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Positioning NK- κ B in multiple myeloma

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The non-canonical NF- κ B pathway and/or the canonical pathway, is aberrantly activated in 17% of patients with Multiple Myeloma (MM) and 40% of myeloma cell lines (MMCLs) through various gene abnormalities that result in NF- κ B inducing kinase (NIK) stabilization mostly¹.

The NF- κ B (nuclear factor kappa-light-chain-enhancer of activated B cells) pathway involves dimers of transcription factors (TFs) of the Rel/NF- κ B family, comprising RelA/P65, c-rel, RelB, NF- κ B1 (p50 and its p105 precursor) and NK- κ B2 (p52 and its precursor p100)². NF- κ B pathway plays a central role in infection and inflammation, and in lymphopoiesis, particularly in normal B cell and plasma cell development. These TFs are normally kept inactive in the cytoplasm through binding with inhibitors called I κ B (inhibitor of κ B) or unprocessed NF- κ B1 or NF- κ B2. Cell activation may result in activation of I κ B kinases (IKKs), mainly IKK β in the canonical pathway and IKK α in the non-canonical pathway. Activation of IKK β results in the phosphorylation, ubiquitination and degradation of I κ B α and other I κ Bs, releasing TF dimers (mainly P65/P50) to the nucleus that activate NF- κ B target genes. In the non-canonical pathway, NIK induces phosphorylation and activation of IKK α dimers, resulting in phosphorylation and proteasome processing of NK- κ B2 precursor into P52 subunit. P52 form active dimers with RelB that migrate to the nucleus and activate NK- κ B target genes. The NK- κ B canonical pathway is activated via numerous receptors, namely TNF receptor, T cell receptor, Toll like receptors activated by bacterial products, and BCMA and TACI receptors for BAFF or APRIL growth factors. The non-canonical NK- κ B pathway is activated by a more restricted sets of stimuli activating namely CD40, Rank, lymphotoxin- α receptor, or BAFF-receptor.

Demchenko et al. have further documented the mechanisms of action of the gene abnormalities targeting the regulatory elements of canonical and non-canonical NK- κ B pathway in MMCLs¹. These gene abnormalities were initially reported in 2007 in 40% of MMCLs and primary myeloma cells of 17% of patients with MM^{3,4}. These gene abnormalities target genes encoding either for products amplifying one of the 2 NK- κ B pathways (*TACI*, *CD40*, *LTB-R*, *NK- κ B1*, *NIK*) or for inhibitors of the canonical (*CYLD*) or non-canonical NK- κ B pathway (*clAP1&2*, *TRAF2*, *TRAF3*, *NK- κ B2*). Indeed in inactive cells, NIK is rapidly degraded through ubiquitination by clAP1-2 ubiquitinases. NIK binds TRAF3 first and the NIK/TRAF3 complex is recruited to clAP1-2/TRAF2 via TRAF3 binding to TRAF2. Activation of receptors by external stimuli induces recruitment of c-IAP1-2/TRAF3/TRAF3 complex to the activated receptors, increasing ubiquitination and degradation of TRAF3. TRAF3 decrease makes it possible to increase NIK and phosphorylation and activation of IKK α dimers. In MM, the most frequent mutations involve *TRAF3* gene located on chromosome 14q32, but also *TRAF2* or *clAP1-2*, resulting in increased NIK expression.

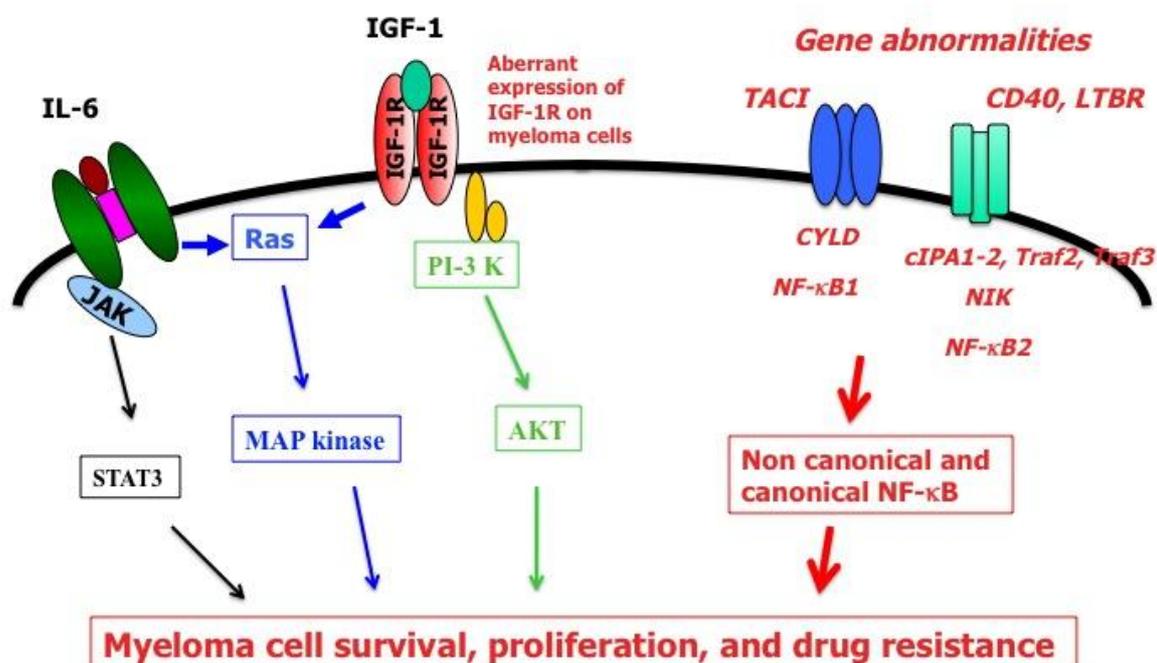
Demchenko et al. have shown that nuclear p52 and RelB are detected in MMCLs

with abnormalities targeting the non-canonical pathway, but also nuclear P65 and p50 that are indicators of canonical pathway activation. A deletion of cIAP or TRAF2 results in loss of NIK downregulation, whereas TRAF3 mutations or deletions partly remove NIK control. Actually, Demchenko et al. have shown that TRAF2 can directly interact with NIK and recruit cIAP to NIK, promoting NIK ubiquitination and degradation in the absence of TRAF3. In addition, Demchenko et al. have shown that an overexpression or knockout of NIK in 2 MMCLs result in upregulation or downregulation of non-canonical NK- κ B pathway (increased nuclear P52 and RelB) but also of the canonical NK- κ B pathway (increased nuclear P65 and P50). Finally, Demchenko et al. show that the overexpression of IKK β or NIK in one MMCL resulted in the same pattern of gene expression in one MMCL, but less overlapping in the second one.

The importance of NK- κ B pathway deregulation in MM disease is a consequence of its pivotal role in the process of generation of normal plasma cells. Activation of CD40 B cell molecules by the CD40ligand on cognate T cells and of Toll like receptor by bacterial products induce non-canonical and canonical NF- κ B pathway activation resulting in IRF4 expression, downregulation of Bcl6 B cell TF and upregulation of Blimp1 and XBP1 plasma cell TFs ⁵. The generated plasmablasts exit the lymph node and have to home to bone marrow or mucosa and find an empty niche that will provide further differentiation and survival signal. Normal plasma cells may survive for many years in the bone marrow and mucosa niches provided the right survival factors are provided. Among numerous survival factors, BAFF and APRIL are important ones since treatment of mice with a BAFF/APRIL receptor TACI fused with Fc fragment abrogate plasma cell survival in mice and partly in humans ^{6,7}.

But it is important to emphasize that a cooperation of NK- κ B pathway with other pathways should be critical to promote MMC and normal plasma cell survival. As an example, TNF α is a moderate growth factor for myeloma cell lines whereas it induces a strong activation of canonical NF- κ B pathway in MMCLs ⁸. BAFF and APRIL are frequently mentioned as activator of NF- κ B pathways, but it is worthwhile to recall that they efficiently induce MAPK and AKT phosphorylation in myeloma cell lines ⁹. Primary MMC express aberrantly the receptor for insulin growth factor type 1 in association with a poor prognosis. IGF-1 is the major growth factor for myeloma cell lines ¹⁰, but it activates MAPK and PI-3/AKT pathways, unlike NF- κ B pathway. The same holds true for IL-6 that activates JAK/STAT3, MAPK, unlike NF- κ B pathway or PI3/AKT pathway.

Strategies to block NF- κ B pathway should consider resistance mechanisms linked to the activation of these other potent pathways in MMC and the association of drugs targeting these different pathways.



References

1. Demchenko YN, Glebov OK, Zingone A, Keats JJ, Bergsagel PL, Kuehl WM. Classical and/or alternative NF-κB pathway activation in multiple myeloma. *Blood*.
2. Vallabhapurapu S, Karin M. Regulation and Function of NF-κB Transcription Factors in the Immune System. *Annual Review of Immunology*. 2009;27:693-733.
3. Annunziata CM, Davis RE, Demchenko Y, et al. Frequent Engagement of the Classical and Alternative NF-κB Pathways by Diverse Genetic Abnormalities in Multiple Myeloma. *Cancer Cell*. 2007;12:115-130.
4. Keats JJ, Fonseca R, Chesi M, et al. Promiscuous Mutations Activate the Noncanonical NF-κB Pathway in Multiple Myeloma. *Cancer Cell*. 2007;12:131-144.
5. Tarlinton D, Radbruch A, Hiepe F, Dörner T. Plasma cell differentiation and survival. *Current Opinion in Immunology* Lymphocyte Development - Edited by Klaus Rajewsky and Harald von Boehmer / Tumour Immunology - Edited by Haruo Sugiyama. 2008;20:162-169.
6. O'Connor BP, Raman VS, Erickson LD, et al. BCMA is essential for the survival of long-lived bone marrow plasma cells. *J Exp Med*. 2004;199:91-98.
7. Rossi JF, Moreaux J, Hose D, et al. Atacicept in relapsed/refractory multiple myeloma or active Waldenstrom's macroglobulinemia: a phase I study. *Br J Cancer*. 2009;101:1051-1058.

8. Jourdan M, Moreaux J, Vos JD, et al. Targeting NF-kappaB pathway with an IKK2 inhibitor induces inhibition of multiple myeloma cell growth. *Br J Haematol.* 2007;138:160-168.
9. Moreaux J, Legouffe E, Jourdan E, et al. BAFF and APRIL protect myeloma cells from apoptosis induced by interleukin 6 deprivation and dexamethasone. *Blood.* 2004;103:3148 - 3157.
10. Sprynski AC, Hose D, Caillot L, et al. The role of IGF-1 as a major growth factor for myeloma cell lines and the prognostic relevance of the expression of its receptor. *Blood.* 2009;113:4614-4626.

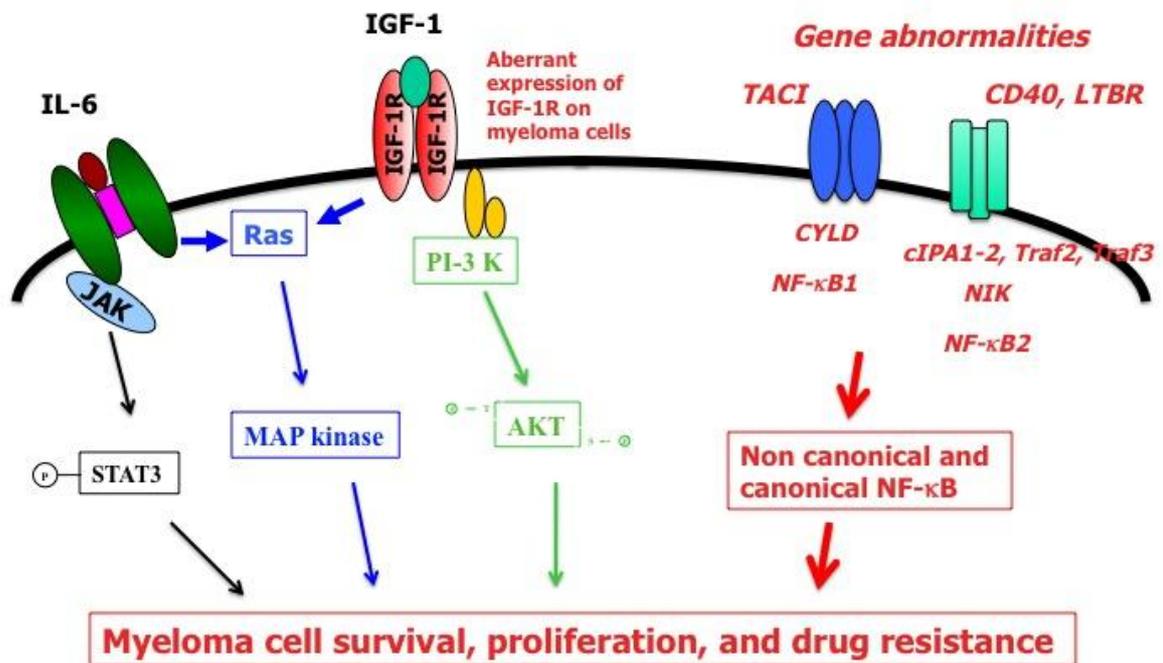


Figure : The main gene abnormalities targeting NF-κB pathway are displayed in red and italic font.

They result in aberrant activation of non-canonical and canonical NF-κB pathway that together with other signaling pathway activation (JAK/STAT, MAPkinase, PI3K/AKT) trigger myeloma cell survival, proliferation and drug resistance.