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Antonia Tabernero, Kristina Schoonjans, Laurence Jesel, Irina Carpusca, Johan Auwerx, et al.. Activation of the peroxisome proliferator-activated receptor alpha protects against myocardial ischaemic injury and improves endothelial vasodilatation.. *BMC Pharmacology*, 2002, 2, pp.10. inserm-00124786

HAL Id: inserm-00124786

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Submitted on 16 Jan 2007

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Research article

Activation of the peroxisome proliferator-activated receptor α protects against myocardial ischaemic injury and improves endothelial vasodilatation

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Published: 9 April 2002

BMC Pharmacology 2002, 2:10

Received: 7 December 2001

Accepted: 9 April 2002

This article is available from: <http://www.biomedcentral.com/1471-2210/2/10>

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Abstract

Background: The peroxisome proliferator-activated receptor α (PPAR α) plays an important role in the metabolism of lipoproteins and fatty acids, and seems to protect against the development of atherosclerosis. To evaluate the possible protective role of PPAR α on cardiovascular function, the effect of the PPAR α agonist, fenofibrate was assessed with respect to ischaemia/reperfusion injury and endothelial function in mice.

Results: Fenofibrate treatment reduces myocardial infarction size and improves post-ischaemic contractile dysfunction. Hearts from PPAR α null mice exhibit increased susceptibility to ischaemic damages and were refractory to protection by fenofibrate treatment suggesting that the beneficial effects of fenofibrate were mediated *via* PPAR α . Furthermore, fenofibrate improves endothelium- and nitric oxide-mediated vasodilatation in aorta and mesenteric vascular bed. A decreased inhibitory effect of reactive oxygen species in the vessel wall accounts for enhanced endothelial vasodilatation. However, the latter cannot be explained by an increase in nitric oxide synthase expression nor by an increase sensitivity of the arteries to nitric oxide.

Conclusions: Altogether the present data suggest that fenofibrate exerts cardioprotective effect against ischaemia and improves nitric oxide-mediated response probably by enhancing antioxidant capacity of the vessel wall. These data underscore new therapeutic perspectives for PPAR α agonists in ischaemic myocardial injury and in cardiovascular diseases associated with endothelial dysfunction.

Background

The peroxisome proliferator-activated receptor α (PPAR α) is a member of the nuclear hormone receptor superfamily.

PPAR α is activated by several fatty acids and is also the molecular target of hypolipidemic fibrates. PPAR α plays an important role in the liver, where it controls the metab-

olism of lipoproteins and fatty acids [1,2]. In addition, PPAR α is widely expressed in the entire cardiovascular system, including the heart and the different cells that compose the vascular wall, such as endothelial, vascular smooth muscle, and monocyte/macrophage cells. Activation of PPAR α increases uptake of fatty acids and activates their β -oxidation as a result of the induction of many genes including some coding for lipid binding proteins, apolipoproteins and those involved in peroxisomal and mitochondrial fatty acid β -oxidation [2-4]. Fatty acid β -oxidation seems to play a critical role not only in normal cardiac metabolic homeostasis but its dysregulation also contributes to the pathogenesis of a variety of disease states such as cardiac hypertrophy and heart failure. Recent studies have suggested that PPAR α agonists, in addition to their beneficial effects on lipid homeostasis, might reduce the progression of atherosclerotic lesions by a direct anti-atherogenic and anti-inflammatory action [5]. Indeed, PPAR α activation results in the induction of antioxidant enzymes and in the inhibition of the expression of pro-inflammatory genes, such as interleukin-6, cyclooxygenase 2, and endothelin within vascular cells *via* negative regulation of signalling pathways, controlled by the nuclear factor κ B (NF κ B) and the activator protein-1 (AP-1). Furthermore, PPAR α inhibits the expression of monocyte-recruiting proteins, such as vascular cell adhesion molecules [6], induces apoptosis in monocyte-derived macrophages [7], and reduces tissue factor expression and activity in monocytes [8,9], accounting for the reduction of the thrombogenicity of atherosclerotic lesions.

We analyzed here the effect of the PPAR α agonist, fenofibrate on the regulation of cardiovascular function with respect to ischaemia-reperfusion myocardial injury and endothelial dysfunction. Evidence is provided that treatment of mice with fenofibrate *via* PPAR α reduces myocardial infarction size and improves post-ischaemic contractile dysfunction, after an ischaemic period. Furthermore, we demonstrate that fenofibrate improves endothelial nitric oxide (NO)-mediated vasodilatation by increasing the antioxidant defense system of the vessel wall.

Results

Fenofibrate treatment reduces ischaemia-induced myocardial infarct

Administration of the PPAR α agonist, fenofibrate, for 10 days to male C57BL/6 mice resulted in a significant decrease in body weight as compared to control mice treated with placebo (Control: 30.1 ± 0.26 , $n = 21$; Fenofibrate: 26.8 ± 0.47 , $n = 19$, $P < 0.001$). At the end of the treatment period, hearts from fenofibrate-treated mice weighed significantly less than those from control mice (Control: 0.135 ± 0.003 g, $n = 9$; Fenofibrate: 0.113 ± 0.05 g, $n = 8$; $P < 0.05$). However the heart/body weight ratio was not

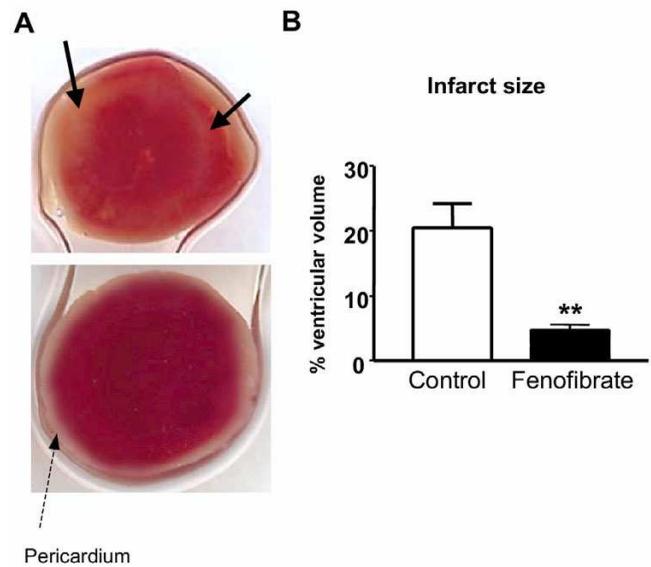


Figure 1

Fenofibrate treatment reduces ischaemia-induced myocardial infarct. (A) Representative images of transversal slices from ischaemic hearts from control (upper panel) and fenofibrate-treated (lower panel) mice stained with triphenyltetrazolium (TTC). The arrows represent unstained areas (white) showing irreversible damaged myocardial tissue. (B) Volume of infarcted myocardium. Data are the mean \pm SEM of 6 experiments. ** $P < 0.01$

altered after fenofibrate treatment (Control: 0.48 ± 0.006 %, $n = 9$; Fenofibrate: 0.46 ± 0.11 %, $n = 8$).

To analyze the effects of PPAR α activation on myocardial infarction, hearts were subjected to 30 min global, no-flow ischaemia at 37°C, followed by 1 h reperfusion. Myocardial infarct size was evaluated morphologically using TTC-staining as an index of irreversible myocardial injury (Fig. 1A). Fenofibrate treatment significantly reduced ischaemia-induced myocardial infarction size by more than 3-fold relative to control mice ($P < 0.01$) (Fig. 1B).

Fenofibrate improves the post-ischaemic recovery of cardiac function

To determine whether treatment of mice with fenofibrate protects against ischaemia/reperfusion-induced functional injury, ventricular parameters such as developed force, heart rate, and coronary flow were studied in control and fenofibrate-treated animals. Basal parameters were not significantly different between the two groups of mice at the end of the equilibration period before ischaemia as (Table 1). After the ischaemic insult, all the hearts from fenofibrate-treated mice recovered both contractile force and heart rate whereas only 58% of the hearts in control mice showed an improvement (Fig. 2A). For the following

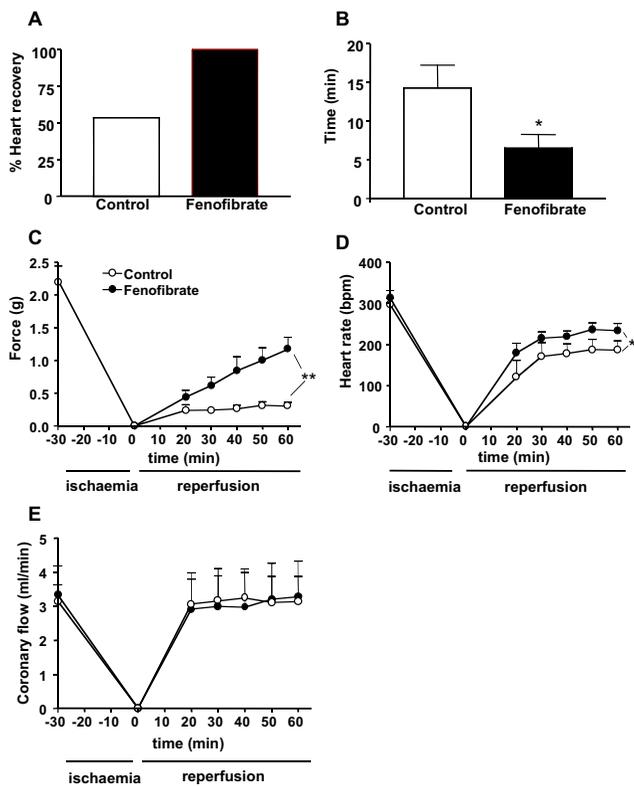


Figure 2
Fenofibrate improves the post-ischaemic recovery of cardiac function. Percentage of hearts that recover after ischaemic condition (A), time of recovery (B) and postischaemic recovery of force (C), heart rate (D), and coronary blood flow (E) in hearts from control and fenofibrate-treated mice. Data are the mean \pm SEM of 7–10 experiments. * $P < 0.05$, ** $P < 0.01$.

measured parameters, only the values of heart that recovered for the ischaemic insult have been included. Also the time of recovery of force and heart rate after the ischaemia was significantly shorter in hearts from fenofibrate-treated as compared to control mice (Fig. 2B). Recovery of contractile force in hearts from control group was stabilized after 20 min reperfusion (Fig. 2C). The level of the contractile force recovered was, however, very low (*i.e.* 13%), when compared with the contractile force measured before the induction of the ischaemia. By contrast in hearts from fenofibrate-treated mice, the recovery in cardiac contractile force was dramatically enhanced (60 % of the initial force), and did not yet reach maximal values 1 h after reperfusion. Thus at the end of the reperfusion period, the developed force was significantly greater in fenofibrate-treated mice than in control mice. Heart rate recovered rather quickly and reached a steady state level 30 min after reperfusion in both groups of mice. The level of heart rate recovery was, however, significantly higher in fenofibrate-treated mice when compared to that of control mice (Fig.

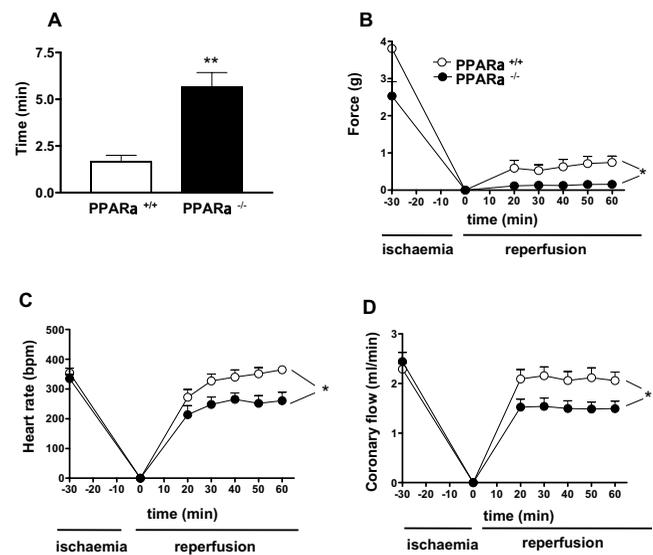


Figure 3
Hearts from $PPAR\alpha^{-/-}$ mice were more susceptible to post-ischaemic damages than those from $PPAR\alpha^{+/+}$ mice. Time of recovery (A) and post-ischaemic recovery of force (B), heart rate (C), and coronary blood flow (D) in hearts from $PPAR\alpha^{+/+}$ and $PPAR\alpha^{-/-}$ mice. Data are the mean \pm SEM of 6 experiments. * $P < 0.05$, ** $P < 0.01$.

Table 1: Ventricular functional parameters measured before ischaemia in control and fenofibrate-treated mice.

	Force (g)	Heart rate (beats/min)	Coronary flow (ml/min)
Control (n = 7)	2.19 \pm 0.66	296 \pm 42	3.03 \pm 1.15
Fenofibrate (n = 10)	3.01 \pm 1.09	312 \pm 59	3.03 \pm 1.29

2D). The coronary flow rate returned after 20 min of reperfusion to its initial value before the ischaemic condition in the two groups of animals and it remained stable until the end of the reperfusion period (Fig. 2E). Altogether, these data suggest that fenofibrate treatment markedly improves myocardial contractile function following ischaemia/reperfusion.

Hearts from mice lacking $PPAR\alpha$ ($PPAR\alpha^{-/-}$) were more susceptible to post-ischaemic damages and were refractory to myocardial protection by fenofibrate treatment

To test the role of $PPAR\alpha$, ischaemia/reperfusion-induced functional injury was conducted in hearts from both $PPAR\alpha^{+/+}$ and $PPAR\alpha^{-/-}$ mice. After the ischaemic insult, the percentage of hearts recovered from both contractile force

and heart rate were not different between PPAR $\alpha^{+/+}$ and PPAR $\alpha^{-/-}$ mice being 75% versus 87% respectively. However, the time of recovery of force and heart rate after the ischaemia was significantly shorter in hearts from PPAR $\alpha^{+/+}$ as compared to hearts from PPAR $\alpha^{-/-}$ mice (Fig. 3A). Recovery of contractile force in hearts from control group was stabilized after 20 min reperfusion (Fig. 3B). The level of the contractile force recovered was, however, very low (*i.e.* 22%), when compared with the contractile force measured before the induction of the ischaemia. Interestingly in hearts from PPAR $\alpha^{-/-}$ mice, the recovery in cardiac contractile force was dramatically decreased (8 % of the initial force), and did not improve 1 h after reperfusion. Thus at the end of the reperfusion period, the developed force was significantly lower in hearts from PPAR $\alpha^{-/-}$ than in those from PPAR $\alpha^{+/+}$ mice. Heart rate recovered rather quickly and reached a steady state level 30 min after reperfusion in both groups of mice. The level of heart rate recovery was, however, significantly higher in heart from PPAR $\alpha^{+/+}$ mice when compared to that of PPAR $\alpha^{-/-}$ mice (Fig. 3C). The coronary flow rate returned after 20 min of reperfusion to its initial value before the ischaemic condition in PPAR $\alpha^{+/+}$ mice and it remained stable until the end of the reperfusion period (Fig. 3D). In PPAR $\alpha^{-/-}$ mice, the coronary flow did not returned to its initial values (*i.e.* 60%). Furthermore, the level of coronary flow recovery in PPAR $\alpha^{-/-}$ mice was significantly lower than the level obtained in heart from PPAR $\alpha^{+/+}$ mice.

Finally, to test whether the protective effect of fenofibrate treatment against ischaemia/reperfusion-induced functional injury was due to PPAR α activation, experiments were carried out in heart from mice lacking PPAR α treated with fenofibrate. Interestingly, no beneficial effect of fenofibrate on PPAR $\alpha^{-/-}$ mice was observed and in fact 5 out of 6 hearts were dead after ischaemia.

Altogether, these data suggest that hearts from PPAR $\alpha^{-/-}$ mice were more susceptible to myocardial dysfunctions following ischaemia/reperfusion than those from PPAR $\alpha^{+/+}$ mice. In addition, the beneficial effects of fenofibrate treatment on myocardial functions following ischaemia reperfusion were lost and even worsened in PPAR $\alpha^{-/-}$ mice leading us to conclude that this effect was mediated through PPAR α .

The hypothesis that nitric oxide (NO) may play a role in the fenofibrate treatment-induced cardioprotection was also tested by analyzing the expression of endothelial NO synthase (eNOS) in hearts taken from either control or fenofibrate treated mice. Western blot analysis showed that fenofibrate treatment did not modify eNOS protein expression in the heart (Fig. 4A). Likewise, fenofibrate treatment did not affect the expression of the RNA of the

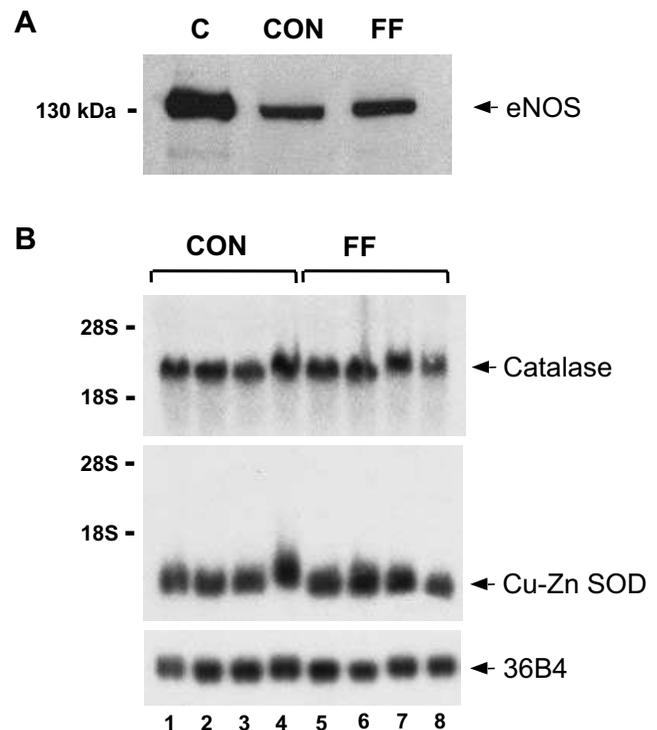


Figure 4
Fenofibrate does not modified eNOS, catalase or Cu-Zn SOD expression in the heart. Representative Western-blot (A) of eNOS protein expression in hearts from control mice (CON) or mice treated with fenofibrate (FF). Positive control (C) is the eNOS from bovine endothelial cells. Similar results were obtained in three separate experiments. Effect of fenofibrate on catalase and Cu-Zn SOD mRNA expressions in heart (B). Expression of catalase mRNA, Cu-Zn SOD mRNA and the human acidic ribosomal phosphoprotein 36B4.

antioxidant genes Cu-Zn superoxide dismutase (Cu-Zn SOD) or catalase (Fig. 4B,4C).

Fenofibrate enhances the participation of NO in modulating vasoconstriction

We next analyzed contractile responses in the aorta and in the mesenteric vascular bed. In mesenteric vascular bed, no significant differences in the changes in perfusion pressure produced by the vasoconstrictor agonist, phenylephrine were observed between control and fenofibrate-treated mice (Table 2). Likewise, neurogenic vasoconstriction of the mesenteric bed produced by electrical stimulation was not different between the two groups of mice. In the aorta, the thromboxane A₂ analogue, U46619, produced a concentration-dependent contraction that was not different between both groups of animals. This led us to conclude that fenofibrate treatment did not modify the contractile responses to vasoconstrictor agonists or to

Table 2: Maximal effect (E_{max}) and EC_{50} values for phenylephrine- and electrical stimulation induced increases in perfusion pressure in mesenteric bed and for U46619-induced contraction in aorta from control and fenofibrate-treated mice.

	Control	Fenofibrate
MESENTERIC BED		
Phenylephrine		
E_{max} (mmHg)	22.14 ± 7.03	21.73 ± 5.55
EC_{50} (M)	1.46 ± 0.58 10 ⁻⁵ (n = 9)	2.12 ± 0.55 10 ⁻⁵ (n = 9)
Electrical stimulation		
E_{max} (mmHg)	14.17 ± 1.88 (n = 7)	15.15 ± 2.06 (n = 9)
AORTA		
U46619		
E_{max} (mN/mm)	4.91 ± 0.34	5.16 ± 0.37
EC_{50} (M)	2.95 ± 0.85 10 ⁻⁸ (n = 6)	2.8 ± 0.83 10 ⁻⁸ (n = 6)

electrical stimulation in the two vascular preparations studies, mesenteric bed and the aorta (Table 2).

In the aorta, the NO-synthase inhibitor, L-NAME (300 μ M) was without effect on U46619-induced contraction in aorta rings from control mice (Fig. 5A), whereas it significantly enhanced the sensitivity of the vasoconstrictor agonist in aortas taken from fenofibrate-treated mice (Fig. 5B).

Fenofibrate improves endothelial function as a result of an increased anti-oxidant capacity of the vessel wall

The hypothesis that fenofibrate treatment improves endothelial function was tested next by looking at the endothelium-dependent relaxation to acetylcholine both in the aorta and the mesenteric vascular bed. Interestingly, both the mesenteric vascular bed (Fig. 6A) and the aorta (Fig. 6B) from mice treated with fenofibrate displayed enhanced endothelium-dependent relaxation to acetylcholine, as compared to vascular preparations from control mice.

The common endothelial factor released by the two vascular preparations is NO. Therefore, we investigated whether the improvement of acetylcholine-induced relaxation in vessels from fenofibrate treated animals could be explained by an enhanced participation of endothelial NO. Western blot analysis showed that eNOS protein expression was not modified by the fenofibrate treatment both in the mesenteric vascular bed and in the aorta (Fig. 6C,B,D), suggesting that the enhanced relaxation to acetylcholine cannot be attributed to an increased expression

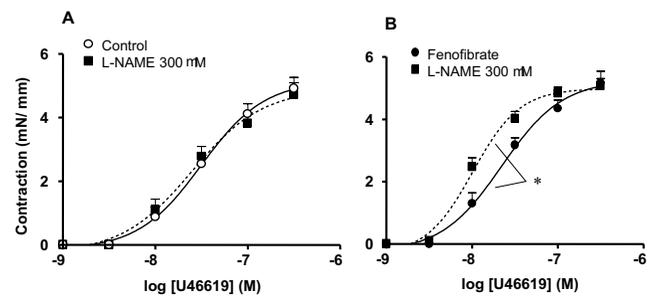


Figure 5
Fenofibrate enhances the participation of NO in modulating vasoconstriction. Concentration-effect curves for U46619-induced vasoconstriction obtained in thoracic aorta rings of control (A) and fenofibrate-treated (B) mice in the absence or presence of N^w-nitro-L-arginine methyl ester (L-NAME, 300 μ M). Data are the mean \pm SEM of 6–7 experiments. *P < 0.05.

of eNOS protein in the two vascular preparations. Relaxation in response to the NO-donor, SNAP, was hence studied in both the mesenteric bed and the aorta, in order to test the hypothesis of an increased response to NO or decreased of NO breakdown by reactive oxygen species. In the two vascular preparations from fenofibrate-treated mice, SNAP-mediated relaxation was significantly greater than the relaxation obtained in vessels from control mice (Fig. 7A,B). Interestingly, the combined addition of the superoxide anion and the hydrogen peroxide scavengers, SOD (200 U/ml) and catalase (750 U/ml) increased relaxation to SNAP in the aorta taken from control, but not in that from fenofibrate-treated mice (Fig. 7C). The resulting concentration-response curves to SNAP were hence not significantly different in the aorta from the two groups of mice in the presence of reactive oxygen scavengers. These data suggest that the antioxidant capacity of aortic wall from fenofibrate-treated mice was already sufficiently high. In contrast to the situation in the controls, supplementation with reactive oxygen scavengers (SOD and catalase) could not further enhance the protection against NO breakdown and the relaxation to SNAP. Increased antioxidant capacity of the vessel wall from fenofibrate-treated mice was not associated with an increased expression of mRNA for Cu-Zn SOD or catalase in the aorta and mesenteric bed (data not shown).

Discussion

In the present study, we demonstrate that fenofibrate treatment reduces the size of the myocardial infarction and improves post-ischaemic contractile dysfunction. This effect of fenofibrate is mediated via PPAR α activation. In addition we report that fenofibrate improves endothelial NO-mediated vasodilatation both in the aorta (conductance vessel) and in the mesenteric vascular bed

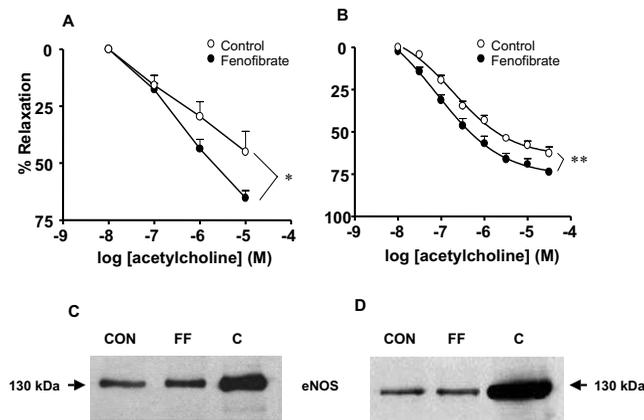


Figure 6
Fenofibrate improves endothelial function. Acetylcholine-induced relaxation in mesenteric bed (A) and thoracic aorta (B). Results were expressed as a percent of the initial phenylephrine- or U46619-induced pre-contraction level. The values of contraction obtained with phenylephrine (10 μ M) in mesenteric bed from control and fenofibrate-treated mice were 10.8 ± 1.1 mmHg ($n = 9$) and 11.44 ± 1.5 mmHg ($n = 9$), respectively. The values of contraction obtained with U46619 (30 nM) in aorta from control and fenofibrate-treated mice were 3.26 ± 0.2 mN/mm ($n = 8$) and 3.01 ± 0.2 mN/mm ($n = 8$) respectively. Data are the mean \pm SEM of 8–9 experiments. * $P < 0.05$, ** $P < 0.01$. Representative Western-blot of endothelial NO-synthase (eNOS) protein expression in mesenteric bed (C) and thoracic aortic segments (D) from control (CON) and fenofibrate-treated (FF) mice. Positive control (C) is the eNOS from bovine endothelial cells. Similar results were obtained in three separate experiments.

(which contains small arteries that participate actively in the regulation of peripheral resistance and thus blood pressure). Increased anti-oxidant capacity of the vessel wall, but not increased eNOS expression or increased sensitivity to NO of the vascular smooth muscle accounts for this enhancement of endothelial vasodilatation.

The function of PPAR α in the heart with respect to cardioprotection against severe ischaemia has not yet been documented. Here, we provide evidence that fenofibrate treatment reduces the infarct size after ischaemia. Furthermore, fenofibrate treatment shortened the time of recovery of the heart after an ischaemic insult and increased the percentage of heart that recovered from ischaemia. Moreover, fenofibrate treatment markedly improved contractile function. It should be noted that fenofibrate treatment did not produce any beneficial effect in heart from PPAR α ^{-/-} mice. Finally, heart from PPAR α ^{-/-} mice displayed increased susceptibility to ischaemia/reperfusion-induced functional injury *versus* those from PPAR α ^{+/+} mice. Altogether these results show that fenofibrate through the activation of PPAR α exerts a cardio-protective effect. The

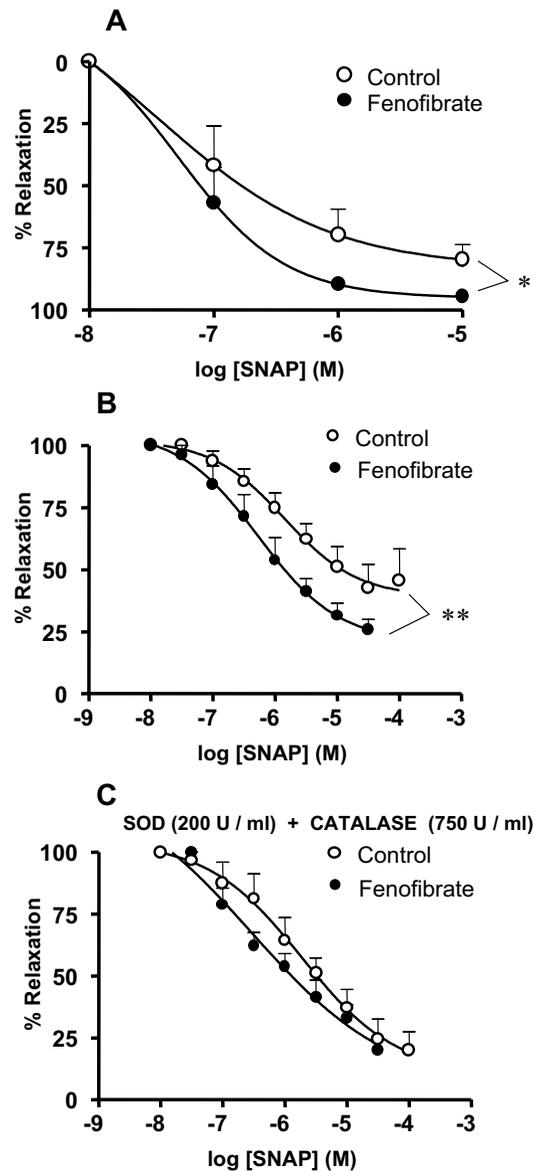


Figure 7
Fenofibrate increases anti-oxidant capacity of the vessel wall. Concentration-effect curves to the NO-donor S-N-acetylpenicillamine (SNAP) obtained in mesenteric bed (A) and thoracic aorta rings (B). Concentration-effect curves to SNAP obtained in aorta rings in the presence (C) of superoxide dismutase (SOD, 200 U/ml) plus catalase (750 U/ml). Results were expressed as a percent of the initial U46619-induced precontraction level. The values of contraction obtained with U46619 (0.3 μ M for mesenteric bed and 30 nM in thoracic aorta) were 22.4 ± 5.6 mmHg ($n = 5$) and 28 ± 4.9 mmHg ($n = 6$) in mesenteric bed and 3.55 ± 0.15 mN/mm ($n = 7$) and 3.37 ± 0.2 mN/mm ($n = 6$) in aorta from control and fenofibrate-treated mice, respectively. Pretreatment with SOD and catalase did not affect contractile responses to U46619 (data not shown). Data are the mean \pm SEM of 5–7 experiments. * $P < 0.05$, ** $P < 0.01$.

mechanisms that may explain these effects of fenofibrate are not fully understood. In the heart, PPAR α has been reported to regulate cardiac lipid and energy metabolism, by controlling genes encoding the mitochondrial fatty acid β -oxidation pathway, which provides the major source of energy under the form of ATP to the heart muscles [4]. The use of PPAR α ^{-/-} mice has provided evidence for the role of PPAR α in the control of cardiac lipid utilization. Although PPAR α ^{-/-} mice appear normal under basal conditions, the constitutive expression of protein and enzymes involved in fatty acid transport and metabolism are all significantly lower in heart and liver compared to age matched control [10–14]. The reduction in fatty acid transport and metabolism may hence explain the increased accumulation of lipid, the decreased energy consumption, and the reduction ATP content in hearts from PPAR α ^{-/-} mice. These above reports are supported by our data showing that hearts from PPAR α ^{-/-} mice were more susceptible to ischaemic insults when compared to those from PPAR α ^{+/+} animals. Indeed, the levels of post-ischaemic recovery of contractile force, heart rate and coronary flow were lower in hearts from PPAR α ^{-/-} versus PPAR α ^{+/+} mice. In mice in which PPAR α gene is valid, PPAR α agonists such as fenofibrate would favour the maintenance of adequate energy levels, to protect against the fall in ATP during ischaemia by increasing the expression of genes involved in fatty acid utilization, and therefore the content of ATP. This may explain the improved survival of cardiomyocytes and their resistance to ischaemia, as well as the preservation of cardiac contractile function. Most of ischaemia-reperfusion injury is ultimately due to a burst of reactive oxygen species that occurs upon readmission of oxygen [15,16]. One of the potential mechanisms, which contribute to protection against ischaemia-reperfusion damage by fenofibrate, may be an increase in the antioxidant capacity of the heart due to the induction of antioxidant enzymes. Indeed, under certain conditions, activation of PPAR α *in vivo* has been reported to cause an induction of mRNA and protein levels of antioxidant enzymes, such as catalase, Cu-Zn SOD, and mediators of glutathione pathway [5,17,18] which may be able to counterbalance, at least partly, the deleterious effects of oxidative stress during the reperfusion. In the present study, no increased expression of Cu-Zn SOD or catalase mRNA was observed within the heart. Finally, PPAR α activation decreases inflammatory gene response by inhibiting NF κ B and AP-1 activity, and by establishing a new redox balance caused by oxidant stress-related disease states [5,17,18]. The later would be responsible for the increase in defence mechanisms towards the myocardial infarction and reduced myocardial effects of ischaemia.

PPAR α has also been identified in different vascular cell types including endothelial, smooth muscle cells and monocyte/macrophages [3,19]. There is growing evidence

that the effect of PPAR α activation may not solely be restricted to an improvement in the atherogenic lipoprotein profile, but may also be caused by a direct beneficial effect on vessel wall [20]. The early stages of atherosclerosis may result from an insult to the endothelium, a reduction in the production of protective and relaxant factors (*i.e.* NO, prostacyclin or endothelium-dependent hyperpolarizing factor), or both factors combined. This may consequently induce other changes, such as the expression of adhesion molecules, the reduction of barrier function, and the uptake of lipoproteins into the sub-endothelial matrix. Although inhibition of NF κ B and AP-1 could limit the secretion of endothelin-1 by endothelial cells or reduce vascular cell adhesion molecule-1 expression, no direct studies have looked at the beneficial effect of fenofibrate treatment on endothelial function. In the present study, we demonstrated that fenofibrate treatment improved acetylcholine-induced relaxation in the aorta and in the mesenteric vascular bed with regard to endothelium-dependent vasodilatation. A reduced contractility of the vessels to vasoconstrictor agonists cannot account for the enhanced relaxation, because the levels of pre-contractions were matched between vascular beds from either control or fenofibrate-treated mice. Furthermore, fenofibrate treatment did not affect the response elicited by nerve stimulation nor that produced by the α_1 -adrenoceptor agonist, phenylephrine at least in the mesenteric bed. Interestingly, the use of the NO synthase inhibitor unmasked the enhanced participation of NO in the regulation of vasoconstrictor activity in the aorta from fenofibrate-treated mice. The most relevant hypothesis to explain the enhanced relaxation to acetylcholine upon fenofibrate treatment might be an increase in either generation (*i.e.* synthesis or release) of, or response to endothelial relaxant factors. Alternatively, an alteration in the balance between endothelial relaxing and constricting factors can be invoked.

One of the major endothelial vasodilator factor and which is also common to the two vascular preparations studied (*i.e.*, aorta and mesenteric bed) is NO, the reduced participation of which has been implicated in diseases linked to endothelial dysfunction [21]. Furthermore, NO has been shown to inhibit platelet aggregation [22], to limit the flux of the atherogenic plasma proteins into the artery walls [23] and in the longer term, to induce the expression of genes protective to the cardiovascular system [24]. Fenofibrate-induced increased participation of endothelial NO cannot be attributed to an increased expression of the eNOS protein, both in aorta and mesenteric bed, even though an increased enzymatic activity cannot be fully ruled out. The increased relaxation to acetylcholine may, however, result from an increased sensitivity of the smooth muscle cell to NO, as shown by the improvement of the vasodilator effect of the NO donor, SNAP, in

vessels taken from fenofibrate-treated mice. It is interesting to note that the superoxide anion scavengers, SOD and catalase, were able to potentiate the effect of SNAP-induced relaxation in the aorta from control mice, but not in those from fenofibrate-treated mice. Thus, the vasodilatory response to the NO donor was not different in the two groups after blockade of superoxide anion production. This would imply that the antioxidant capacity of the aortic wall from fenofibrate-treated mice was sufficiently high, that the supplementation with reactive oxygen scavengers could not produce any further protection against NO breakdown nor a further increase in relaxation to SNAP. Therefore, the most and parsimonious hypothesis to explain the enhanced endothelium-dependent relaxation to acetylcholine after fenofibrate treatment is an increased antioxidant defence of the vessel wall, that preserves endothelial NO from breakdown by the superoxide anions. Up-regulation of mediators of the glutathione pathway [5,17,18] or down regulation of the superoxide generating enzyme, NADPH oxidase [18] might account for the effect fenofibrate since we did not observe any changes in the expression of Cu-Zn SOD or catalase in the aorta and the mesenteric bed.

In conclusion, the present study proves that treatment with the PPAR α agonist, fenofibrate, has protective effects against myocardial infarction induced by severe ischaemia and significantly improves endothelial vasodilatation. All the above effects of PPAR α agonists open new therapeutic perspectives for such compounds in ischaemic myocardial injury and in cardiovascular diseases associated with endothelial dysfunction, secondary to pro-inflammatory or oxidant stress-related disease states.

Materials and methods

Animals and general issues

All animal experimentation was performed in accordance with institutional guidelines, and protocols were approved by the French Animal Care Committee in accordance with European regulations.

This study was performed in male adult mice (C57BL/6 strain, 13–15 week-old). Mice were housed at 20°C with a 12 h light cycle, and allowed free access to food and water. One group of animals (control) received placebo and the second received during 10 days, fenofibrate mixed in the food at 0.5 % w/w (15 mg/day) [2]. The dose used is in the range of that commonly reported in mice in which no toxic effect of fenofibrate has been observed as assessed by alteration in body weight at least in PPAR α ^{+/+} mice [2,25–27]. To test whether the effect of fenofibrate was due to activation of PPAR α , a group of PPAR α ^{-/-} mice (25% C57BL/6, 75% SV-129) also received fenofibrate for 10 days. To further study the role of PPAR α , on ischaemic insult, comparisons were made between PPAR α ^{+/+} versus

PPAR α ^{-/-} mice (25% C57BL/6, 75% SV-129). Prior to experimentation, mice were anaesthetised with ketamine (100 mg/kg i.p.), medetomidine (50 μ g/kg i.p.) and heparinized (500 U/kg i.p.).

Isolated perfused heart

The heart was excised, cannulated and perfused according to Langendorff at 37°C and pH 7.4 with Krebs-Henseleit solution containing (mM) NaCl 118, NaHCO₃ 24, KCl 4, MgCl₂ 1.2, NaH₂PO₄ 1, CaCl₂ 2.5, Na₂EDTA 0.5 and glucose 10, gassed with 95%O₂/5% CO₂. Perfusion pressure was constant and equivalent to 80 cm H₂O. The heart was stretched to a force of 1 g. The preparation was equilibrated for 30 min, then subjected to 30 min global no-flow ischaemia at 37°C.

Morphological analysis of cardiac ischaemic damage by triphenylene-tetrazolium chloride (TTC)-staining

After the ischaemic insult followed by 1 h reperfusion, hearts were stored at -20°C during 24 h. After this period, the hearts were sliced transversally (2 mm thickness) and incubated at 37°C with TTC for 15 min. Slices were then fixed with 10% formalin for 20 min. After scanning the slice, measurements of stained and unstained TTC areas of the hearts were carried out by the use of a planimeter. The infarct size was calculated as a percentage of the total ventricular volume minus cavities.

Evaluation of cardiac function and post-ischaemic recovery

Mechanical activity of hearts from control and fenofibrate-treated mice were studied at baseline condition and during 60 min reperfusion following 30 min ischaemia. Cardiac parameters were monitored continuously and included cardiac developed force (g), heart rate (beats/min) and coronary flow (ml/min).

Vascular reactivity experiments

Mesenteric vascular perfused bed

The mesenteric bed was perfused via the main mesenteric artery with physiological salt solution (PSS) with the following composition in mM: NaCl 119; KCl 4.7; KH₂PO₄ 0.4; NaHCO₃ 15; MgSO₄ 1.17; CaCl₂ 2.5 and glucose 5.5, and was mounted in a perfusion system. Flow rate was adjusted at 2 ml/min with PSS maintained at 37°C and continuously gassed with 95%O₂/5%CO₂ (pH 7.4). Agonists were administered intraluminally. Changes in perfusion pressure (Δ P, mmHg) were measured with a pressure transducer and recorded by a physiograph. Concentration-response curves to the vasodilator agents acetylcholine or the NO donor, S-nitroso-N-acetylpenicillamine (SNAP; Sigma, Grenoble, France) were performed on mesenteric bed pre-contracted with either phenylephrine or the thromboxane A₂ analogue 9,11-dideoxy-9 α ,11 α -methanoepoxy Prostaglandin F_{2 α} (U46619; Cayman Chemical Company, USA). Also, concentration-response

curves to phenylephrine were constructed. Finally, the mesenteric bed was subjected to nerve stimulation using a tungsten bipolar electrode. Rectangular pulses (0.2 ms, 100 mA) were applied at 7 min intervals for 10 s at various frequencies (5, 10, 15 and 20 Hz).

Aortic ring preparations

Segments of 2 mm of intact aorta were mounted on a wire myograph [28] filled with PSS continuously kept at 37°C and gassed with 95% O₂/5%CO₂ (pH 7.4) and mechanical activity was recorded isometrically. Concentration-response curves were constructed by cumulative application of U46619 in the absence or in the presence of the NO-synthase inhibitor, N ω nitro-L-arginine methyl ester (L-NAME, 300 μ M). In another set of experiments, endothelium-dependent relaxation to acetylcholine (0.01–100 μ M) or endothelium-independent relaxation to the exogenous NO-donor, SNAP (0.01–100 μ M) were examined in vessels pre-contracted with U46619. Concentration-response curves to SNAP were also constructed in the absence or in the presence of the superoxide anion and the hydrogen peroxide scavengers, superoxide dismutase (SOD, 200 U/ml) and catalase (750 U/ml). The inhibitors were added into the bath 25 min prior to the pre-contraction with U46619.

Western blot analysis

Cytosolic proteins either from heart, mesenteric bed or aorta were subjected to SDS-PAGE using 7% gels. After electrophoresis, proteins were transferred to nitrocellulose membranes and were probed with a monoclonal mouse anti-endothelial NO-synthase (eNOS) antibody (Transduction Laboratories, Montluçon, France). Bound antibodies were detected with a secondary peroxidase-conjugated anti-mouse antibody (Bio-Rad, Ivry sur Seine, France). The bands were visualized using the enhanced chemiluminescence system (ECL, Amersham, Buckinghamshire, UK) and quantified by densitometry.

RNA analysis

RNA preparation, northern blotting and quantification of total cellular RNA derived from heart, aorta and mesenteric bed were performed as described previously [2]. Blots of heart tissues were hybridized with 32P-labeled cDNAs corresponding to full length mouse Cu-Zn SOD (GenBank Accession Number X06683) and mouse catalase (GenBank Accession Number L25069, nt 1109–1504). The human acidic ribosomal phosphoprotein 36B4 probe was used as a marker for equal loading [29]. Reverse transcription of RNA was performed at 37°C for 1 h using M-MLV reverse transcriptase (Life Technologies) and oligo dT, followed by a 15 min inactivation at 70°C. Quantification of aortic and mesenteric bed mRNA for Cu-Zn SOD and catalase was carried out by real time PCR using a

LightCycler and the DNA double-stranded specific SYBR Green I dye for detection (Roche, Basel, Switzerland).

Data analysis and statistics

All results are expressed as mean \pm SEM of *n* experiments, *n* representing the number of mice. The sensitivities to vasoconstrictor agonists are expressed as EC₅₀ values which represent the half maximally effective molar concentration. Statistical evaluation was carried out using two-way analysis of variance (ANOVA) or unpaired Student's *t*-test. The level of significance was *P* < 0.05.

Authors' contributions

AT carried out organ bath and Western blot experiments, and participated in the design of the study. KS carried out RTPCR experiments, and participated in the design of the study. LJ and IC carried out organ bath experiments (heart and blood vessels). JA participated in the design of the study and in the redaction of the manuscript. RA conceived of the study, and participated in its design, redaction and coordination.

Acknowledgements

Antonia Taberero was supported by the "Fondation pour la Recherche Médicale". This work was supported by "Fonds de Recherche Hoechst Marion Roussel (GIP HMR)": Exploration Fonctionnelle et Analyse Globale de l'Expression des Gènes.

The authors would like to thank Dr Gonzalez F.J for providing the PPAR alpha knock out mice (Mol. Cell. Biol. 15: 3012-3022, 1995).

References

1. Kersten S, Desvergne B, Wahli W: **Roles of PPARs in health and disease.** *Nature* 2000, **405**:421-424
2. Schoonjans K, Peinado-Onsurbe J, Lefebvre AM, Heymann RA, Briggs M, Deeb S, Staels B, Auwerx J: **PPARalpha and PPARgamma activators direct a distinct tissue-specific transcriptional response via a PPRE in the lipoprotein lipase gene.** *Embo J* 1996, **15**:5336-5348
3. Bishop-Bailey D: **Peroxisome proliferator-activated receptors in the cardiovascular system.** *Br. J. Pharmacol.* 2000, **129**:823-834
4. Brandt JM, Djouadi F, Kelly DP, Brandt JM, Djouadi F, Kelly DP: **Fatty acids activate transcription of the muscle carnitine palmitoyltransferase I gene in cardiac myocytes via the peroxisome proliferator-activated receptor alpha.** *J. Biol. Chem.* 1998, **273**:23786-92
5. Delerive P, De Bosscher K, Besnard S, Vanden Berghe W, Peters JM, Gonzalez FJ, Tedgui A, Haegeman G, Staels B: **Peroxisome proliferator-activated receptor alpha negatively regulates the vascular inflammatory gene response by negative cross-talk with transcription factors NF-kappaB and AP-1.** *J. Biol. Chem.* 1999, **274**:32048-32054
6. Jackson SM, Parhami F, Xi XP, Berliner JA, Hsueh WA, Law RE, Demer LL: **Peroxisome proliferator-activated receptor activators target human endothelial cells to inhibit leukocyte-endothelial cell interaction.** *Arterioscler Thromb Vasc* 1999, **19**:2094-2104
7. Chinetti G, Griglio S, Antonucci M, Torra IP, Delerive P, Majd Z, Fruchart JC, Chapman J, Najib J, Staels B: **Activation of proliferator-activated receptors alpha and gamma induces apoptosis of human monocyte-derived macrophages.** *J. Biol. Chem.* 1998, **273**:25573-25580
8. Neve BP, Corseaux D, Chinetti G, Zawadzki C, Fruchart JC, Duriez P, Staels B, Jude B: **PPAR-alpha agonists inhibit tissue factor expression in human monocytes and macrophages.** *Circulation* 2001, **103**:207-212

9. Marx N, Mackman N, Schonbeck U, Yilmaz N, Hombach VV, Libby P, Plutzky J: **PPAR-alpha activators inhibit tissue factor expression and activity in human monocytes.** *Circulation* 2001, **103**:213-219
10. Aoyama T, Peters JM, Iritani N, Nakajima T, Furihata K, Hashimoto T, Gonzalez FJ: **Altered constitutive expression of fatty acid-metabolizing enzymes in mice lacking the peroxisome proliferator-activated receptor alpha (PPARalpha).** *J. Biol. Chem.* 1998, **273**:5678-5684
11. Djouadi F, Brandt JM, Weinheimer CJ, Leone TC, Gonzalez FJ, Kelly DP: **The role of the peroxisome proliferator-activated receptor alpha (PPAR alpha) in the control of cardiac lipid metabolism.** *Prostaglandins Leukot. Essent. Fatty Acids* 1999, **60**:339-343
12. Watanabe K, Fujii H, Takahashi T, Kodama M, Aizawa Y, Ohta Y, Ono T, Hasegawa G, Naito M, Nakajima T, et al: **Constitutive regulation of cardiac fatty acid metabolism through peroxisome proliferator-activated receptor alpha associated with age-dependent cardiac toxicity.** *J. Biol. Chem.* 2000, **275**:22293-22299
13. Barger PM, Kelly DP: **PPAR signaling in the control of cardiac energy metabolism.** *Trends Cardiovasc. Med.* 2000, **10**:238-245
14. Barger PM, Brandt JM, Leone TC, Weinheimer CJ, Kelly DP: **Deactivation of peroxisome proliferator-activated receptor-alpha during cardiac hypertrophic growth.** *J. Clin. Invest.* 2000, **105**:1723-1730
15. Wang P, Zweier JL: **Measurement of nitric oxide and peroxynitrite generation in the postischemic heart. Evidence for peroxynitrite-mediated reperfusion injury.** *J. Biol. Chem.* 1996, **271**:29223-29230
16. Yasmin W, Strynadka KD, Schulz R: **Generation of peroxynitrite contributes to ischemia-reperfusion injury in isolated rat hearts.** *Cardiovasc. Res.* 1997, **33**:422-432
17. Poynter ME, Daynes RA: **Peroxisome proliferator-activated receptor alpha activation modulates cellular redox status, represses nuclear factor-kappaB signaling, and reduces inflammatory cytokine production in aging.** *J. Biol. Chem.* 1998, **273**:32833-32841
18. Inoue I, Goto S, Matsunaga T, Nakajima T, Awata T, Hokari S, Komoda T, Katayama S: **The ligands/activators for peroxisome proliferator-activated receptor alpha (PPARalpha) and PPARgamma increase Cu²⁺, Zn²⁺-superoxide dismutase and decrease p22phox message expressions in primary endothelial cells.** *Metabolism* 2001, **50**:3-11
19. Fruchart JC, Duriez P, Staels B: **Peroxisome proliferator-activated receptor-alpha activators regulate genes governing lipoprotein metabolism, vascular inflammation and atherosclerosis.** *Curr. Opin. Lipidol.* 1999, **10**:245-57
20. Buchan KW, Hassall DG: **PPAR agonists as direct modulators of the vessel wall in cardiovascular disease.** *Med. Res. Rev.* 2000, **20**:350-366
21. Cai H, Harrison DG: **Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress.** *Circ. Res.* 2000, **87**:840-844
22. Yao SK, Ober JC, Krishnaswami A, Ferguson JJ, Anderson HV, Golino P, Buja LM, Willerson JT: **Endogenous nitric oxide protects against platelet aggregation and cyclic flow variations in stenosed and endothelium-injured arteries.** *Circulation* 1992, **86**:1302-1309
23. Cardona-Sanclemente LE, Born GV: **Effect of inhibition of nitric oxide synthesis on the uptake of LDL and fibrinogen by arterial walls and other organs of the rat.** *Br. J. Pharmacol.* 1995, **114**:1490-1494
24. Bogdan C: **Nitric oxide and the regulation of gene expression.** *Trends Cell Biol.* 2001, **11**:66-75
25. Berthou L, Duverger N, Emmanuel F, Langouët S, Auwerx J, Guillouzo A, Fruchart JC, Rubin E, Denèfle P, Staels B, Branellec B: **Opposite regulation of human versus mouse apolipoprotein A-I by fibrates in human apolipoprotein A-I transgenic mice.** *J. Clin. Invest.* 1996, **97**:2408-2416
26. Kockx M, Gervois PP, Poulain P, Derudas B, Peters JM, Gonzalez FJ, Princen HMG, Kooistra T, Staels B: **Fibrates suppress fibrinogen gene expression in rodents via activation of the peroxisome proliferator-activator receptor- α .** *Blood* 1999, **93**:2991-2998
27. Bouly M, Masson D, Gross B, Jiang XC, Fievet C, Castro G, Tall AR, Fruchart JC, Staels B, Lagrost L, Luc G: **Induction of the phospholipid transfer protein gene accounts for the high density lipoprotein enlargement in mice treated with fenofibrate.** *J. Biol. Chem.* 2001, **276**:25841-25847
28. Mulvany MJ, Halpern W: **Contractile properties of small arterial resistance vessels in spontaneously hypertensive and normotensive rats.** *Circ. Res.* 1977, **41**:19-26
29. Masiakowski P, Breathnach R, Bloch J, Gannon F, Krust A, Chambon P: **Cloning of cDNA sequences of hormone-regulated genes from the MCF-7 human breast cancer cell line.** *Nucleic Acids Res.* 1982, **10**:7895-7903

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