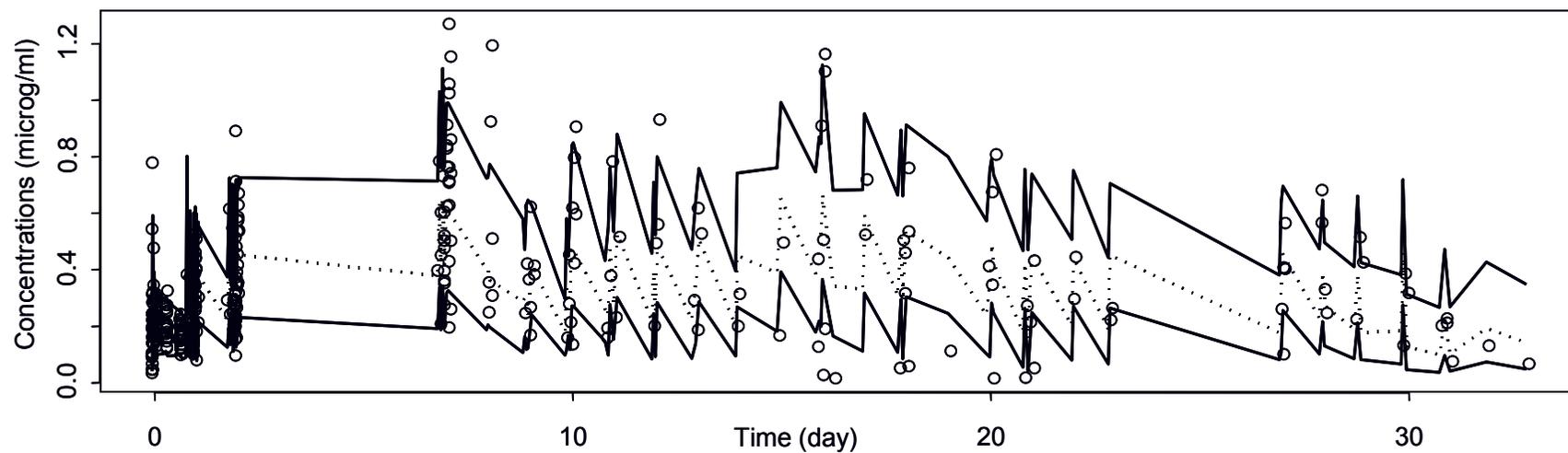


Figure 2. Visual predictive check of final PK model from 200x21 simulated patients. In order to normalize the scale, all concentrations (observed and predicted) are divided by actual received doses. Black lines: 80% confidence interval of predicted concentrations. Dotted line: median of predicted concentrations. Black circles: observed concentrations.

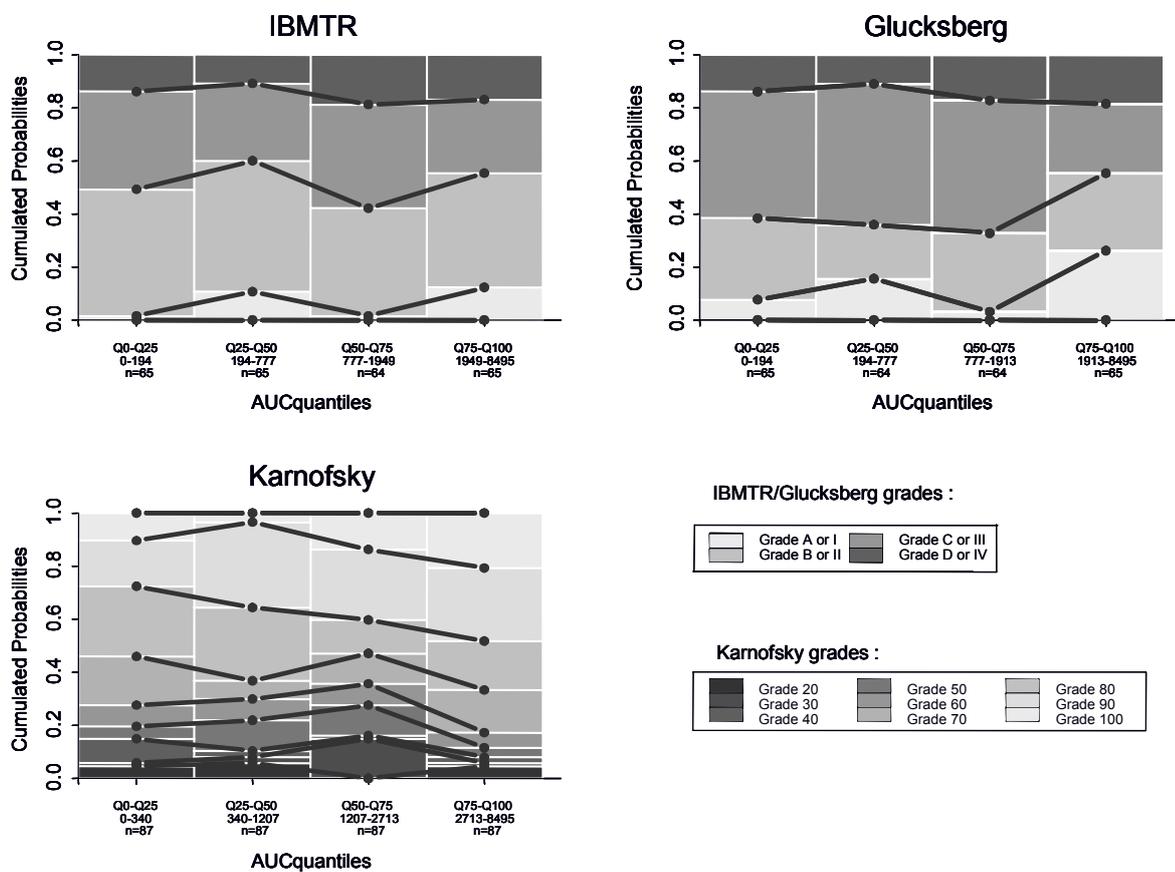


Figure 3. Observed cumulative probabilities of composite scores (IBMTR, Glucksberg, and Karnofsky) function of predicted cumulative AUC. Data are split in 4 intervals according to quantiles 25, 50 and 75%. It allows for a sufficient number of data (at least n=65), to represent evolution of probability to observe each grade versus different values of predicted cumulative AUC. Solid black line: link between observed cumulative probabilities of a same grade. Black, gray and white bars: observed cumulative probabilities ($P(Y \leq j)$).

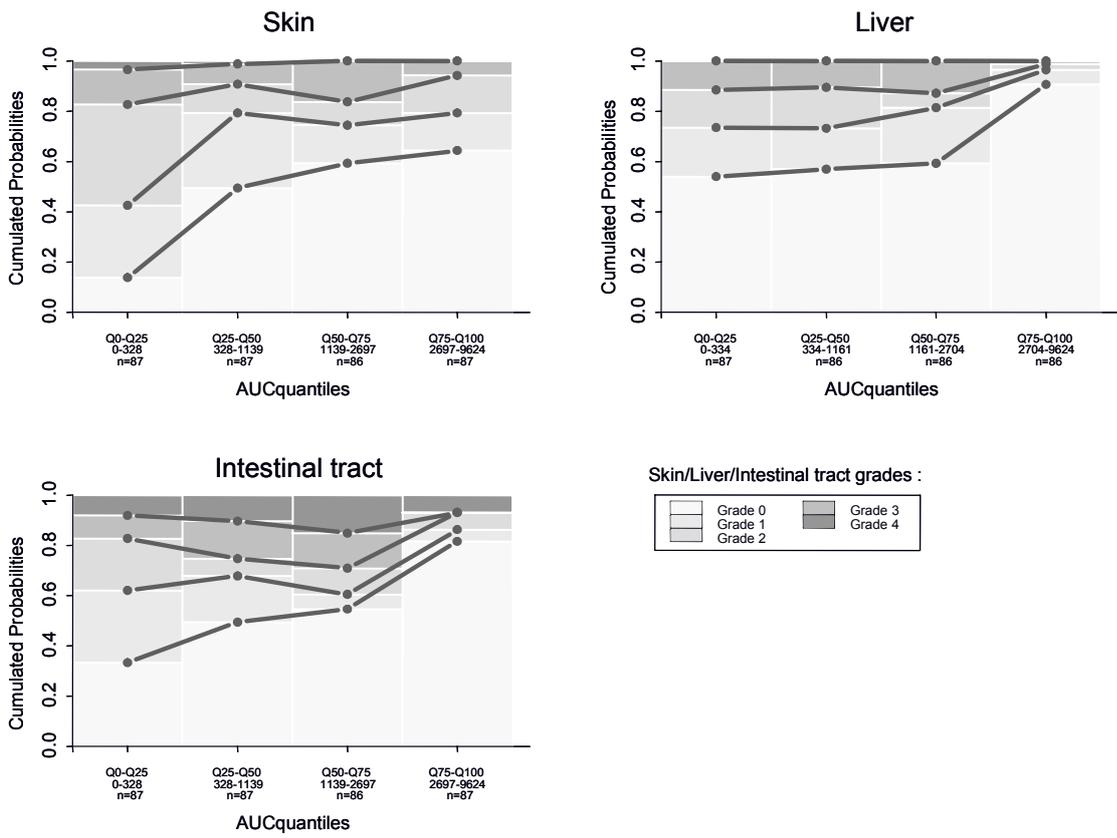


Figure 4. Observed cumulated probabilities of organs scores (Skin, Liver , and Intestinal tract) function of predicted cumulated AUC. For other details, see legend Fig. 4..

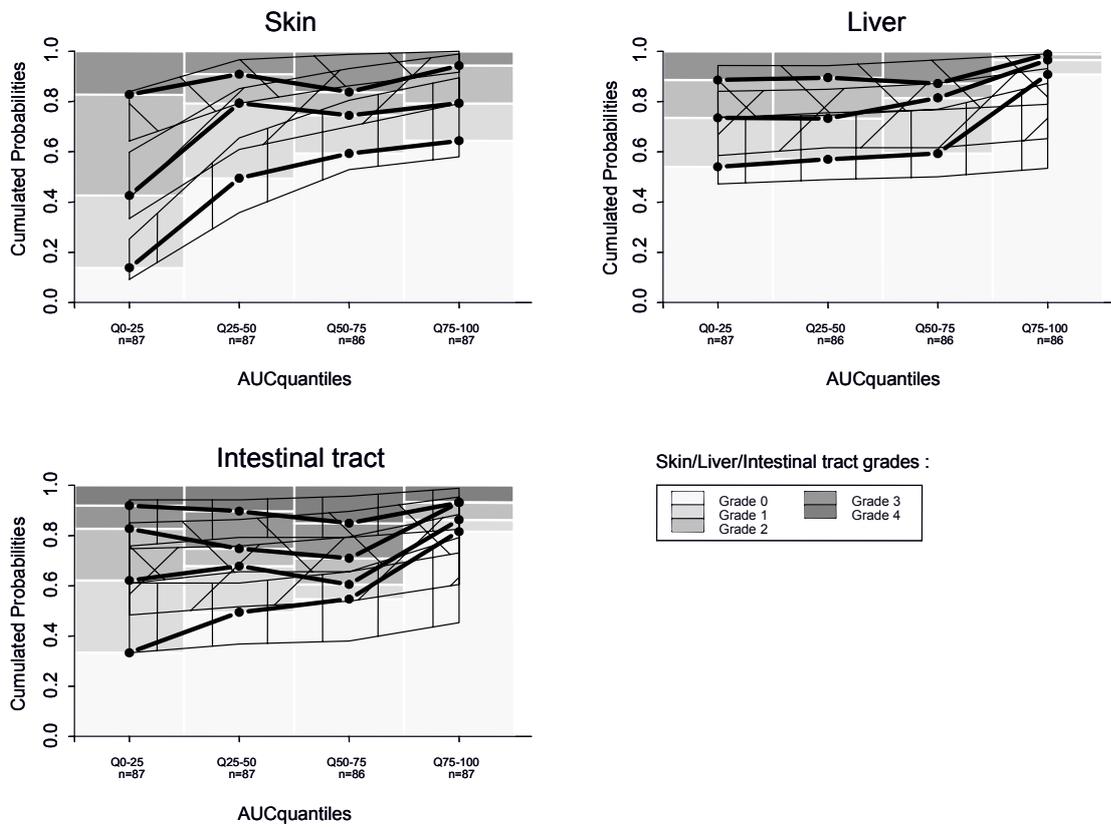


Figure 5. Visual predictive check of final PK-PD models from 200x21 simulated patients.

Observations plotted in Figure 4 are compared with model predictions. Thin, solid and black line: prediction and 80% confidence interval of predicted cumulated probabilities.

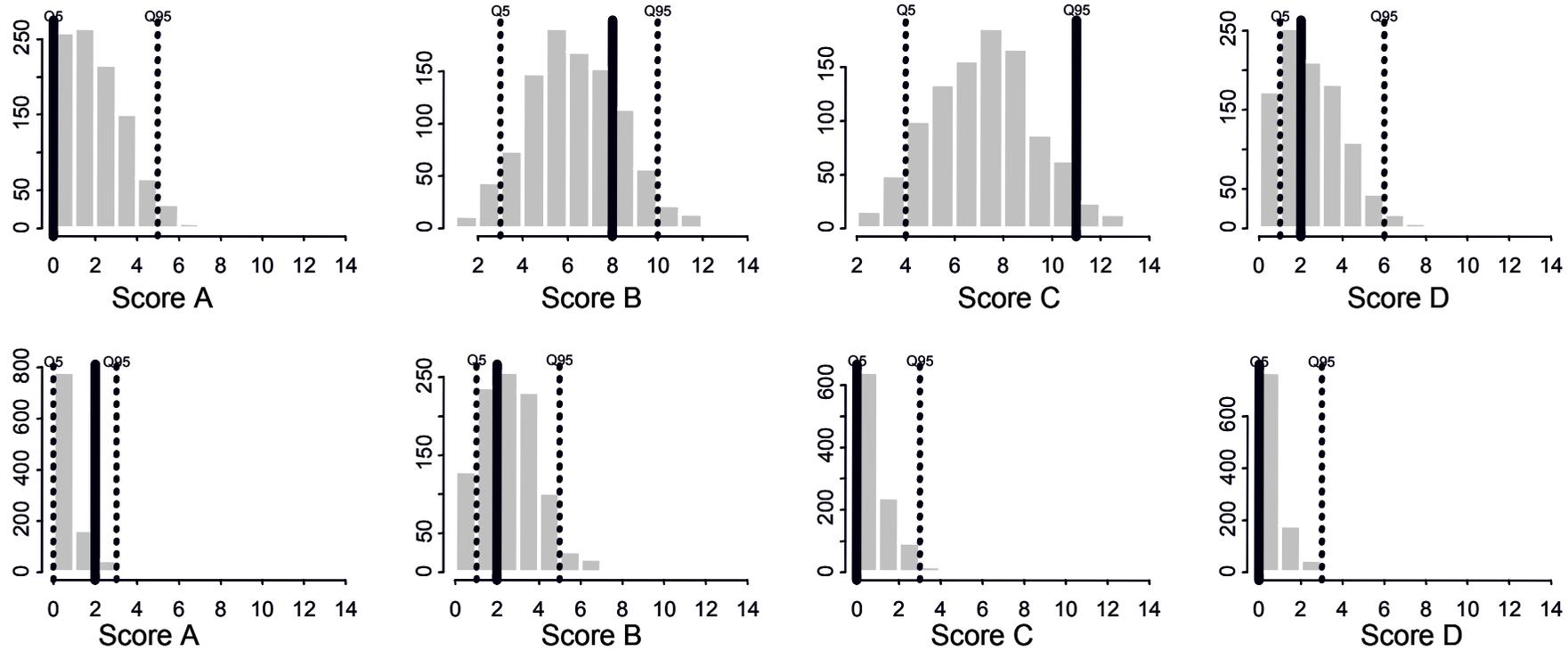


Figure 6. Predictive check of final PK-PD models. Histogram of IBMTR scores from 1000x21 simulated patients at treatment start (top row) and from 1000x11 simulated patients still in the study at day 28 (bottom row). Dotted line: 90% confidence interval of simulated IBMTR. Black line: observed IBMTR in 21 patients at treatment start (top row) and in 11 patients still in the study at day 28 (bottom row).

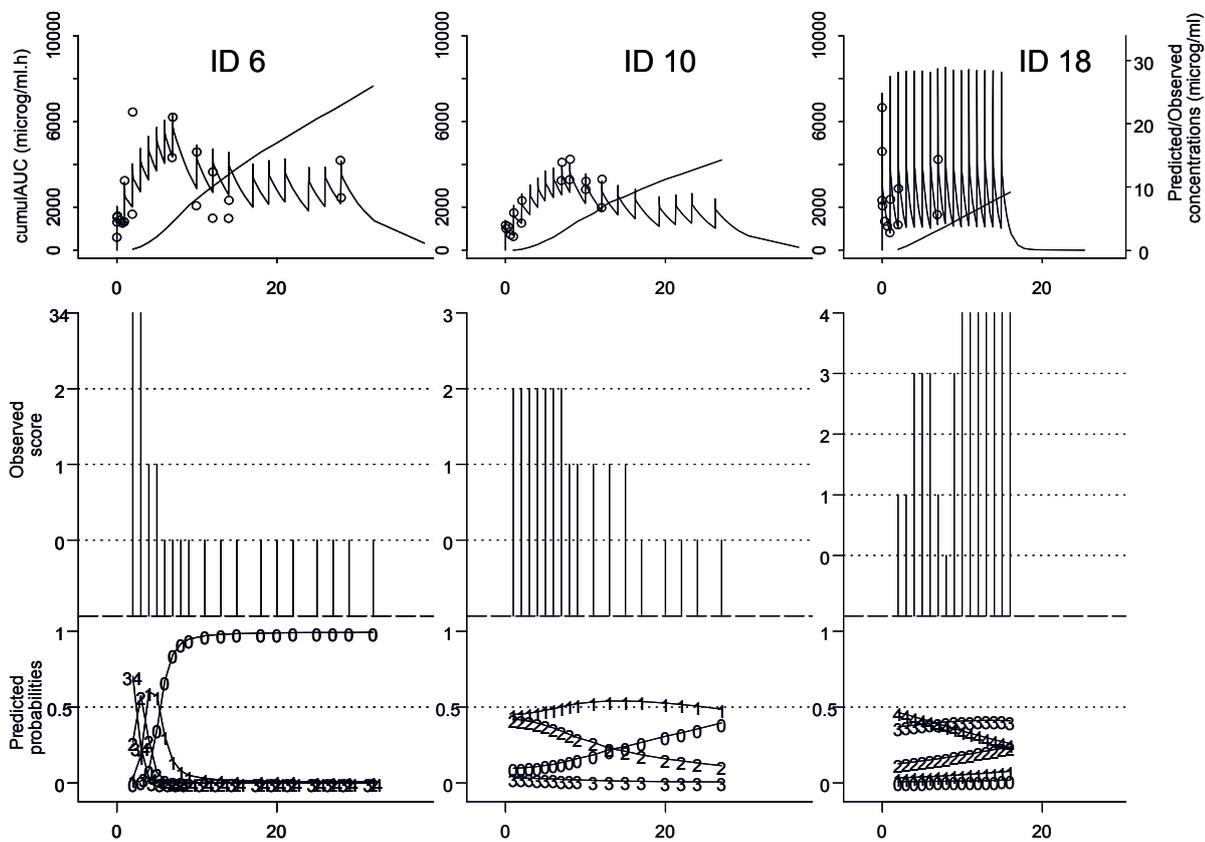


Figure 7. Observations and individual predictions versus time of final PK and PKPD models for 3 patients enrolled in the clinical trial. On the top: Observed concentrations (O) and PK predictions as well as predicted corresponding cumulated AUC (-). In the middle: observed PD grades. In the bottom: predicted probability of a grade observation. For ID 6, PD results for the skin, for ID 10, PD results for the liver, and for ID 18, PD results for intestinal tract.