

Transforming growth factor- β released by apoptotic white blood cells during red blood cell storage promotes transfusion-induced alloimmunomodulation.

Romain Vallion, Francis Bonnefoy, Anna Daoui, Loredane Vieille, Pierre Tiberghien, Philippe Saas, Sylvain Perruche

► **To cite this version:**

Romain Vallion, Francis Bonnefoy, Anna Daoui, Loredane Vieille, Pierre Tiberghien, et al.. Transforming growth factor- β released by apoptotic white blood cells during red blood cell storage promotes transfusion-induced alloimmunomodulation.: Apoptotic WBC limit RBC alloimmunization. Transfusion, Wiley, 2015, [Epub ahead of print]. <inserm-01146033>

HAL Id: inserm-01146033

<http://www.hal.inserm.fr/inserm-01146033>

Submitted on 27 Apr 2015

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **TGF- β released by apoptotic white blood cells during red blood cell storage promotes**
2 **transfusion-induced alloimmunomodulation**

3 Romain Vallion^{1,2,3,4}, Francis Bonnefoy^{1,2,3,4}, Anna Daoui^{1,2,3,4}, Loredane Vieille^{1,2,3,4}, Pierre
4 Tiberghien^{1,2,3,4}, Philippe Saas^{1,2,3,4} and Sylvain Perruche^{1,2,3,4}

5 ¹INSERM, UMR1098, Besançon, France

6 ²Université de Bourgogne Franche-Comté, UMR1098, Besançon, France

7 ³EFS Bourgogne Franche-Comté, UMR1098, Besançon, France

8 ⁴LabEx LipSTIC, ANR-11-LABX-0021, FHU INCREASE, Besançon, France

9

10 Correspondence to Sylvain Perruche, INSERM UMR1098, 8 rue du Dr Girod, F-25020

11 Besançon, France; Tel: +33.3.81.615.615, Fax +33.3.81.615.800; sylvain.perruche@inserm.fr

12 Reprints will not be available from the author

13

14 Source of support: EFS (#2010-07 and #2011-05 to SP), *Association Recherche et Transfusion*

15 (#53-2009 to SP), *Agence Nationale de la Recherche* (Labex LipSTIC, ANR-11-LABX-0021),

16 *Conseil Régional de Franche-Comté* (“soutien au LabEX LipSTIC” 2013/2014 to PS).

17

18 The authors declare that they have no conflicts of interest

19

20 Word count: 3972

21

22 Running head: Apoptotic WBC limit RBC alloimmunization

1 **Abstract**

2 BACKGROUND: Red blood cell alloimmunization is a major immunological risk of transfusion.
3 However, RBC storage facilitates WBC apoptosis and apoptotic cells have immunomodulatory
4 properties. We investigated the behavior of WBCs, and apoptosis in particular, in RBC units
5 during storage, and then studied the impact of WBC-apoptosis on the modulation of post-
6 transfusion alloimmunization in RBC products stored short term. STUDY DESIGN AND
7 METHODS: We used a mouse model of alloimmunization to transfused HOD surface antigen
8 expressed specifically on RBCs. The presence of circulating anti-HOD IgG detected by FACS
9 confirmed immunization to HOD⁺ RBCs. WBC apoptosis and factors released by apoptotic
10 WBC during storage were determined and in particular the role of TGF- β was assessed on RBC
11 alloimmunization. RESULTS: In blood stored 72 hours, 30% of WBCs were apoptotic, and
12 transfusion of short-term stored blood resulted in lesser immunization than did fresh blood or
13 stored leukoreduced RBCs (LR RBCs). WBC undergoing apoptosis released during short term
14 storage factors modulating RBC alloimmunization. Indeed apoptotic cell-released factors
15 modulate alloimmunization whereas exogenous apoptotic cells directly transfused with LR RBC
16 did not. While microparticles released during RBC storage had no immunomodulatory role,
17 TGF- β found in the supernatant of stored blood demonstrated the capacity to favor Treg
18 polarization of naïve CD4⁺CD25⁻ T cells *in vitro* and limited RBC alloimmunization *in vivo*.
19 Indeed, addition of recombinant TGF- β to stored LR RBC transfusion strongly limited post-
20 transfusion RBC alloimmunization. CONCLUSION: Our findings show that short-term storage
21 of non-leukoreduced blood facilitates WBC apoptosis therefore releasing TGF- β that modulates
22 post-transfusion RBC alloimmunization.

23 **Key words:** apoptotic WBC, RBC, alloimmunization, transfusion, TGF- β

24

1 INTRODUCTION

2 Post-transfusion alloimmunization is the chief immune-mediated adverse effect of transfusion.¹
3 Alloantibodies to RBC antigens are responsible for transfusion-related hemolytic reactions with
4 potentially lethal consequences, transfusion failure when compatible RBC units cannot be found,
5 and hemolytic disease of the newborn in women who were immunized prior to pregnancy.
6 Alloimmunization to RBC occurs in <10% of recipients and can be triggered by immunogenetic
7 predispositions² and other factors such as the inflammatory status of the recipient at the time of
8 transfusion. The number (<40) or the intensity of transfusions do not seem to increase the risk of
9 RBC alloimmunization.³ Storage of RBC units and leukoreduction may however, have a
10 significant impact on RBC alloimmunization. Indeed, experimental rodent models have shown
11 that immunogenicity of RBC units increases with storage time⁴. In Humans, blood storage has
12 been associated with increased inflammatory cytokine levels (TNF and IL-6) and higher priming
13 capacities of neutrophils (PMN).⁵⁻⁷ Leukoreduction reduced PMN activation by RBC
14 supernatants, activation which increases with prolonged storage,⁷ suggesting that WBC make
15 stored RBCs immunogenic. This has been indirectly addressed in humans in a study that
16 investigated the rate of alloimmunization to RhD –the most immunogenic RBC antigen– during
17 different storage lengths.⁸ The authors were unable to associate anti-D alloimmunization with
18 RBC unit storage duration.⁸
19 RBC integrity can also be affected during storage,⁹⁻¹¹ notably through membrane-encapsulated
20 hemoglobin iron that promotes immunogenicity of RBCs as demonstrated in an experimental
21 model.¹² Interestingly, supernatants from stored RBCs did not produce the same inflammatory
22 signal *in vivo* as stored RBCs.¹² Of note, in that study, RBC units were leukoreduced.¹² WBCs
23 may also dramatically affect RBC alloimmunization. Indeed, WBC apoptosis during storage has

1 been described¹³ and apoptotic cells are known to have immunomodulatory properties.¹⁴⁻¹⁶ Some
2 authors have speculated that in addition to soluble factors,^{17,18} apoptotic WBCs may be essential
3 to transfusion-related immunomodulation.¹⁹

4 To address the role of WBCs in RBC alloimmunization, we first studied WBC behavior during
5 storage in RBC units, notably apoptosis, and then the impact of apoptotic WBCs on RBC
6 alloimmunization using an experimental mouse model of post-transfusion alloimmunization.

7 8 **MATERIALS AND METHODS**

9 **Mice**

10 C57Bl/6, BALB/c and FVB mice were purchased from Charles River laboratories (L'Arbresle,
11 France). Membrane-bound hen egg lysozyme (mHEL) C57Bl/6 and HEL-ovalbumin-duffy
12 (HOD) FvB transgenic mice were kindly provided by Dr J Zimring (Puget Sound Blood Center
13 Research Institute, Seattle, WA, USA) and bred in our specific pathogen-free rodent facility
14 (agreement #D25-056-7) with food and water available *ad libitum*. Experimentation (#10004R)
15 was approved by the local ethics committee (#58, approved by the French Ministry of Higher
16 Education and Research [*Ministère de l'Enseignement Supérieur et de la Recherche*]) and
17 conducted in accordance with the European Union's Directive 2010/63.

18 **RBC preparation and transfusion**

19 Blood was collected by retro orbital bleeding into ethylenediaminetetra acetic acid (EDTA) or
20 acid citrate dextrose (ACD) tubes (BD Biosciences) and leukoreduced (=LR RBC) or washed 3
21 times (=blood) to reduce platelet count (15 min, 300 g): $628 \times 10^6 \pm 189 \times 10^6$ of CD61⁺
22 platelets/mL in whole blood, $4.3 \times 10^6 \pm 0.4 \times 10^6$ platelets /mL in washed blood ($P < 0.05$ vs

1 whole blood), $2.7 \times 10^6 \pm 0.2 \times 10^6$ platelets /mL \pm in LR RBCs ($P < 0.05$ vs whole blood; $P < 0.05$
2 vs washed blood); n=3 independent experiments pooled together; Student's unpaired *t* test.

3 ~~Washed blood was then leukoreduced (=RBCs) or not (=non-LR blood or blood)~~ Leukoreduction
4 ~~was performed~~ using a RN1 filter (Haemonatics, Plaisir, France) ~~and included 3 washes times~~ to
5 remove platelets and plasma ~~and resuspended in PBS or saline-adenine-glucose-manitol (SAGM)~~
6 ~~solution, and used or stored (+4°C)~~. Leukoreduction was quantified by FACS (CantoII, BD
7 Biosciences, Le Pont de Claix, France) using Trucount tubes (BD Biosciences), as previously
8 described.²⁰ WBC counts were greatly decreased from $6.3 \times 10^6 \pm 0.3 \times 10^6$ to $7.0 \times 10^3 \pm 4.5 \times$
9 10^3 WBC/mL (n=4; $P < 0.001$, Student's paired *t*-test). Blood or LR RBCs (100 μ L) ~~were~~
10 ~~adjusted for RBC number and volume~~ and then transfused *via* the lateral tail vein (500 μ L in
11 PBS or SAGM). Hematocrits were controlled using micro hematocrit tubes (Brand GMBH,
12 Wertheim, Germany). Mice received 100 μ g of poly(I:C) ip (Amersham Biosciences,
13 Piscataway, NJ, USA) diluted in PBS, 4 h before transfusion as priming, a prerequisite for high
14 alloantibody titers. To mimic bedside leukoreduction, stored blood supernatant was collected
15 before leukoreduction to re-suspend stored RBCs in their own supernatant. Apoptosis and
16 membrane-bound TGF- β expression (using latency-associated peptide [LAP] antibody, clone
17 TW7-16B4, BD Biosciences) were evaluated in RBC units by FACS using APC-conjugated
18 CD45_{LCA} antibody, PE-conjugated Annexin-V and 7-AAD (BD Biosciences) according to the
19 manufacturer's procedure, on gated CD45_{LCA}⁺ cells, CD45_{LCA}⁺GR1⁺CD11b⁺ neutrophils,
20 CD45_{LCA}⁺GR1⁻CD11b⁺ monocytes/macrophages, CD45_{LCA}⁺CD19⁺ B cells and CD45_{LCA}⁺CD3⁺
21 T cells. HOD⁺ RBC survival was assessed in peripheral blood at different time-points post-
22 transfusion, by HOD⁺ RBC detection by FACS ~~cross-match using plasma from mice immunized~~
23 ~~to HEL protein, followed by APC-conjugated goat anti-mouse IgG Ab staining.~~

1 **Alloimmunization evaluation**

2 Circulating anti-HOD IgG levels were quantified by FACS in the plasma of mice two weeks
3 after transfusion. Plasma was diluted 1:4 in PBS and incubated with HOD⁺ RBCs for 30 min,
4 then washed and incubated with conjugated secondary anti-mouse IgG APC (SouthernBiotech,
5 Birmingham, Alabama, USA). Finally, APC-intensity was evaluated by FACS on gated RBCs
6 confirming the presence of anti-HOD Ab. Mean fluorescence intensity (MFI) ratio (MFI R)
7 corresponded to MFI from sample/mean MFI from controls (transfused with vehicle). Anti-HOD
8 Ab-positive plasma from mice immunized with HEL and ovalbumin proteins in complete
9 Freund's adjuvant were used as crossmatch positive controls.

10 **Induction of apoptotic leukocytes**

11 WBCs or thymic cells were subjected to a 35-Gray X-ray radiation (Raycell blood irradiator,
12 Best Theratronic Ltd, Ottawa, Canada) and cultured 6 h in complete DMEM (10⁶ cells/mL) to
13 allow apoptosis.²¹ Apoptosis was confirmed by FACS using Annexin-V staining (BD
14 Biosciences) and 7-AAD exclusion. Cells were considered as apoptotic when positive for
15 Annexin-V and negative for 7-AAD staining. Annexin-V and 7-AAD positive cells were
16 considered as necrotic. Apoptotic cell used were early-stage apoptotic (70-85% Annexin-V⁺7-
17 AAD⁻ cells and <10% 7-AAD⁺ cells). Apoptotic cells were washed in PBS and injected (5.10e6
18 cells/mouse) with LR RBCs, stored for 72 h at 4°C and injected with LR RBCs, or stored for 72
19 h at 4°C to collect supernatant and inject it with LR RBCs.

20 **Cell cultures with RBC supernatants**

21 Spleen cells or 90% enriched CD4⁺CD25⁻ T cells (using Miltenyi Biotec isolation kit) were
22 cultured (10⁶ cells/mL) for 72 h in complete DMEM with soluble anti-CD3ε mAb (0.5 μg/mL;
23 clone 145-2C11; eBioscience) or plate-bound anti-CD3ε mAb (5 μg/mL) and soluble anti-CD28

1 mAb (2 µg/mL; clone 37.51; eBioscience), respectively. Supernatant from fresh or stored LR
2 RBCs or blood were added to the culture (20% final volume). Recombinant TGF-β1 was used as
3 a Foxp3 induction-positive control (5 ng/mL; R&D Systems, Minneapolis, MN, USA). After
4 culture, cells were stained for CD4 and CD25 molecules and intracellular Foxp3 transcription
5 factor, according to the manufacturer's instructions (eBioscience), and analyzed by FACS.

6 **TGF-β measurement and blockage**

7 Total and/or latent forms of TGF-β were quantified in RBC unit supernatants by ELISA
8 following the manufacturer's instructions (Promega, Madison, WI, USA; Biolegend, Ozyme,
9 Saint-Quentin-en-Yvelines, France). Anti-TGF-β monoclonal mAb (clone 2G7) was used (50
10 µg/mL) to neutralize TGF-β in stored RBC 30 min before transfusion. TGF-β-depletion in RBC
11 units was performed using anti-TGF-β (clone 2G7) mAb coated on goat anti-mouse IgG BioMag
12 beads (50 µg of mAb + 0.5 mL beads for 1 mL of RBC unit) according to the manufacturer's
13 instructions (Polysciences, Warrington, PA, USA).

14 **Statistics**

15 Group comparisons of continuous data were made using Student's *t*-test or the Mann-Whitney
16 Rank Sum test. Statistical analyses were performed using GraphPad Prism 5 software (GraphPad
17 software, San Diego, CA, USA). *P*-values <0.05 were considered significant.

18

19 **RESULTS**

20 **WBC underwent apoptosis in RBC units upon storage**

21 Storage of RBC units triggers WBC apoptosis and/or necrosis.¹³ WBC-behavior was evaluated
22 post-puncture in blood stored at +4°C in PBS or SAGM for 14 days. Apoptosis of WBCs,

1 hematocrits, *in vivo* survival and ultimately alloimmunization were assessed at different time-
2 points. WBC numbers decreased progressively from Day 0 to Day 14, when the number of
3 WBCs remaining was less than 10^6 cells/mL (Fig. 1A). This was around 20% of the initial
4 number of WBCs in fresh blood. Soon after storage, WBCs began to undergo apoptosis and this
5 proportion (around 20 to 30% of WBC) remained constant over the storage-time (Fig. 1A,B). In
6 contrast, we observed that WBCs slowly underwent necrosis after storage but at around Day 6-8,
7 necrotic cell death became more marked until Day 14 when necrotic WBCs represented approx.
8 60% of all WBCs (Fig. 1A). WBC apoptosis or necrosis was not influenced by the medium used
9 for storage (PBS vs SAGM) (Fig. 1C), nor by the anticoagulant solution used (EDTA vs ACD)
10 (Fig. 1D). Hematocrits were found to be slightly unchanged throughout storage time and
11 independently of the anticoagulant solution used (Fig. 1E). After 3 days of storage, SAGM
12 seemed to better protect RBCs from hemolysis; this difference was not observable at Day 14.
13 Hemolysis was associated with our 3 washes step procedure (data not shown). Survival of RBCs
14 was not found to be significantly different according to the storage medium used; although
15 storage itself seemed to affect *in vivo* RBC survival (Fig. 1F). Since WBC apoptosis was not
16 influenced by experimental settings at Day 3 of storage, we then addressed how WBC apoptosis
17 influenced RBC alloimmunization.

18

19 **Presence of apoptotic WBCs influenced post-transfusion alloimmunization**

20 The presence of apoptotic WBCs in RBC units presumably decreases immunogenicity because
21 apoptotic cells have immunomodulatory properties.^{15,16} Therefore, in an experimental model of
22 post-transfusion alloimmunization, we investigated whether the presence of apoptotic WBCs
23 affects RBC alloimmunization. We compared transfusion of fresh blood with that of blood stored

1 for 72 h since this time point presented the best apoptotic/necrotic cell ratio (Fig. 1A,B) since at
2 day 3 of storage, stored blood products exhibit apoptotic WBCs (Fig. 1A,B). RBC stored for 72
3 hours is roughly equivalent to storage of human RBCs for 6-7 days.²² WBCs are extremely
4 sensitive to cold temperatures and rapidly undergo apoptosis with DNA laddering during routine
5 72-h storage at 4°C.²³ We used blood from HOD transgenic mice –only RBCs express
6 membrane-bound HOD– that was either leukoreduced, stored for 72 h, or both.²⁴ Post-
7 transfusion alloimmunization was evaluated two weeks post-transfusion through the detection of
8 anti-HOD alloantibodies. First, we observed that unprimed mice transfused with fresh blood
9 demonstrated stronger alloimmunization than mice receiving fresh LR RBCs ($P<0.01$; Fig.
10 2A,B), and this was also manifest in poly(I:C)-primed transfused mice ($P<0.01$; Fig. 2C,D). This
11 suggested that leukoreduction may have decreased immunogenicity of fresh RBCs by removing
12 immunogenic WBCs. In contrast, when blood and LR RBCs were stored for 72 h before
13 transfusion, leukoreduction appeared to promote alloimmunization since mice receiving short-
14 term stored blood demonstrated a lower magnitude of immunization than those receiving stored
15 LR RBCs (Fig. 2A,B). This observation was significant in primed mice ($P<0.001$; Fig. 2C,D)
16 and demonstrated that reducing WBC numbers before short-term storage increases
17 immunogenicity of stored RBCs. Our findings showed that after short-term storage, without
18 WBCs, RBCs displayed enhanced immunogenicity whereas in the presence of WBCs, they
19 became less immunogenic, possibly due to WBC apoptosis. In addition, LR RBC
20 immunogenicity observed after storage might be related to altered RBC integrity due to
21 leukoreduction. Because 72 h-stored blood contained apoptotic WBCs (Fig. 1A,B), we
22 hypothesized that apoptotic WBCs may modulate stored-blood alloimmunization.

23

1 **Addition of exogenous apoptotic cells to leukoreduced-RBC transfusion did not modulate**
2 **post-transfusion alloimmunization**

3 Apoptotic cells are endowed with immunomodulatory properties that can control humoral
4 immune response²⁵⁻²⁹ independently of their strain origin.^{30,31} We investigated whether addition
5 of apoptotic cells would limit RBC alloimmunization. Alloimmunization was evaluated 2 weeks
6 after transfusion of HOD LR RBCs plus apoptotic cells from different donor origins (FvB,
7 C57Bl/6 or BALB/c origin); we observed no modulation of alloimmunization when apoptotic
8 cells were transfused concomitantly with LR RBCs (Fig. 3A). Alloimmunization was not
9 ameliorated either when the number of apoptotic cells was adapted to the number of WBCs
10 injected into 72 h-stored blood (0.3×10^6 apoptotic WBCs/transfusion) (Fig. 3B) nor when the
11 time of injection was delayed from the time of transfusion to facilitate elimination of apoptotic
12 cells by macrophages (Fig. 3C). Indeed, macrophages are essential to apoptotic cell-induced
13 immunomodulation^{21,29,32,33} and RBCs may compete with apoptotic cells, limiting both apoptotic
14 cell removal by macrophages, and apoptotic cell-induced immunomodulation. Using mHEL⁺
15 apoptotic cells (Figure 3 D) or apoptotic WBCs, as compared to apoptotic thymocytes (Fig. 3E),
16 post-transfusion alloimmunization was not alleviated either. We also injected apoptotic cells with
17 fresh or stored LR RBCs or blood, with no impact on alloimmunization (Fig. 3F). These data
18 suggest that injecting apoptotic cells at the time of transfusion may not modulate RBC
19 alloimmunization.

20

21 **Stored blood contained soluble factors limiting post-RBC-transfusion alloimmunization**

22 We then investigated whether the modulation of alloimmunization observed after transfusion of
23 stored blood was related to the secretion of immunomodulatory factors by apoptotic cells within

1 the blood unit rather than to the presence of apoptotic cells. Firstly, to rule out the role of WBCs
2 at the time of transfusion, we performed post-storage leukoreduction to eliminate WBCs,
3 including apoptotic WBCs but not WBC-released soluble factors. This mimics bedside
4 leukoreduction. In this setting, we observed that post-storage leukoreduction did not inhibit
5 modulation associated with short-term stored-blood transfusion (Fig. 4A). This highlights the
6 immunomodulatory role of soluble factors released by WBCs during storage. To confirm this,
7 we used the supernatant from 72 h-stored blood to modulate stored-LR RBC alloimmunization;
8 addition of stored-blood supernatant modulated stored-LR RBC alloimmunization (Fig. 4B).
9 This confirms that the factors released by WBCs during short-term storage modulate
10 alloimmunization. Transfusion of stored blood that was washed to eliminate soluble factors thus
11 restoring RBC alloimmunization was used as a control (Fig. 4B).

12 To reconcile our data showing that addition of apoptotic cells to RBC transfusion did not
13 ameliorate alloimmunization, with our data showing the critical role of stored blood-derived
14 soluble factors on the amelioration of alloimmunization, we then investigated whether incubation
15 of apoptotic cells with LR RBCs for 72 h would limit stored-LR RBC alloimmunization. We
16 observed that mice transfused with LR RBCs stored with apoptotic cells demonstrated less
17 alloimmunization than mice transfused with stored LR RBCs (Fig. 4C). This demonstrates that
18 soluble factors released by apoptotic cells during 3-day storage modulated alloimmunization.
19 Control mice transfused with stored LR RBCs plus 72 h-stored apoptotic cell supernatant again
20 demonstrated reduced levels of RBC alloimmunization (Fig. 4C). Lastly, adding washed stored
21 apoptotic cells to stored LR RBC transfusion did not modulate alloimmunization (Fig. 4C).
22 These data confirm that WBCs undergoing apoptosis release soluble factors that reduce RBC
23 immunogenicity.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23

Stored blood contained TGF-β

WBC-derived products are known to modulate immunogenicity of stored RBCs;^{13,34} we therefore investigated the role of microparticles in stored-blood supernatant. Microparticles isolated from stored blood did not decrease stored-LR RBC alloimmunization (Fig. 4D). However, addition of the stored-blood supernatant remaining fraction –without microparticles– decreased stored-LR RBC alloimmunization (Fig. 4D). To determine which factors other than microparticles could be responsible for reducing alloimmunization, we studied anti-inflammatory cytokines in stored-blood supernatant using T-cell polarization assays. Spleen cells were cultured in the presence of supernatants from stored and fresh LR RBCs or blood, and whereas differences were minimal in terms of Th1- or Th17-commitment of T cells (data not shown), stored-blood supernatant induced significantly greater expression of the Foxp3 transcription factor than other supernatants (Fig. 5A). This increase was equivalent to that observed with recombinant TGF-β as a control (Fig. 5A), suggesting the presence of TGF-β in stored blood. The same assay was performed using naïve CD4⁺CD25⁻ T cells; surprisingly, we did not observe any significant increase in Foxp3 expression in T cells cultured with stored-blood supernatant (Fig. 5B). To reconcile our data, latent TGF-β, which can be activated by spleen cells but not by naïve CD4⁺ T cells, was quantified in stored blood by ELISA. We only observed the latent form in the tested supernatants (Fig. 5C) and latent TGF-β levels were significantly high only in stored-blood supernatants (Fig. 5C). Likewise, addition of blocking anti-TGF-β mAb to stored-blood supernatant in spleen cell culture strongly inhibited Foxp3 expression induced by stored-blood supernatant (Fig. 5D). Similar results were obtained using stored-blood supernatant depleted for TGF-β using magnetic beads (Fig. 5D). Our findings

1 confirm the presence of latent TGF- β in stored blood, which may modulate RBC
2 alloimmunization.

3

4 **WBCs undergoing apoptosis during RBC storage secreted TGF- β**

5 Since our blood products were reduced in platelets, TGF- β must have been secreted by WBCs.
6 We measured expression of membrane-bound TGF- β on leukocytes and subsets from fresh or 3
7 day-stored blood. Whereas T and B cells did not increase membrane-bound TGF- β expression
8 over time, monocyte/macrophage populations, and neutrophils demonstrated slight and strong
9 increases in membrane-bound TGF- β expression, respectively (Fig. 6A,C). Cells undergoing
10 apoptosis are known to release TGF- β ;¹⁴ we therefore investigated leukocyte apoptosis.
11 Leukocytes and neutrophils in particular, demonstrated more apoptosis after storage (Fig. 6B,C).
12 Apoptotic leukocytes were positive for membrane-bound TGF- β -expression and as expected, this
13 effect was strongly increased after 72 hours' storage (Fig. 6D,E). Necrotic cells demonstrated
14 non-specific positivity for LAP expression. Our findings show that stored blood contains TGF- β
15 released by WBCs undergoing apoptosis, mostly by the myeloid lineages.

16

17 **TGF- β contained in stored blood limited stored-RBC alloimmunization**

18 Apoptotic WBCs in stored blood released TGF- β , which may limit post-transfusion
19 alloimmunization. Thus, TGF- β was neutralized using a blocking antibody in stored blood before
20 transfusion and abrogated the modulation of stored-blood alloimmunization (Fig. 7A). However,
21 residual anti-TGF- β antibody was also transfused. To avoid such a bias, we depleted TGF- β from
22 stored blood using magnetic beads, thus achieving TGF- β -specific deletion before transfusion. In
23 this setting, TGF- β -depleted stored blood no longer modulated alloimmunization (Fig. 7B). **To**

1 further confirm the key role of TGF- β , we first increased apoptotic cell-derived TGF- β content in
2 stored apoptotic cell supernatants by storing 1, 5 or $15 \cdot 10^6$ apoptotic cells per 400 μ L of PBS that
3 were then added (after cell elimination) to 100 μ L of stored LR RBCs, as done in Figure 4C.
4 Indeed, TGF- β content was increased in such supernatants from 53 ± 21 pg/mL for $1 \cdot 10^6$
5 apoptotic cells to 709 ± 8 pg/mL for $15 \cdot 10^6$ apoptotic cells. When those supernatants were
6 transfused with stored LR RBCs, post-transfusion RBC alloimmunization was strongly reduced
7 with $5 \cdot 10^6$ apoptotic cell supernatant, and was almost prevented with $15 \cdot 10^6$ apoptotic cell
8 supernatant (Fig. 7C). Then we used recombinant TGF- β in quantities similar to those quantified
9 in stored apoptotic cell supernatants, to limit stored LR RBC-induced alloimmunization.
10 Interestingly, recombinant TGF- β was also able to decrease post-transfusion alloimmunization
11 when used at 500 pg/mL of transfused blood, and to prevent alloimmunization when used at
12 1500 pg/mL (Fig. 7D). Our data indicate that apoptotic WBC-derived TGF- β can modulate post-
13 transfusion alloimmunization and even prevent it when used at 1500 pg/mL.
14 Likewise, since Day 3 of storage was selected for the best apoptotic/necrotic cell ratio in RBC
15 units, we observed that stored blood induced less immunization than stored LR RBCs only when
16 storage lasted three days, and not six, eight or 14 days (Fig. 7E). Membrane-bound TGF- β^+
17 (=LAP $^+$) apoptotic WBC levels in blood were also seen to be higher after three days *versus*
18 longer periods of storage, but no differences in terms of RBC alloimmunization modulation was
19 observed after 3 days of storage (Fig. 7F). TGF- β levels in stored blood showed that low levels
20 of latent TGF- β were present in LR RBCs (Fig. 7G). Whereas 3-day stored blood demonstrated
21 high levels of latent TGF- β , significant amounts were also observed in blood stored longer (Fig.
22 7G), but with no effect on modulation of RBC alloimmunization (Fig. 7E,F), suggesting that
23 other factors, possibly related to WBC necrosis, contributed to the higher immunogenicity of

1 RBCs stored long term. Our data highlight the key role of 3-day stored apoptotic WBC-derived
2 TGF- β to limit post-transfusion RBC alloimmunization.

3

4 **DISCUSSION**

5 In this study, we describe how WBCs that undergo apoptosis during storage, and more
6 specifically apoptotic WBC-derived TGF- β , are associated with modulation of alloimmunization
7 following transfusion of stored RBC. This is the first time that an experimental model of
8 transfusion-induced alloimmunization has shown that apoptotic WBC-derived soluble factors
9 induced by short-term blood storage, and in particular TGF- β , are responsible for
10 immunomodulation of RBC immunogenicity. Fresh blood, LR RBCs and stored LR RBCs did
11 not demonstrate such properties. Apoptotic WBC-derived soluble factors can negatively
12 modulate transfusion-related alloimmunization. These data also highlight the crucial role of
13 apoptotic WBCs in modulating immunogenicity of blood products thus confirming the role of
14 TGF- β derived from apoptotic WBCs in the immunomodulation associated with transfusion
15 TRIM effect.

16 Viable leukocytes in transfused blood products have already been shown to be deleterious for
17 recipients, especially for the incidence of alloimmunization.^{35,36} Indeed, in our study, fresh LR
18 RBC was less immunogenic than fresh blood, reinforcing the idea that WBCs promote both class
19 I MHC immunization as well as RBC antigen immunization. However, after a short storage of 72
20 h, corresponding to approximately 6-7 days' storage in human settings, the presence of WBCs in
21 stored blood modulated RBC immunogenicity. Our data suggest that with respect to minimizing
22 post-transfusion alloimmunization, short-term blood storage might be beneficial when short term
23 blood storage is being considered, leukoreduction should be done at the bedside and not before

1 storage. Firstly, blood preparation procedures might alter RBC integrity and LR RBCs became
2 more immunogenic with time of storage. Secondly, when WBCs were kept in blood units, they
3 become apoptotic and released TGF- β modulating stored-blood immunogenicity whereas after
4 long period of storage, WBCs become necrotic and source of immunogenic signals. Loss of
5 RBC integrity was not address in this study. We used the now-well-established HOD mouse
6 model to address the role of apoptotic WBCs on erythrocyte allo-immunization. For this purpose,
7 we washed blood in order to avoid the influence of platelets. This step, not performed in human
8 blood processing, influenced RBC integrity (data not shown) but similarly in all of our
9 conditions, without affecting HOD expression on RBCs.

10 Our data highlight the crucial role of apoptotic WBC-derived immunomodulatory factors.
11 Indeed, whereas injection of apoptotic cells has been shown to induce immunomodulation in
12 inflammatory settings,^{15,16} in our model of post-transfusion alloimmunization, it had no impact.
13 ~~We expected the apoptotic cell mediated effect to be limited by RBCs because once injected,~~
14 ~~RBCs would saturate macrophages thus eliminating apoptotic cells and mediating~~
15 ~~immunomodulation less efficiently.~~ Apoptotic cells were injected at different time points from
16 transfusion but with no benefit on RBC alloimmunization. However, when apoptotic cells were
17 stored with RBCs, the former had time to release TGF- β into the RBC unit thus reducing RBC
18 immunogenicity. A similar effect was mimicked using the supernatant from stored apoptotic
19 cells transfused with LR RBCs. TGF- β was shown to be the key cytokine in this effect.

20 Prolonged storage increases immunogenicity of RBCs. This increase can be associated with LR
21 affecting RBC integrity and/or with significantly greater WBC necrosis. The high rate of WBC
22 necrosis in our setting can be related to our blood preparation procedure since 3 washes step
23 facilitated cell necrosis particularly after 6 days of storage (data not shown). Necrotic WBCs

1 stimulate inflammation,³⁷ and several studies have highlighted the harmful effects of prolonged
2 storage on post-transfusion alloimmunization.^{12,38} We elected to use blood stored 72 hours and
3 studied WBC apoptosis-related but not necrosis-related effects that are easier to assess with
4 blood stored 14 days.

5 Apoptosis of WBCs during storage is well known,^{23,39} and may explain the anti-inflammatory
6 effect of blood transfusion.^{19,34} Our result supports this hypothesis since the presence alone of
7 apoptotic WBCs within blood modulates RBC alloimmunization. Apoptotic WBCs have to age
8 within RBCs to release anti-inflammatory cytokines such as TGF- β . Indeed, addition of
9 apoptotic leukocytes to LR RBCs at the time of transfusion did not limit post-transfusion
10 alloimmunization whereas addition of stored apoptotic cell supernatant did. Thus, TGF- β ,
11 released by apoptotic WBCs,¹⁴ is responsible for modulating RBC alloimmunization. The role of
12 TGF- β in this setting was further confirmed by depleting TGF- β from stored blood, which
13 reestablished immunogenicity to stored RBCs. Furthermore, addition of recombinant TGF- β
14 (1500 pg/mL) to stored LR RBC transfusion can prevent post-transfusion alloimmunization in
15 almost all mice. TGF- β is a key cytokine for Foxp3⁺ Treg induction, both in humans and
16 rodents.^{40,41} Treg are known to suppress T-cell activation and initiation of the immune response.
17 Recently, Treg have also been shown to directly abrogate humoral responses by inhibiting both
18 B cell Ig-production and class switching,⁴² killing B cells,⁴³ and preventing long-lived plasma
19 cell commitment.⁴⁴ Moreover, one study reported that Treg injection prior to transfusion totally
20 suppresses alloimmunization.⁴⁵ In our study, whereas stored-blood supernatant triggered Treg
21 polarization *in vitro*, no differences in terms of Treg percentages or numbers at one, two or 14
22 days post-transfusion were observed (data not shown). The mechanisms behind
23 alloimmunization modulation by stored WBC-derived TGF- β have yet to be elucidated. We can

1 hypothesize that TGF- β directly affects and limits APC to create an efficient humoral response to
2 RBC antigens. Our data suggested that TGF- β modulated APC engulfment of RBCs Ag,
3 processing and presentation that limited immune response to the RBC's Ag.
4 Finally, this is the first demonstration of the relation between storage-induced apoptosis of
5 WBCs and modulation of transfusion immunogenicity. Our study highlights the tolerogenic role
6 immunomodulatory properties of apoptotic WBCs and apoptotic WBC-derived TGF- β in
7 modulating post-transfusion alloimmunization. Apoptotic WBC-derived TGF- β represents only
8 the tip of the iceberg and many other environmental factors, including the inflammatory status of
9 the recipient, may influence post-transfusion alloimmunization and other adverse effects of
10 human transfusion. In such settings, however, the presence of apoptotic WBCs in blood after
11 short-term storage limits RBC immunogenicity. Human transfusion procedures should include
12 LR at patients' bedside for blood units stored short term (6-7 days) and include LR for blood
13 units selected for long-term storage.
14

1 **Acknowledgments:** Thanks to Prof Lucienne Chatenoud (INSERM U1013, Paris, France) and
2 James Zimring (Puget Sound Blood Center Research Institute, Seattle, WA, USA) who kindly
3 provided us with 2G7 hybridoma cell line and HOD plus mHEL transgenic mice, respectively.
4

1 References

- 2 1. Vamvakas EC, Blajchman MA. Transfusion-related mortality: the ongoing risks of
3 allogeneic blood transfusion and the available strategies for their prevention. *Blood*
4 2009;**113**: 3406-17.
- 5 2. Noizat-Pirenne F, Tournamille C, Bierling P, Roudot-Thoraval F, Le Pennec PY, Rouger
6 P, Ansart-Pirenne H. Relative immunogenicity of Fya and K antigens in a Caucasian
7 population, based on HLA class II restriction analysis. *Transfusion* 2006;**46**: 1328-33.
- 8 3. Zalpuri S, Middelburg RA, Schonewille H, de Vooght KM, le Cessie S, van der Bom JG,
9 Zwaginga JJ. Intensive red blood cell transfusions and risk of alloimmunization.
10 *Transfusion* 2014;**54**: 278-84.
- 11 4. Hendrickson JE, Hod EA, Spitalnik SL, Hillyer CD, Zimring JC. Storage of murine red
12 blood cells enhances alloantibody responses to an erythroid-specific model antigen.
13 *Transfusion* 2010;**50**: 642-8.
- 14 5. Shanwell A, Kristiansson M, Remberger M, Ringden O. Generation of cytokines in red
15 cell concentrates during storage is prevented by prestorage white cell reduction.
16 *Transfusion* 1997;**37**: 678-84.
- 17 6. Chin-Yee I, Keeney M, Krueger L, Dietz G, Moses G. Supernatant from stored red cells
18 activates neutrophils. *Transfus Med* 1998;**8**: 49-56.
- 19 7. Sparrow RL, Patton KA. Supernatant from stored red blood cell primes inflammatory
20 cells: influence of prestorage white cell reduction. *Transfusion* 2004;**44**: 722-30.
- 21 8. Yazer MH, Triulzi DJ. Receipt of older RBCs does not predispose D-negative recipients
22 to anti-D alloimmunization. *Am J Clin Pathol* 2010;**134**: 443-7.
- 23 9. Bennett-Guerrero E, Veldman TH, Doctor A, Telen MJ, Ortel TL, Reid TS, Mulherin
24 MA, Zhu H, Buck RD, Califf RM, McMahan TJ. Evolution of adverse changes in stored
25 RBCs. *Proc Natl Acad Sci U S A* 2007;**104**: 17063-8.
- 26 10. Tinmouth A, Chin-Yee I. The clinical consequences of the red cell storage lesion.
27 *Transfus Med Rev* 2001;**15**: 91-107.
- 28 11. Relevy H, Koshkaryev A, Manny N, Yedgar S, Barshtein G. Blood banking-induced
29 alteration of red blood cell flow properties. *Transfusion* 2008;**48**: 136-46.
- 30 12. Hod EA, Zhang N, Sokol SA, Wojczyk BS, Francis RO, Ansaldi D, Francis KP, Della-
31 Latta P, Whittier S, Sheth S, Hendrickson JE, Zimring JC, Brittenham GM, Spitalnik SL.
32 Transfusion of red blood cells after prolonged storage produces harmful effects that are
33 mediated by iron and inflammation. *Blood* 2010;**115**: 4284-92.
- 34 13. Saas P, Angelot F, Bardiaux L, Seilles E, Garnache-Ottou F, Perruche S.
35 Phosphatidylserine-expressing cell by-products in transfusion: A pro-inflammatory or an
36 anti-inflammatory effect? *Transfus Clin Biol* 2012;**19**: 90-7.
- 37 14. Chen W, Frank ME, Jin W, Wahl SM. TGF-beta released by apoptotic T cells contributes
38 to an immunosuppressive milieu. *Immunity* 2001;**14**: 715-25.
- 39 15. Poon IK, Lucas CD, Rossi AG, Ravichandran KS. Apoptotic cell clearance: basic biology
40 and therapeutic potential. *Nat Rev Immunol* 2014;**14**: 166-80.
- 41 16. Saas P, Kaminski S, Perruche S. Prospects of apoptotic cell-based therapies for
42 transplantation and inflammatory diseases. *Immunotherapy* 2013;**5**: 1055-73.
- 43 17. Vamvakas EC, Blajchman MA. Transfusion-related immunomodulation (TRIM): an
44 update. *Blood Rev* 2007;**21**: 327-48.

- 1 18. Ghio M, Contini P, Ubezio G, Mazzei C, Puppo F, Indiveri F. Immunomodulatory effects
2 of blood transfusions: the synergic role of soluble HLA Class I free heavy-chain
3 molecules detectable in blood components. *Transfusion* 2008;**48**: 1591-7.
- 4 19. Dzik S, Mincheff M, Puppo F. Apoptosis, transforming growth factor-beta, and the
5 immunosuppressive effect of transfusion. *Transfusion* 2002;**42**: 1221-3.
- 6 20. Perruche S, Kleinclauss F, Lienard A, Robinet E, Tiberghien P, Saas P. A single-platform
7 approach using flow cytometry and microbeads to evaluate immune reconstitution in
8 mice after bone marrow transplantation. *J Immunol Methods* 2004;**294**: 53-66.
- 9 21. Bonnefoy F, Perruche S, Couturier M, Sedrati A, Sun Y, Tiberghien P, Gaugler B, Saas
10 P. Plasmacytoid dendritic cells play a major role in apoptotic leukocyte-induced immune
11 modulation. *J Immunol* 2011;**186**: 5696-705.
- 12 22. Gilson CR, Kraus TS, Hod EA, Hendrickson JE, Spitalnik SL, Hillyer CD, Shaz BH,
13 Zimring JC. A novel mouse model of red blood cell storage and posttransfusion in vivo
14 survival. *Transfusion* 2009;**49**: 1546-53.
- 15 23. Frabetti F, Musiani D, Marini M, Fanelli C, Coppola S, Ghibelli L, Tazzari PL, Bontadini
16 A, Tassi C, Conte R. White cell apoptosis in packed red cells. *Transfusion* 1998;**38**:
17 1082-9.
- 18 24. Hendrickson JE, Desmarests M, Deshpande SS, Chadwick TE, Hillyer CD, Roback JD,
19 Zimring JC. Recipient inflammation affects the frequency and magnitude of
20 immunization to transfused red blood cells. *Transfusion* 2006;**46**: 1526-36.
- 21 25. Perruche S, Kleinclauss F, Bittencourt Mde C, Paris D, Tiberghien P, Saas P. Intravenous
22 infusion of apoptotic cells simultaneously with allogeneic hematopoietic grafts alters
23 anti-donor humoral immune responses. *Am J Transplant* 2004;**4**: 1361-5.
- 24 26. Notley CA, Brown MA, Wright GP, Ehrenstein MR. Natural IgM is required for
25 suppression of inflammatory arthritis by apoptotic cells. *J Immunol* 2011;**186**: 4967-72.
- 26 27. Xia CQ, Peng R, Qiu Y, Annamalai M, Gordon D, Clare-Salzler MJ. Transfusion of
27 apoptotic beta-cells induces immune tolerance to beta-cell antigens and prevents type 1
28 diabetes in NOD mice. *Diabetes* 2007;**56**: 2116-23.
- 29 28. Gray M, Miles K, Salter D, Gray D, Savill J. Apoptotic cells protect mice from
30 autoimmune inflammation by the induction of regulatory B cells. *Proc Natl Acad Sci U S*
31 *A* 2007;**104**: 14080-5.
- 32 29. Wang Z, Larregina AT, Shufesky WJ, Perone MJ, Montecalvo A, Zahorchak AF,
33 Thomson AW, Morelli AE. Use of the inhibitory effect of apoptotic cells on dendritic
34 cells for graft survival via T-cell deletion and regulatory T cells. *Am J Transplant* 2006;**6**:
35 1297-311.
- 36 30. Bittencourt MC, Perruche S, Contassot E, Fresnay S, Baron MH, Angonin R, Aubin F,
37 Herve P, Tiberghien P, Saas P. Intravenous injection of apoptotic leukocytes enhances
38 bone marrow engraftment across major histocompatibility barriers. *Blood* 2001;**98**: 224-
39 30.
- 40 31. Zhang M, Xu S, Han Y, Cao X. Apoptotic cells attenuate fulminant hepatitis by priming
41 Kupffer cells to produce interleukin-10 through membrane-bound TGF-beta. *Hepatology*
42 2011;**53**: 306-16.
- 43 32. Perruche S, Zhang P, Liu Y, Saas P, Bluestone JA, Chen W. CD3-specific antibody-
44 induced immune tolerance involves transforming growth factor-beta from phagocytes
45 digesting apoptotic T cells. *Nat Med* 2008;**14**: 528-35.

- 1 33. Kleinclaus F, Perruche S, Masson E, de Carvalho Bittencourt M, Biichle S, Remy-
2 Martin JP, Ferrand C, Martin M, Bittard H, Chalopin JM, Seilles E, Tiberghien P, Saas P.
3 Intravenous apoptotic spleen cell infusion induces a TGF-beta-dependent regulatory T-
4 cell expansion. *Cell Death Differ* 2006;**13**: 41-52.
- 5 34. Dzik WH. Apoptosis, TGF beta and transfusion-related immunosuppression: Biologic
6 versus clinical effects. *Transfus Apher Sci* 2003;**29**: 127-9.
- 7 35. Blumberg N, Heal JM, Gettings KF. WBC reduction of RBC transfusions is associated
8 with a decreased incidence of RBC alloimmunization. *Transfusion* 2003;**43**: 945-52.
- 9 36. Blajchman MA. The clinical benefits of the leukoreduction of blood products. *J Trauma*
10 2006;**60**: S83-90.
- 11 37. Kono H, Rock KL. How dying cells alert the immune system to danger. *Nat Rev*
12 *Immunol* 2008;**8**: 279-89.
- 13 38. Hendrickson JE, Chadwick TE, Roback JD, Hillyer CD, Zimring JC. Inflammation
14 enhances consumption and presentation of transfused RBC antigens by dendritic cells.
15 *Blood* 2007;**110**: 2736-43.
- 16 39. Mincheff M. Changes in donor leukocytes during blood storage. Implications on post-
17 transfusion immunomodulation and transfusion-associated GVHD. *Vox Sang* 1998;**74**
18 **Suppl 2**: 189-200.
- 19 40. Chen W, Jin W, Hardegen N, Lei KJ, Li L, Marinos N, McGrady G, Wahl SM.
20 Conversion of peripheral CD4+CD25- naive T cells to CD4+CD25+ regulatory T cells
21 by TGF-beta induction of transcription factor Foxp3. *J Exp Med* 2003;**198**: 1875-86.
- 22 41. Yamagiwa S, Gray JD, Hashimoto S, Horwitz DA. A role for TGF-beta in the generation
23 and expansion of CD4+CD25+ regulatory T cells from human peripheral blood. *J*
24 *Immunol* 2001;**166**: 7282-9.
- 25 42. Lim HW, Hillsamer P, Banham AH, Kim CH. Cutting edge: direct suppression of B cells
26 by CD4+ CD25+ regulatory T cells. *J Immunol* 2005;**175**: 4180-3.
- 27 43. Zhao DM, Thornton AM, DiPaolo RJ, Shevach EM. Activated CD4+CD25+ T cells
28 selectively kill B lymphocytes. *Blood* 2006;**107**: 3925-32.
- 29 44. Jang E, Cho WS, Cho ML, Park HJ, Oh HJ, Kang SM, Paik DJ, Youn J. Foxp3+
30 regulatory T cells control humoral autoimmunity by suppressing the development of
31 long-lived plasma cells. *J Immunol*; **186**: 1546-53.
- 32 45. Yu J, Heck S, Yazdanbakhsh K. Prevention of red cell alloimmunization by CD25
33 regulatory T cells in mouse models. *Am J Hematol* 2007;**82**: 691-6.

34

35

36

1 **Figure Legends**

2 **Figure 1. WBCs undergo apoptosis during blood storage independently of experimental**

3 **settings, not influencing RBC hematocrits or RBC *in vivo* survival.** WBC numbers (white

4 circles), apoptosis (grey bars) and necrosis (black bars) were evaluated in blood from HOD mice

5 stored at +4°C on different days as indicated (A). Representative data pooled from two out of six

6 independent experiments. Percent of WBC apoptosis was compared between Day 0 and Day 3 of

7 storage (B). Data pooled from 16 independent experiments. ***= $P < 0.001$ (Student's *t* test).

8 Blood from HOD mice was stored at +4°C in PBS (open symbols) or SAGM (black symbols)

9 and apoptotic (squares), necrotic (triangles) and live (circles) WBCs were evaluated at different

10 days as indicated (C). The same experiment was done to assess the influence of the anticoagulant

11 used for blood puncture, EDTA (open symbols) or ACD (black symbols) (D). Data from one of

12 two representative experiments. Hematocrits were evaluated in RBC units upon storage in

13 SAGM (black symbols) or PBS (open symbols), punctured using EDTA- (circles) or ACD-

14 coated tubes (squares) (E). Data from one of three representative experiments. Survival of RBCs

15 (squares) or LR RBCs (circles), fresh (open symbols) or after 72-h storage (black symbols) in

16 SAGM (left panel) or PBS (right panel) was evaluated *in vivo* after transfusion by the detection

17 of HOD+ RBCs within total RBCs in transfused wild type mice (F). Data from one of two

18 representative experiments showing mean \pm SEM, five mice/group.

19

20 **Figure 2. RBC alloimmunization is influenced by both leucoreduction and storage.** Post-

21 transfusion alloimmunization was demonstrated by anti-HOD IgG antibody detection by cross-

22 match in the plasma of unprimed (A,B) or poly(I:C) primed (C,D) mice two weeks after

23 receiving vehicle, fresh or 72-h stored LR RBCs or blood. Data were expressed as MFI ratio (to

1 vehicle) (A,C) or MFI (B,D). Im. HEL group corresponded to plasma from mice immunized
2 against HEL Ag. Representative histograms from flow cytometry cross-match analysis showing
3 mouse serum are given in **B and D**: light grey=mice receiving vehicle, dark grey=mice
4 transfused with the indicated RBC. Numbers given correspond to MFI of mice transfused with
5 RBC units as indicated. Data from three to 11 independent experiments, 15 to 55 mice/group,
6 showing mice individually and mean \pm SEM. *= $P < 0.05$; **= $P < 0.01$; ***= $P < 0.001$ (Student's
7 *t* test).

8
9 **Figure 3. Apoptotic cell injection with leukoreduced RBC transfusion does not modulate**
10 **post-transfusion alloimmunization.** Alloimmunization (*i.e.*, presence of circulating anti-HOD
11 IgG) was evaluated by cross-match in the plasma of primed C57Bl/6 mice, two weeks after
12 receiving leukoreduced HOD RBCs (LR RBCs) with PBS or 5×10^6 apoptotic cells of FvB (FvB
13 Apo) or C57Bl/6 (C57Bl/6 Apo) or BALB/c (BALB/c Apo) origin (A). Data from five
14 independent experiments, 6 to 25 mice/group, showing mice individually and mean \pm SEM.
15 Apoptotic leukocytes (Apo) were also injected in different quantities: 0.1, 1 and 5×10^6
16 cells/mouse and post-transfusion alloimmunization evaluated (B). Data from two independent
17 experiments, 4 to 8 mice/group, showing mice individually and mean \pm SEM. Apoptotic cell
18 injection was also delayed (-2 and +2 h) or not, from LR RBC transfusion and post-transfusion
19 alloimmunization evaluated (C). Data from two to four independent experiments, 8 to 19
20 mice/group, showing mice individually and mean \pm SEM. Primed mice were transfused with
21 HOD⁺ LR RBCs, with or without 5×10^6 apoptotic cells from mHEL transgenic mice (mHEL
22 Apo). One group of mice received apoptotic cells alone. Post-transfusion alloimmunization was
23 evaluated by cross-match and confirmed by the presence of Ab directed against HOD antigen

1 (D). Data from one experiment showing mice individually and mean \pm SEM, 2 to 5 mice/group.
2 $*=P<0.05$; $***=P<0.001$ (Student's *t* test). Apoptotic cells prepared from thymus (thymo. Apo)
3 or WBCs (WBC Apo) were compared to modulate alloimmunization when injected with LR
4 RBCs (E). Data from one experiment showing mice individually and mean \pm SEM, 2 to 5
5 mice/group. Apoptotic cells were also injected in addition to fresh or stored LR RBCs or blood
6 (+Apo = black squares) and RBC alloimmunization evaluated (F). Data from one experiment, 2
7 to 5 mice/group, showing mice individually and mean \pm SEM. $*=P<0.05$ (Student's *t* test). Im.
8 HEL=control plasma from mice immunized against HEL Ag.

10 **Figure 4. Stored blood contains soluble factors limiting post-transfusion RBC**

11 **alloimmunization.** Post-transfusion alloimmunization (*i.e.*, presence of circulating anti-HOD
12 IgG) was evaluated by cross-match of the plasma of primed mice, two weeks after receiving
13 fresh or stored, leukoreduced (LR) RBCs or blood, or stored blood subjected to LR after storage
14 (stored blood + LR) (A). Data from two independent experiments showing individual mouse and
15 mean \pm SEM, 4 to 10 mice/group. In addition to stored LR RBCs or stored blood, other mice
16 were transfused with stored blood that was washed to eliminate soluble factors, or stored LR
17 RBCs plus stored-blood supernatant (sup) (B). Alloimmunization was determined as previously
18 described. Data from three independent experiments showing mice individually and mean \pm
19 SEM, 8 to 15 mice/group. Mice were transfused with stored LR RBCs or blood, or LR RBCs
20 stored with apoptotic cells (stored [LR RBCs+Apo]), stored LR RBCs plus stored apoptotic cells
21 (+ stored Apo) or stored LR RBCs + the supernatant from stored apoptotic cells (+ stored Apo
22 sup) (C). Alloimmunization was determined as previously described. Data from two independent
23 experiments showing mice individually and mean \pm SEM, 4 to 10 mice/group. To evaluate the

1 role of microparticles in RBC units, mice were transfused with stored LR RBCs or blood, stored
2 blood washed to eliminate soluble factors, or stored LR RBCs plus stored-blood supernatant (+
3 stored blood sup), stored-blood microparticles (+ stored blood μ P), stored-blood supernatant
4 exempt of microparticles (+ stored blood sup wo μ P) or both of the latter (+ stored blood μ P +
5 stored blood sup wo μ P) (**D**). Alloimmunization was determined as described previously. Data
6 from one experiment showing mice individually and mean \pm SEM, five mice/group. $*=P<0.05$;
7 $**=P<0.01$; $***=P<0.001$ (Student's *t* test). Im. HEL=control plasma from mice immunized
8 against HEL Ag.

9
10 **Figure 5. Stored-blood supernatant contains TGF- β .** Spleen (**A**) or CD4⁺CD25⁻ (**B**) cells were
11 cultured for three days in the presence or not (open bars) of CD3- or CD3/CD28-specific Abs
12 (black bars), respectively, and with fresh or stored LR RBCs or blood supernatants. Polarization
13 toward a regulatory T-cell profile was evaluated by FACS in CD4⁺ T cells by measuring the
14 expression of CD25 molecule and Foxp3 transcription factor. Data showing three pooled
15 independent experiments mean \pm SEM. Total (black bars) and active (open bars) forms of TGF- β
16 were quantified by ELISA in corresponding supernatants (**C**). Mean \pm SEM of duplicates from
17 one of two representative experiments. Spleen cells were cultured as indicated in (**A**) in the
18 presence of stored-blood supernatant and with or without (-) anti-TGF- β blocking antibody (+
19 α TGF- β), its corresponding isotype (+ iso) or with stored-blood supernatant depleted of TGF- β
20 (TGF- β -depleted) (**D**). Polarization toward a regulatory T-cell profile was evaluated similarly.
21 Data from two pooled experiments, showing means of triplicates \pm SEM. $*=P<0.05$; $**=P<0.01$;
22 $***=P<0.001$ (Student's *t* test).

23

1 **Figure 6. WBCs express membrane-bound TGF- β on apoptosis during RBC storage**
2 Monocytes/macrophages (CD11b⁺), neutrophils (GR1⁺), B (CD19⁺) and T (CD3⁺) cells were
3 gated in CD45_{LCA}⁺ leukocytes for their expression of membrane-bound TGF- β (= LAP
4 expression) (A,C) or apoptosis (B,C) by FACS during storage on leukocytes (diamonds),
5 mono/macrophages (circles), neutrophils (triangles), T (crosses) and B (squares) cells in RBC
6 units during storage as indicated. Leucocytes were also examined for the expression of
7 membrane-bound TGF- β (= LAP expression) by FACS during storage of blood depending on
8 their survival status (alive [7-AAD⁻Annexin V⁻], triangles; apoptotic [7-AAD⁻Annexin V⁺],
9 circles; necrotic [7-AAD⁺Annexin V⁺], squares) (D,E). Percentages of cells of interest are shown
10 on dot plots. Representative dot plots from one of three independent experiments showing
11 similar results (A,B,D). Data from three pooled experiments showing mean \pm SEM (C,E).

12
13 **Figure 7. TGF- β contained in stored blood modulates stored RBC alloimmunization.** Post-
14 transfusion alloimmunization (*i.e.*, presence of circulating anti-HOD IgG) was evaluated by
15 cross-match in the plasma of primed mice, two weeks after receiving fresh or stored LR RBCs or
16 blood. In addition, some mice also received stored blood in which TGF- β had been neutralized
17 prior to infusion using anti-TGF- β Ab (+ α TGF- β), or stored blood plus corresponding isotype
18 (+ iso.) (A), or stored blood depleted from TGF- β using microbeads (stored blood TGF- β
19 depleted) (B). Data from one to three independent experiments showing mice individually and
20 mean \pm SEM, five to 15 mice/group. Stored blood or LR RBC were transfused alone and stored
21 LR RBCs were also transfused with apoptotic cell-derived TGF- β (+stored 1.10⁶ Apo sup.,
22 +stored 5.10⁶ Apo sup., +stored 1.10⁶ Apo sup.) (C) or with recombinant TGF- β (150, 500 and
23 1500 pg/mL of transfused blood) (E) and alloimmunization was detected in plasma two weeks

1 after transfusion. Data from two experiments showing mice individually and mean \pm SEM, 4 to
2 10 mice/group. Blood or LR RBCs were transfused into mice either fresh (0 days) or after three,
3 six, eight or 14 days' storage, and alloimmunization was detected in plasma (E). Data from one
4 experiment showing mice individually and mean \pm SEM, five mice/group. Alloimmunization to
5 LR RBCs or blood was evaluated according to the duration of storage or the number of LAP⁺
6 WBCs in blood (F). Data from one of two independent experiments showing mean \pm SEM, five
7 mice/group. Latent TGF- β was quantified by ELISA in blood and LR RBCs, fresh or stored as
8 indicated (G). Data from one of two representative experiments showing mean of duplicates \pm
9 SEM. *= P <0.05; **= P <0.01; ***= P <0.001 (Student's t test). Im. HEL=control plasma from
10 mice immunized against HEL Ag.