

Novel alpha interferon (IFN-alpha) variant with improved inhibitory activity against hepatitis C virus genotype 1 replication compared to IFN-alpha2b therapy in a subgenomic replicon system.

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1 ABSTRACT

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3 Hepatitis C treatment is based on the association of pegylated IFN- α and ribavirin. To improve the level of sustained virological response to treatment, especially in patients 4 5 infected with HCV genotype 1, new IFNs with improved efficacy and toxicity profile may be 6 developed. In this report we show that a novel and naturally occurring human IFN- α 17 7 variant, GEA007.1, which was discovered using an original population genetics-based drug 8 discovery approach, inhibits HCV genotype 1 RNA replication more efficiently than IFN-a 9 2b, in the BM4-5 cell line harbouring an HCV subgenomic replicon. Moreover, we show that 10 the complete viral clearance is obtained in BM4-5 cells after long-term treatment with 11 GEA007.1, when HCV subgenomic RNA is still detected in cells treated with other IFN-a 12 variants or with standard IFN- α 2b. Eventually, we demonstrate that the better inhibitory 13 activity of GEA007.1 is likely to be due to a stronger and faster activation of the JAK-STAT 14 signalling pathway and to a broader expression of IFN- α responsive genes in cells, compared 15 to standard IFN-α. Our results demonstrate a superior inhibitory activity of GEA007.1 over 16 IFN- α 2b in the HCV replicon system. Clinical trials are required to determine if GEA007.1 17 could be a potent 'next generation' IFN for the treatment of HCV infection, especially in non-18 responders or relapsing patients infected with HCV genotype 1 who currently represent a 19 clinical unmet need.

20

1 INTRODUCTION

2 Hepatitis C virus infects 170 million people worldwide and leads, in approximately 70% of 3 cases, to chronic infection (3, 24, 26). Current treatments are based on the association of pegylated-interferon-alpha 2a or 2b (IFN-a 2a or IFN-a 2b) and ribavirin (5, 8, 20). This 4 5 treatment induces a sustained virological response (SVR) in approximately 60% of cases. The 6 rate of SVR varies from 30-40% in patients infected with genotype 1, to 80-90% for those 7 infected with genotype 2 or 3. Despite a rather good efficacy, this treatment is often poorly 8 tolerated because of side effects (5). It is therefore important to improve current molecules 9 and carry on searching for novel antiviral agents with an enhanced anti-HCV activity to 10 overcome the failure of current treatments, especially in patients infected with HCV genotype 11 1 (4).

Until the very recent development of a complete and robust HCV replication system (19, 27, 12 13 28), hepatoma cell lines harbouring HCV subgenomic replicon were a relevant model to study 14 some aspects of HCV biology as well as the effect and mechanism of action of antiviral 15 agents against HCV replication (1). Various interferons, including IFN- α , β , and γ , were 16 studied using this model, thus providing some insight on their antiviral effect (6, 7, 10, 18), 17 and mechanism of action (6, 11, 12, 15). Overall it was shown that the transduction pathway 18 following the interaction of exogenous IFNs with their cellular receptors was functional in 19 most cases, although slight differences could exist between different Huh7 cell lines 20 harbouring HCV subgenomic replicon. These differences may account for the variability 21 observed in the amount of interferon used to obtain a given inhibition. Altogether, HCV 22 replicon systems were validated as a suitable tool for the study of the efficacy and mechanism 23 of action of interferon molecules. Interferon- α is known to exhibit several independent biological activities, including immunomodulatory, antiproliferative, and antiviral activities 24 25 (14, 22, 23). In the context of an antiviral strategy, this pleiotropic effect of IFN- α may 1 account for adverse effects that are observed *in vivo* which are a major cause of either non 2 compliance or premature interruption of the treatment (5). One possible approach to improve 3 the currently used IFN- α is to identify novel IFN- α entities with increased antiviral activities 4 and potentially decreased other activities.

5 In this study, we evaluated and compared to standard IFN- α 2b (i.e. IFN- α 2 wild-type) the 6 inhibitory effect of original and functionally-relevant variants of different IFN- α subtypes 7 (IFNv), against the replication of the hepatitis C virus (HCV) using a cell line harbouring a 8 HCV genotype 1b subgenomic replicon (BM4-5 cells). One of the studied variants, 9 GEA007.1, was found seven times more efficient than the IFN- α 2WT molecule to inhibit 10 HCV RNA synthesis in BM4-5 cells. Interestingly, long-term treatment with GEA007.1 was 11 able to cure BM4-5 cells from HCV replicon in contrast to the standard IFN- α 2WT molecule 12 tested in the same conditions. Moreover, based on mechanistic studies, we propose that the 13 increased efficacy of the novel IFN- α 17 variant GEA007.1 against HCV replication is likely 14 to be due to a more potent activation of the JAK-STAT transduction pathway in BM4-5 cells, 15 following the interaction of GEA007.1 with type 1 IFN receptor.

16

1 MATERIALS AND METHODS

Identification and production of IFN- α variants. Original genetic variants of 14 different 2 human IFN- α protein subtypes were provided by GenOdyssee. Discovery of natural cytokine 3 4 mutants with improved therapeutic utility was performed using the following process: a 5 genomic DNA library was constituted with genomic DNA from 239 different individuals representing altogether 85% of the human ethnic diversity of the worldwide population, and 6 7 screened for non-synonymous single nucleotide polymorphisms in the coding sequence of the 14 different human IFN- α gene subtypes. The identification and selection of IFN- α protein 8 mutants with functionally-relevant amino-acid mutations, was then carried out using 9 bioinformatics tools, including sequence/structure analysis and molecular modelling. IFN-a 10 11 variants (IFNv), named thereafter GEA007.1, GEA009.2, GEA011.1, GEA013.1, derived from 12 such procedure, are respectively natural mutants of the human IFN- α 17, 21, 5, and 7 (SwissProt primary accession numbers P01571, P01568, P01569, P01567). All carry a unique 13 14 amino-acid mutation (G45R for GEA007.1, D95N for GEA013.1, C122S for GEA011.1, and 15 K179E for GEA009.2), with changes in three-dimensional structure and electrostatic 16 isopotentials at the receptor-binding surface, based on molecular modelling. The recombinant human IFN- α variants (IFNv) used in this study were produced in the methylotropic yeast, 17 18 Pichia pastoris, with a yield of approximately 10-20 mg/l, following protocols and recommendations provided by Research Corporation Technologies, Inc. IFNv proteins were 19 secreted into the culture medium and purified using Cibacron and ion exchange 20 21 chromatography to obtain pure preparations of the recombinant human IFNv in a phosphate 22 buffered solution (PBS). The purity of each preparation was greater than 95% and endotoxin 23 content, determined by the LAL test, was less than 5IU/µg. IFN- α 2b, named IFN- α 2WT thereafter, in order to be an relevant control was synthesized and purified using the same 24 25 process. In our experiments 1mg of IFN-a produced in yeast was equivalent to 3x10e8

1 international unit (IU), (i.e. 1 IU is equivalent to 3.33 pg). Stock solutions were stored at -2 80°C. Dilutions of these stock solutions were done in PBS extra-temporally. Prior to activity testing of the different IFN preparations against HCV in the replicon model, the different IFN 3 4 preparations were tested for their specific activity and standardized in vitro using a gene-5 reporter assay as described elsewhere (17). It was shown that the inhibitory activities of IFN-6 α 2WT produced in *Pichia Pastoris* and manufactured IFN-α 2b (Intron A®, Schering Plough 7 Corporation) produced in E. Coli are similar, indicating that the yeast Pichia Pastoris 8 production system does not affect the activity of IFN- α .

9 *Cell Culture*. BM4-5 cells, a Huh7 cell line harbouring an HCV genotype 1b subgenomic 10 replicon, that were kindly provided by Dr C. Seeger (Fox Chase Cancer Center, Philadelphia, 11 USA), were cultured in DMEM medium (Invitrogen) complemented with 10% FCS (Foetal 12 Calf Serum; Perbio), antibiotics, and 500 μ g/mL of geneticin (G418 Sulfate; Invitrogen) as 13 previously described (10).

14 Protocols of administration of IFN. BM4-5 cells were seeded in 6-well plates at a density of 15 2.5x10e5 cells/well, sixteen hours before starting treatments. IFNv (GEA007.1, GEA009.2, 16 GEA011.1, and GEA013.1) and IFN- α 2WT were administrated to cells in complete medium 17 in the absence of geneticin. For short-term treatments, the administration of each drug (at 18 concentrations ranging from 0 to 3333 pg/ml) was renewed every day for three consecutive 19 days. For long-term treatments, IFN-α 2WT or GEA007.1 was administered every day at 333 20 pg/mL for 21 days. In order to maintain cells in proliferation/division, cells were trypsinized 21 and diluted four times every four days. A total of 5 passages was performed during treatment. 22 At the end of this period of treatment, the administration of IFN- α 2WT and GEA007.1 was 23 interrupted and cells were kept for 5 further passages in medium containing or not geneticin 24 to monitor a potential rebound of HCV subgenomic RNA replication.

1Analysis of IFN toxicity. Cells were seeded in 96-well plates at a density of 12,500 cells/well.2They were treated by IFNs with the same concentrations and conditions than those used for3short-term inhibitory assays. The cell viability was measured by both neutral red and MTS4(CellTiter 96 Aqueous One Solution Cell Proliferation Assay, Promega) tests. The cytotoxic5concentration 50% (CC₅₀) is defined as the concentration of interferon leading to a 50%6reduction in the absorbance value compared to non treated cells.

7 Analysis of HCV subgenomic RNA synthesis. Total RNA was extracted with the "Extract 8 all" reagent according to manufacturer's instructions (Eurobio). Northern Blot analysis was 9 then performed using the NorthernMaxTM-Gly kit according to manufacturer's instructions 10 (Ambion). Briefly, 5 µg of total RNA were denaturated in glyoxal buffer at 50°C for 30 11 minutes, separated on 1,1% agarose gel, and transferred over night by capillarity onto a 12 charged nylon membrane (HybondN+, Amersham). Hybridization was carried out with three different [³²P]CTP-labelled riboprobes obtained by *in vitro* transcription (Riboprobe in vitro 13 14 transcription system, Promega). Two probes complementary to the NS5A region were used to 15 detect either the negative or positive HCV subgenomic RNA strands. A third probe was 16 complementary to the β-actin mRNA and obtained by *in vitro* transcription using the plasmid 17 pTRI beta actin human (Ambion). First, the blot was hybridized for 16 hours at 68°C with the 18 two riboprobes directed against the negative strand of HCV subgenomic RNA and β-actin 19 mRNA, respectively. After hybridization, the membrane was washed twice in stringent 20 conditions, then exposed sequentially to X-ray film and a phosphor screen for quantification. 21 The amount of β-actin mRNA was used as an internal loading control to standardize the 22 amount of HCV subgenomic RNA detected. The same membrane was subsequently 23 hybridized with the negative sense riboprobe to visualise and quantify the level of positive 24 HCV subgenomic RNA strand, using the same procedure.

1 Analysis of the signal transduction pathway induced by IFN- α treatment in BM4-5 cells. 2 To evaluate the activation of intracellular signaling pathways by IFN-α 2WT and IFNv, BM4-3 5 cells (75x10e3 cells/well in 24 wells plate) were transfected with an Interferon Stimulating 4 Response Element (ISRE) -luciferase plasmid (Stratagene) by using the TransIT-TKO reagent 5 (Mirus) following the manufacturer's recommendations. Twenty-four hours after transfection, 6 duplicate cultures in 24 well plates were either left untreated or treated for 24 hours with a 7 range of IFN- α 2WT and IFNv of 0.3, 3, 33, 333 and 3333 pg/ml. Cells were then washed 8 with cold phosphate-buffered saline (PBS), lysed by passive lysis buffer (Promega) and the 9 luciferase activity measured as recommended by the manufacturer (Promega). The luciferase 10 activity induced by IFN- α 2WT for each concentration was taken as 100% and other 11 luciferase activity were normalized accordingly.

12 Western Blot analysis. IFN- α 2WT or GEA007.1 were administered at a concentration of 13 3333 pg/mL or at 33 pg/mL on BM4-5 cells for 30 min, 1 h, 2h, 4h, 8h, 12h, 24h, 48h, and 14 72h. Administration was renewed each 24h. At the indicated time, cells were washed twice 15 with cold PBS, then lysed using a buffer (0,2% NP40, 150 mM NaCl, 20 mM Tris, 2 mM 16 EDTA, 0,1% glycerol, 10 mM dithiothreitol) containing serine/threonine phosphatases 17 inhibitors (50 mM NaF, 200 µM orthovanadate), and protease inhibitor (complete protease 18 inhibitor, Roche). Proteins of cell lysates were denaturated at 100°C in Laemmli buffer (50 19 mM Tris pH 6.8; 2% SDS; 10% glycerol; 0,1% bromophenol blue; 1.25% 2-20 mercaptoethanol), separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis 21 (SDS-PAGE), and transferred onto polyvinylidene difluoride (PVDF) membranes (Amersham 22 Pharmacia Biotech). Non-specific binding sites were blocked for 2 h at room temperature 23 (RT) with 5% milk, 0,1% Tween 20 in PBS. The membrane was then incubated overnight at 24 4°C with monoclonal antibodies raised against PKR (Santa Cruz Biotechnology), p48 (Beckton Dickinson) or β -actin (Sigma). Membranes were then washed three times, and 25

1 incubated at RT for 1 hour with horseradish-peroxidase-conjugated goat anti-mouse or anti-2 rabbit immunoglobulins (Immunotech) at 1/10 000. Bound antibodies were detected using 3 ECL technologies, ECL+ for p48 and ECL for PKR and β -actin, as specified by the 4 manufacturer (Amersham Biosciences).

5 Analysis of IFN- α specific gene expression using GEArray \mathbb{T}_Q Series Array. BM4-5 cells 6 were incubated in 6 well plates at 80% confluence. The cells were either left untreated or were 7 incubated with IFN-2WT or GEA007.1 at the same concentration of 33 pg/mL. After 8 hr of 8 incubation, total RNA was extracted with Extract all reagent (Eurobio) and stored at -80°C. 9 For one set of experiment, six arrays (GEArrayTM Q series Human Interferon α , β Response 10 Gene Array-SuperArray, Bioscience Corporation) were used following manufacturer's 11 recommendation. For each array, 96 gene-specific cDNA fragments are printed on a 3,8 x 12 4,8cm nylon membrane. The list of the genes is accessible on the SuperArray web site 13 (www.superarray.com; Ref. HS-054). A total of 3µg of each RNA sample was used as a 14 template to generate cDNA by reverse transcription. cDNA was then amplified and labelled 15 with biotin-16-dUTP (Roche) by a Linear Polymerase Reaction (LPR) (AmpoLabeling-LPR 16 Kit). After a step of pre-hybridization, the membranes were hybridized overnight with the 17 different probes, corresponding to the different treatments. The signal was revealed by 18 chemiluminescent reaction using alkaline phosphatase-conjugated streptavidin, and detected 19 by a scanner camera (Biorad). All the results were obtained from the analysis of non saturated 20 images, after 2 min of exposure. Spot quantification was performed with Quantify-One 21 software (BioRad). The mean intensity of blank spots served as negative controls and was substracted from other values. The mean intensity of β -actin gene served as a positive control 22 23 and was used to normalize intensities of the spots to allow relative comparisons between the different arrays. To determine which genes were up regulated, intensities of spots resulting 24 25 from hybridization with cDNA obtained from total RNA extracted from untreated cells were

used to establish the basal expression level. The up-regulation of genes under interferon
 treatment was reported as positive fold increase compared to untreated control samples.

1 **RESULTS**

2 Identification of novel IFN- α variants with improved inhibitory properties. Based on the 3 hypothesis that natural evolution of the human genome has likely created in human 4 populations a series of unpredictable and functionally-relevant genetic variants with various 5 structural and biological properties, non-synonymous single nucleotide polymorphisms were 6 searched in the genomes of 239 individuals representing altogether 85% of the current human 7 population ethnic diversity to identify interesting novel alleles of IFN-α genes coding for 8 IFN- α proteins with superior or novel therapeutic properties, compared to standard IFN- α 2a 9 or 2b. Several variants of IFN- α 17, 21, 5, and 7, referred thereafter as GEA007.1, GEA009.2, 10 GEA011.1, and GEA013.1 (IFNv), were selected following this approach, for their 11 significantly improved (GEA007.1, GEA013.1) or decreased (GEA009.2, GEA011.1) 12 inhibitory properties, compared to standard IFN-a 2WT, based on preliminary data obtained 13 with both an *in vitro* model of vesicular stomatitis virus (VSV)-infected WISH cells and an *in* 14 vivo model of EMCV infected-mice (data not shown, manuscript in preparation by Tovey et 15 al.).

16 Anti-HCV effect of novel IFN- α variants. To evaluate and quantify the inhibitory effect of 17 these novel IFNv, BM4-5 cells were treated for three days with increasing concentrations of 18 GEA007.1, GEA009.2, GEA011.1, GEA013.1, as well as with IFN-α 2WT used as reference. 19 Total RNA was extracted and subjected to Northern Blot analysis with three different 20 riboprobes designed to detect both negative and positive HCV subgenomic RNA strands and 21 β -actin mRNA (Figure 1). A dose dependent reduction in the amount of HCV subgenomic 22 RNA was observed with all IFNv tested, and the greatest effect was obtained with GEA007.1. 23 Three independent experiments were performed to obtain statistically relevant data. The 24 quantification of the relative intensity of HCV subgenomic RNA and control β-actin signals

1 after exposure to phosphor screens allowed the calculation of the effective concentration to 2 inhibit 50%, defined as the concentration required to induce a reduction of 50% of the amount 3 of HCV subgenomic RNA detected by Northern Blot after normalisation, for each IFN tested 4 (Table 1). Two IFNv, GEA007.1, and GEA013.1 to a lesser extent, had a better EC₅₀ than IFN-a 2WT, suggesting higher efficacy of these molecules compared to control. The better 5 6 inhibitory activity of GEA007.1 (x7) and GEA013.1 (x1.5) over IFN-a 2WT remained 7 significant when the drugs EC_{90} were concerned (Table 1). This inhibitory activity was not 8 due to toxicity as all IFNs tested did not show any cytotoxic effect on the BM4-5 cells, with 9 CC_{50} (i.e. cytotoxic concentration 50%) always superior to 3333 pg/mL, the highest dose used 10 in our experiments (Table 1).

11 Long-term administration of GEA007.1 allows to cure cells from HCV genotype 1b replicon subgenome. Having established that GEA007.1 had the best inhibitory activity 12 13 compared to IFN- α 2WT, with a very good EC₉₀, we asked whether a prolonged 14 administration of this IFNv could induce a better sustained inhibitory effect than IFN- α 2WT 15 and lead to the rapid eradication of HCV replication in cells. BM4-5 cells were treated during 16 20 days though 5 passages, with either IFN- α 2WT or GEA007.1 at a single concentration of 17 333 pg/mL in medium without geneticin as described in materials and methods. The results of 18 a Northern Blot analysis presented in Figure 2 shows that HCV subgenomic RNA synthesis 19 was inhibited more intensively by GEA007.1 than IFN- α 2WT, thus confirming the better 20 inhibitory activity of GEA007.1. Moreover, after 20 days of treatment and 5 passages, HCV subgenomic RNAs (both strands) were no more detectable in cells treated with 333 pg/mL of 21 22 GEA007.1, whereas the positive strand HCV subgenomic RNA was still detectable in 23 standard IFN-α 2WT treated cells. To monitor the potential rebound of HCV replication after 24 cessation of treatment, cells were kept untreated for at least 4 additional passages either in 25 absence or presence or geneticin. The presence of geneticin was meant to select back more

1 efficiently HCV replication from the very low level of residual replication after 20 days of 2 treatment with IFN. In the absence of geneticin, HCV subgenomic RNA remained 3 undetectable after 5 passages post-treatment in GEA007.1 treated cells, whereas a low amount 4 of positive strand HCV subgenomic RNA was maintained in IFN-α 2WT treated cells (Figure 5 2). When cells were cultured in a medium with geneticin after treatment cessation, the cells 6 that had been treated with IFN- α 2WT could survive, but it took five more days for these cells 7 to reach confluence, compared to untreated cells. Interestingly, cells that had been treated 8 with GEA007.1 were no more able to survive in the presence of geneticin, indicating that 9 GEA007.1 was able to cure the cells from HCV subgenomic RNA (Figure 2). In our 10 experimental conditions, GEA007.1, but not IFN- α 2WT, was able to eliminate HCV 11 replication from BM4-5 cells, thus indicating once again the better efficacy of this variant 12 molecule over the wild type.

13 Stronger and faster expression of proteins of type 1 IFN signal transduction pathway in 14 BM4-5 cells is obtained with GEA007.1, compared to IFN- α 2WT. Having established that 15 GEA007.1 had better anti-HCV properties than IFN- α 2WT in the replicon system, we sought 16 to determine whether the higher activity of GEA007.1 was associated with a better activation of key cell signalling proteins involved in type 1 IFN transduction pathways. To this end two 17 18 set of experiments were performed. First, BM4-5 cells were transfected with a reporter 19 plasmid containing an Interferon Stimulating Response Element upstream driving the 20 expression of the firefly luciferase. Twenty-four hours after transfection, culture were either 21 left untreated or treated with a range of IFN- α 2WT and IFNv of 0.3, 3, 33, 333 and 3333 22 pg/ml. Luciferase activity was measured 24 hours after and results obtained with IFNv for 23 each concentration were normalized to IFN- α 2WT induced activity (Figure 3). The results 24 showed an increased activation of the IFN stimulating pathway with GEA007.1 compared to 25 the others IFNs with the following order GEA007.1> GEA013.1 > IFN- α 2WT = GEA011.1

1 > GEA009.2. A 3 and 2 fold increased activation of the IFN signaling pathways was observed 2 with GEA007.1, by comparison with IFN- α 2WT, at the concentrations of 33 and 333 pg/ml 3 respectively. At higher concentrations, i.e. 3333 pg/ml, the difference of activation was not 4 significant most likely because of the saturation of the IFN receptor. Further experiments 5 were therefore performed with low IFN concentrations of 33pg/ml (± equivalent to 10 IU) to 6 clearly see differences between IFN- α 2WT and GEA007.1. Western Blot analyses were 7 performed to further characterize the expression of proteins involved in the IFN signaling 8 pathway. BM4-5 cells were treated with either GEA007.1 or IFN-α 2WT at 33 pg/mL for 30 9 minutes, 1, 2, 4, 8, 12, 24, 48, and 72 hours and proteins were extracted and subjected to 10 Western Blot analysis, using antibodies directed against p48 (IRF-9) and PKR, two important 11 proteins of the type 1 IFN transduction pathways (25). With these conditions, GEA007.1 12 triggered a faster and stronger expression of PKR and recruitment of p48 proteins in BM4-5 13 cells compared to IFN- α 2WT. The difference in activation of IFN inducible proteins was 14 more marked for p48. In our experimental conditions, the low dose of IFN-a 2WT do not allow to visualize a significantly increase of expression of p48 and PKR proteins (Figure 4). 15

16 IFN responsive genes expression is stronger and broader after treatment with GEA007.1 than after treatment with IFN- α 2WT. To broaden results obtained by Western blotting 17 18 analysis and gain a global view of the differences in transduction pathway activation activities 19 elicited by GEA007.1 and IFN-a 2WT in BM4-5 cells, the expression of IFN-a-stimulated 20 genes (ISGs) were analysed using cDNA expression arrays. BM4-5 cells were left untreated 21 or treated with either 33 pg/mL of GEA007.1 or IFN-a 2WT for 8 hours. RNA were then 22 extracted and used for RT-PCR. Labelled cDNA were used for the hybridization on arrays as 23 described in materials and methods. Relative intensities, for the different genes and conditions 24 of treatment, were calculated, and results from significant up-regulation in BM4-5 cells 25 induced by the administration of either IFN are reported in Table 2. A more than 2 fold

1 increase in gene expression was considered as significant. Two IFN-responsive genes, MX1 2 and ISG56 (25), were found significantly up-regulated following IFN- α 2WT treatment, 3 while a stronger and a broader up-regulation of genes (MX1, ISG56, ISG15, ISG60, ADAR, 4 KIAA1268, STAT1) was found associated with GEA007.1 inhibitory activity in BM4-5 cells. The dose of IFN-α 2WT chosen in this experiment to point out differences with GEA007.1 5 6 was not high enough to trigger strong activation of gene expression, in particular for PKR and 7 p48, at least in our experimental conditions. This result is in good correlation with the lack of 8 increase of protein amount observed by western blotting (Figure 4). Collectively, these results 9 confirm a better activation of type 1 IFN signal transduction in these cells with GEA007.1, 10 compared to IFN- α 2WT at the same concentration and time of exposure. This better ability 11 to induce a stronger, faster and broader cell signalling response was therefore associated with 12 a superior inhibitory property of GEA007.1 over that of IFN-a 2WT observed in cells 13 harbouring the HCV genotype 1b replicon.

14

1 **DISCUSSION**

2 While waiting for the development and clinical use of new anti-HCV molecules, interferon 3 alpha remains a major component of the anti-HCV therapy. Its use may also remain 4 mandatory in combination with the novel HCV inhibitors to prevent resistance to the latter drugs (8). However, currently used interferon- α 2a and 2b are not efficient in all cases. Non 5 6 responding and relapsing patients are observed with current pegylated IFN- α /ribavirin 7 combination therapies. Many factors may explain this resistance of the virus to short- or long-8 term viral clearance activity of IFN- α 2, including host genetic and virological factors. For 9 instance, it is well known that patients infected with HCV genotype 1 have a significantly 10 lower response rate (5, 6), and that genetic host factors may influence the response rate to 11 interferon based therapies (2, 16).

12 The main objective of this work was to determine whether better anti-HCV genotype 1 IFN 13 molecules, compared to clinically used interferon alpha 2a and 2b, could be identified 14 amongst the novel interferon- α variants, GEA007.1 and GEA013.1, with improved inhibitory 15 properties on VSV and EMCV (manuscript in preparation, Tovey et al.,). For this study, IFN-16 α 2b (IFN- α 2WT) served as standard IFN- α and additional IFN- α variants with low 17 inhibitory properties, GEA009.2 and GEA011.1, were included as negative controls. 18 GEA007.1, GEA013.1, GEA009.2, and GEA011.1 were all identified following an original 19 approach of natural genetic variation of cytokine genes. Basically, the genomes of 239 20 individuals representing altogether 85% of the human ethnic diversity were searched for new 21 and natural functionally-relevant genetic mutants of the different human interferon- α subtypes 22 and variants with potential superior activities compared to IFN- α 2 were selected. BM4-5 cells, a Huh7 cell line harbouring an HCV genotype 1b subgenomic replicon, were used to 23 24 test the inhibitory properties of IFN- α variants. Interestingly, among the four interferon

1 variants tested in this study, GEA007.1 and to a lesser extent GEA013.1, were found 2 significantly more potent than IFN- α 2WT for inhibiting the replication of an HCV genotype 3 1b subgenomic replicon in BM4-5 cells. GEA007.1 was the best molecule tested with an EC_{50} of 22.7 pg/mL, which is 7 times lower than the EC₅₀ obtained with standard IFN- α 2WT (i.e. 4 5 154.2 pg/mL) in the same conditions (Table 1). Moreover, we found that GEA007.1, administered at 333 pg/mL every day for 20 days, was able to clear HCV subgenomic 6 7 replicon from BM4-5 cells, while HCV subgenomic RNA was still detected in cells treated 8 with standard IFN- α 2WT in the same conditions (Figure 2). This result demonstrates the 9 superiority of GEA007.1 over interferon- α 2b in an HCV genotype 1b subgenomic replicon 10 system. However, the results obtained in the HCV subgenomic replicon system may not 11 translate into antiviral activity in vivo. Therefore, our results warrant further evaluation of 12 GEA007.1 in vitro, using systems allowing a complete multiplication of HCV (19, 27, 28), 13 and in vivo, in the frame of clinical trials.

14 The biological activity of IFN- α is related to its capacity to induce type1 IFN signal 15 transduction pathways after interaction with its cellular receptor. Different mechanisms have 16 been proposed to explain the inhibitory effects of IFN- α against HCV replication, including 17 non-cytopathic and cytopathic processes (12). It was shown that the *in vitro* non cytopathic 18 inhibition of HCV RNA replication by IFN-a in Huh7 cells is dependent on functional JAK-19 STAT pathway and proteasome. However, the dsRNA-dependent antiviral pathways, such as 20 PKR and Rnase L pathways, were shown not to be involved in IFN-induced inhibition of 21 HCV replication in Huh7 cells (12). The IFN- α induced antiviral program could i) affect the 22 HCV IRES-directed translation and therefore cause a reduction in viral protein, and ii) could 23 in turn inhibit viral RNA amplification as a result of a tight linkage between the translation, 24 assembly of replication complexes and viral RNA synthesis (11). Interestingly, using an ISRE 25 reporter assay, we showed an increased activation of the IFN stimulating pathway by

1 GEA007.1 over IFN- α 2WT, which was associated with a better inhibitory activity on HCV 2 subgenomic RNA synthesis. Then we analysed key cell signalling proteins involved in type 1 3 IFN-transduction pathway after BM4-5 cells exposure to GEA007.1 or IFN- α 2WT. Western 4 Blot analyses were performed to monitor the level of expression of some key proteins like 5 p48 or PKR, and cDNA expression array experiments were performed to monitor the 6 expression of IFN- α responsive genes. GEA007.1 induced a stronger expression of p48 than 7 IFN- α 2WT. This result suggests that p48 may be recruited more efficiently by the STAT1^P and STAT2^P heterodimer complex to form the ISGF3 complex (25), after treatment with 8 9 GEA007.1 than after treatment with IFN- α 2WT. Our results are in accordance with previous 10 works showing that type 1 IFN signalling pathways are qualitatively functional in Huh7 and 11 BM4-5 cell lines, as persistent HCV replication does not modify gene expression patterns 12 after IFN- α treatment (13, 21). In addition, in our experimental conditions, IFN-13 α responsive genes activated in Huh7 cells harbouring HCV subgenomic replicon by both 14 GEA007.1 and IFN- α 2WT did not differ from those described and reported in other studies 15 (13, 21). Lastly, a stronger and broader expression of IFN- α responsive genes was seen in BM4-5 cells treated with GEA007.1, compared to IFN-a 2WT. Amongst most activated 16 17 genes were found Mx1, ISG56, ISG15, ISG60, ADAR, KIAA1268, and STAT1. Mx1 (also 18 named MxA) was the most induced gene in our system. Mx1 was also found previously as an 19 important IFN- α inducible gene in IFN- α -treated replicon cell lines (13). A stronger 20 expression of Mx1 was observed in cells treated with GEA007.1 which in turn could have 21 contributed to the better inhibitory effect of GEA007.1 compared to IFN- α 2WT. In view of 22 the results obtained by others, showing the absence of inhibition of HCV RNA replication by 23 Mx1 (6), it might be hypothesized that the activation of the different genes described can be 24 only a reflect of the activation of the Jak-Stat pathway, without predicting a direct inhibitory 25 role for a specific gene. Moreover, it is not possible at this stage to rule out the possibility that

1 GEA007.1 may also induce unknown IFN inducible genes responsible for an enhanced 2 inhibitory activity. The ability of GEA007.1 to trigger a strong antiviral response in HCV 3 infected BM4-5 cells is most likely due to either a better interaction of this IFN to the cellular 4 receptor or a better activity of the complex IFN-receptor in transmitting the antiviral signal. 5 Affinity of IFN to its receptor is a major issue in terms of efficacy. For instance, it was shown 6 that different pegylation patterns of IFN could lead to different efficacy of binding to its 7 cellular receptor and activation of the corresponding cell signalling pathway (9). One may 8 hypothesize that GEA007.1 has a better affinity to the type 1 IFN receptor (IFNAR) and/or 9 that the complex formed between GEA007.1 and IFNAR has a better capacity of signal 10 transduction than the complex formed between IFN- α 2 and IFNAR. Work is in progress to 11 confirm this hypothesis. The difference in binding properties of GEA007.1 compared to 12 standard IFN- α 2WT could be due to the differences observed in the amino acid sequence of 13 GEA007.1. In particular, GEA007.1 possesses a unique G45R mutation with significant 14 change in three-dimensional structure and electrostatic isopotentials at the receptor-binding 15 site, compared to IFN- α 17WT protein or IFN- α 2WT protein.

16 In conclusion, this study shows that the IFN- α variant GEA007.1 exhibits a more potent and 17 sustained inhibitory activity on HCV genotype 1b subgenomic replican replication than 18 standard IFN- α 2b. This enhanced inhibitory activity is associated with a more efficient 19 induction of the JAK STAT cell signalling pathway. These results warrant further evaluation 20 to determine if this enhanced activity in the HCV subgenomic replicon system could translate 21 into a clinical benefit, especially in naïve HCV genotype 1b infected patients who do not 22 respond well to the currently available IFN- α 2/ribavirin combination therapies as well as in 23 non responder patients who represent a growing population of difficult to treat patients. 24 Clinical trials are required to determine the potential of GEA007.1 to fulfil this unmet clinical 25 need. It will be also interesting to see the inhibitory effect of GEA007.1 on other HCV

genotypes both *in vitro* and *in vivo*. Furthermore, an IFN-α variant with an enhanced anti HCV genotype 1 activity may be of utmost clinical relevance in the perspective of the new
 generation treatment regimens, combining IFNs with specific small molecule HCV inhibitors
 in current development (4).

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1 Tables

	EC ₅₀ (pg/mL)	p ^a	EC ₉₀ (pg/mL)	p ^a	CC ₅₀ (pg/mL)
IFN-α 2WT	154.2 ± 8.8		3080 ± 406		>> 3333
GEA007.1	22.7 ± 9.4	< 0.05	$471~\pm 276$	< 0.05	>> 3333
GEA009.2	483.3 ± 86.6	< 0.05	> 3333	>0.05	>> 3333
GEA011.1	187.9 ± 57.5	>0.05	> 3333	>0.05	>> 3333
GEA013.1	95.6 ± 86.6	< 0.05	2156 ± 300	>0.05	>> 3333

2 Table 1: Inhibitory features and toxicity of various interferons

3 Indicated EC_{50} , EC_{90} , and $CC_{50} \pm$ standard deviation values are the means of at least three independent 4 experiments. ^a The Mann-Whitney test was used to determine whether the activities of GEA007.1, 5 GEA009.2, GEA011.1 and GEA013.1 on inhibiting HCV subgenomic replicon were significantly different

- 6 from IFN-α 2WT treatment.
- 7

8 Table 2. Relative Gene induction in BM4-5 cells after 8 hours treatment with IFN-α

9 **2WT or GEA007.1**

Gene name	IFN-α 2WT Fold increase compared to no treatment	GEA007.1 Fold increase compared to no treatment
MX1	17.83	25.92
ISG-56	2.055	4.455
ISG-15	1.7	4.125
ISG-60	1.27	3.985
ADAR	1.38	2.91
KIAA1268	1.19	2.35
IFI6-16	0.795	2.13
STAT1	1.38	2.08
MAPKK1/MEK1	0.65	1.72
PKR	0.89	1.535
P48/IRF-9	0.82	1.365

10 Human Interferon α, β Response Gene Arrays (SuperArray) were performed with a concentration of

11 33pg/ml of IFN. Indicated values are the mean of two independent experiments.

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13

1 Figures

Figure 1. GEA007.1 has improved inhibitory properties compared to IFN- α 2WT and other IFN- α variants in the replicon system. BM4-5 cells were treated during three days by IFN- α 2WT or GEA007.1. or GEA009.2 or GEA011.1 or GEA013.1 at 0, 3, 17, 33, 333, 3333 pg/mL. Total RNA was isolated from the cells and analysed by Northern Blot for negative and positive HCV subgenomic RNA. The β -actin serves as internal control for loading of cellular RNA.



Figure 2. GEA007.1 is able to clear HCV from BM4-5 cells after prolonged administration

3 BM4-5 cells were left untreated or treated every day with either IFN- α 2WT or GEA007.1 at 4 333 pg/mL in the absence of geneticin (upper panel left). At confluence, cells were passaged 5 by 1:4 dilution. After five passages, the treatment was stopped, and cells were further passaged by 1:4 dilution in medium free of geneticin (upper panel right) or containing 6 7 geneticin (lower panel right). In the latter case no result are presented for GEA007.1 as cells 8 died in the presence of geneticin (See text for detail). At each passage, total RNA was isolated 9 from cells, separated on agarose gel, and analysed by Northern Blot for the detection of 10 negative and positive RNAs. The β -actin served as internal loading control. P means cell 11 passage, O means no IFN treatment, 2wt means treatment with IFN-a 2WT, 7.1 means 12 treatment with GEA007.1.



Figure 3. GEA007.1 is a better inducer of the IFN signaling pathways compared with IFN-α 2WT. BM4.5 cells were transfected with an ISRE luciferase construct. 24 h after transfection the cells were incubated without or with IFN-α 2WT t and GEA007.1 for 24h. The cells were then harvested and assayed for luciferase activity. The luciferase activity induced by IFN-a 2WT for each concentration was taken as 100% and other luciferase activity were normalized accordingly. Mean values \pm S.D. of three independent experiments are shown. The absolute luciferase values obtained with increasing concentration of IFN-a 2WT (0 - 0,3 - 3 - 33 - 333 - 3333 pg/ml) are: 205 - 234 - 226 - 217 - 798 - 2004, respectively.



IFN induced luciferase activity (percentage of IFN-a 2WT)

Figure. 4. Effect of GEA007.1 on PKR and p48 (IRF-9) protein expression. BM4-5 cells were cultured in absence or presence 33 pg/mL of IFN-a 2WT or GEA007.1. At different time points following IFNs administration, total cellular proteins were extracted and subjected to Western Blot analysis using anti-PKR or anti-p48 antibodies. As loading control, Western Blot was performed with anti-actin antibodies.



