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Research article

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Syndecan-3 and syndecan-4 are enriched in Schwann cell perinodal processes

Laurence Goutebroze*, Michèle Carnaud, Natalia Denisenko, Marie-Claude Bouterin and Jean-Antoine Girault

Address: INSERM U536, UPMC, Institut du Fer à Moulin, 17 rue du Fer à Moulin, 75005 Paris, France

Email: Laurence Goutebroze* - goutebro@fer-a-moulin.inserm.fr; Michèle Carnaud - mcarnaud@fer-a-moulin.inserm.fr; Natalia Denisenko - nehrbass@fer-a-moulin.inserm.fr; Marie-Claude Bouterin - boutteri@fer-a-moulin.inserm.fr; Jean-Antoine Girault - girault@fer-a-moulin.inserm.fr

* Corresponding author

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Abstract

Background: Nodes of Ranvier correspond to specialized axonal domains where voltage-gated sodium channels are highly concentrated. In the peripheral nervous system, they are covered by Schwann cells microvilli, where three homologous cytoskeletal-associated proteins, ezrin, radixin and moesin (ERM proteins) have been found, to be enriched. These glial processes are thought to play a crucial role in organizing axonal nodal domains during development. However, little is known about the molecules present in Schwann cell processes that could mediate axoglial interactions. The aim of this study is to identify by immunocytochemistry transmembrane proteins enriched in Schwann cells processes that could interact, directly or indirectly, with axonal proteins.

Results: We show that syndecan-3 (S3) and syndecan-4 (S4), two proteoglycans expressed in Schwann cells, are enriched in perinodal processes in rat sciatic nerves. S3 labeling was localized in close vicinity of sodium channels as early as post-natal day 2, and highly concentrated at nodes of Ranvier in the adult. S4 immunoreactivity accumulated at nodes later, and was also prominent in internodal regions of myelinated fibers. Both S3 and S4 were co-localized with ezrin in perinodal processes.

Conclusions: Our data identify S3 and S4 as transmembrane proteins specifically enriched in Schwann cell perinodal processes, and suggest that S3 may be involved in early axoglial interactions during development.

Background

Clustering of voltage-gated Na⁺ channels at nodes of Ranvier is an essential aspect of the fast saltatory propagation of action potential along myelinated axons. The nodes are devoid of myelin and flanked by paranodes where lateral loops of glial cells are tightly attached to the axon by septate-like junctions. Thus, axoglial interactions generate distinct axonal domains characterized by specific multi-

molecular complexes (see [1] for a review). Several proteins are highly concentrated in the nodal axolemma, including Na⁺ channels, comprised of an α subunit, Nav1.2, replaced by Nav1.6 later during development, and a β subunit [2,3]. Nodal Na⁺ channels interact with ankyrin G [4], which is associated with β IV-spectrin [5], and with two cell adhesion molecules, Nr-CAM and the 186 kDa isoform of neurofascin [6]. These nodal axonal

proteins are similar to those found at axonal initial segments and are thought to form a highly interconnected multimolecular meshwork. However, in contrast to initial segments, enrichment of these proteins at nodes of Ranvier is dependent on myelinating glial cells in both central and peripheral nervous systems [7-11]. In the central nervous system where there is no direct contact between oligodendrocytes and the nodal axolemma, a diffusible factor may trigger the clustering of nodal proteins [3,8]. In contrast, in peripheral nerves, the nodal axolemma is covered by microvilli emanating from myelinating Schwann cells, and a direct contact with Schwann cells may be required for node formation [7,10,11]. However, little is known about the molecules present in Schwann cell processes that could mediate the organization of axonal nodal proteins. Ezrin, radixin and moesin (ERM proteins), three homologous proteins that are enriched in microvilli of epithelial cells, are also concentrated in Schwann cell microvilli [12,13]. They co-localize with ezrin-binding protein 50 kDa (EBP-50) [12]. Since clustering of these proteins occurs early during development at the time of the initial accumulation of ankyrin G and Na⁺ channels in the axon, a direct or indirect role of microvilli in the organization of nodal regions is very likely. In support of this hypothesis a recent work has demonstrated that myelinating Schwann cells emit long processes at the tip of which ERM proteins are enriched in a Rho A-dependent fashion [14]. These tips overlap with zones of ankyrin G accumulation in the axon. ERM proteins are regulated cytoplasmic cross-linkers between actin cytoskeleton and transmembrane proteins [15], and cannot mediate direct nodal axoglial contacts. Therefore it is of interest to identify transmembrane proteins enriched in Schwann cell perinodal processes that could interact with axonal proteins, either directly or indirectly.

Syndecans are a family of proteoglycans (syndecan 1-4, here abbreviated as S1-4) containing heparan sulfate and chondroitin sulfate sugar chains [16,17]. They are type I membrane proteins, with an ectodomain containing several glycosaminoglycan attachment sites, a transmembrane domain and a short C-terminal cytoplasmic tail. Their extracellular chains bind to a variety of growth factors and extracellular matrix proteins. Many cytoplasmic partners of syndecans have also been isolated, with scaffolding or signaling properties [18,19]. Here we have investigated the localization of S3, known to be expressed in Schwann cells [20], and S4, an ubiquitous protein, in rat sciatic nerve. Using specific antibodies we found that both are enriched in Schwann cell perinodal processes.

Results

Production of specific antibodies against S3 and S4

To detect the membrane-associated forms of S3 and S4 we raised antibodies against their cytoplasmic domains. As

the cytoplasmic tail of each syndecan encompasses two highly conserved regions, C1 and C2, flanking a variable region (V) unique to each syndecan (Fig. 1A), sera were affinity-purified using another fusion protein encompassing only the variable part of the cytoplasmic region of the cognate protein to avoid cross-reactivity. The specificity of the purified antibodies was tested using GFP-fusion proteins encompassing the intracellular region of the various syndecans and expressed in COS-7 cells (Fig. 1B,1C,1D,1E). Both S3 and S4 antibodies were selective for their cognate antigen by immunoblotting (Fig. 1B,1D) and by immunofluorescence (Fig. 1C, 1E).

Distribution of S3 in sciatic nerve

S3 is the most abundant syndecan in the central and peripheral nervous systems, with a peak of expression in the first postnatal week [21]. In the adult central nervous system it is present along axons [22]. S3 is also expressed in Schwann cells [20] in which it allows adhesion to α 4 Type V collagen [23]. In sections of neonatal rat sciatic nerves, S3 was reported to be present in membranes of Schwann cells and/or neurons [20], but its localization was not investigated at later stages. We studied the distribution of S3 immunoreactivity in sections of adult rat sciatic nerve, in parallel with that of Na⁺ channels (Nav), and *shaker*-type K⁺ channels (Kv1.1) enriched in juxtaparanodal regions [24]. S3 labeling was concentrated in narrow bands corresponding to nodal gaps (Fig. 2A). This localization was almost identical to that of Nav (Fig. 2B). However, in sections perpendicular to the nerve axis, the ring of S3 immunoreactivity was wider than the ring of Nav and appeared to surround it (Fig. 2C). Thus, the two proteins were not co-localized in the axon, indicating the presence of S3 in the Schwann cell perinodal processes. In teased fibers from rat sciatic nerve, a similar distribution was observed (Fig. 2D). In this preparation S3 labeling was clearly located around the nodal axon. In sciatic nerve sections as well as in teased fibers, all nodes that were identifiable by Nav immunoreactivity were stained with anti-S3 antibodies.

Distribution of S4 in sciatic nerve

S4 is ubiquitously expressed and cooperates with integrins to form focal adhesions [18,19]. However, its localization in nerves is not known. In adult rat sciatic nerve, S4 antibodies labeled several structures (Fig. 3). A diffuse and irregular labeling was observed along myelinated fibers at the level of internodes, whereas paranodal regions were consistently immunonegative (Fig. 3A, 3B arrowheads). Although S4 labeling often appeared more pronounced in the vicinity of paranodal regions, suggesting a possible enrichment at juxtaparanodes, it did not co-localize with Kv1.1 (Fig. 3B). S4 immunoreactivity was also concentrated at nodes, identified by Nav co-staining (Fig. 3A). Higher magnification confocal study in sections parallel

A.

		TM	Cytoplasmic region			
			C1	V	C2	
		(aa)				(aa)
Syndecan-1 (S1)	280	RMKKKDEGSY	SLEEPKQANGGAYQK-PTKQE	EFYA	313	
Syndecan-2 (S2)	170	RMRKKDEGSY	DLGERKPSS-AAAYQKAPT---	EFYA	201	
Syndecan-3 (S3)	410	RMKKKDEGSY	<u>TLEEPKQAN-VTYQK-PDKQE</u>	EFYA	442	
Syndecan-4 (S4)	171	RMKKKDEGSY	<u>DLG-KKP----IYKKAPT---</u>	EFYA	198	

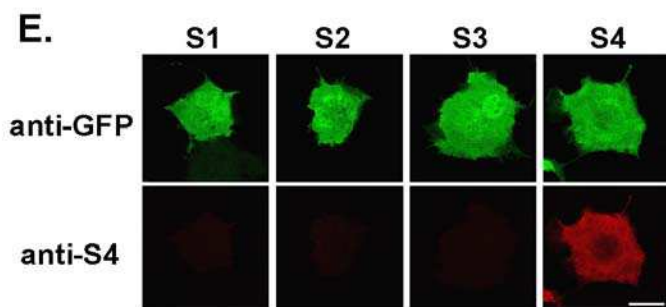
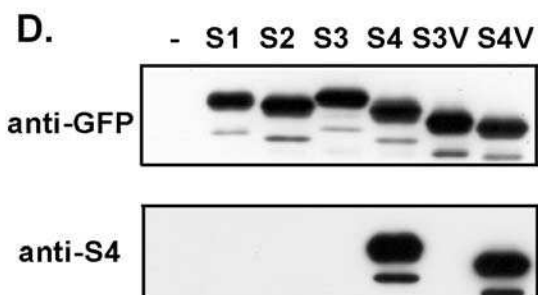
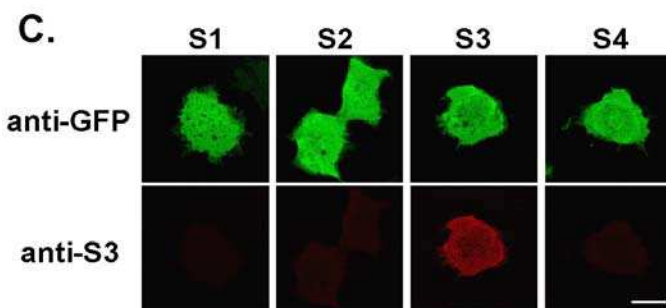
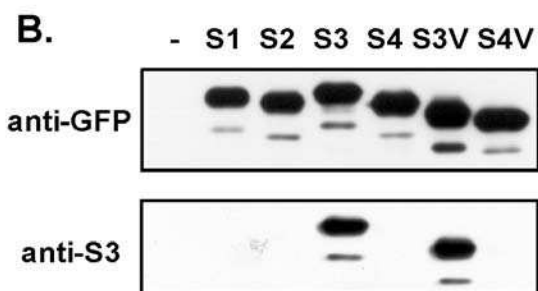


Figure 1
Characterization of antibodies against S3 and S4. (A) Organization in three sub-regions, C1, C2 and V (upper panel), and amino acid alignment of the cytoplasmic C-terminal regions of human S1–S4. Antibodies were raised against the entire cytoplasmic tails of S3 or S4, and purified by affinity on peptides encompassing only the variable (V) regions (underlined peptide sequences). Dashes represent gaps in the sequence introduced to optimize alignment. The position of the residues in the entire sequence of the proteins is indicated (aa). TM, transmembrane region. (B–E) Specificity of S3 and S4 antibodies was tested by immunoblotting (B, D) and immunocytochemistry (C, E) using COS-7 cells transfected with GFP-fusion proteins expressing either the intracellular regions of S1 to S4 (B–E, S1–S4) or only the variable regions of S3 and S4 (B and D, S3V and S4V). Antibodies directed against GFP (anti-GFP) were used to verify the expression of the proteins. COS-7 cells transfected with the vector alone were used as mock control in immunoblotting experiments (-). Anti-S3 and anti-S4 antibodies recognized specifically GFP-fusion proteins encompassing S3 and S4 intracellular region, respectively. Scale bars: 20 μm.

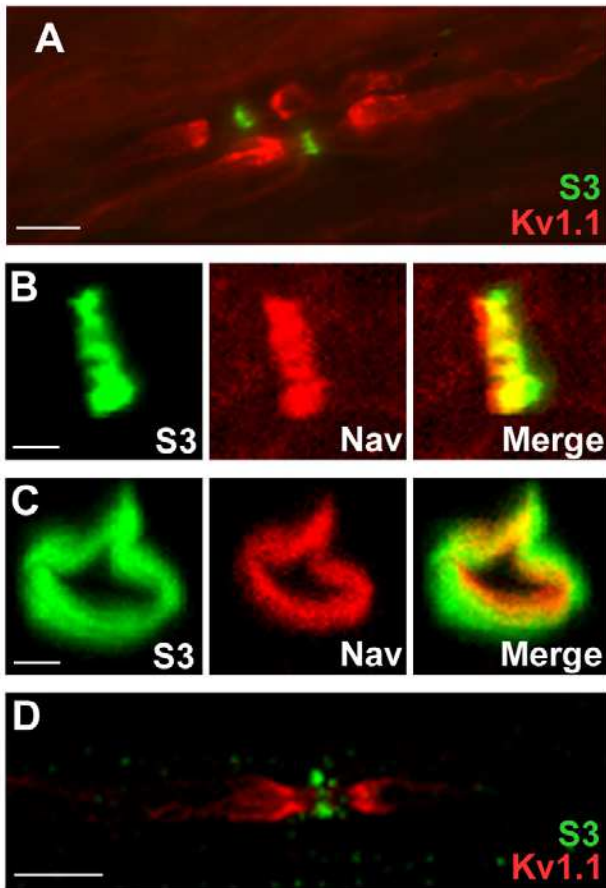


Figure 2
Localization of S3 in nodal regions of the PNS. (A-C) Sections through rat sciatic nerve at P30 were labeled with antibodies to S3 (green) and K⁺ channels (Kv1.1, A, red) or Na⁺ channels (Nav, B-C, red). Superposition of the two labels is shown (merge). Note that on sections perpendicular to nerve axis, S3 immunoreactivity is peripheral to Nav labeling. **(D)** Teased fiber from rat sciatic nerve stained for S3 (green) and Kv1.1 (red) showing that S3 immunoreactivity is on both sides of the node. B-D: confocal stacks. Scale bars: 10 μm (A, D), 1 μm (B, C).

(Fig. 3C) or perpendicular (Fig. 3D) to the nerve axis demonstrated that nodal S4 immunofluorescence was located peripherally to Na⁺ channels, indicating its presence in the Schwann cells perinodal processes and not in the axon. Although S4 nodal labeling was less conspicuous than that of S3, due to the immunoreactivity of myelinated fibers, careful examination of the sections showed that all nodes, identified by Nav staining, were also labeled by S4 antibodies.

Colocalization of S3 and S4 with ezrin

We compared the distribution of S3 and S4 to that of ezrin in adult rat sciatic nerves. As previously described, ezrin immunoreactivity was concentrated in perinodal processes (Fig. 4A), the position of which was easily identified by double labeling with antibodies to paranodin/Caspr, a marker of paranodal regions [25]. Co-immunostaining revealed a complete colocalization of ezrin and S3 (Fig. 4B) or S4 (Fig. 4C). These results strongly indicate that S3 and S4 are co-localized with ezrin in the microvilli of Schwann cells.

Distribution of S3 and S4 in sciatic nerve during development

Aggregates of nodal proteins start to appear early during myelination of peripheral nerves [11]. We examined the distribution of S3 and S4 with respect to the localization of Nav during the development of sciatic nerve in rat pups, from post-natal days 2 to 21 (P2 to P21, Fig. 5). At P2, Nav clusters were readily detectable (Fig. 5A.a, arrows). In the same sections, spots of concentrated S3 immunoreactivity were clearly distinguishable from the labeling of the rest of the fibers (Fig. 5A.b, arrows). However, S3 and Nav labeling did not exactly overlap, but appeared to be in very close contact with each other (Fig. 5A.a-c, insets). At P6, as the number of presumptive nodes increased, S3 and Nav immunoreactivities were always closely associated (Fig. 5A.d-f). This pattern remained similar at later stages, as nodal aggregates enlarged and acquired their mature form (Fig. 5A.g-l). When pairs of heminodes were observed [11], both were stained for S3 and Nav (Fig. 5A.g-i, arrowheads). As development proceeded, the immunoreactivity along nerve fibers decreased, and at p21 S3 staining appeared virtually restricted to the nodes of Ranvier (Fig. 5A.j-l), as in the adult (Fig. 2).

The developmental pattern of S4 immunoreactivity was also examined in rat sciatic nerves. At P2 although some S4 labeling was visible, with a few cells more intensely stained, no overlap of labeling with Nav was observed and there was no indication of the accumulation of S4 at presumptive nodes (Fig. 5Ba-c). Some nodal labeling was visible at P6 (Fig. 5B.d-f), and became more frequent at P12 and P21 (Fig. 5B.g-l). A general increase of S4 immunoreactivity in internodal regions of myelinated fibers occurred during development (Fig. 5B), leading to a pattern similar to that observed in adult rats (Fig. 3).

Discussion

Our results identify two proteoglycans, S3 and S4, as novel components of nodal regions in rat sciatic nerve. There are similarities between the distributions of the two proteins. Both S3 and S4 were found concentrated in Schwann cells perinodal processes, co-localized with

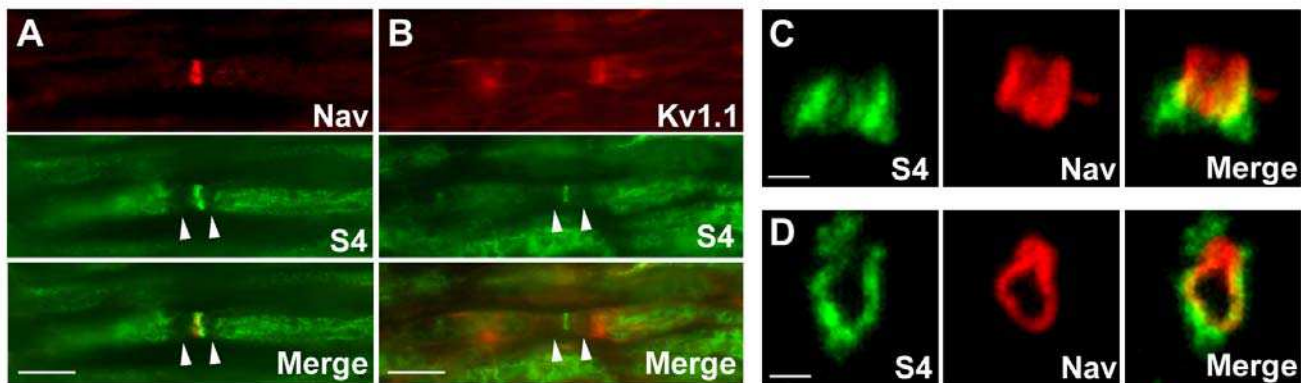


Figure 3

Localization of S4 in nodal regions of the PNS. Sections through rat sciatic nerve at p30 were labeled with antibodies to S4 (A-D, green) and Na⁺ (Nav, A, C-D, red) or K⁺ (Kv1.1, B, red) channels. Superposition of the labels is shown (merge). The paranodal regions are not labeled with antibodies against S4 (A, B, arrowheads). On sections perpendicular to nerve axis (D), S4 immunoreactivity is peripheral to Nav labeling. C, D: confocal stacks. Scale bars: 10 μm (A, B), 1 μm (C, D).

ezrin, surrounding the sodium channel clusters. S3 and S4 are therefore likely to be both localized in Schwann cells microvilli. There were also marked differences between the distributions of the two proteins. S3 was detected in the vicinity of sodium channels aggregates very early in development, at a time when ezrin is also enriched in Schwann cells processes [12,14], whereas accumulation of S4 near sodium channels occurred progressively during the first postnatal weeks. In adult nerves S3 was highly concentrated at nodes as compared with any other cellular structures in the sciatic nerve. In contrast, S4 was also detected in the internodal region of myelinated fibers. These results show that at least two proteoglycans of the syndecan family are transmembrane components of Schwann cells microvilli. Since these molecules are known to have a number of intracellular and extracellular partners, it is likely that they play similar and perhaps complementary roles in the structural organization of nodal regions, as well as in signal transduction in glial processes.

Syndecans are thought to play a role in the regulation of cytoskeleton organization (for a recent review see [26]). S2 can regulate the actin cytoskeleton in different cell types, and S4 is present in focal adhesions of a number of cell lines [18]. S1 ectopically expressed in primary Schwann cells transiently co-localizes with actin filaments during cell spreading [27]. In the same transfected cells, S1 crossed-linked with anti-S1 antibodies co-localizes with actin filaments [28]. This localization is dependent on a specific region of the cytoplasmic region of the protein [29]. Several studies have suggested that signaling

through the core protein of syndecans could regulate cytoskeleton organization through their clustering, binding to cytoplasmic proteins, and regulation of intracellular protein phosphorylation (for review see [26]). In this context, the ERM proteins may function as adaptors that couple syndecans to cytoskeletal proteins since S2 has been shown to interact with ezrin through a motif conserved in the others syndecans [30,31]. All these observations raise the interesting possibility that both S3 and S4 might provide plasma membrane anchors for ERM proteins in Schwann cells perinodal processes [12-14], and therefore be indirectly linked to actin microfilaments that are also enriched in microvilli [32]. In addition, the conserved C-terminal motif of syndecans can interact with PDZ (PSD-95 / discs large / ZO-1) domain-containing scaffold proteins syntenin [33], CASK/LIN [34,35], and synectin [36]. These interactions may coordinate clustering of receptors and also connection to the actin cytoskeleton [18,19]. It is not known whether syntenin, synectin or CASK/LIN are enriched in nodes of Ranvier, but it seems likely that these or other PDZ-containing proteins could be involved in the organization of perinodal glial processes.

S3 heparan sulphate chains bind to fibroblast growth factor-2 (FGF-2) [37] and heparin-binding growth-associated molecule (HB-GAM) also known as pleiotrophin or midkine [38]. FGF-2 and its high affinity receptor [39], as well as HB-GAM [40], are expressed by Schwann cells. In neuroblastoma cells S3 can activate neurite outgrowth in response to HB-GAM through a cortactin/src kinase-mediated pathway [41]. This raises the possibility that S3 might

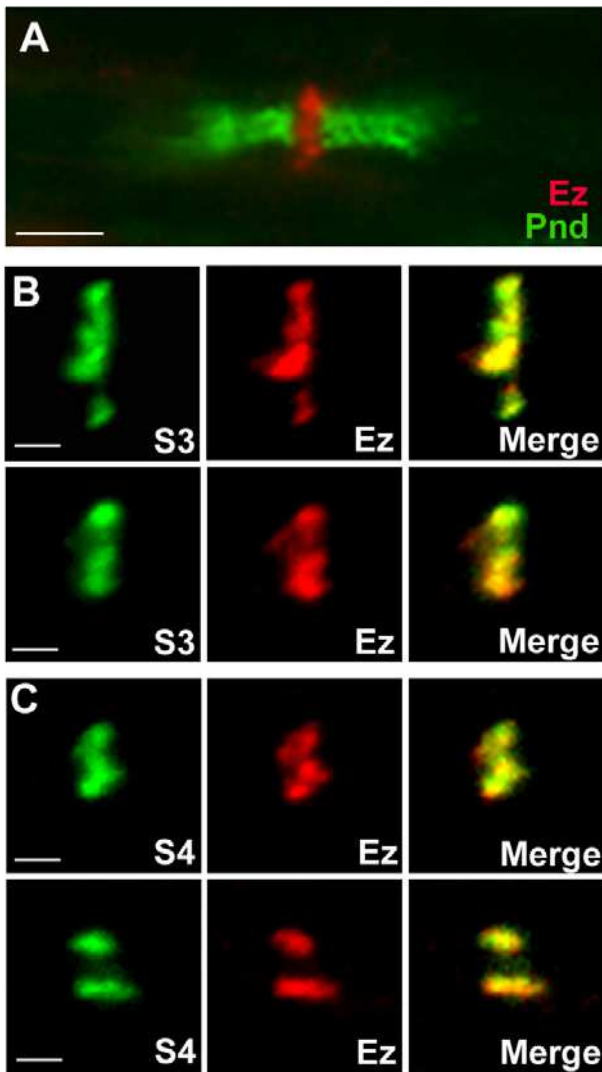


Figure 4
S3 and S4 co-localize with ezrin in nodal regions of PNS. (A) Sections through rat sciatic nerve at p30 were labeled with antibodies to ezrin (Ez, red) and to paranodin/Caspr (Pnd, green), a marker of paranodal regions. (B, C) Sections were double labeled with antibodies to S3 (B, green) or S4 (C, green) and antibodies to ezrin (Ez, red). Superposition of syndecan and ezrin labelings is shown (merge). Two different nodes are shown in each case. B, C: confocal stacks. Scale bars: 4 μ m (A), 1 μ m (B, C).

be involved in the formation and growth of Schwann cells nodal processes [14]. On the other hand, S4 binds to fibronectin and drives the formation of focal adhesions [42], while interaction of S4 with tenascin-C decreases adhesion [43]. A similar role of S4 could be important in the nodal area where loose axoglial contacts contrast with

the tight septate-like junctions of neighbouring paranodes. S4 plays also a role in signaling through its cytoplasmic tail that binds and activates the catalytic domain of PKC α in the presence of phosphatidylinositol-4,5-bisphosphate [44,45]. Moreover, S4 regulates signaling by Rho family GTPases and focal adhesion kinase [46], and is able to bind to α -actinin [47]. All these interactions that have been described in the context of focal adhesions, may also have functional relevance for the organization of cortical cytoskeleton in Schwann cells microvilli.

Whether S3 and S4 could interact with axonal membrane proteins and contribute to the organization of the nodal region remains to be determined. Although S3 and S4 null mutant mice display relatively mild phenotypes [48-50], suggesting some redundancy between proteoglycans, their peripheral nerves have not been studied. It will be interesting to examine carefully the axoglial interactions and the molecular organization at nodes of Ranvier in these mice, as well as in double mutant mice.

Conclusions

In summary our findings show the enrichment of two syndecans in the perinodal processes of Schwann cells. These proteoglycans are receptors for extracellular proteins and growth factors. Syndecans are able to organize scaffolding proteins and activate signaling pathways involved in the reorganization of actin cytoskeleton, through tyrosine phosphorylation and activation of Rho family GTPases. Thus, they may be pivotal in the organization of Schwann cells growing tips and microvilli that seem to play an important role in the early interaction with nodal axonal proteins [14]. Since S3 accumulates earlier than S4, it is a candidate for being involved in the earliest steps of nodes of Ranvier formation.

Methods

Antibodies and constructs

Monoclonal antibodies sources were as follows: voltage-gated Na⁺ channel α subunit (PAN Nav, clone K58/35) [51] and ezrin (clone 3C12) [52], Sigma; Kv1.1 (clone K20/78) [53], Upstate Biotechnology; green fluorescent protein (GFP), Roche. Polyclonal antiserum against paranodin/Caspr (L-51) was described previously [25]. Alexa Fluor 488- and 594-conjugated goat anti-rabbit and anti-mouse antibodies were from Molecular Probes (Leiden, Netherlands). HRP-conjugated goat anti-mouse and anti-rabbit antibodies were from Amersham. Antibodies against S3 and S4 were generated by immunizing rabbits with the intracellular region of the human proteins (S3: residues 410-442; S4: residues 171-198, Fig. 1) fused to glutathione-S-transferase (GST), and affinity-purified on GST-fusion proteins encompassing only the variable region of the respective syndecans (S3: residues 420-439; S4: residues 181-194). Constructs for GST- and GFP-

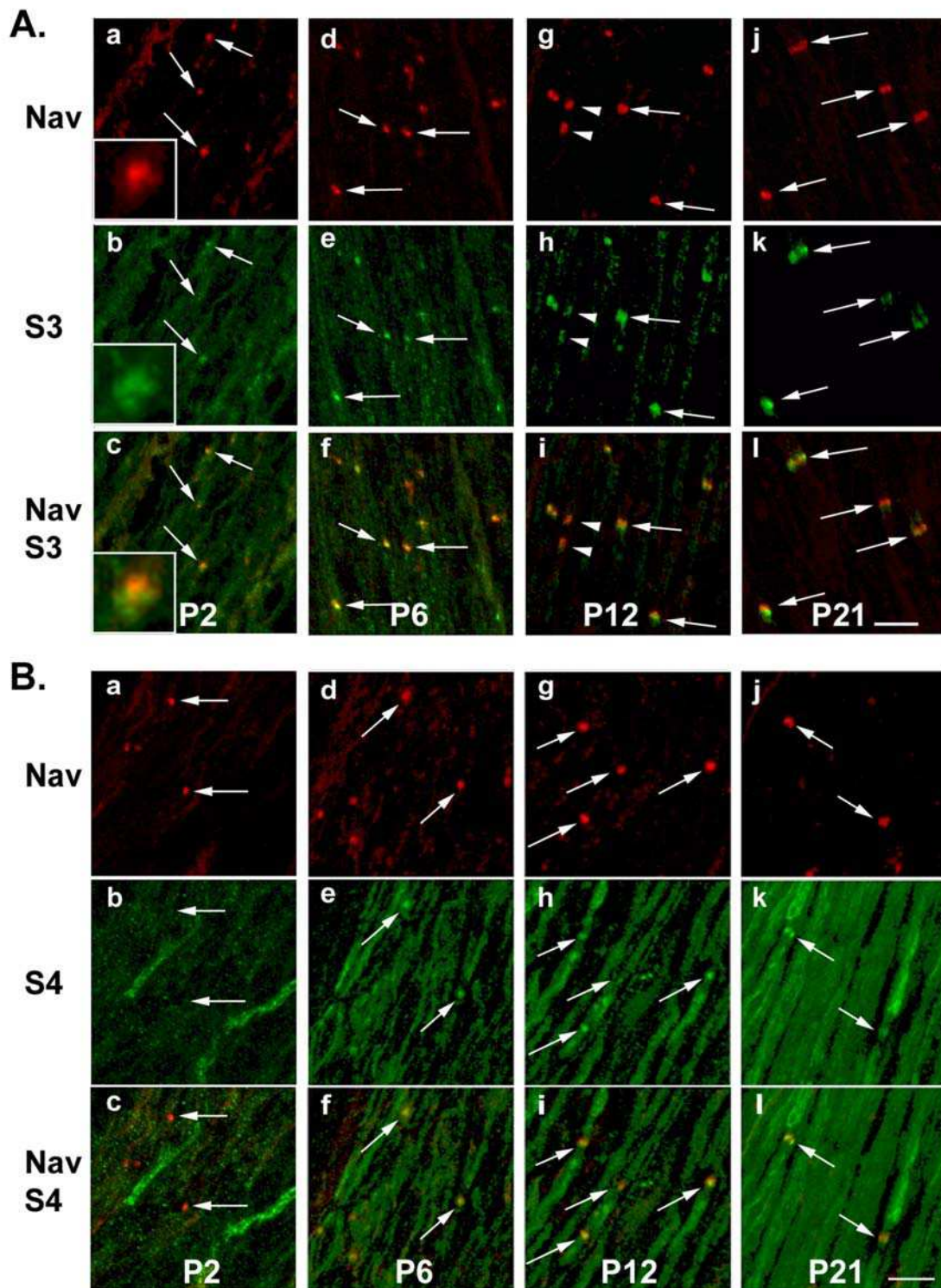


Figure 5
Developmental profile of S3 and S4 in rat sciatic nerve. Sections through rat sciatic nerve at P2, P6, P12, and P21 were labeled with antibodies to S3 (A, green) or S4 (B, green) and Na⁺ channels (Nav, A, B, red). Superposition of the two labels is indicated by arrows. At P2, S3 and Nav labelings appear to be in very close contact with each other (A.a-c, insets). For S3, a pair of heminodes is indicated by arrowheads (A.g-l). A, B: confocal stacks. Scale bars: 10 μm.

fusion proteins were generated by RT-PCR, verified by sequencing, and expressed using pGEX (Pharmacia) and pEGFP vectors (Clontech), respectively. For S1 and S2, the sequences fused to GFP encompassed residues 280–313 and 170–201 of rat S1 and human S2, respectively.

Cell culture, immunoblotting and immunocytochemistry

COS-7 cells transfection and immunoblotting were done as described [54]. For immunocytochemistry, cells were fixed for 20 min in 4% PFA/PBS, permeabilized with 0.2% Triton X-100 in PBS for 10 min, and incubated sequentially with primary and secondary antibodies for 30 min at room temperature. Immunostaining of cryostat sections (10 μ m) of sciatic nerve tissues was performed essentially as previously described [9]. Sections were fixed for 20 min in methanol/acetone (50/50 vol/vol) at -20°C (for labeling with Nav antibodies) or in 4% PFA/PBS at room temperature (for labeling with Kv1.1, ezrin and paranodin antibodies), pre-incubated for 1 hour at room temperature in 0.2% porcine skin gelatin and 0.25 % Triton X-100 in PBS (PGT buffer), before incubation with primary antibodies (diluted in PGT) overnight at 4°C. After washing with PBS, coverlips were incubated for 2 hours at room temperature with secondary antibodies (diluted in PGT), washed again with PBS, and mounted in Vectashield. Images were acquired using a Leica epifluorescence microscope equipped with a CCD camera or a Leica SP laser Scanning microscope. Teased fibers were prepared from sciatic nerve as described [55]. Briefly, twenty-one days-old rats were perfused with 2% PFA in phosphate buffer (PB) at 4°C. Nerves were dissected out, post-fixed with 2% PFA in PB for 30 minutes at 4°C, and washed 1 hour in PB, before teasing and spreading on slides. Immunostaining was performed as for cryostat sections, except that slides were blocked and incubated with antibodies in PB, 0.2% porcine skin gelatin, 0.25 % Triton X-100 and 10% goat serum. Antibodies dilutions were: anti-S3 (1:300); anti-S4 (1:100); anti-Nav (1:100); anti-Kv1.1 (1:200); anti-paranodin (1:1000); anti-ezrin (1:400); anti-GFP (1:200); secondary antibodies (1:400).

Abbreviations

S: syndecan

ERM: ezrin, radixin, moesin

Nav: voltage-gated sodium channel α subunit

Kv: *shaker*-type K⁺ channels

GST: glutathione-S-transferase

GFP: green fluorescent protein

PDZ: PSD-95 / discs large / ZO-1

Authors' contributions

LG conceived of the study, produced, purified and characterized the antibodies, performed image analysis; and participated in the redaction of the manuscript. MC performed immunocytochemistry experiments on sciatic nerves. ND carried out immunocytochemistry on teased fibers. MCB participated in the purification of the antibodies. JAG participated in the design and coordination of the study, and drafted the manuscript. All authors read and approved the manuscript.

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