Prothrombotic markers and early spontaneous recanalization in ST-segment elevation myocardial infarction.
Marie-Geneviève Huisse, Emilie Lanoy, Didier Tcheche, Laurent Feldman, Annie Bezeaud, Eduardo Anglès-Cano, Murielle Mary-Krause, Dominique De Prost, Marie-Claude Guillin, Ph.Gabriel Steg

To cite this version:

HAL Id: inserm-00160732
http://www.hal.inserm.fr/inserm-00160732
Submitted on 22 Oct 2007

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.
PROTHROMBOTIC MARKERS AND EARLY SPONTANEOUS RECANALIZATION IN ST-SEGMENT ELEVATION MYOCARDIAL INFARCTION

Journal: *Thrombosis and Haemostasis*

Manuscript ID: TH-06-11-0621.R3

Manuscript Type: Basic/Clinical Studies: cardiovascular biology and cell signalling

Date Submitted by the Author: 21-May-2007

Complete List of Authors: Huisse, Marie-Genevieve; Hopital Bichat, Haematology; Lanoy, Emilie; INSERM U720; Tchetch, Didier; Hopital Bichat, Cardiology; Feldman, Laurent; Hopital Bichat, Cardiology; Bezeaud, Annie; INSERM U698; Angles-Cano, Eduardo; INSERM U698; Mary-Krause, Murielle; INSERM U720; de Prost, Dominique; INSERM U698; Guillin, Marie-Claude; Hopital Bichat, Haematology; INSERM U698; Steg, Ph. Gabriel; Hopital bichat, Cardiology

Keywords: Thrombin, Tissue factor / factor VII, Microparticles, Acute myocardial infarction, Risk factors
PROTHROMBOTIC MARKERS AND EARLY SPONTANEOUS RECANALIZATION IN ST-SEGMENT ELEVATION MYOCARDIAL INFARCTION

Marie-Geneviève Huisse\textsuperscript{1,4,5}, Emilie Lanoy\textsuperscript{3,6}, Didier Tcheche\textsuperscript{2}, Laurent J. Feldman\textsuperscript{2,5}, Annie Bezeaud\textsuperscript{4,5}, Eduardo Anglès-Cano\textsuperscript{5}, Murielle Mary-Krause\textsuperscript{3,6}, Dominique de Prost\textsuperscript{4,5}, Marie-Claude Guillin\textsuperscript{1,4,5}, Ph. Gabriel Steg\textsuperscript{2,5}.

AP-HP, Hôpital Bichat, Departments of \textsuperscript{1}Haematology and \textsuperscript{2}Cardiology, \textsuperscript{4}CIB PhenoGen; \textsuperscript{5}INSERM U698 and \textsuperscript{3}U720; University Paris\textsuperscript{7}-Denis Diderot, \textsuperscript{6}University Pierre et Marie Curie-Paris 6, Paris, France,

Correspondence to: Dr Marie-Geneviève HUISSE

Service d’Hématologie, Hôpital Bichat
46 rue Henri Huchard, 75018, Paris, France
Tel 33.1.40.25.85.21
Fax 33.1.40.25.88.53
e-mail marie-genevieve.huisse@bch.aphp.fr

Running Title: Thrombin and Plasmin Generation in early Recanalization

Financial support: This work was supported by funds from Fondation de France. Additional support was provided by Diagnostica Stago.
Abstract

We tested the hypothesis that selected prothrombotic biomarkers might be associated with early spontaneous coronary recanalization in patients with ST-segment elevation acute myocardial infarction (STEMI).

We prospectively enrolled 123 patients with STEMI including 53 patients with spontaneous coronary recanalization (Cases) and 70 patients with persistent occlusion (Controls) at the time of emergent coronary angiography and before angioplasty. All had received aspirin and heparin. Blood samples were collected immediately before angioplasty to measure soluble P-selectin, circulating microparticles originating from platelets (PMPs), granulocytes (GMPs), endothelial cells (EMPs); tissue factor-associated MP (TF-MP); soluble platelet glycoprotein V (sGPV) and prothrombin F1+2; tissue plasminogen activator (tPA), plasminogen activator inhibitor (PAI-1) and plasmin-antiplasmin (PAP). A sub-group of 70 patients (35 cases, 35 controls) was available for flow cytometry analysis of platelet P-selectin and activated GPIIb-IIIa. Baseline clinical characteristics did not differ between groups except for more frequent hypertension and dyslipidemia in Controls. Platelet activation markers and PMP did not differ between the two groups. Controls had higher numbers of EMPs and GMPs compared to Cases, but the difference was no longer significant when corrected for risks factors. Controls differed from Cases by higher plasma levels of sGPV [64 (47-84) ng/ml vs 53 (44-63) ng/ml] and PAP [114(65-225) ng/ml vs 88 (51-147) ng/ml]. The difference persisted after adjustment for risks factors (p=0.031 and 0.037, respectively). Persistent occlusion of the infarct related artery is associated with some markers related to higher thrombin (sGPV) and plasmin (PAP) production but is not associated with markers of platelet activation.
KEYWORDS: myocardial infarction, thrombin, soluble Glycoprotein V, endothelial injury, plasmin-antiplasmin.

Introduction

Acute myocardial infarction (AMI) results from the disruption of unstable atheromatous plaques exposing thrombogenic material to blood flow and initiating the
formation of an occluding arterial thrombus. Platelet activation is triggered by contact with collagen in the extracellular matrix of the plaque, while tissue factor produced by macrophages and smooth muscle cells induces thrombin formation (1). Thrombin amplifies the activation of platelets and converts fibrinogen to fibrin, yielding the characteristic arterial thrombus formed of platelets entrapped in fibrin. Furthermore, microparticles (MPs) resulting from cell activation/apoptosis within the atheromatous plaques (2) contribute to plaque thrombogenicity and may disseminate blood-borne tissue factor activity and procoagulant phospholipids upon plaque rupture (3). Natural fibrinolytic mechanisms (plasminogen activators tPA and uPA) contribute to the dissolution of arterial thrombi. However, high levels of the plasminogen activator inhibitor PAI-1 (associated with several genetic or environmental factors) limit the efficacy of spontaneous fibrinolysis and this factor has been recognized to contribute to cardiovascular risk (4).

Primary percutaneous coronary intervention (PCI) is now established as the reference therapy for the management of ST segment elevation AMI (5). Before PCI, approximately 15-18 % of patients present with angiographically proven spontaneous patency of the infarct artery. These patients have less myocardial damage and a better outcome than patients with occluded arteries (6, 7). In the present prospective study, we sought to determine whether patients with spontaneous recanalization of the infarct related artery differ from patients with persistent occlusion at the time of initial angiography, before PCI, in terms of selected biomarkers of cell stimulation, coagulation and/or fibrinolysis activation. This analysis may provide insight into the optimal pathways for improving current pharmacologic therapies designed to recanalize infarct arteries.

**METHODS**

**Study design**
We undertook a prospective case-control study comparing patients who presented with patent artery at the time of emergency coronary angiography (cases) and patients with occluded artery (controls).

**Study patients:**

**Inclusion and exclusion criteria**

Patients were eligible for inclusion if they were between 18-80 year old, with symptoms of acute coronary syndrome within the past 12 hours, ST-segment elevation of greater than 1 mm on their electrocardiogram and creatine kinase twice the upper limit of normal. Patients were excluded if they presented with life-threatening arrhythmia, hemodynamic instability or shock or if they had received within 30 days (including the present episode), thrombolitics, GpIIb/IIIa receptor blockers, ticlopidine or clopidogrel, or had undergone angioplasty in the preceding 6 months. From October 2000 to December 2003, we enrolled a total of 123 patients who were triaged to primary percutaneous transluminal coronary angioplasty (PTCA). Patients were transported promptly to the catheterization laboratory and underwent immediate coronary angiography. Flow was assessed at the first contrast injection via the guiding catheter before any wire crossing. All patients had received sublingual or IV nitroglycerin.

**Definition of Cases and Controls**

Coronary flow was evaluated quantitatively according to the Thrombolysis In Myocardial Infarction (TIMI) grading system (8). Fifty three patients had a grade TIMI 2-3 and constituted the cases whereas 70 patients had a grade TIMI 0-1 and constituted the controls. All the patients had received 250 to 500 mg aspirin and 5000 UI of unfractionated heparin before blood collection and angiography. The vast majority received abciximab just before angiography but after blood collection was performed (Table I) The protocol was
approved by the local Ethics Committee. Written informed consent was obtained from all patients.

**Healthy volunteers**

Normal reference values for each prothrombotic markers was determined in 24 healthy volunteers 24-65 year old (median 45), 12 females and 12 males, free of known cardiovascular risks factors. Reference values for plasmin-antiplasmin complexes were determined by one of the co-authors (EAC) in a large cohort including 125 healthy volunteers 21-80 year old (median 52) comprising 56 females and 69 males. All these individuals have given their informed consent.

**Blood sampling and processing:**

Venous peripheral blood was collected from all patients on admission, just before angiography and treatment with abciximab. Blood was drawn in evacuated tubes (Vacutainer®, Becton-Dickinson) containing 0.129 M trisodium citrate (1 vol / 9 vol blood), for all assays but plasma soluble GPV (sGPV) measurement (tubes contained CTAD: 0.109M sodium citrate, citric acid, theophylline, adenosine, and dipyridamole) and fibrinolysis parameters analysis (tubes contained CTAD, 100 nM PPACK and 10 UI/ml aprotinin). Blood was centrifuged within one hour either at 2,500 g for 20 minutes at 14°C for plasma proteins assays or at 1,500 g for 20 minutes at 20°C for microparticles (MPs) isolation. Plasma was kept frozen at – 80°C until analysis.

**Whole blood platelet activation markers:**

Platelet surface P-Selectin (CD62P, Immunotech, Le Pont-de-Chaix, France) and activated GPIIbIIIa (PAC-1, IgMκ, Beckton-Dickinson) were analyzed in whole blood by flow cytometry within two hours of blood collection. Preliminary experiments demonstrated
that these parameters were stable up to two hours after blood collection which limited the
flow cytometry study to a sub-group of 35 TIMI 2-3 patients and 35 TIMI 0-1 patients
corresponding to patients included when flow cytometry could be rapidly performed. Whole
blood samples (5μl) diluted 1:10 in PBS were incubated with saturating concentrations of
fluorescein-conjugated antibodies CD62P, PAC-1 or isotype-matched controls for 30 minutes
in the dark and after addition of 1 ml PBS, immediately analyzed by flow cytometry.
Percentage of positive platelets were determined as compared to isotype-matched controls.
Normal values were less than 2 % for each marker.

**Microparticles (MPs) isolation and characterization:**

Microparticles were isolated according to Nieuwland et al (9) and analyzed in a
coulter epics XL™ with expo 32 software (beckman coulter). In brief, MPs were extracted
from plasma within 2 months of storage at –80°C by centrifugation at 18 000g for 20 min at
RT, the pellet being washed once in working buffer (WB:10 mM Hepes pH 7.35, 136 mM, 5
mM KCl, 2 mM MgCl2) containing either 5 mM EDTA (first washing) or no EDTA (for the
second washing). The pellet was finally resuspended in WB and directly analyzed by flow
cytometry. Extracted MPs (5 μl) were incubated for 30 min in the dark with 45 μl WB
containing 2 mM CaCl2 and 5 μl fluorochrome-conjugated probes, consisting of fluorescein-
isothiocyanate (FITC)-annexin V (immunotech ) and phycoerythrin(PE)-conjugated specific
monoclonal antibodies (MoAbs). MoAbs included anti-CD15 (Lewisx, clone 80175, IgM),
anti-CD41(GPⅠb, clone P2, IgG1), anti-CD106 (VCAM1, clone 5110 C9, IgG1) or isotype
controls IgG1 (MOPC21) or IgM (G155-228) (from Immunotech, and PharMingen, San José,
CA). Normal values [median(IQR)] of annexinV-positive MPs (total MPs) in 24 normal
healthy adult volunteers were 365 (281-596) /μl, originating from platelets (PMPs: 87%),
granulocytes (GMPs: 5%) and endothelial cells (EMPs: 1.5%).
Microparticle-linked tissue factor activity.

Tissue factor (TF) activity associated to MPs (TF-MP) was determined through the ability of MPs to promote the activation of FX by FVIIa, as described by Key et al (10). In brief, the MPs suspension was mixed with 1 nM recombinant FVIIa (gift from Novo Nordisk, Måløv, Denmark) and 250 nM human FX (Enzyme Research Laboratory, South Bend, IN, USA). After 3 minutes incubation at 37°C, normal plasma containing 12.5 μM phospholipids (20% phosphatidyl-serine / 80% phosphatidyl-choline) was added and the clotting time was recorded after addition of 5 mM CaCl₂. A standard curve was constructed using relipidated human recombinant TF (American Diagnostica, Greenwich, CT, USA), and results were expressed as pg/ml of TF-MP. The specificity of the assay was ascertained by the use of inactivated FVIIa (VIIai, Novo Nordisk,) or a blocking anti-TF MoAb (American Diagnostica) which both completely abolished FXa generation in presence of TF. Normal values [median (IQR)] established in 20 healthy adult volunteers were 35.5 (29-50) pg/ml.

Soluble P-selectin (sPselectin), soluble GPV (sGPV) and Prothrombin fragment 1+2 (F 1+2)

Plasma levels of sPselectin, sGPV and F 1+2 were measured using enzyme-linked immunosorbent assays (ELISA) (sPselectin, R&D systems Europe, Lille; sGPV, Serbio, Gennevilliers, France and Enzygnost F 1+2, Dade-Behring, Marburg, Germany, respectively). Normal values for plasma sGPV [median (IQR)] were 25.1 (14.8 – 39.9) ng/ml, very similar to the values recently reported in 300 healthy blood donors (11). Normal values indicated by the manufacturer for F 1+2 (median and 5th to 95th percentile) was 115 (69-229) pmol/L.

Biomarkers of fibrinolysis.
The mass concentrations of tPA and PAI-1 were measured by ELISA using commercially available reagents from Serbio, France. Plasmin-\(\alpha\)2 antiplasmin complexes (PAP) were measured using a local ELISA, using specific antibodies as described by Montes et al (12). Normal values [median (IQR)] established from 125 healthy controls were 24.8 (7.3-35.4) ng/ml.

**Statistical analysis**

Continuous variables were expressed as median and interquartile range (IQR), and distributions of qualitative variables were presented with number of patients and percentages. All STEMI patients were included in the analyses (except for platelet P-selectin and activated GpIIb-IIIa which were available in 35 cases and 35 controls). We first tested the relation between the baseline characteristics and the status of the patients: case (i.e spontaneous coronary recanalization) or control (persistent occlusion) in an univariate analysis with the Mann-Whitney test (continuous variables), Chi square test or Fischer exact test as appropriate (qualitative variable).

When biological markers associated with the case versus control status with a p value \(\leq 0.20\) in the univariate analysis, they were included simultaneously in a multivariate logistic regression model adjusted on the known coronary risk factors (sex, age, smoking status, hypertension, diabetes mellitus, dyslipidemia, anterior MI, delay of revascularization) to evaluate their specific effects. The presence of an interaction between selected biological markers and each of the clinical variables, was also tested in the logistic regression model. All analyses were performed with SPSS statistical software version 11.0 (SPSS Inc., Illinois, Chicago).

A Spearman correlation was computed between biological variables and TF activity.
RESULTS

Patients characteristics

Among the 123 patients enrolled in this prospective study, 53 achieved TIMI 2 to 3 flow (15 TIMI 2 and 38 TIMI 3 flow) and 70 had TIMI 0-1 flow on baseline (pre-intervention) coronary angiography. Patients with TIMI grade 2 patency did not differ from patients with TIMI 3 flow in terms of biomarkers and the data were therefore pooled together (TIMI 2-3) for comparison with TIMI 0-1 patients. Killip class I was not different between groups (88.7% v 91.4% respectively in cases v controls), as well as Killip class II (11.3% v 8.6% in cases v controls). Systolic (SBP) and Diastolic Blood pressure (DBP) did not differ either between groups [125(113-152) mmHg, median(IQR) v 128(112-145)] and [76(69-90) mmHg v 75(68-90)] in cases v controls respectively. The baseline characteristics are listed in Table 1 and were similar between groups, with the exception of dyslipidemia and hypertension, which were less frequent in patients with TIMI 2-3 flow (spontaneous recanalization) than in TIMI 0-1 patients (occluded artery). The time delay from symptoms onset and recanalization (spontaneous or instrumental) did not differ between the two groups.

Cell activation markers.

As previously observed by others (13), platelet surface expression of P-selectin and activated GPIIb/IIIa was increased in patients compared to healthy controls. However, no significant difference was observed between TIMI 0-1 and 2-3 patients (Table 2). Soluble P-selectin was not different between groups, 42 (31.9-54.2) ng/ml, median (IQR) and 40 (33.3-49.2) ng/ml in TIMI 0-1 and TIMI 2-3 respectively. Likewise, the platelet-derived MPs (PMPs) were not different in the two groups of patients (Table 3). In contrast, the numbers of both GMPs and EMPs were significantly lower in patients with patent infarct arteries compared to patients with occluded vessels (Table 3, p=0.021 and 0.002, respectively). The
difference disappeared after adjustment for baseline risk factors. EMPs in TIMI 2-3 patients correlated with SBP (r=0.307, p=0.038).

**Tissue factor.**

Tissue factor activity associated to MPs (TF-MP) did not differ from normal values in TIMI 2-3 patients, but was significantly increased in TIMI 0-1 patients compared to healthy controls (p=0.0012). However, although levels of TF-MP tended to be lower in patients with patent vs occluded vessels (Table 3), the difference did not reach significance. TF-MP activity correlated with PMPs numbers (r=0.287, p = 0.0012).

**Prothrombin F1+2 and sGPV**

Plasma levels of prothrombin F1+2 were not significantly different between the two patients’ groups [136 (60-211) pM/l and 151 (57-246) pM/l, median (IQR), for TIMI 2-3 and TIMI 0-1 patients, respectively]. We observed a negative correlation between F1+2 levels and anti-Xa activity (r= -0.254, p=0.0052).

As previously reported by others (14, 15), increased plasma levels of sGPV were observed in our patients, reflecting the important role of platelet activation by thrombin in the acute phase of myocardial infarction. Moreover, sGPV was significantly (p= 0.031) lower in TIMI 2-3 patients [53 (44-63) ng/ml] compared to TIMI 0-1 patients [64 (47-84) ng/ml] (fig. 1). The difference persisted after adjustment for risk factors (p=0.002). A weak correlation was observed between sGPV and TF-MP (r=0.293, p=0.031).

**Biomarkers of fibrinolysis.**

Plasma levels of tPA antigen were significantly (p=0.011) lower in TIMI 2-3 [7 (6-11) ng/ml] than in TIMI 0-1 patients [10 (7-13) ng/ml]. The difference disappeared after
adjustment for cardiovascular risks factors (p=0.303). Plasma levels of PAI-1 were similar in TIMI 2-3 patients [22 (11-44) ng/ml] and TIMI 0-1 patients [22 (12-45) ng/ml]. Plasma levels of PAP complexes were increased in the two groups of patients (fig.2), but significantly lower in TIMI 2-3 patients [88 (51-147) ng/ml] compared to TIMI 0-1 patients [114 (65-225) ng/ml] (p=0.041). The difference persisted after adjustment for risk factors (p=0.037) (fig.2).

**DISCUSSION**

Our prospective study aimed to use spontaneous recanalization of the infarct vessel (in patients who received standardized antithrombotic therapy with aspirin and unfractionated heparin) as a paradigm for antithrombotic therapy in STEMI. Investigating differences in circulating markers of platelet activation, tissue factor expression, thrombin generation and fibrinolysis may be valuable to guide selection of additional antithrombotic therapy. The main result of the present analysis is that there are little differences between patients with occluded versus patent vessels in STEMI with respect to markers of platelet activation, except sGPV. In contrast, significant differences were observed in terms of leukocytes and endothelial cells activation. However, these differences disappear after adjustment on risk factors, indicating that the latter are causal. In addition, PAP and tPA levels were lower, paradoxically, in patients with patent versus those with occluded vessels.

Platelet-dependent thrombosis is known to play a critical role in patency and recanalization of the infarct artery in acute myocardial infarction. Yet our study failed to show differences in terms of platelet activation (platelet surface P-selectin and activated GPIIb-IIIa, or PMPs) between patients with patent versus occluded vessels. In contrast, Yip et al (13) have recently showed that platelet surface P-selectin was independently associated with the extent of myocardial necrosis in patients with AMI. We cannot exclude that antithrombotic treatment (aspirin + heparin) already introduced at the time of blood sampling in the present
study has masked potential differences in the extent of platelet activation between the two
groups of patients. Previous observations have shown that unfractionated heparin decreases
levels of circulating P-selectin and platelet activation in vivo (16). Aspirin influences P-
selectin expression on platelets and inhibits baseline reactivity in patients with AMI (17, 18).
However, it is questionable if this treatment could have influenced PMPs numbers.

As previously described by others (2, 15, 19), acute myocardial infarction was
associated with an increase shedding of MPs originating from endothelial cells (EMP).
Interestingly, patients with early spontaneous coronary recanalization exhibited significantly
lower numbers of circulating EMPs than those with persistent occlusion. EMPs are also
associated with multiple concommitant risk factors, in particular with hypertension (20, 21).
In the present work and consistent with a previous study (21), we found a correlation between
EMP and SBP, but only in cases. This apparent paradox might indicate that the
mechanism(s) involved in EMPs generation in patients with persistent occlusion would be
different and probably more complex than in patients with early spontaneous coronary
recanalization. We also observed an increase in GMPs in our patients, which was significantly
lower in TIMI 2-3 versus TIMI 0-1 patients. The present finding extend previous observations
demonstrating the involvement of leukocyte-derived MPs in the thrombus growth (22) and
plaque burden (23). The difference in EMPs and GMPs levels was no more significant after
adjustment for baseline risk factors, indicating that hypertension and dyslipidemia, which
were less prevalent in TIMI 2-3 patients, contribute to the differences in MPs shedding from
endothelial cells and leukocytes. EMPs and GMPs constitute reliable hallmarks of vascular
injury (24) and inflammatory response, which suggests that an inverse relationship between
the severity of vascular and inflammatory cells damage and early spontaneous recanalization
is highly probable. Alternatively, MPs can act as diffusible messengers, transporting bioactive
agents (25, 26), and high levels of EMPs and GMPs could contribute to the persistence of the coronary occlusion.

Tissue factor is the initial activator of the blood coagulation pathway that leads to thrombin generation and culminates in the fibrin clot formation. Elevated intra-vascular TF, blood-borne or shedded from ruptured plaque, has been reported in MI (27, 28) and MPs contribute at least in part to this activity (2). The correlation between TF activity and PMPs, although weak, may suggest that TF-MP is in part supported by PMPs, probably resulting from multiple fusions and exchanges between leukocyte, endothelial and platelet plasma or MP membranes (29) or resulting from platelet TF synthesis upon activation (30). We found a trend for lower levels of TF-MP in patients with early spontaneous recanalization compared to patients with persistent occlusion. It is unclear whether this merely reflects lack of power of our small study or a true lack of difference. In addition, TF activity on MPs is modulated by its inhibitor TFPI (31), which is susceptible to proteolysis thereby limiting its inhibitory activity (32, 33).

A reduced generation of thrombin was also associated with spontaneous coronary reperfusion, as indicated by a lower level of sGPV in TIMI 2-3 patients. However, the causal role of TF-MP in sGPV shedding could not be directly established in the present study since the two markers were weakly correlated.

Platelet GPV is directly cleaved by thrombin during platelet activation and sGPV is an indirect but exquisitely sensitive marker of thrombin presence (34). This study indicates that sGPV represents a more sensitive marker of thrombin-induced platelet activation than PMPs. The increased levels of sGPV in patients with occluded infarct arteries thus suggest increased presence of thrombin compared to patients with patent arteries. However, in contrast to sGPV, prothrombin F$_{1+2}$ levels were similar in the two groups of patients, all of which were treated with unfractionated heparin. The influence of this treatment is highly suggested by the
correlation between $F_{1+2}$ levels and anti-Xa activity as already reported (35). We hypothesize that sGPV, which has a longer half-life than $F_{1+2}$, represents a better marker of thrombin generation in patients receiving heparin (34, 36).

In addition to thrombin, TNF-alpha converting enzyme (TACE or ADAM17), a metalloproteinase that is present in platelets, is capable of cleaving GPV (37) upon platelet activation. The ELISA used for the sGPV assay does not discriminate between both fragments released. Consequently, we cannot eliminate the impact of this mechanism in the shedding process, although its contribution would be improbable since we did not evidence differences in platelet activation between the two groups of patients.

Baseline levels of the fibrinolytic components tPA and PAI-1 are recognized biomarkers linked to the risk of major cardiovascular events (38). Lower levels of tPA may appear paradoxical in recanalized patients. Since the tPA assay measures, to a large extent, inactive tPA/PAI complexes, increased tPA antigen levels may be viewed as a correlate of increased PAI-1 activity (39). Plasma levels of tPA antigen are relatively constant over time while PAI-1 antigen levels fluctuate markedly (39), perhaps explaining why the former is a more sensitive marker than the latter. We hypothesize that lower tPA levels in recanalized patients may be related to less endothelial injury mainly modulated by baseline risk factors. The lower level of PAP observed in TIMI 2-3 patients suggests that less plasmin is generated in recanalized patients at the time of blood sampling, which may be the direct consequence of low levels of fibrin-bound tPA able to transform plasminogen into plasmin. Alternatively, the higher levels of PAP in TIMI 0-1 patients could result from plasminogen activation by uPA as a consequence of the greater granulocyte activation (40, 41) observed in this group of patients.

Our results suggest that there may be differences in the arterial lesions and/or the hemostatic system between patients with STEMI: less endothelial injury and granulocyte
stimulation, lower levels of thrombin and plasmin generation appear to correlate with a greater chance of early recanalization of the infarct vessel in patients receiving heparin, and antiplatelet therapy. In addition, our results also show the impact of classical risk factors on persistent coronary thrombosis. Indeed, in the present study, hypertension and dyslipidemia appeared to play a critical role in differences in markers of cell activation, in accordance with previous observations (42, 21). Our results indicate that antithrombotic treatment with molecules active on thrombin generation or activity such as the pentasaccharide (43) or bivalirudin (44, 45) might favor spontaneous recanalization and deserve to be tested.

Study Limitations

A potential limit of our study is the selection of cases and controls towards inclusion of the most stable patients. The small size of the study could have also underpowered differences between groups.

Flow cytometry on platelets was performed only in 35 patients in each group which corresponded to patients included during the day-time. It is possible that this selection has introduced a bias in platelet activation markers (46).

Aknowledgements.

We wish to thank L. Venisse, S. Loyau, C. Bousquet, N. Belgueirma, Y. Baudoin, P. Cornelie for their expert technical assistance. We also wish to thank A. Dauphin for collecting clinical data.
References


Legends to Figures and Tables

Figure 1.
Soluble GPV (sGPV) in 70 controls (TIMI 0-1, grey bars) and 53 cases (TIMI 2-3, clear bars). P value was calculated after adjustment on baseline cardio-vascular risk factors. In these plots, lines within boxes represent median values, the lower and upper lines of the boxes represent the 25th and 75th percentiles, respectively, and the lower and upper bars outside the boxes represent the 10th and 90th percentiles, respectively.

Figure 2.
Plasmin-antiplasmin (PAP) complexes in 70 controls (TIMI 0-1, grey bars) and 53 cases (TIMI 2-3, clear bars) patients. P value was calculated after adjustment on baseline cardio-vascular risk factors. Plots are outlined as in figure 1.

Table 1
Qualitative variables are expressed as number (observed number/total number) and (%), and quantitative variables are expressed with median and range (minimum-maximum).

Table 2
Platelet P-selectin and activated GpIIb-IIIa (PAC-1) expression as measured by flow cytometry in 35 controls (TIMI 0-1 flow) and 35 cases (TIMI 2-3 flow). Data are expressed as per cent positive platelets as compared to isotype controls.

Table 3.
Microparticles (MPs) values are expressed as number per μl, median (interquartile range, IQR); PMPs: platelet derived MPs; GMPs: granulocyte-derived MPs; EMPs: endothelial
derived MPs; TF-MP: Tissue factor associated with microparticles. TF-MP activity is expressed as pg/ml, median (IQR).

P value according to Mann-Whitney U-test. P* value of logistic regression after correction for risk factors – age, smoking, hypertension, dyslipidemia, diabetes mellitus, anterior location of MI, delay between onset of pain and recanalization.

Table 1. Patients Baseline Characteristics

<table>
<thead>
<tr>
<th>Variables</th>
<th>TIMI 2/3 patients (n=53)</th>
<th>TIMI 0/1 patients (n=70)</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Women</td>
<td>3/53 (5.8)</td>
<td>11/70 (15.7)</td>
<td>0.088</td>
</tr>
<tr>
<td>Age (years), median (range)</td>
<td>53 (34-80)</td>
<td>57 (29-80)</td>
<td>0.385</td>
</tr>
<tr>
<td>Age &gt; 70 years</td>
<td>8/53 (15.1)</td>
<td>11/70 (15.7)</td>
<td>0.611</td>
</tr>
<tr>
<td>Hypertension</td>
<td>12/53 (22.6)</td>
<td>30/69 (43.5)</td>
<td>0.016</td>
</tr>
<tr>
<td>Prior or current smoking</td>
<td>46/52 (88.5)</td>
<td>54/69 (78.2)</td>
<td>0.321</td>
</tr>
<tr>
<td>Dyslipidemia</td>
<td>18/53 (34.0)</td>
<td>36/69 (52.2)</td>
<td>0.045</td>
</tr>
<tr>
<td>Diabetes mellitus</td>
<td>7/53 (13.2)</td>
<td>8/70 (11.4)</td>
<td>0.765</td>
</tr>
<tr>
<td>Delay from symptoms onset to recanalization (hours), median (range)</td>
<td>3.0 (1.3-10.9)</td>
<td>3.5 (0.4-9.4)</td>
<td>0.377</td>
</tr>
<tr>
<td>Delay from symptoms onset to recanalization &gt;2 hours</td>
<td>47/53 (88.7)</td>
<td>52/63 (82.5)</td>
<td>0.352</td>
</tr>
<tr>
<td>Anterior myocardial infarction</td>
<td>22/53 (41.5)</td>
<td>23/70 (32.8)</td>
<td>0.324</td>
</tr>
<tr>
<td>Abciximab before coronarography</td>
<td>43/53 (81.1)</td>
<td>62/70 (88.6)</td>
<td>0.248</td>
</tr>
</tbody>
</table>
Table 2. Platelet P-selectin and PAC-1 expression by flow cytometry. Per cent positive platelets, median(IQR)

<table>
<thead>
<tr>
<th></th>
<th>TIMI 0/1</th>
<th>TIMI 2/3</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>N=35</td>
<td>N=35</td>
<td></td>
<td></td>
</tr>
<tr>
<td>P-selectin</td>
<td>3.6 (2-6.1)</td>
<td>5.2 (1.6-10.3)</td>
<td>0.173</td>
</tr>
<tr>
<td>PAC-1</td>
<td>12 (6-21)</td>
<td>13.2 (4-33)</td>
<td>0.411</td>
</tr>
</tbody>
</table>
Table 3. Microparticles quantification, cellular origin and associated-TF activity in patients with AMI according to their flow grade.

<table>
<thead>
<tr>
<th>Microparticles</th>
<th>TIMI 0/1</th>
<th>TIMI 2/3</th>
<th>P</th>
<th>p*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N=70</td>
<td>N=53</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Origin</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total MPs</td>
<td>507 (186-738)</td>
<td>312 (152-751)</td>
<td>0.331</td>
<td>1.000</td>
</tr>
<tr>
<td>PMPs</td>
<td>386 (112-677)</td>
<td>251 (104-459)</td>
<td>0.126</td>
<td>1.000</td>
</tr>
<tr>
<td>GMPs</td>
<td>38 (13-112)</td>
<td>27 (10-62)</td>
<td>0.021</td>
<td>0.995</td>
</tr>
<tr>
<td>EMPs</td>
<td>14 (8-26)</td>
<td>11 (4-15)</td>
<td>0.002</td>
<td>0.991</td>
</tr>
<tr>
<td>Activity</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TF-MP pg/ml</td>
<td>72 (31-151)</td>
<td>40 (22-101)</td>
<td>0.09</td>
<td>0.996</td>
</tr>
</tbody>
</table>