

# Intracellular Pharmacokinetics of Antiretroviral Drugs in HIV-Infected Patients, and their Correlation with Drug Action.

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# **Intracellular pharmacokinetics of antiretroviral drugs in HIV-infected patients and correlation with drug action**

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## Table of Contents

<b>ABSTRACT .....</b>	<b>5</b>
<b>1 INTRODUCTION.....</b>	<b>7</b>
<b>2 CLINICAL PHARMACOKINETICS OF ARVS.....</b>	<b>8</b>
<b>2.1 ENTRY INHIBITORS .....</b>	<b>8</b>
2.1.1 CCR5 INHIBITORS .....	8
2.1.2 ENFUVIRTIDE.....	9
<b>2.2 NUCLEOSIDE AND NUCLEOTIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE .....</b>	<b>9</b>
<b>2.3 NON NUCLEOSIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE.....</b>	<b>10</b>
<b>2.4 INTEGRASE INHIBITORS .....</b>	<b>10</b>
<b>2.5 PROTEASE INHIBITORS .....</b>	<b>10</b>
<b>3 METHODOLOGICAL CONSIDERATIONS .....</b>	<b>11</b>
<b>3.1 CELL COLLECTION .....</b>	<b>11</b>
<b>3.2 ESTIMATION OF CELL NUMBER .....</b>	<b>12</b>
<b>3.3 ANALYTICAL METHODS FOR INTRACELLULAR ASSAYS .....</b>	<b>12</b>
3.3.1 NUCLEOSIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE .....	13
3.3.2 NON NUCLEOSIDE ANALOGS AND PROTEASE INHIBITORS .....	15
<b>4 MECHANISMS INFLUENCING INTRACELLULAR ACCUMULATION .....</b>	<b>16</b>
<b>4.1 GENERAL PRINCIPLES .....</b>	<b>16</b>
<b>4.2 NUCLEOSIDE AND NUCLEOTIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE .....</b>	<b>17</b>
<b>4.3 NON NUCLEOSIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE.....</b>	<b>18</b>
<b>4.4 PROTEASE INHIBITORS .....</b>	<b>18</b>
<b>4.5 IMPORTANCE OF GENETIC POLYMORPHISM.....</b>	<b>19</b>
<b>5 CLINICAL STUDIES WITH INTRACELLULAR CONCENTRATIONS .....</b>	<b>20</b>
<b>5.1 RELATIONSHIP BETWEEN INTRACELLULAR AND PLASMA PHARMACOKINETICS .....</b>	<b>20</b>
<b>5.2 DRUG-DRUG INTERACTIONS AT THE INTRACELLULAR LEVEL .....</b>	<b>21</b>
<b>5.3 RELATIONSHIP BETWEEN INTRACELLULAR CONCENTRATIONS AND EFFICACY.....</b>	<b>26</b>
<b>5.4 RELATIONSHIP BETWEEN INTRACELLULAR CONCENTRATIONS AND TOXICITY .....</b>	<b>30</b>
<b>6 CONCLUSION .....</b>	<b>33</b>

1 **Figure captions**

2

3 **Fig. 1.** Host-cell-mediated sequential enzymatic phosphorylation steps required for  
4 activating the nucleotide- and nucleoside-analogue reverse-transcriptase inhibitors  
5 (NRTIs) to the triphosphate moiety (Reproduced from Anderson et al.<sup>[17]</sup>). ABV =  
6 abacavir; AMPD = adenosine monophosphate deaminase; AMPK = adenosine  
7 monophosphate kinase (adenylate kinase); APT = adenosine phosphotransferase;  
8 CBV = carbovir; dCK = deoxycytidine kinase; dCMPK = deoxycytidine  
9 monophosphate kinase; ddA = 2',3'-dideoxyadenosine; ddl, = didanosine; DP, =  
10 diphosphate; d4T = stavudine; FTC = emtricitabine; gK = guanylate kinase; MP  
11 monophosphate; PMPA = tenofovir (PMPA DP is a triphosphate analogue); TFV =  
12 tenofovir; TP = triphosphate; ZDV = zidovudine; 3TC = lamivudine; 5'NDPK = 5'  
13 nucleoside diphosphate kinase; 5'NT = 5' nucleotidase.

14

15 **Fig. 2.** Schematic representation of uptake and efflux transporters that may influence  
16 intracellular concentrations of antiretroviral drugs in peripheral blood cells.  
17 Transporters are named by gene and proteins (Adapted from Ford et al.<sup>[53]</sup> and  
18 updated<sup>[55-57]</sup>). OCT = Organic Cation Transporters, hCNT = Concentrative  
19 Nucleoside Transporter, ENT = Equilibrative Nucleoside Transporter, P-gp = P-  
20 glycoprotein, MRP = Multidrug Resistance Protein, BCRP = Breast Cancer  
21 Resistance protein.

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## 1 **Abstract**

2 In patients infected by human immunodeficiency virus (HIV), the efficacy of highly  
3 active antiretroviral therapy (HAART) through the blockade of different steps of this  
4 retrovirus life-cycle is now well established. As HIV is a retrovirus which replicates  
5 within the cells of the immune system, intracellular drug concentrations are important  
6 to determine antiretroviral efficacy and toxicity. Indeed, nucleoside reverse  
7 transcriptase inhibitors (NRTI), non nucleoside reverse transcriptase inhibitors  
8 (NNRTI), newly available integrase inhibitors and protease inhibitors (PI) act on  
9 intracellular targets. NRTIs are prodrugs that require intracellular anabolic  
10 phosphorylation to be converted into their active form: the triphosphorylated drug  
11 metabolite (NRTI-TP), half-life of which being longer than plasma half-life of the  
12 parent compound for most. Activity of intracellular kinases, expression of uptake  
13 transporters which may be dependent upon cell functionality or their activation state  
14 may greatly influence intracellular concentrations of NRTI-TP. In contrast, NNRTIs as  
15 well as PIs are not prodrugs and exert their activity by inhibiting directly enzyme  
16 targets. All PIs, are substrates of CYP3A, which explains most of them display poor  
17 pharmacokinetic properties with intensive pre-systemic first pass metabolism and  
18 short elimination half-lives. There are evidences that intracellular concentration of PIs  
19 depends on P-gp and/or other efflux transporters activity, which is modulated by  
20 genetic polymorphism and co-administration of drugs with inhibiting or inducing  
21 properties. Assaying adequately the intracellular concentrations of antiviral (ARVs)  
22 drugs is still a major technical challenge, together with the isolation and the counting  
23 of peripheral blood mononuclear cells (PBMCs). Furthermore, intracellular drug could  
24 be bound to cell membranes or proteins; the amount of intracellular ARV available for  
25 antiretroviral effectiveness is never measured which is a limitation of all published  
26 studies. In this review, we summarized the results of thirty-one articles that provided  
27 results of intracellular concentrations of ARVs in HIV-infected patients. Most studies  
28 also measured plasma concentrations but few of them studied the relationship  
29 between plasma and intracellular concentrations. For NRTIs, most studies could not  
30 established significant relationship between plasma and triphosphate concentrations.  
31 Only eight published studies reported an analysis of the relationships between  
32 intracellular concentrations and virological or immunological efficacy of antiretroviral  
33 drugs in HIV patients. In prospective studies well designed and with a reasonable

1 number of patients, a significant correlation between virological efficacy and  
2 intracellular concentrations of NRTIs was found with no influence of plasma  
3 concentration. For PIs, the only prospectively design trial on lopinavir found both the  
4 influence of trough plasma and intracellular concentrations. ARVs are known to  
5 produce important adverse effects through their interferences with cellular  
6 endogenous processes. The relationship between intracellular concentrations of  
7 ARVs and their related toxicity were investigated in only four articles. For zidovudine,  
8 the relative strength of the association between haemoglobin decrease and plasma  
9 zidovudine compared to intracellular zidovudine-triphosphate is still unknown.  
10 Similarly, for efavirenz and neuropsychological disorder methodological differences  
11 penalize the comparison between studies. In conclusion, intracellular concentrations  
12 of ARVs play a major role in their efficacy and toxicity and are influenced by  
13 numerous factors. However the number of published clinical studies in that area is  
14 limited; most studies were small and not always adequately designed. In addition,  
15 standardization of assays and PBMC counts are warranted. Larger and prospectively  
16 designed clinical studies are needed to further investigate the links between  
17 intracellular concentrations of ARVs and clinical endpoints.

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20  
21

# 1 Introduction

2 Human Immunodeficiency Virus (HIV) is a retrovirus, which replicates within the cells  
3 of the immune system. The efficacy of highly active antiretroviral therapy (HAART), is  
4 now well established, and has provided extraordinary benefits to many patients with  
5 HIV infection.<sup>[1]</sup> The morbidity and mortality related to HIV infection have dramatically  
6 decreased in countries in which HAART has been available, turning HIV infection into  
7 a chronic manageable disease.<sup>[2]</sup> Life-long antiretroviral treatment seems necessary,  
8 as viral replication and loss of CD4 cells resume when HAART is interrupted. HAART  
9 regimens have shown some limitations, the major one being the failure to eradicate  
10 HIV even after several years of therapy. One of the reasons is that despite potent  
11 antiretroviral (ARV) treatment, compartments of replication-competent virus persist,  
12 suggesting that ARVs do not reach all the infected cells: however, there are no data  
13 to support this theoretical assumption. This article will focus on pharmacologic  
14 principles that govern intracellular concentrations of antiretroviral drugs and on  
15 clinical studies which aimed at assessing whether intracellular concentrations of  
16 ARVs could be a useful parameter to predict efficacy or toxicity of antiretroviral drug  
17 regimen.

18 The different steps of HIV replication are now well identified and understood. A  
19 number of antiretroviral drugs are now available and are grouped in five  
20 pharmacologic classes according to their mechanism of action. These drugs target  
21 essential receptors or enzymes at different steps of the life cycle of the virus and will  
22 block the production of infectious retroviral particles from the cell.<sup>[3]</sup> However virus  
23 eradication cannot be achieved with the available treatments because of the pool of  
24 latently infected CD4 cells.<sup>[4]</sup>

25 HAART is the standard of care to avoid selection of viral mutations. Selection of  
26 drugs for treatment naïve patients and experienced patients take into account the risk  
27 benefit ratio and the viral genotype. Current guidelines recommend in  
28 treatment-naïve patients a combination of a ritonavir boosted PI or a NNRTI plus two  
29 NRTIs and in treatment experienced patients a combination of at least two active  
30 ARV drugs from different classes based on viral genotype.<sup>[5-7]</sup>

31 Besides entry inhibitors which act on receptors located on cell surface, most ARV  
32 drugs inhibit viral replication inside the cell, therefore intracellular concentration  
33 should be a reliable parameter to consider when relating pharmacokinetics and



1 efficacy. Results from several *in vitro* studies also exist in this area. But, because of  
2 the difficulties of extrapolating the results from *in vitro* to *in vivo* studies, in this paper,  
3 we focus only on *in vivo* studies.

4 This article summarizes clinical trials where intracellular concentrations were  
5 measured and related to plasma concentrations, virological efficacy or toxicity. Prior  
6 to this presentation and to understand limitations of such studies, the following topics  
7 are presented and discussed: clinical pharmacokinetics of ARVs, intracellular drug  
8 assays and mechanisms influencing intracellular diffusion and accumulation.

9

## 10 **2 Clinical pharmacokinetics of ARVs**

11 Pharmacokinetic parameters of ARVs are summarized in table I. <sup>[6, 8-13]</sup>

### 12 **2.1 Entry inhibitors**

13 Entry inhibitors block the virus attachment on receptor of the cell surface. They have  
14 an extracellular mode of action and therefore differ from other available classes of  
15 antiretroviral agents. Two drugs of this class are available, enfuvirtide and maraviroc,  
16 considering their mechanism of action, they are outside the scope of this review. To  
17 have an exhaustive overview of ARVs, their pharmacological properties are briefly  
18 summarized below.

#### 19 **2.1.1 CCR5 inhibitors**

20 CCR5 or CXCR4 chemokine co-receptor antagonists were promising entry inhibitors.  
21 Maraviroc is the first approved drug of this new class. Maraviroc inhibits CCR5  
22 chemokine co-receptor preventing HIV binding to cell membrane. Pharmacokinetic  
23 characteristics have been summarized elsewhere.<sup>[14]</sup> In brief, maraviroc is a CYP3A  
24 substrate and dosing differs according to combined drugs (150 mg bid with ritonavir  
25 boosted PI, 600 mg bid when combined with drugs with enzyme inducing properties  
26 such as efavirenz and 300 mg bid when combined with nucleoside analogs).  
27 Maraviroc is a P-gp substrate, which limits intracellular concentrations.  
28 Concentrations in cervico-vaginal fluid and vaginal tissue are higher than in plasma.

1 **2.1.2 Enfuvirtide**

2 Enfuvirtide (T20) is a HIV-1 fusion inhibitor, which prevents fusion of HIV-1 and host  
3 cell membranes. It is a synthetic peptide (4492 Da), which is not bioavailable when  
4 taken orally and is administered subcutaneously twice daily (90 mg bid) which is  
5 obviously a limitation to its long-term use. Pharmacokinetic properties have been  
6 previously reported.<sup>[15]</sup>

7 **2.2 Nucleoside and nucleotide analog inhibitors of reverse**  
8 **transcriptase**

9 Zidovudine (ZDV) is the oldest antiretroviral drug; since a number of nucleoside  
10 analogs were developed (zalcitabine, didanosine (ddI), stavudine (d4T) lamivudine  
11 (3TC), emtricitabine (FTC), abacavir (ABC)). Tenofovir (TFV) is a nucleotide analog  
12 obtained after drug administration of tenofovir disoproxil (TDF), its ester prodrug.  
13 Apricitabine is a new NRTI under development.

14 Although absolute bioavailability is unknown, bioavailability is supposed to be high for  
15 most nucleoside analogs but ddI, which is degraded at acid pH, and TDF. None of  
16 these drugs are highly protein bound. Elimination of parent compound occurs as  
17 unchanged drug via the kidney or non-CYP drug metabolizing enzymes, therefore  
18 potential for drug-drug interaction is low, although TDF was demonstrated to inhibit  
19 ddI metabolism.<sup>[16]</sup> Triphosphate (TP) metabolites are the active component of all  
20 nucleoside analogs. They also inhibit to varying degrees human mitochondrial  
21 polymerase  $\gamma$ . Phosphorylation steps occur within the cell and involved kinases,  
22 which are listed in figure 1.<sup>[17]</sup> Half-life of the active moiety is longer than plasma half-  
23 life of the parent compound for all nucleoside analogs. Long half-lives of TP  
24 metabolites favor once daily dosing for most of nucleoside analogs except ZDV and  
25 d4T, which are administered on a twice-daily basis. TFV is a nucleotide analog for  
26 which the active form is a diphosphate (DP). All NRTIs compete with endogenous  
27 analogs and stop DNA elongation. Nucleosides such as ABC (carbovir (CBV)) and  
28 TFV are much less apt to cause mitochondrial toxicity compared with d4T.<sup>[18]</sup>

29

### 1 **2.3 Non nucleoside analog inhibitors of reverse transcriptase**

2 NNRTIs do not require phosphorylation to inhibit reverse transcriptase. Nevirapine  
3 (NVP) and efavirenz (EFV) are the most commonly used. Delavirdine is available in  
4 some countries, and etravirine is a new NNRTI recently approved in the European  
5 Union and in the United States. NVP and EFV have long half-life after single dose  
6 administration. They are metabolized through CYP3A and CYP2B6 and a genetic  
7 polymorphism has been described which explains at least part of interindividual  
8 variability of their total clearance. They both have enzyme inducing and autoinducing  
9 properties, which explains drug-drug interactions and their non linear  
10 pharmacokinetics.<sup>[19]</sup>

11

### 12 **2.4 Integrase inhibitors**

13 Integrase inhibitors represent a new class. These drugs inhibit the integration of HIV-  
14 DNA into the host genome. Raltegravir was approved in early 2008 and elvitegravir is  
15 under development. Raltegravir is rapidly absorbed and plasma concentrations  
16 decline with a terminal half-life of 7 to 12 h which supports a twice daily dosing.<sup>[20]</sup>  
17 Plasma protein binding is 83%. Biotransformation pathway involved UGT1A1  
18 therefore drug-drug interactions are limited.<sup>[21]</sup> ATV which inhibits UGT1A1, increases  
19 raltegravir concentrations modestly.<sup>[22]</sup> Inducers such as EFV, TPV or rifampin  
20 decrease raltegravir concentrations although the clinical consequences are currently  
21 unclear.<sup>[21]</sup>

22

### 23 **2.5 Protease inhibitors**

24 Protease inhibitors (PI) prevent cleavage of viral precursor protein into the subunits  
25 required to form new virions. Approved PIs include amprenavir (APV), fosamprenavir,  
26 atazanavir (ATV), darunavir (DRV), indinavir (IDV), lopinavir/ritonavir (LPV/r),  
27 nelfinavir (NFV), ritonavir (RTV), saquinavir (SQV), and tipranavir (TPV).

28 They all are substrate and inhibitor of CYP3A, which explains part of their poor  
29 pharmacokinetic properties: pre-systemic first pass metabolism, variable plasma  
30 concentrations and short half-life in the 7 to 15 h range. RTV, which is the most  
31 potent CYP3A inhibitor, is combined to all PIs but NFV to improve their  
32 pharmacokinetic properties, increase plasma exposure and /or decrease the

1 administered dose.<sup>[23, 24]</sup> As basic organic chemicals they all are bound to plasma  
2 proteins,  $\alpha$ 1-glycoprotein acid and albumin. They differ for some pharmacokinetic  
3 parameters, extent of first pass metabolism, extent of protein binding (IDV 60%, LPV  
4 98-99%) and some of them such as APV have inducing properties which make drug-  
5 drug interaction prevision very difficult.

6

### 7 **3 Methodological considerations**

8 All intracellular assays described to date do not discriminate between drug localised  
9 in cell membrane or in cytoplasm, either bound to intracellular proteins or truly  
10 unbound which should be the effective antiretroviral moiety. Measurement of total cell  
11 concentrations is somehow of limited value.

#### 12 **3.1 Cell collection**

13 Isolation of PBMCs is the first step before analyzing the intracellular concentrations  
14 of either NRTIs, NNRTIs or/and PIs. PBMCs can be isolated either using  
15 conventional Ficoll gradient centrifugation or using cell preparation tubes (CPTs)  
16 (from Becton Dickinson). The two procedures were compared by Becher et al.<sup>[25]</sup> on  
17 phosphorylated anabolites of two NRTIs and were shown to give identical results.  
18 However using CPTs was found to be easier, less time-consuming and therefore  
19 quicker which in the case of d4T-TP was most important as the drug was shown  
20 instable in the cell ring of the Ficoll gradient (40% loss within 40 min) and lead the  
21 authors to collect the ring in less than 10 min. However before this isolation step the  
22 stability of the phosphorylated anabolites, that of NNRTIs and PIs in blood should be  
23 considered.

24 Regarding d4T-TP, its stability has been checked in blood before PBMCs isolation  
25 and the authors recommend to perform the isolation within 6 h after sampling.<sup>[26]</sup>  
26 Similar results were obtained with 3TC-TP and ZDV-TP.<sup>[27]</sup> It therefore seems that for  
27 the phosphorylated anabolites the storage of blood samples in CPTs before isolation  
28 could last 6 h, although it has not been thoroughly investigated for CBV-TP and TFV-  
29 DP.

30 Other issues during cell processing are to avoid contamination by red cells which  
31 may phosphorylate some nucleoside analogs<sup>[28]</sup> and efflux of PIs and NNRTIs out of  
32 cells. In contrast the NRTI-TPs are ion-trapped intracellularly. For NNRTIs and PIs

1 intracellular measurements, the collection of PBMCs has not been systematically  
2 studied. The authors mentioned that samples should be immediately taken to  
3 laboratory (within 5 min) and that all the procedures should be performed at 4°C to  
4 inhibit enzymatic activity and to prevent active drug efflux, the time between blood  
5 sampling and the cell isolation and extraction procedure should be less than 1 h.<sup>[29, 30]</sup>  
6

### 7 **3.2 Estimation of cell number**

8 Since the number of cells normalizes intracellular concentration, a critical step in the  
9 process of intracellular assay is the determination of the number of cells from  
10 which the compounds were quantified.

11 In most studies when the information is indicated the cells were determined on a  
12 small aliquot with a Coulter Counter, or using a Malassez cell and a microscope.  
13 However this last procedure may suffer from insufficient accuracy and precision,  
14 specifically when multiple sites are involved which explains that a biochemical test  
15 was developed based on the relationship between DNA content and cells count by  
16 Malassez cell.<sup>[31]</sup> This test could be performed in the analytical laboratories where  
17 there is no Coulter Counter available.

18 The concentration is therefore expressed as amounts per  $10^6$  cells and can be  
19 converted in amount per volume on the approximation that the PBMC volume is  
20 0.4 pL in order to compare intracellular and plasma concentrations.<sup>[32]</sup> The accuracy  
21 of this volume may be questionable as it varies according to the state of the cells  
22 (quiescent or stimulated) or to the nature of the cells (cell volume of human  
23 lymphoblast : 2.1 pL).<sup>[33]</sup> This highlights the pitfalls of the conversion. However the  
24 0.4 pL volume is mostly used.<sup>[29, 34]</sup> This calculation step is critical for the comparison  
25 of the results from different teams, and a standardized procedure should therefore be  
26 chosen.

### 27 **3.3 Analytical methods for intracellular assays**

28 The approaches regarding the analysis of the intracellular drugs due to the difference  
29 in their concentrations (low about fmol/ $10^6$  cells for intracellular TP anabolites and  
30 ng/ $3 \times 10^6$  cells i.e. about pmol/ $10^6$  cells for PIs) are quite different.

31

### 1 3.3.1 Nucleoside analog inhibitors of reverse transcriptase

2 The major problem in measuring intracellular TP anabolites is the small amount  
3 present in cells of patients and the presence of the endogenous intracellular  
4 nucleotides able to interfere. Thus, selective and sensitive analytical methodologies  
5 should be developed.

6 Rodriguez et al.,<sup>[35]</sup> in 2000 reviewed the latest information regarding the intracellular  
7 *in vivo* quantification of NRTI-TP. The authors described the first methods used and  
8 pointed out all their drawbacks (lack of sensitivity, cumbersome assays, inability to  
9 differentiate NRTI-TP from the endogenous nucleotides, lack of internal standard).  
10 More recent approaches were based on the same first steps, i.e. separation of ZDV-  
11 anabolites using anion-exchange cartridges, cleavage of the phosphate group using  
12 acid phosphatase, addition of an internal standard after enzymatic digestion,  
13 desalting and quantification by HPLC-MS/MS. Moreover the calibration curve was  
14 prepared from ZDV-TP contrary to the previous procedures, which used the parent  
15 compound. The limit of detection was 4.0 fmol/10<sup>6</sup> cells. The authors applied the  
16 same procedure to the simultaneous determination of ZDV-TP and 3TC-TP.<sup>[36]</sup> The  
17 limit of quantification was 0.1 pmol and 4.0 pmol for ZDV-TP and 3TC-TP,  
18 respectively. Moore et al.,<sup>[37]</sup> improved this procedure describing an analytical  
19 method which allows to measure simultaneously intracellular 3TC-TP, d4T-TP and  
20 ZDV-TP with HPLC-MS/MS. The limits of detection were 5, 25, 25 pg on column for  
21 3TC-TP, d4T-TP, ZDV-TP respectively. Similar methods were applied by King et  
22 al.,<sup>[38]</sup> to measure TFV-DP and by Robbins et al.,<sup>[39]</sup> to measure simultaneously ZDV-  
23 TP, TFV-DP and 3TC-TP in PBMC i.e. isolation by anion exchange, addition of a  
24 stable labeled isotope, dephosphorylation, desalination and detection by LC MS/MS.  
25 The lower limit of quantification were 10 fmol/10<sup>6</sup> cells for TFV-DP<sup>[38]</sup> and 0.11  
26 pmol/10<sup>6</sup> cells, 2 fmol/10<sup>6</sup> cells and 3.75 fmol/10<sup>6</sup> cells for 3TC-TP, ZDV-TP and TFV-  
27 DP respectively for a sample size of 10<sup>6</sup> cells.<sup>[39]</sup> Most of these indirect methods are  
28 quite labor-intensive, involving multiple steps, which may restrict their use to  
29 specialized laboratories.

30 New methodologies were described based on direct HPLC-MS/MS determination on  
31 the cellular extracts without dephosphorylation. However these processes need the  
32 use of ion pairing agents to circumvent the poor retention of the nucleotides which  
33 most of them are incompatible with ionisation mass spectrometry. Pruvost et al.<sup>[26]</sup>

1 described the direct determination of d4T-TP as well as that of the endogenous  
2 competitor deoxythymidine triphosphate (dT-TP). Just before cell lysis, an internal  
3 standard was added. The instrument was operated in the electrospray negative ion  
4 mode under MS/MS conditions. The limit of quantification was 9.8 fmol/10<sup>6</sup> cells i.e.  
5 20 pg injected for d4T-TP. In this article the authors focus on the stability of the d4T-  
6 TP at the different steps (in blood, in the cells ring, in dry cells at 4°C, after cell lysis  
7 at 4°C, in the injection solvent at room temperature). This procedure is very simple to  
8 perform as it does not need any extraction step. However due to the very high pH of  
9 the mobile phase (ion pairing agent: 1,5-Dimethylhexylamine) the column was  
10 changed every two weeks.<sup>[40]</sup>

11 With slight modifications regarding the internal standard and the chromatographic  
12 column, the same authors were able to measure simultaneously d4T-TP, 3TC-TP  
13 and ddA-TP (active anabolite of ddi) with their corresponding natural nucleotides in  
14 the same run.<sup>[25]</sup> However regarding ZDV-TP, a massive and tailing peak was  
15 observed near the retention time of ZDV-TP, which precludes the analysis of ZDV-TP  
16 simultaneously with d4T-TP, 3TC-TP and ddA-TP. To overcome this problem Becher  
17 et al.,<sup>[41]</sup> developed a specific extraction of ZDV-TP using immunoaffinity and  
18 detection of ZDV-TP using LC-MS/MS. More recently the same group improved the  
19 specificity and obtained a slightly better sensitivity for 3TC-TP, CBV-TP and TFV-TP  
20 using a positive electrospray ionization mode.<sup>[42]</sup>

21 Although the direct methods should be faster and more precise King et al.,<sup>[43]</sup> were  
22 unsuccessful in reproducing these methodologies. In particular they pointed out the  
23 difficulty in analyzing ZDV-TP due to the large amount of ATP and to the interference  
24 with dGTP, these latter compounds having the same precursor ion and the same  
25 product ion. This was evidenced by Compain et al.,<sup>[44]</sup> who developed an improved  
26 method to determine ZDV-TP. The authors chose a minor but specific fragment ion  
27 and had to spike their sample with a constant amount of ZDV-TP to allow the signal  
28 to emerge from background in order to increase the sensitivity.

29 HPLC-MS/MS is susceptible to matrix effects, i.e. co-eluting matrix components that  
30 affect the ionization of the target analyte, resulting in ion suppression, or, in some  
31 cases ion enhancement.<sup>[45]</sup> For intracellular assay the main parameter to study is the  
32 influence of the number of cells in the sample, as it cannot be fixed. The matrix effect  
33 plus recovery was tested by Becher et al.,<sup>[46]</sup> on d4T-TP and ddA-TP and the  
34 influence of the cell number was evidenced. The use of an appropriate internal

1 standard controlled the influence of the matrix effect between  $7$  and  $14 \times 10^6$  cells for  
2 the simultaneous assay of d4T-TP and ddA-TP. However the use of stable isotope  
3 analog as internal standard would be the best choice to control the influence of the  
4 matrix effect.

5 Monitoring the very low intracellular concentrations of these active anabolites  
6 remains an analytical challenge. All the methods described have their drawbacks.  
7 However they are all based on sophisticated methods, which can be hardly  
8 reproduced, so each laboratory favors the analytical procedure in which it is familiar.

9 Whatever the choice regarding the procedure, indirect or direct, it appears that the  
10 quantification of the compounds using HPLC coupled to MS/MS is very specific and  
11 may circumvent all the drawbacks due to the multiple natural nucleotides that are  
12 found in the complex mixture of the intracellular medium that can interfere with the  
13 determination of intracellular phosphorylated anabolites of NRTIs.

14

### 15 **3.3.2 Non nucleoside analogs and protease inhibitors**

16 The measurement of intracellular concentrations of NNRTIs and PIs could be  
17 obtained using HPLC-UV detection as was it published for EFV<sup>[34]</sup> and for 10 of them  
18 (NVP, DLV, APV, IDV, metabolite of NFV (M8), RTV, LPV, EFV, SQV, NFV)<sup>[47]</sup>.

19 However most reported data were obtained using LC-MS/MS methods either for one  
20 drug<sup>[48]</sup> or for the simultaneous measurement of several of them. The method

21 involved automated solid-phase extraction<sup>[48]</sup>, liquid-liquid extraction ((APV, LPV,  
22 SQV, EFV)<sup>[49]</sup>, NVP<sup>[30]</sup>, (LPV, RTV)<sup>[50]</sup>) or single-step extraction ((NVP, DLV, APV,  
23 IDV, M8, RNV, LPV, EFV, SQV, NFV)<sup>[47]</sup>, (IDV, APV, SQV, RTV, NFV, LPV, ATV,  
24 EFV)<sup>[29]</sup>). Few quantitative immunoassays were published for the intracellular

25 determination of LPV and ATV.<sup>[51, 52]</sup> These methods imply the preparation of a  
26 polyclonal antibody obtained with a synthetic antiretroviral drug derivative coupled to  
27 hemocyanin or serum albumin as the immunogen and the chemical synthesis of an  
28 enzyme tracer. Obviously these methods are not available in most laboratories,  
29 which preclude their use as useful tools to study the intracellular concentrations of  
30 NNRTIs and PIs.

31



## 4 Mechanisms influencing intracellular accumulation

### 4.1 General principles

As already stated, most ARVs acting on cell receptors need to enter the cell to bind to antiretroviral targets, reverse transcriptase, integrase or protease. In general, disposition from systemic circulation and capillary lumen to the extravascular compartment occurs by diffusion or involves active transporters

Simple diffusion is generally the most common mechanism for transmembrane movement of xenobiotics in the body. The rate of diffusion is defined by Fick's law and accordingly the small, lipophilic, unionised and unbound molecules readily diffuse across the membrane. Difference in the pH gradient between plasma and lymphocytes could explain ion trapping. As reported by Ford et al.<sup>[53]</sup>, the pH gradient between plasma and lymphocytes is subject to change depending on the membrane potential. Binding of drugs to plasma proteins may slow diffusion rate as only free unbound drug will cross biological membranes. However basic drugs which have higher affinity for cells or tissues proteins than for plasma proteins may leave very rapidly the blood stream and protein binding is not a limiting factor; for such drugs, volume of distribution is high, the amount of drug in plasma small compared to the amount in tissues and cells and small changes in plasma protein binding will not affect the amount in extra vascular compartments.<sup>[54]</sup> Membrane transporters (efflux, influx) are now recognized to play an important role in drug absorption and disposition and to explain, at least in part, the broad interindividual variability in intracellular concentrations of drugs. Figure 2 summarizes the different carrier proteins determining intracellular concentrations within a typical immune cell.<sup>[53, 55-57]</sup>

Efflux transporters which operate at the expense of adenosine triphosphate (ATP) hydrolysis are members of ATP-binding cassette (ABC)-type transport proteins and are now well studied. P-gp was first described for its ability to reduce intracellular concentrations of anticancer compounds. Other multidrug resistance proteins (MDR) have been isolated since. They are expressed in the apical membrane of many barrier tissues such as the intestine, liver, kidney, blood-brain-barrier, placenta, testis and in immune cells. Relevance for pharmacotherapy of expression of ABC drug transporters in peripheral blood cell have been reviewed recently.<sup>[57]</sup> The Breast Cancer Resistance Protein (BCRP) was found to play a major role in nucleoside

1 efflux.<sup>[58, 59]</sup> Although influx transporters are not as well studied, several proteins have  
2 been identified for nucleosides transport (Solute Carrier, SLC); they differ by their  
3 mechanism of action. Some are powered by electrochemical gradient (Concentrative  
4 Nucleoside Transporter hCNT), others are Equilibrative Nucleoside Transporter  
5 (ENT), Organic Cation Transporters (OCT) or Organic Anion transporters (OAT),  
6 although this later was not found to be expressed in immune cells.<sup>[53, 60]</sup>  
7 Several factors may influence transporters expression within cell membrane, cell  
8 subsets and functionality, activation state of cells and polymorphism of coding genes.  
9 Polymorphism in the coding region of the transporter genes has been evidenced  
10 which lead to produce functional changes in the encoded transporter protein and  
11 result in variation in drug disposition and response, however the studies are  
12 scanty.<sup>[55]</sup> Several factors which could affect intracellular concentrations of ARVs  
13 have to be taken into consideration: drug affinity and expression of the transporters  
14 according to different cells or tissues, and many of these transporters are known to  
15 be modulated by co-administrated ARVs.

16

## 17 **4.2 Nucleoside and nucleotide analog inhibitors of reverse** 18 **transcriptase**

19 Data on nucleoside analogs (purine or pyrimidine base coupled to a sugar) cellular  
20 penetration are scarce. As they are more hydrophilic compounds, it was suggested  
21 that they could be substrates of the endogenous nucleoside transporters.<sup>[61, 62]</sup>  
22 Although studies demonstrated that cerebral penetration occurs mainly by passive  
23 diffusion and that the low concentration of nucleoside in brain is the consequence of  
24 active efflux transporters<sup>[63]</sup>, expression of uptake transporters in lymphocytes could  
25 favor high intracellular concentrations. It was evidenced that ZDV-TP and 3TC-TP  
26 concentrations were effluxed by MRP4 and BCRP. TFV as a nucleotide has an  
27 ionized phosphate group, which confers acidic properties. It was demonstrated that  
28 TFV uptake in the kidney proximal tubule basolateral membrane is mediated via  
29 OAT1 and cellular efflux into the urine via MRP2 and MRP4.<sup>[64]</sup> TP concentrations  
30 differ according to cell type most likely as a consequence of influx and efflux  
31 transporters expression. In healthy volunteers, 3TC-TP concentrations were close in  
32 PBMCs and purified CD4 cells, whereas ZDV-TP concentrations were lower in CD4  
33 cells than in PBMCs.<sup>[65]</sup> Concentrations of TFV-DP were compared in PBMC, lymph

1 node tissue and digestive lymphatic tissue and were higher in PBMCs than in other  
2 tissues.<sup>[66]</sup> These data strongly suggest that transporters localisation may differ  
3 according to cell functionality. Kinase activity could also influence the intracellular  
4 concentration of TP. *In vitro* experiments suggested this activity varies greatly and is  
5 lower in resting cells than in activated PBMC.<sup>[67, 68]</sup> This could have important  
6 consequences as kinases activity will govern the intracellular level of both  
7 endogenous triphosphates and NRTI-TP which compete at the level of HIV-reverse  
8 transcriptase. All NRTIs have been demonstrated to be more effective in monocyte  
9 derived macrophages which are important HIV1 reservoirs than in CD4+T  
10 lymphocytes.<sup>[69]</sup> These could also well explain the differences in NRTI-TP intracellular  
11 concentrations according to different cell types and different activation state.<sup>[28]</sup>  
12

### 13 **4.3 Non nucleoside analog inhibitors of reverse transcriptase**

14 NNRTIs are weakly acidic and predominantly bind to albumin. Neither EFV nor NVP  
15 were thought to be substrate of P-gp.<sup>[70]</sup> In a limited number of patients, Almond et  
16 al.,<sup>[34]</sup> demonstrated a relationship between intracellular concentration of EFV and  
17 % bound EFV in plasma. Such data are in contrast with those obtained with NVP by  
18 the same team.<sup>[30]</sup> They demonstrated that intracellular concentrations of NVP are far  
19 below those measured in plasma. Intracellular concentration was negatively related  
20 to P-gp expression, but not related to plasma % unbound NVP.<sup>[30]</sup> To explain these  
21 data, the authors suggest that NVP could induce P-gp or co-regulated efflux  
22 transporter. Clearly, to understand all mechanisms, which are involved in intracellular  
23 concentrations of NNRTIs, further studies are needed.  
24

### 25 **4.4 Protease inhibitors**

26 The intracellular pharmacology of PIs has been carefully reviewed by Ford et al.<sup>[53]</sup>  
27 The PI physio-chemical properties are in favour of passive transfer:  
28 - The transfer is in agreement with lipophilicity measured by the n-octanol to water  
29 partition coefficient. Accumulation of PIs in lymphocytes reflects the rank order of  
30 lipophilicity: the less lipophilic PI being IDV and the most lipophilic NFV.  
31 - PIs are weak bases and are mostly unionized in a basic environment; intracellular  
32 sequestration is dependant upon pH gradient between plasma and cells.

1 - Protein binding of PIs to  $\alpha$ 1-acid glycoprotein ranged from 60% for IDV to 97-99%  
2 for RTV, LPV, SQV and NFV. However protein binding per se is not a limiting factor  
3 to intracellular diffusion as IDV, which is 60% bound, has lower intracellular  
4 concentrations than other PIs more highly bound. Within the cell PIs are bound to  
5 cell proteins and HIV proteases and therefore their relative affinity for each protein  
6 may influence their dynamic equilibrium.<sup>[53]</sup>  
7 However active transport may play a role in the intracellular accumulation. It is now  
8 well established that PIs are substrates of P-gp and others efflux transporters such  
9 as MRP1<sup>[71]</sup> or MRP2<sup>[72]</sup>. P-gp is expressed in the gastro-intestinal tract and the liver  
10 and act with CYP3A to reduce their bioavailability. RTV combined to most PIs as a  
11 pharmacologic enhancer inhibits both CYP3A and P-gp and markedly increases the  
12 bioavailability of PIs. Such transporters are expressed on lymphocytes and may  
13 reduce cellular accumulation. Meaden et al.,<sup>[73]</sup> found a relationship between  
14 combined expression of P-gp and MRP1 on PBMCs of HIV-infected patients and  
15 intracellular accumulation of SQV and RTV. In summary, there are evidences that  
16 intracellular concentration of PIs depends on P-gp and/or other efflux transporters  
17 activity, which is modulated by genetic polymorphism and coadministration of drugs  
18 with inhibiting or inducing properties. How these transporters will control intracellular  
19 concentrations of PIs need further studies.  
20

#### 21 **4.5 Importance of genetic polymorphism**

22 At evidence, the role of transporters and their genetic polymorphism in drug  
23 disposition should be considered<sup>[74]</sup> and reviews have summarized findings from  
24 recent pharmacogenetics studies.<sup>[55, 75]</sup>

25 La Porte et al.,<sup>[76]</sup> studied the relationship between ABCB1 (MDR1) genetic  
26 polymorphism, P-gp expression and SQV or SQV/r pharmacokinetics in 150 healthy  
27 volunteers. No relationship was found between the C3435T, G2677T/A or C1236T  
28 polymorphisms of the ABCB1 gene and the pharmacokinetics of SQV or the  
29 expression and activity of P-gp in PBMCs. Seventy one HIV-infected children treated  
30 with a NFV backbone antiretroviral drug regimen were evaluated for MDR1  
31 polymorphism (MDR1 C3435T), NFV plasma concentrations, CD4 cell count and  
32 HIV-RNA.<sup>[77]</sup> Children with the C/T genotypes had higher 8h post dose NFV  
33 concentrations and more rapid response to HAART. Unfortunately, intracellular

1 concentrations of PIs were not measured in these studies. In contrast, in 12 HIV-  
2 infected patients, Ford et al.,<sup>[78]</sup> could not evidenced higher intracellular  
3 concentrations of NFV or its M8 metabolite and lymphocyte cell surface expression of  
4 P-gp. In a cohort of 47 patients treated with PI boosted or not by RTV, Chaillou et  
5 al.,<sup>[79]</sup> demonstrated that intracellular concentration of RTV was related to  
6 undetectable plasma HIV-RNA, which was not related to MDR1 gene expression.  
7 Interestingly the importance of MRP4 carrier was evidenced by Anderson et al.,<sup>[72]</sup>,  
8 as they demonstrated that patients carrying MRP4 T4131G had elevated 3TC-TP  
9 concentrations and patients with MRP4 G3724A had a trend for elevated ZDV-TP.  
10 They also found that IDV clearance was faster in patients expressing CYP3A5 and in  
11 patients carrying the MRP2-24C/T variant<sup>[72]</sup>; whether this latter may contribute to  
12 lower intracellular concentrations remains to be established.  
13 Recently, Kiser et al.,<sup>[80]</sup> demonstrated that intracellular concentrations of TFV-DP  
14 were higher, first with decrease in kidney glomerular filtration rate and consequently  
15 total and renal clearance of TFV (p=0.04) and second in presence of the ABCC4  
16 3463 A>G variant (p=0.04 after adjustment for race, treatment group and glomerular  
17 filtration rate). The authors pointed out the limitation of this small sample size study  
18 for genetic association and thus results should be confirmed in larger study. Whether  
19 those exploratory data could be extrapolated to intracellular concentration of TFV-DP  
20 within renal proximal tubule cells is presently unknown.  
21 At evidence, the control of intracellular concentrations of ARVs is complex and  
22 dependant on many factors and more work is needed in this area taking into account  
23 the differences in cell biology.

24

## 25 **5 Clinical studies with intracellular concentrations**

### 26 ***5.1 Relationship between intracellular and plasma*** 27 ***pharmacokinetics***

28 Clinical studies performed in HIV-infected patients reporting intracellular  
29 concentrations are summarized in table II.<sup>[9-13, 30, 34, 72, 78, 79, 81-101]</sup> They are displayed  
30 by antiretroviral class, then within an antiretroviral class they are listed by molecule  
31 with respect to the date of Health Authority approval. For each molecule, the more  
32 recent studies are presented first. All those studies were published after 2000, except

1 for ZDV for which intracellular concentrations have been studied since 1994. Most  
2 studies reported both plasma and intracellular concentrations but only few of them  
3 studied the relationship between them.

4 It can be seen from table II that plasma pharmacokinetic parameters are rather  
5 similar across studies but some differences are observed for intracellular parameters.  
6 For NRTIs, most studies did not establish significant relationship between plasma  
7 and TP concentrations. In contrast for NNRTIs and PIs results are more conflicting,  
8 some studies evidencing correlation, while others could not. These results support  
9 the use of plasma concentration of NNRTIs or PIs but not NRTIs to monitor antiviral  
10 efficacy.

11 Such results are not surprising knowing first that PBMCs collection, preparation and  
12 quantification are not an easy task (see section 3) and second that many factors  
13 influence intracellular drug penetration and among them genetic polymorphism of  
14 influx and efflux carriers (see section 4). Moreover, these intracellular studies have  
15 been carried out in relatively few patients and larger studies would be needed to  
16 address consistently the relationship between plasma and intracellular  
17 concentrations of ARVs.

18 It is also important to note that there are potential methodological problems when  
19 studying relationship between concentrations observed at single time points, as it is  
20 done in a number of studies. Indeed, plasma and intracellular half-life are very  
21 different. It is more adequate to assess the relationship through pharmacokinetic  
22 parameters such as AUC. Surprisingly, most studies reported concentrations at some  
23 time points or PK parameters obtained by non-compartmental analysis. Population  
24 approaches were never used to analyse intracellular concentrations and their link  
25 with plasma concentrations, although this approach seems more appropriate as it  
26 allows to analyze sparse measurements.

## 27 **5.2 Drug-drug interactions at the intracellular level**

28 On a theoretical point of view, changes in the intracellular concentration of  
29 antiretroviral drugs can be secondary to modifications of (i) plasma concentration of  
30 the drug and/or the prodrug, (ii) activity of the enzymes responsible for drug  
31 anabolism/metabolism at the cellular level, (iii) activity of membrane transporters  
32 involved in cellular uptake or efflux. Since the intracellular amount of the active drug

1 is responsible for treatment efficacy, interactions leading to changes in intracellular  
2 concentrations are a relevant issue regarding the virologic outcome.

3 The clinical impact of these interactions was first evidenced by the poor efficacy of  
4 therapies combining ZDV with d4T.<sup>[102]</sup> Though the likely mechanism of this result,  
5 competitive inhibition of d4T phosphorylation by ZDV, was assessed only by *in vitro*  
6 experiments, this phenomenon highlighted the necessity to investigate the possible  
7 alteration in the intracellular concentrations of ARVs due to drug association.<sup>[103, 104]</sup>

8 Potential interactions involving NRTIs at the intracellular level were therefore  
9 investigated in several studies. Hawkins et al.,<sup>[82]</sup> evaluated whether the high rate of  
10 virological failure observed in patients receiving a triple NRTIs combination including  
11 TDF could be explained by modifications in the intracellular anabolism of these  
12 compounds.

13 So, intracellular levels of TFV-DP, CBV-TP and 3TC-TP were measured in 15 HIV-  
14 infected patients receiving a triple NRTI combination (TDF-ABC-3TC or TDF-ABC-  
15 d4T), before and after replacement of TDF or ABC by a NNRTI or a PI. No  
16 modification in the intracellular concentrations of the active anabolites of the  
17 remaining NRTIs was observed, which suggested the lack of significant interaction  
18 between the investigated drugs. Another recent study confirmed these results on 27  
19 patients.<sup>[105]</sup> Taken together, these results suggest the clinical failure that was  
20 observed with the triple NRTI (ABC/TDF/3TC) regimen was not due to drug  
21 interactions but was more likely the consequence of lack of intrinsic power.<sup>[106]</sup> This  
22 latter study also evidenced a significant 50% increase in the intracellular  
23 concentration of TFV-DP when TFV was combined to LPV/r. However, this result  
24 could simply be the intracellular reflection of the systemic interaction between these  
25 two drugs.<sup>[107]</sup> This study found no significant difference in the intracellular  
26 concentrations of CBV-TP and 3TC-TP with respect to LPV/r use, despite a 46%  
27 decrease in ABC plasma concentration in the LPV/r group. Last, nevirapine was also  
28 found not to significantly modify the intracellular concentrations of TFV-TP, CBV-TP,  
29 and 3TC-TP.

30 Hoggard et al.,<sup>[108]</sup> investigated whether prior exposure to ZDV could subsequently  
31 inhibit d4T phosphorylation. The rationale for this study came from the observation  
32 that naïve patients receiving a d4T -3TC combination experienced a further one log<sub>10</sub>  
33 decrease in viral DNA compared to patients previously treated by ZDV.<sup>[109]</sup> A  
34 subsequent inhibition of d4T phosphorylation due to a down regulation of thymidine

1 kinase induced by ZDV was one of the hypotheses raised to explain this result.  
2 However, the cellular concentration of d4T-TP measured in 7 ZDV-experienced  
3 patients was not different to the concentration measured in 20 ZDV-naïve subjects.  
4 Furthermore, the ability of PBMCs to phosphorylate d4T was not different between  
5 ZDV-experienced and ZDV-naïve subjects.<sup>[108]</sup> Similarly, no influence of prior  
6 exposure to ZDV on ZDV phosphorylation was observed during a 12 months period  
7 on 23 HIV-infected patients.<sup>[83]</sup> It is therefore likely that the decrease in efficacy  
8 observed in ZDV-experienced patients was due to the acquisition of resistance  
9 mutations rather to a modification in intracellular metabolism.

10 By measuring the TP moieties of ZDV and 3TC in the PBMCs of 8 patients, Fletcher  
11 et al.,<sup>[84]</sup> found a strong correlation between the intracellular concentrations of ZDV-  
12 TP and 3TC-TP. If this result suggested the existence of interplay among the cellular  
13 anabolism and/or metabolism of these drugs, its precise mechanism and possible  
14 consequences have still not been elucidated.

15 TDF is known to increase the plasma concentration of ddl, the most likely  
16 mechanism for this interaction being the inhibition by TVF of the enzyme responsible  
17 for the hydrolysis of guanosine and adenosine analogues, the purine nucleoside  
18 phosphorylase.<sup>[16]</sup> This interaction is clinically relevant since it is responsible for  
19 adverse effects<sup>[110-113]</sup> or treatment failure<sup>[114]</sup> which may be secondary to didanosine  
20 overexposure. Because of this, this association is currently not recommended for the  
21 initiation of HAART, but is nevertheless not contraindicated for ulterior lines of  
22 treatment.<sup>[6]</sup> Pruvost et al.,<sup>[85]</sup> investigated the possible consequences of this  
23 systemic interaction on the intracellular concentrations of the active moieties.  
24 Intracellular concentrations of ddA-TP and TFV-DP were compared between 14  
25 patients receiving the ddl/TDF (250 mg/300 mg) combination and 16 patients  
26 receiving ddl (400 mg) without TDF or 14 patients receiving TDF (300 mg) without  
27 ddl. The measured concentrations were found to be comparable between the groups  
28 which validated the strategy consisting in decreasing ddl dose from 400 to 250 mg  
29 when it is combined with TDF.<sup>[115]</sup>

30 Apricitabine, a novel deoxycytidine analog currently under investigation, shares its  
31 initial phosphorylation pathway by deoxycytidine kinase with 3TC and FTC. The  
32 potential interaction between apricitabine (600 mg bid) and 3TC (300 mg qd) was  
33 evaluated in a crossover study performed on 21 healthy volunteers who received  
34 sequentially each drug separately and the combination of both. No significant



1 modification in the plasma pharmacokinetics of 3TC, or in the cellular  
2 pharmacokinetics of its active TP moiety was observed during the combination  
3 compared to the monotherapy period. However, if co-administration with 3TC had no  
4 influence on apricitabine plasma pharmacokinetics, cellular concentration of  
5 apricitabine TP dropped by 85% during the same period.<sup>[116]</sup> These findings strongly  
6 suggested that apricitabine should not be co-administered with deoxycitidine  
7 analogues.

8 Hydroxyurea is an antiproliferative drug that was shown to provide a further 0.7 log<sub>10</sub>  
9 reduction in plasma HIV RNA when combined with ddl compared to patients  
10 receiving ddl alone.<sup>[117]</sup> By measuring intracellular deoxyadenosine triphosphate (dA-  
11 TP) in 69 HIV-infected subjects, it was evidenced that patients receiving the  
12 hydroxyurea-ddl combination achieved significantly lower dA-TP concentrations than  
13 patients under ddl or hydroxyurea monotherapy, whereas no modification in the  
14 plasma pharmacokinetics of the two drugs was observed.<sup>[118]</sup> If the precise  
15 mechanism of this interaction is still unknown, the likely explanation for the  
16 enhancement of ddl's efficacy is the decrease in the intracellular dA-TP/ddA-TP ratio,  
17 which would facilitate the incorporation ddA-TP in the replicating viral DNA.

18 Similarly to hydroxyurea, mycophenolic acid, an immunosuppressive agent, is known  
19 to decrease the intracellular concentration of an endogenous nucleotide, the  
20 deoxyguanosine triphosphate (dG-TP), which could enhance the antiviral activity of  
21 abacavir by decreasing the dG-TP/CBV-TP ratio.<sup>[119]</sup> Since this ratio could not be  
22 measured to date in patients receiving the mycophenolate mofetil–ABC combination,  
23 this hypothesis still needs to be confirmed. Nevertheless, the lack of influence of  
24 mycophenolic acid on 3TC phosphorylation was suggested by the similar intracellular  
25 concentration of 3TC-TP observed in patients receiving 3TC with or without  
26 mycophenolate mofetil.<sup>[86]</sup>

27 Ribavirin is a nucleoside analogue used for the treatment of hepatitis C virus (HCV)  
28 infection. Although its mechanism of action is still not fully understood, it involves at  
29 least in part an intracellular transformation into a TP moiety.<sup>[120]</sup> Thus, because  
30 ribavirin is used in HIV/HCV coinfecting patients, its potential interactions with NRTIs  
31 were investigated in several studies.

32 Rodriguez-Torres et al.,<sup>[121]</sup> evaluated the combination of ribavirin with 3TC, d4T, or  
33 ZDV in HIV/HCV coinfecting patients. Plasma concentrations of ZDV, d4T, 3TC and  
34 intracellular concentrations of ZDV-TP, d4T-TP, 3TC-TP were measured in 31

1 patients receiving concomitant ribavirin and compared to the concentrations obtained  
2 in 25 patients receiving a placebo instead of ribavirin. No significant difference in  
3 plasma and cellular concentrations of the measured compounds was observed,  
4 suggesting ribavirin does not modify the plasma pharmacokinetics and the  
5 intracellular phosphorylation of ZDV, d4T, 3TC. The lack of interaction between  
6 ribavirin and ZDV was confirmed in another study performed on 14 HIV-infected  
7 subjects.<sup>[122]</sup>

8 It is noteworthy these results are discrepant with *in vitro* data which evidenced an  
9 inhibition of the phosphorylation of ZDV<sup>[123, 124]</sup> and d4T<sup>[104]</sup> by ribavirin. However it is  
10 still unexplained whether these discrepancies are due to a poor ability of *in vitro*  
11 models to predict *in vivo* phenomenon or to some methodological drawbacks in the  
12 *ex-vivo* quantification of intracellular TP moieties.

13 Oppositely, ribavirin was found to potentiate *in vitro* the phosphorylation of ddl via the  
14 inhibition of inosine 5-monophosphate dehydrogenase.<sup>[125]</sup> However, despite its  
15 potential virologic interest, this interaction is also characterized by a high risk of  
16 mitochondrial toxicity<sup>[126, 127]</sup>, so the ribavirin-ddl association is not recommended.

17 Concerning PIs, the influence of ATV on the plasma and intracellular  
18 pharmacokinetics of SQV and RTV was investigated in 9 HIV-infected patients who  
19 received the SQR/RTV (1600/100 mg qd) combination with and without ATV (200 mg  
20 qd). ATV was found to significantly increase both plasma and intracellular  
21 concentrations of SQV by a similar factor of approximately 4, but had no effect on  
22 RTV concentrations.<sup>[128]</sup> Interestingly, the cellular half-life of SQV was unaffected by  
23 ATV, which suggested the increase in the intracellular concentration was secondary  
24 to the increase in plasma concentration rather to the inhibition of a cellular  
25 transporter.

26 The possible modification of plasma and cellular concentrations of SQV by quercetin,  
27 a bioflavonoid displaying inhibitory properties on CYP3A4 and P-gp, was investigated  
28 on 10 healthy adults who received SQV alone (1200 mg bid) for 11 days followed by  
29 the association SQV/quercetin (1200 mg bid/500 mg bid) during the next three  
30 days.<sup>[129]</sup> If no change was observed for SQV plasma concentration, its intracellular  
31 concentration surprisingly decreased by almost 50% when combined with quercetin.  
32 However, the important intra and intersubject variability of the intracellular  
33 concentrations prevented to draw conclusions from this result.

1 The accumulation ratio, equal to the cellular concentration divided by the plasma  
2 concentration, of some PIs was found to be modified by low doses of RTV in HIV-  
3 infected subjects.<sup>[79]</sup> APV and IDV accumulation ratios in presence of 100 mg or 400  
4 mg of RTV were indeed increased three-fold and five-fold respectively. However,  
5 conflicting results were obtained in another study which found that RTV did not  
6 increase the accumulation ratio of SQV and IDV.<sup>[130]</sup>  
7 Despite these discrepancies, assessing the consequences of drug interactions at the  
8 cellular level is of great concern in order to validate new combinations. Some recent  
9 surprising results, like the possible decrease in the efficacy of HCV therapy due to  
10 ABC<sup>[131]</sup>, evidenced the need for a better understanding of drug interactions.

### 12 **5.3 Relationship between intracellular concentrations and efficacy**

13 Only eight studies were published reporting an analysis of the relationships between  
14 intracellular concentrations and virological or immunological efficacy of antiretroviral  
15 drugs in HIV patients. Five studied intracellular NRTIs and three PIs. These articles  
16 are summarized in table III by molecule from the most recent to the oldest one.<sup>[8, 79, 84,</sup>  
17 <sup>87-89, 122, 132]</sup> Of note, five were prospective study and interestingly found a significant  
18 relationship between higher intracellular concentrations and virological response.<sup>[8, 84,</sup>  
19 <sup>87, 88, 132]</sup> The correlation with plasma concentration was not always studied but was  
20 mainly not or less significant. The three studies with non significant results were  
21 cross-sectional studies not designed for that purpose.<sup>[88, 89, 122]</sup> We start as in table II  
22 by the results on NRTIs and then on PIs, there is no such study for NNRTIs. With  
23 nucleoside analogues, it is important to notice that the relevant determinant of  
24 pharmacodynamic response is the ratio between drug triphosphate and endogenous  
25 nucleoside triphosphates rather than the absolute intracellular concentration.<sup>[133]</sup>  
26 The study by Moore et al.,<sup>[87]</sup> was a substudy of the ACTG 862, a prospective trial  
27 where naïve patients were starting a dual NRTI therapy. A significant correlation  
28 between change in viral load between week 0 and week 28 and intracellular  
29 concentrations was found for 3TC-TP ( $R^2= 0.62$ ) in the 39 patients receiving either  
30 3TC-ZDV or 3TC-d4T, and for ZDV-TP ( $R^2= 0.28$ ) in the 10 patients receiving 3TC-  
31 ZDV. No significant correlation was found for d4T-TP in the 15 patients receiving  
32 3TC-d4T and no significant relationship was found for any drug when studying  
33 change in CD4 cells. The authors did not study the relationships between efficacy

1 and plasma concentration but showed a very large interpatient variability in the  
2 intracellular to plasma concentration ratio. The authors also showed that there was  
3 an important increase of intracellular concentration between first dose and week 28  
4 only for 3TC-TP.

5 The study of Aweeka et al.,<sup>[122]</sup> was performed in HCV or HBV co-infected HIV  
6 patients and its primary objective was to study the influence of ribavirin on ZDV  
7 plasma and intracellular concentrations by measuring the AUC in patients before and  
8 after introduction of ribavirin. Different regimens were allowed and all patients had to  
9 receive ZDV for at least 4 weeks. In the cross-sectional analysis performed in 13  
10 patients before receiving ribavirin, no significant relationships was found between  
11 AUC of ZDV-TP and the CD4 cell count measured the same day.

12 The studies by Anderson et al.,<sup>[8]</sup> and Fletcher et al.,<sup>[84]</sup> are substudies of the well  
13 known concentration-controlled randomised trial by Fletcher et al.,<sup>[134]</sup>. In this trial  
14 patients received a tritherapy with ZDV, 3TC and IDV either with a fixed dose or with  
15 a dose adapted to get trough concentrations in a define range. Intracellular  
16 concentrations of ZDV-TP and 3TC-TP were measured 2 h after dose administration  
17 at the three pharmacokinetics visits scheduled at weeks 2, 26 and 52 and at variable  
18 time post dose at the nine bimontly visits. For each patient the median intracellular  
19 concentration from all measurements was considered for the analysis of the link with  
20 efficacy. Unfortunately the authors did not analyse inpatient variability of these  
21 concentrations.

22 In their first study<sup>[84]</sup>, only 8 patients were studied and the efficacy criteria were the  
23 change between baseline and week 24 HIV RNA or the percent change between  
24 baseline and week 24 CD4 cell count. For ZDV-TP a significant correlation with  
25 intracellular concentrations was found both for changes in HIV RNA ( $R^2=0.54$ ) and  
26 CD4 cells ( $R^2=0.84$ ). For 3TC-TP a significant correlation with intracellular  
27 concentrations was found for changes in HIV RNA ( $R^2=0.79$ ) but not for CD4 cells  
28 ( $P=0.07$ ,  $R^2=0.44$ ). There was no significant correlation between efficacy and steady-  
29 state plasma concentration of these drugs.

30 In the article by Anderson et al.,<sup>[8]</sup>, 33 patients were studied; the analysis was very  
31 thorough with various efficacy endpoints and with several multivariate analyses. After  
32 a first simple correlation study, the authors defined thresholds for intracellular  
33 concentrations as the first quartiles, which are of 30 fmol/ $10^6$  cells for ZDV-TP and of  
34 7017 fmol/ $10^6$  cells for 3TC-TP. They then studied the impact on several efficacy

1 endpoints of having a median intracellular concentration below or above those  
2 thresholds. The first efficacy endpoint was the time to plasma HIV RNA less than 50  
3 cp/mL, using a survival analysis. For ZDV-TP, the median time to less than 50 cp/mL  
4 is significantly reduced in patients with ZDV-TP above the threshold than in patients  
5 below as shown.<sup>[8]</sup> A significant relationship was also found for 3TC-TP but only ZDV-  
6 TP remained in the multivariate analysis. The second efficacy endpoints were the  
7 virological status (< 50 cp/mL) at week 24 and at week 52 after starting ARVs drug  
8 regimen. For ZDV-TP, 92% of patients with concentrations above threshold were  
9 undetectable at week 24, this proportion was significantly lower (44%, P=0.009) for  
10 patients below the threshold; but this relationship was not significant at week 52. For  
11 3TC-TP, 96% of patients with concentrations above threshold were undetectable at  
12 week 24, proportion that was significantly higher than for patients below the threshold  
13 (37.5 %, P=0.002); similarly at week 52, 91.7% of patients above threshold were  
14 undetectable which is significantly higher than for patients below the threshold (25%,  
15 P=0.0008). Another efficacy endpoint was the time to rebound (two HIV RNA greater  
16 than 50 cp/mL), this time was shorter for patients with ZDV-TP and 3TC-TP lower  
17 threshold; in the multivariate analysis only 3TC-TP remains significant (P=0.009). It is  
18 interesting to note that ZDV-TP was mostly associated with the initial viral load  
19 decrease whereas 3TC-TP with sustained response. No significant relationship was  
20 found between CD4 cell count and intracellular concentrations. However, the authors  
21 did not study the change in CD4 cell count but performed cross-sectional analyses  
22 looking for correlation at week 24 and week 52 between CD4 cell counts and  
23 corresponding intracellular concentrations. Link between plasma concentrations of  
24 ZDV and 3TC and efficacy was not studied and no significant relationships were  
25 found with IDV concentrations. The authors also found that intracellular  
26 concentrations were significantly higher in female than in male: 2.3 folds for ZDV-TP  
27 (P<0.0001) and 1.6 fold for 3TC-TP (P<0.0001), whereas no influence of gender was  
28 found on plasma concentrations. These results suggest NRTI phosphorylation  
29 differences between genders.

30 The first ever published article that analyzed the link between intracellular  
31 concentration and efficacy was the pharmacokinetic trial reported by Stretcher et  
32 al.,<sup>[132]</sup> where ZDV was given five times a day as a single therapy (500 mg/day) in 21  
33 patients followed during 24 weeks. AUC of total ZDV-phosphates (ZDV-P) was  
34 evaluated at week 4 and week 24 from five samples. Efficacy was analysed mainly

1 through CD4 cell count, %CD4 cells and CD4/CD8 ratio. Unfortunately, in the  
2 analysis of the correlation of ZDV-P AUC and immunological efficacy, the authors  
3 pooled the observations made at week 4 and week 24, not taking into account the  
4 correlation induced by the repetition within patients. Here, only results on the  
5 analyses performed separately at week 4 and week 24 are reported. At week 4, the  
6 authors found a significant correlation of ZDV-P AUC both with %CD4 cells change  
7 from baseline ( $R^2=0.06$ ,  $P=0.029$ ) and with CD4/CD8 change from baseline ( $R^2=0.06$ ,  
8  $P=0.028$ ) but not with the value measured at week 4. These correlations were no  
9 longer significant at week 24. No significant correlation was found with plasma AUC.  
10 With respect to studies with PIs, the main objective of the prospective trial reported  
11 by Lamotte et al.,<sup>[89]</sup> was to investigate the concept of a once daily administration of  
12 the new galenic formulation soft gel capsule of SQV in association with RTV in PI-  
13 experienced HIV patients. The evaluation of the link between SQV intracellular  
14 concentrations and virological efficacy in 13 patients was explored as one of the  
15 secondary objectives. No significant correlation was obtained between trough SQV  
16 intracellular concentration at weeks 2, 4 or 12 and variation of plasma HIV RNA  
17 between week 0 and week 12. No significant correlation was found also for plasma  
18 SQV concentrations.  
19 The main objective of the prospective study reported by Breilh et al.,<sup>[88]</sup> was the  
20 impact on virological success, defined as HIV less than 50 cp/mL, of intracellular and  
21 plasma trough concentrations of LPV in 38 patients receiving LPV/RTV based  
22 regimen. They found that trough intracellular concentrations of LPV at week 4 were  
23 significantly higher (12.7  $\mu\text{g/mL}$ ) in patients achieving virological success before  
24 week 4 than in others (4.8  $\mu\text{g/mL}$ ) ( $P<0.0002$ ). Similarly intracellular concentrations of  
25 LPV at week 24 were significantly higher (10.5  $\mu\text{g/mL}$ ) in patients achieving  
26 virological success before week 24 than in others (4.6  $\mu\text{g/mL}$ ) ( $P<0.002$ ).<sup>[88]</sup>  
27 Virological success was also significantly associated with higher plasma trough  
28 concentration at week 4 ( $P=10^{-5}$ ) and 24 ( $P=0.05$ ) and with the genotype inhibitory  
29 quotient at week 4 ( $P=10^{-6}$ ) and 24 ( $P=0.0004$ ). In a multivariate analysis of  
30 virological success at week 24, the authors found the effect of baseline LPV  
31 mutations, plasma concentration at week 4 and intracellular concentration at week  
32 24. The authors defined thresholds of 4  $\mu\text{g/mL}$  and 8  $\mu\text{g/mL}$  for plasma and  
33 intracellular concentration, respectively; they suggested combining plasma and

1 intracellular concentration of LPV for therapeutic drug monitoring. The authors  
2 derived cellular accumulation ratio but it was not used in the analysis of link with  
3 efficacy.

4 The cross-sectional study by Chaillou et al.,<sup>[79]</sup> included 49 patients with antiretroviral  
5 combinations containing various protease inhibitors. The first objective was to study  
6 the relationship between MDR-1 gene expression and intracellular PI concentrations  
7 and then to evaluate the correlation of PI intracellular concentrations with virological  
8 response. Efficacy was defined as undetectable HIV RNA load (<40 cp/mL) at day of  
9 study. As various PIs were analyzed, to normalize concentrations authors studied the  
10 influence of the ratio of intracellular to plasma concentrations that they defined as  
11 accumulation but which is not a measure showing the amount of drug in body. They  
12 did not find any significant correlation on the main PI. The only parameter  
13 significantly linked with efficacy was the intracellular presence of RTV (P=0.04). For  
14 the 19 patients receiving RTV as a booster, patients with undetectable HIV viral load  
15 had significantly higher RTV intracellular accumulation than patients with detectable  
16 HIV RNA (P=0.029).

17 In conclusion, in prospective studies well designed and with a reasonable number of  
18 patients, all authors found a significant correlation between virological efficacy and  
19 intracellular concentrations of NRTIs with no influence of plasma concentration. For  
20 PIs, there is only one well-designed prospective trial on LPV, which found both the  
21 influence of trough plasma and intracellular concentrations at different week after  
22 treatment initiation. From these results, it is difficult to know whether the primary  
23 association is with plasma or intracellular concentrations. These findings obtained in  
24 only 38 patients should be confirmed by other studies.

25

#### 26 **5.4 Relationship between intracellular concentrations and toxicity**

27 ARVs are known to produce important adverse effects which are the main drawback  
28 of HAART.<sup>[135]</sup> The toxicity related to ARVs is indeed an important cause of poor  
29 compliance, which is in return the main cause of treatment failure.<sup>[136]</sup> Besides, a viral  
30 rebound can be associated to the acquisition of mutation resistances by the virus,  
31 which can critically penalize the choice of the subsequent therapeutic strategy.<sup>[137-139]</sup>  
32 Most toxicities displayed by ARVs are typical of a pharmacological class excepted for  
33 NRTIs which trend to have their own toxicities. For instance, PIs are known to induce

1 digestive troubles and metabolic disorders, such as hyperlipidaemia, insulin  
2 resistance, diabetes mellitus, peripheral lipodystrophy, central adiposity<sup>[140-143]</sup>. NRTIs  
3 can be responsible for lipodystrophy<sup>[144]</sup>, neuropathy<sup>[145]</sup> (ddl, d4T), myopathy<sup>[146]</sup>,  
4 pancreatitis<sup>[147]</sup> (ddl,d4T), anaemia and neutropenia<sup>[148]</sup>,(ZDV), renal impairment and  
5 fanconi syndrome<sup>[149]</sup> (TFV), hepatic steatosis, and lactic acidosis<sup>[144]</sup>; whereas  
6 NNRTIs can provide neuropsychological disorders for EFV<sup>[150]</sup> and skin or hepatic  
7 toxicity for NVP<sup>[151-153]</sup>.

8 To date, the mechanisms leading to these toxicities are not perfectly understood, but,  
9 the main hypotheses highly suggest interferences with some cellular endogenous  
10 processes. For example, PIs could alter adipose tissue and lipid metabolism by  
11 inhibiting the heterodimeric nuclear receptor complex composed of peroxisome  
12 proliferator activated receptor  $\gamma$  and the retinoid X receptor, the cellular retinoic acid-  
13 binding protein, and the synthesis of cis-9-retinoic acid.<sup>[154]</sup> PIs could also inhibit the  
14 degradation of the sterol element-binding proteins which regulate the transcription of  
15 the LDL receptor gene.<sup>[155, 156]</sup> Diabetes mellitus induced by PIs could be secondary  
16 to the direct inhibition of GLUT4, a transporter that mediates the cellular uptake of  
17 glucose stimulated by insulin.<sup>[157]</sup> Similarly, adverse effects due to NRTIs are thought  
18 to be related to mitochondrial damages, which are a consequence of NRTIs ability to  
19 inhibit the mitochondrial DNA polymerase.<sup>[158]</sup>

20 Despite these elements, the possible relationship between intracellular concentration  
21 of ARVs and their related toxicity was to date investigated for four molecules (ZDV,  
22 3TC, TDF and EFV) only.

23 First, Stretcher et al.,<sup>[132]</sup> evaluated in 13 naïve patients if the intracellular  
24 concentration of ZDV-P in PBMCs was related to some markers of ZDV-induced  
25 toxicity. A negative correlation was found between intracellular ZDV-P and the  
26 decrease in haemoglobin from its baseline level. It is noteworthy that the authors did  
27 not find in this study a significant correlation between plasma ZDV and intracellular  
28 ZDV-P, and did not investigate the possible relationship between plasma ZDV and  
29 haemoglobin decrease. However, other studies evidenced an association between  
30 plasma ZDV and anaemia<sup>[159, 160]</sup>, so the relative strength of the association between  
31 haemoglobin decrease and plasma ZDV compared to intracellular ZDV-P is still  
32 unknown. A different result was nevertheless found in both adults and children.  
33 Indeed, in a study performed by Anderson et al.,<sup>[8]</sup> on 33 naïve adult patients  
34 receiving a ZDV-3TC-IDV regimen, no difference in the intracellular concentrations of



1 ZDV-TP and 3TC-TP was observed between the 14 patients who experienced at  
2 least a grade I biological event and the 19 patients who did not. A similar result was  
3 found in 49 neonates, as the proportion of observed haematological toxicities was not  
4 related to the intracellular concentrations of ZDV-TP and 3TC-TP.<sup>[99]</sup> However,  
5 differences between the pharmacokinetic criteria and the pharmacodynamic  
6 endpoints used (see table IV<sup>[8, 90, 99, 132]</sup>) might explain the inconsistency between  
7 these studies. There are also *in vitro* data suggesting ZDV-MP is the culprit of ZDV-  
8 related anemia<sup>[161]</sup>, so toxicity relationships with ZDV-TP may not be relevant.  
9 Among NNRTIs, in a study performed on 55 patients, Rotger et al.,<sup>[90]</sup> found a  
10 significant correlation between the intracellular concentration of EFV and the risk of  
11 mood disorders. No significant correlation was found with the risk of sleep disorders  
12 and fatigue. In contrast with other studies<sup>[162, 163]</sup>, no significant association was  
13 observed between the plasma concentration of EFV and the neuropsychological  
14 trouble that were investigated. Once again, methodological differences penalize the  
15 comparison between studies.  
16 Last, Izzedine et al.,<sup>[64]</sup> found, without measuring the intracellular concentration of  
17 TFV-DP, that the risk to develop a renal proximal tubulopathy with TDF-containing  
18 treatments was significantly associated with genetic variants in the gene coding for  
19 MRP2, a transporter involved in TVF efflux from tubular cells to the urine. Since  
20 these variants are thought to be associated with a reduced activity of the transporter,  
21 this result could indicate that an accumulation in the tubular cell due to an altered  
22 MRP2-based efflux might be responsible for TDF-induced toxicity.<sup>[64]</sup> However, for  
23 obvious reasons, TFV concentration in the tubular cells could not be investigated, so  
24 this mechanistic explanation remains speculative, as are the role of mitochondria and  
25 TFV-DP in TDF toxicity.  
26 More generally, the impossibility to investigate cellular concentration of ARVs in the  
27 tissues targeted by their toxicity is a major weakness of these studies. However, it is  
28 interesting to note the neuropsychological effect of efavirenz or the anaemia induced  
29 by ZDV are not explained by the diffusion of these compounds to the PBMCs.  
30 Significant correlations indicate that concentration in PBMCs possibly reflects the  
31 diffusion of these drugs to other tissues like central nervous system or bone marrow.  
32 Measuring the concentrations of ARVs in PBMCs could therefore be an interesting  
33 tool to predict and consequently to prevent the appearance of toxicities related to  
34 HAART in a clinical setting. Further studies are therefore warranted to validate

1 PBMCs as a reliable model for investigating the relationship between the intracellular  
2 concentration of ARVs and their toxicity.

3

## 4 **6 Conclusion**

5 In conclusion, intracellular concentrations of ARVs play a major role in their efficacy  
6 and toxicity and are influenced by numerous factors. Although measurement of  
7 intracellular concentrations needs standardisation, this review demonstrates that  
8 relationships between intracellular concentrations of ARVs and their efficacy have  
9 been evidenced. Such relationships should be interpreted with caution as intracellular  
10 concentrations reflect the total amount of drug within the cell and not the effective  
11 unbound fraction. The number of clinical studies in that area is however rather  
12 limited, most studies being small and not always adequately designed. Improved  
13 techniques measuring relevant intracellular concentrations, improved knowledge on  
14 which cells would be the best surrogate marker on antiretroviral therapy including  
15 reservoirs as well as larger and prospectively designed clinical studies are needed to  
16 further investigate the links between intracellular concentrations and clinical  
17 endpoints.

18

19

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1 **Tables**2  
3**Table I.** Summary of pharmacokinetics parameters of available antiretroviral drugs adapted from [6, 8-13]

		Bioavailability (%)	t <sub>max</sub> (h)	Protein Binding (%)	Elimination Pathway	Plasma t <sub>1/2</sub> (h)	Intracellular t <sub>1/2</sub> (h)
<b>Entry Inhibitors</b>							
Enfuvirtide	T20	70 (SC)	7	97	Peptidases -> amino acids	3 - 8	
Maraviroc		25-35%	2	76	25% renal + CYP3A	13	
<b>Nucleosidic Reverse Transcriptase Inhibitors (NRTI)</b>							
Zidovudine	ZDV	60	1	20	20% renal + 80% glucuronidation UGT2B7	1 - 1.5	7-11
Didanosine	ddl	40	1	< 5	50% renal	1-2	15 - 20
Stavudine	d4T	80	1	< 5	80% renal	1 - 1.5	7
Lamivudine	3TC	80	1	< 5	80% renal	2-3	22
Abacavir	ABC	75	1	49	< 5% renal + liver biotransformation	0,8 - 1,5	21
Tenofovir	TFV	40	2-3	< 10	80% renal	14	150-180
Emtricitabine	FTC	90	1	< 5	80% renal	9	39
<b>Non Nucleosidic Reverse Transcriptase Inhibitors (NNRTI)</b>							
Efavirenz	EFV	50	2-5	99.5	< 1% renal + CYP2B6	50	
Nevirapine	NVP	90	4	60	< 15% renal + CYP2B6+3A4	25-30	
<b>Integrase Inhibitor</b>							
Raltegravir		ND	3	83	< 5% renal + UGT1A1	9	
<b>Protease Inhibitors (PI)</b>							
Saquinavir	SQV	4-10	1-2	97	< 5% renal + CYP3A	5	
Indinavir	IDV	60	1	60	10% renal + CYP3A	1.5 - 2	2
Ritonavir	RTV	70	3	99	< 5% renal + CYP3A	3-5	
Nelfinavir	NFV	60-80	3	98	<5% renal + CYP3A	5-6	
Lopinavir/r	LPV/r	ND	5	99	< 5% renal + CYP3A	5-6	
Amprenavir	APV	30-90	2	90	< 5% renal + CYP3A	7-12	
Atazanavir	ATV	ND	2	86	< 10% renal + CYP3A	7	
Darunavir	DRV	ND	1-4	94	< 5% renal + CYP3A	10 -15	
Tipranavir	TPV	ND	3	99	< 5% renal + CYP3A	6 (single dose)	

4  
5**Abbreviations:** SC: subcutaneous administration; r = ritonavir low-dose; CYP = P450 cytochrome; UGT=UDP-glucuronosyltransferase

**Table II.** Intracellular pharmacokinetics of antiretrovirals and relationship with plasma pharmacokinetics

Drug	Dose <sup>a</sup>	Design			Plasma and intracellular pharmacokinetics				Correlation	Ref
		N	Plasma (n / v)	Intracellular (n / v)	Parameters <sup>b</sup>	Plasma	Intracellular	Ratio <sup>c</sup>		
<b>NRTI</b>										
Zidovudine (ZDV)										
8 mg / kg / day	49	1 / 2	1 / 2	C <sub>trough</sub>	0.00 – 1.29 µg/mL	16 – 385 fmol/10 <sup>6</sup> cells	NR	Yes (R <sup>2</sup> = NR)	[99]	
300 mg bid	14	8 / 1	3 / 3	AUC <sub>0-12h</sub>	1.8 – 1.7 h.µg/mL	1241 – 2172 h.fmol/10 <sup>6</sup> cells	NR	No	[81]	
300 mg bid	15	2 / 2	2 / 2	C <sub>1h</sub>	0.01 – 0.96 µg/mL	4 – 53 fmol/10 <sup>6</sup> cells	9 10 <sup>-6</sup> – 4.2 10 <sup>-3</sup>	NR	[87]	
				C <sub>4h</sub>	0.01 – 1.08 µg/mL	5.5 – 50.7 fmol/10 <sup>6</sup> cells	9 10 <sup>-6</sup> – 3.4 10 <sup>-3</sup>	NR		
300 mg bid	26	-	5 / 1	C <sub>trough</sub>	-	7.7 – 23.6 fmol/10 <sup>6</sup> cells	-	-	[91]	
600 mg qd				C <sub>trough</sub>	-	6.4 – 12.9 fmol/10 <sup>6</sup> cells	-	-		
	27			C <sub>24h</sub>	-	4.9 – 13.2 fmol/10 <sup>6</sup> cells	-	-		
200 mg tid	38	7 / 2	4 / 2	AUC <sub>0-8h</sub>					[92]	
				Women	0.82 – 1.34 h.µg/mL	240 – 670 h.fmol/10 <sup>6</sup> cells	NR	NR		
				Men	0.88 – 1.35 h.µg/mL	520 – 1150 h.fmol/10 <sup>6</sup> cells	NR	NR		
300 mg tid	33	-	2 / 3	C <sub>trough</sub> – C <sub>2h</sub>	-	35 – 64 fmol/10 <sup>6</sup> cells	-	-	[72]	
300 mg bid	23	3 / 7	3 / 7	AUC <sub>0-2h</sub>			NR	Yes (R <sup>2</sup> = NR)	[83]	
				Week 0	0.99 ± 0.71 h.µg/mL	110 ± 80 h.fmol/10 <sup>6</sup> cells				
				Week 12	4.21 ± 1.52 h.µg/mL	130 ± 110 h.fmol/10 <sup>6</sup> cells				
				Week 48	1.12 ± 0.41 h.µg/mL	130 ± 120 h.fmol/10 <sup>6</sup> cells				
200 / 300 mg bid <sup>d</sup>	8	1-2 / 12	1-2 / 12	C <sub>2h</sub> – C <sub>8h</sub>	0.18 – 0.32 µg/mL	13.8 – 96.4 fmol/10 <sup>6</sup> cells	NR	No	[84]	
				CL/F	1.16 – 2.53 L/h/kg	NR	-	-		

iv 1 / 2 mg / kg / h <sup>e</sup>	28 <sup>f</sup>	>1 / 1	>1 / 1	C <sub>delivery</sub>	0.19 – 3.66 µg/mL	11 – 127 fmol/10 <sup>6</sup> cells	NR	No	[93]
				CL/F	0.07 – 0.89 L/h/kg	NR	-	-	
100 mg qd	10	6 / 1	6 / 1	AUC <sub>0-12h</sub>	0.38 ± 0.13 h.µg/mL	420 ± 420 fmol/10 <sup>6</sup> cells	NR	No	[94]
300 mg bid				AUC <sub>0-12h</sub>	1.22 ± 0.21 h.µg/mL	610 ± 810 fmol/10 <sup>6</sup> cells			
500 mg qd	21	6 / 2	6 / 2	AUC <sub>0-8h</sub>			NR	No	[12]
				Week 4	0.71 ± 0.31 h.µg/mL	3290 ± 970 h.fmol/10 <sup>6</sup> cells			
				Week ≥24	0.79 ± 0.41 h.µg/mL	2160 ± 1090 h.fmol/10 <sup>6</sup> cells			
500 mg qd	6	6 / 1	6 / 1	AUC <sub>0-8h</sub>	0.74 ± 0.27 h.µg/mL	4200 ± 2720 h.fmol/10 <sup>6</sup> cells	NR	No	[13]
Didanosine (ddl)									
400 mg qd	16	-	4 / 1	C <sub>trough</sub> – C <sub>4h</sub>	-	3.8 – 13.3 fmol/10 <sup>6</sup> cells	-	-	[85]
400 / 250 mg bid	28	1 / 1	1 / 1	C <sub>2.5h</sub> – C <sub>28.5h</sub>	0.00 – 0.16 µg/mL	0 – 23 fmol/10 <sup>6</sup> cells	NR	No	[11]
Stavudine (d4T)									
40 mg bid	19	2 / 2	2 / 2	C <sub>1h</sub>	0.04 – 1.39 µg/mL	3 – 25 fmol/10 <sup>6</sup> cells	5 10 <sup>-6</sup> – 6.5 10 <sup>-5</sup>	NR	[87]
				C <sub>4h</sub>	0.04 – 0.73 µg/mL	3.2 – 18.5 fmol/10 <sup>6</sup> cells	7 10 <sup>-6</sup> – 2.4 10 <sup>-4</sup>	NR	
40 / 30 mg tid	28	1 / 1	1 / 1	C <sub>2.5h</sub> – C <sub>28.5h</sub>	0.04 – 0.67 µg/mL	0 – 99 fmol/10 <sup>6</sup> cells	NR	Yes (R <sup>2</sup> = 0.46)	[11]
Lamivudine (3TC)									
300 mg qd	15	4 / 1	4 / 1	AUC <sub>0-24h</sub>	3.71 – 7.14 h.µg/mL	26910 – 80810 h.fmol/10 <sup>6</sup> cells	-	-	[105]
				C <sub>max</sub>	1.66 – 2.59 µg/mL	-	-	-	
				C <sub>trough</sub>	0.055 – 0.18 µg/mL	6000 – 11460 fmol/10 <sup>6</sup> cells	-	-	
4 mg / kg / day	49	1 / 2	1 / 2	C <sub>trough</sub>	0.00 – 1.16 µg/mL	570 – 38900 fmol/10 <sup>6</sup> cells	NR	Yes (R <sup>2</sup> = NR)	[99]
150 mg bid	14	8 / 1	3 / 3	AUC <sub>0-12h</sub>	4.3 – 6.2 h.µg/mL	78.6 – 164.9 h.fmol/10 <sup>6</sup> cells	NR	NR	[81]
150 mg bid	41	2 / 2	2 / 2	C <sub>1h</sub>	0.06 – 1.42 µg/mL	42 – 4579 fmol/10 <sup>6</sup> cells	0.01 – 1.17	NR	[87]
				C <sub>4h</sub>	0.02 – 1.81 µg/mL	200 – 10730 fmol/10 <sup>6</sup> cells	0.06 – 41.58	NR	



300 mg qd	25	-	4 / 1	$C_{\text{trough}}$	-	$0.7 \cdot 10^6 - 2.4 \cdot 10^6 \text{ fmol}/10^6 \text{ cells}$	-	-	[91]
				$C_{24\text{h}}$	-	$1.0 \cdot 10^6 - 2.5 \cdot 10^6 \text{ fmol}/10^6 \text{ cells}$	-	-	
150 mg bid	27			$C_{\text{trough}}$	-	$0.7 \cdot 10^6 - 4.0 \cdot 10^6 \text{ fmol}/10^6 \text{ cells}$	-	-	
300 mg qd	13	-	1-2 / 6	$C_{\text{trough}}$	-	7060 – 11600 fmol/10 <sup>6</sup> cells	-	-	[82]
150 mg bid	32	-	2 / 3	$C_{\text{trough}} - C_{2\text{h}}$	-	7252 – 9313 fmol/10 <sup>6</sup> cells	-	-	[72]
150 mg bid <sup>e</sup>	8	1-2 / 12	1-2 / 12	$C_{2\text{h}} - C_{8\text{h}}$	0.43 – 0.69 µg/mL	2352 – 13024 fmol/10 <sup>6</sup> cells	NR	Yes	[84]
				CL/F	0.30 – 0.53 µg/mL	NR	-	(R <sup>2</sup> = 0.66)	
Abacavir (ABC)									
300 mg bid	12	4 / 1	4 / 1	$AUC_{0-24\text{h}}$	2.25 – 7.80 h.µg/mL	656 – 2234 h.fmol/10 <sup>6</sup> cells	-	-	[105]
				$C_{\text{max}}$	0.95 – 3.72 µg/mL	-	-	-	
				$C_{\text{trough}}$	0.03 – 0.18 µg/mL	98.3 – 472.8 fmol/10 <sup>6</sup> cells	-	-	
600 mg qd	8	-	1-2 / 6	$C_{\text{trough}}$	-	88 – 200 fmol/10 <sup>6</sup> cells	-	-	[82]
300 mg bid	9	8 / 1	4 / 1	$C_{\text{trough}}$	NQ	NQ	-	-	[86]
				$C_{\text{max}}$	1.48 – 2.35 µg/mL	-	-	-	
600 mg qd	5	-	8 / 1	$C_{\text{trough}}$	-	127 – 575 fmol/10 <sup>6</sup> cells	-	-	[101]
				$C_{1\text{h}}$	-	107 – 670 fmol/10 <sup>6</sup> cells	-	-	
				$C_{12\text{h}}$	-	188 fmol/10 <sup>6</sup> cells	-	-	
				$C_{14\text{h}} - C_{16\text{h}}$	-	101 – 548 fmol/10 <sup>6</sup> cells	-	-	
				$C_{18\text{h}}$	-	113 – 648 fmol/10 <sup>6</sup> cells	-	-	
				$C_{20\text{h}}$	-	80 – 660 fmol/10 <sup>6</sup> cells	-	-	
				$C_{22\text{h}}$	-	105 – 201 fmol/10 <sup>6</sup> cells	-	-	
				$C_{24\text{h}}$	-	62 – 354 fmol/10 <sup>6</sup> cells	-	-	
Tenofovir (TDF)									
300 mg qd	27	4 / 1	4 / 1	$AUC_{0-24\text{h}}$	0.45 – 1.34 h.µg/mL	476.4 – 1386 h.fmol/10 <sup>6</sup> cells	-	-	[105]

				$C_{max}$	0.19 – 0.45 µg/mL	-	-		
				$C_{trough}$	0.03 – 0.119 µg/mL	116.5 – 376.5 fmol/10 <sup>6</sup> cells	-		
300 mg qd	7	-	1-2 / 6	$C_{trough}$	-	85 – 110 fmol/10 <sup>6</sup> cells	-	-	[82]
300 mg qd	8	-	4 / 1	$C_{trough} - C_{4h}$	-	129 – 373 fmol/10 <sup>6</sup> cells	-	-	[85]
300 mg qd	22	7 / 1	3 / 1	$AUC_{0-24h}$	0.002 – 0.003 h.µg/mL	NR	NR	No	[95]
				$C_{1h}$	NR	71 – 130 fmol/10 <sup>6</sup> cells	-	-	
				$C_{4h}$	NR	68 – 130 fmol/10 <sup>6</sup> cells	-	-	
				$C_{24h}$	0.052 – 0.068 h.µg/mL	70 – 123 fmol/10 <sup>6</sup> cells	NR	NR	

#### Emtricitabine (FTC)

25 mg bid	9	8 / 1	5-2 / 2	$C_{1h}$	NR	0 – 1125 fmol/10 <sup>6</sup> cells	-	-	[10,
				$C_{4h}$	NR	0 – 1450 fmol/10 <sup>6</sup> cells	-	-	100]
200 mg qd	8			$C_{1h}$	NR	0 – 2125 fmol/10 <sup>6</sup> cells	-	-	
				$C_{4h}$	NR	0 – 2250 fmol/10 <sup>6</sup> cells	-	-	
100 mg bid	8			$C_{1h}$	NR	0 – 4250 fmol/10 <sup>6</sup> cells	-	-	
				$C_{4h}$	NR	0 – 4625 fmol/10 <sup>6</sup> cells	-	-	
100 mg qd	8			$C_{1h}$	NR	0 – 2675 fmol/10 <sup>6</sup> cells	-	-	
				$C_{4h}$	NR	0 – 4000 fmol/10 <sup>6</sup> cells	-	-	
200 mg bid	8			$C_{1h}$	NR	0 – 4000 fmol/10 <sup>6</sup> cells	-	-	
				$C_{4h}$	NR	0 – 4100 fmol/10 <sup>6</sup> cells	-	-	

#### NNRTI

##### Efavirenz (EFV)

600 mg qd	49	1 / 2	1 / 2	$C_{12h}$					[96]
				Week 4	1.6 – 3.1 µg/mL	2.8 – 11.5 µg/mL	2.2 – 5.4	No	
				Week 24	1.4 – 2.5 µg/mL	3.9 – 8.8 µg/mL	0.5 – 1.8	No	

600 mg qd	10	5 / 1	5 / 1	AUC <sub>0-24h</sub>	36.8 – 131.9 h.µg/mL	29.8 – 176.9 h.fmol/10 <sup>6</sup> cells	0.7 – 3.3	Yes (R <sup>2</sup> = 0.59)	[34]
600 mg qd	55	1 / 1	1 / 1	AUC	7.9 – 63.1 h.µg/mL	6.3 – 794.3 h.µg/mL	NR	Yes (R <sup>2</sup> = 0.24)	[90]
<b>Nevirapine (NVP)</b>									
200 mg bid	10	5 / 1	5 / 1	AUC <sub>0-12h</sub>	0.05 – 0.09 h.µg/mL	0.05 – 2.9 h.µg/mL	0 – 0.05	Yes (R <sup>2</sup> = 0.62)	[30]
400 mg bid	10	1 / 1	1 / 1	AUC	39.8 – 398.1 h.µg/mL	0.82 – 46.0 h.µg/mL	NR	NR	[90]
<b>PI</b>									
<b>Saquinavir (SQV)</b>									
1600 mg qd (+100 mg RTV)	12	4 / 1	4 / 1	AUC <sub>0-24h</sub>	5.7 – 39.3 h.µg/mL	24.7 – 114.6 h.µg/mL	1.5 – 6.7	Yes (R <sup>2</sup> = 0.63)	[97]
1600 mg qd (+100 mg RTV )	13	1 / 3	1 / 3	C <sub>24h</sub>			1.1 – 8.7	Yes (R <sup>2</sup> = 0.31)	[89]
				Week 2	0.04 – 1.43 µg/mL	0.15 – 0.79 µg/mL			
				Week 4	0.04 – 1.97 µg/mL	0.11 – 0.84 µg/mL			
600 mg tid (+100 mg RTV bid)	9	2 / 1	2 / 1	C <sub>trough</sub>	NR	NR	0 – 50	NR	[79]
				C <sub>max</sub>	NR	NR	0 – 20	NR	
<b>Indinavir (IDV)</b>									
800 mg tid	10	6 / 1	6 / 1	AUC <sub>0-8h</sub>	25.1 ± 4.2 h.µg/mL	7.6 ± 1.0 h.µg/mL	NR	No	[9]
800 mg bid (+100 mg RTV)	19	2 / 1	2 / 1	C <sub>trough</sub>	10.7 ± 1.3 µg/mL	3.2 ± 0.7 µg/mL	NR		[79]
				C <sub>trough</sub>	NR	NR	0 – 20	NR	
				C <sub>max</sub>	NR	NR	1 – 5	NR	
400 mg bid (+400 mg RTV)				C <sub>trough</sub>	NR	NR	2.5 – 75	NR	
				C <sub>max</sub>	NR	NR	2.5 – 13	NR	
800 mg tid				C <sub>trough</sub>	NR	NR	0 – 0	NR	
				C <sub>max</sub>	NR	NR	0 – 7.5	NR	

Ritonavir (RTV)										
100 mg bid	11	14 / 1	4 / 1	AUC <sub>0-12h</sub>	2.7 – 4.1 h.µg/mL	9.1 – 14.2 h.µg/mL	3.2 – 7.7	NR	[98]	
100 mg qd	12	4 / 1	4 / 1	AUC <sub>0-24h</sub>	1.5 – 14.6 h.µg/mL	3.2 – 13.7 h.µg/mL	0.8 – 4.2	No	[97]	
400 / 100 mg bid	22	2 / 1	2 / 1	C <sub>trough</sub>	NR	NR	0 – 3.1	NR	[79]	
Nelfinavir (NVF)										
1250 mg bid	12	5 / 1	5 / 1	AUC <sub>0-12h</sub>	5.6 – 50.8 h.µg/mL	5.1 – 60.8 h.µg/mL	NR	Yes (R <sup>2</sup> = 0.45)	[78]	
750 mg tid	12	2 / 1	2 / 1	C <sub>trough</sub>	NR	NR	NR	Yes	[79]	
				C <sub>trough</sub>	NR	NR	0 – 37.5	NR		
				C <sub>max</sub>	NR	NR	0 – 11.3	NR		
Lopinavir (LPV)										
400 / 533 mg bid (+100 / 133 mg RTV)	38	1 / 2	1 / 2	C <sub>trough</sub>					[88]	
				Week 4	0.7 – 5.7 µg/mL	2.9 – 29.0 µg/mL	1.9 – 3.8	Yes (R <sup>2</sup> = 0.72)		
400 mg bid (+100 mg RTV)	11	14 / 1	4 / 1	AUC <sub>0-12h</sub>	61.8 – 82.8 h.µg/mL	63.1 – 113.8 h.µg/mL	0.7 – 2.1	NR	[98]	

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<sup>a</sup> All studies are performed at steady state

<sup>b</sup> Values of the pharmacokinetics parameters as published in the original article: range, IQR or mean ± SD

<sup>c</sup> Ratio defined by intracellular/plasma concentration

<sup>d</sup> Individualized regimen after the second visit

<sup>e</sup> Followed by a continuous infusion of 1 mg/kg/h until delivery

<sup>f</sup> Pregnant women

**Abbreviations:** AUC = area under the concentration time curve; bid = twice a day; C<sub>max</sub> = maximum concentration; CL/F = clearance; C<sub>trough</sub> = trough concentration; NNRTI = Non nucleoside analog inhibitors of reverse transcriptase; N = number of patients; n / v = number of samples per visit / number of visits; NRTI = Nucleoside

1 and nucleotide analog inhibitors of reverse transcriptase; NR = not reported in the article; NQ = not quantifiable; PI = protease inhibitors; qd = once a day; tid = three  
2 times a day.  
3

1 **Table III.** Relationships between intracellular concentrations and efficacy of antiretroviral agents in patients

2

Study	Primary objective (yes/no)	Intracellular moieties	Dosage regimen	Patients	Studied parameters from intracellular concentrations	Efficacy criterion	Results <sup>a</sup>
Type of trial							
<b>NRTI</b>							
Moore et al. <sup>[87]</sup>	Yes Substudy of clinical trial	3TC-TP	3TC 150 mg + ZDV 300 mg bid or 3TC 150 mg + d4T 40 mg bid	39 naïve	W28: Average of 1h and 4h conc. post dose	Change between W0 and W24 in: (1) Log Plasma HIV RNA (2) CD4 cells	(1) P < 0.02 (2) NS
		ZDV-TP	ZDV 300 mg + 3TC 150 mg bid	10 naïve			(1) P < 0.02 (2) NS
		d4T-TP	d4T 40 mg + 3TC 150 mg bid	15 naïve			(1) NS (2) NS
Aweeka et al. <sup>[122]</sup>	No Cross-sectional analysis	ZDV -TP	Any regimen containing ZDV	13 HCV or HBV co-infected patients with stable regimen > 4 weeks	AUC (NCA) from 5 samples: pre dosing and 1,4 ,6 & 8h post dosing	CD4 cell count at time of pharmacokinetic sampling	NS
Anderson et al. <sup>[8]</sup>	Yes Substudy of clinical trial	ZDV-TP	ZDV 300 mg bid + 3TC 150 mg bid + IDV 800 mg tid or Concentration-Controlled ZDV-3TC-IDV regimen	33 naïve	Median conc. above threshold (yes/no) from samples 2h post dose at W2, 28 and 56 and at 2 to 8 h post dose at 9 visits from W8 to W80	(1) Time to reach less than 50cp/mL of HIV RNA (2) Undetectable HIV RNA (< 50 cp/mL) at W24 and W52 (3) Time to loss of virological response in patients achieving undetectable (4) CD4 level at W24 and W48	(1) P = 0.01 (2) W24: P = 0.009 W52: NS (3) P = 0.02 (4) NS
		3TC-TP					Thresholds: ZDV-TP: 30 fmol/10 <sup>6</sup> 3TC-TP: 7017 fmol/10 <sup>6</sup>

Fletcher et al. <sup>[84]</sup>	Yes Substudy of clinical trial	ZDV-TP  3TC-TP	ZDV 300 mg bid + 3TC 150 mg bid + IDV 800 mg tid or Concentration-Controlled ZDV-3TC-IDV regimen	8 naïve	Median conc. from samples 2h post dose at W2, 28 and 56 and at 2 to 8 h post dose at 9 visits from W8 to W80	Change between W0 and W24 in (1) log HIV RNA (2) CD4 cell	(1) P = 0.03 (2) P = 0.001  (1) P = 0.003 (2) NS
Stretcher et al. <sup>[132]</sup>	Yes PK trial	ZDV-P	ZDV only: 800 mg/day 100mg every 4h while awake	21 naïve of ZDV	AUC (NCA) from 6 samples: pre dosing and 1, 2, 4, 6 & 8h post dosing at W4 and >W24	Change between W0 and W4 or W0 and W24 of (1) % CD4 (2) CD4 / CD8 ratio	(1) W4: P = 0.029 W24: NS (2) W4: P = 0.028 W24: NS
<b>PI</b>							
Lamotte et al. <sup>[89]</sup>	No Secondary objective of clinical trial	SQV	SQV1600 mg + RTV 100 mg qd + 2 or 3 NRTI/NNRTI	13 naïve	C <sub>trough</sub> (24h post dose) at W2, W4 or W12	Change in Plasma HIV RNA between W0 and W12	NS for all dates of trough measurements
Breilh et al. <sup>[88]</sup>	Yes Observational study	LPV	LPV 400 mg + RTV 100 mg bid + 2 or 3 NRTI/NNRTI	38 naïve of LPV	C <sub>trough</sub> (12h post dose) at W4 and W24	Virological success: (1) HIV RNA < 50 cp/mL before W4 (2) HIV RNA < 50 cp/mL before W4 and during all follow up until W24	(1) P < 0.0002 (2) P < 0.002
Chaillou et al. <sup>[79]</sup>	No Secondary objective Cross-sectional trial	NFV, IDV, APV, SQV, RTV	NFV 750 mg tid or IDV 800 mg tid or APV 1200 mg bid or IDV 800 mg + RTV 100 mg bid or SQV 600mg + RTV 100mg bid or APV 600 mg + RTV 100 mg bid or IDV 400 mg + RTV 400 mg bid + 2 or 3 NRTI/NNRTI	49 experienced patients	Ratio of intracellular to plasma C <sub>trough</sub> and C <sub>max</sub> (1.5 to 3 h post dose) at day of study: (1) main PI (2) RTV	Undetectable HIV RNA (<40 cp/mL) at day of study	(1) NS for PI ratio (2) P = 0.04 for presence of intracellular RTV P = 0.029 with RTV ratio in 28 patients receiving RTV

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2<sup>a</sup> Significant results always associated better efficacy with higher intracellular concentrations

1 **Abbreviations:** APV = amprenavir; AUC = area under the concentration time curve; bid = twice a day;  $C_{max}$  = maximum concentration;  $C_{trough}$  = trough  
2 concentration; d4T = stavudine; d4T-TP = stavudine triphosphate; IDV = indinavir; LPV = lopinavir ; NCA= non compartmental analysis; NFV = nelfinavir;  
3 NRTI = Nucleoside and nucleotide analog inhibitors of reverse transcriptase; NNRTI = Non nucleoside analog inhibitors of reverse transcriptase; NS = non-  
4 significant ; PI = protease inhibitors; qd = once a day ; PK = pharmacokinetic; RTV = ritonavir ; SQV = saquinavir; tid = three times a day; ZDV = zidovudine;  
5 ZDV-P= total zidovudine phosphates; ZDV-TP = zidovudine triphosphate; 3TC = lamivudine ; 3TC -TP= lamivudine triphosphate; W = week.

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**Table IV.** Relationships between intracellular concentrations and toxicity of antiretroviral agents in patients

Study	Primary objective	Intracellular moieties	Dosage regimen	Patients	Studied parameters from intracellular concentrations	Toxicity criterion	Results <sup>a</sup>
<b>NRTI</b>							
Anderson et al. <sup>[8]</sup>	Yes Substudy of clinical trial	ZDV-TP  3TC-TP	ZDV 300 mg bid + 3TC 150 mg bid + IDV 800 mg tid or Concentration-Controlled ZDV-3TC-IDV regimen	33 naïve	Median conc. above threshold (yes/no) from samples 2h post dose at W2, 28 and 56 and at 2 to 8 hr post dose at 9 visits from W8 to W80  Thresholds: ZDV-TP: 30 fmol/10 <sup>6</sup> 3TC-TP: 7017 fmol/10 <sup>6</sup>	Apparition of a grade I laboratory event (hemoglobin, absolute neutrophil count, aspartate, aminotransferase/alanine)	NS
Stretcher et al. <sup>[132]</sup>	Yes PK trial	ZDV-P	ZDV only: 800 mg / day 100mg every 4 h while awake	21 naïve	AUC (NCA) from 6 samples: pre dosing and 1, 2, 4, 6 & 8h post dosing at W4 and W24	Change between W0 and W4 or W0 and W24 of (1) neutrophils (2) red blood cells (3) hemoglobin	(1) NS (2) NS (3) P = NS
Durand-Gasselinet al. <sup>[99]</sup>	Yes PK trial	ZDV-TP 3TC-TP	ZDV (8 mg/kg/day in 4 daily doses) ± 3TC(4 mg/kg/day in 2 daily doses)	49 neonates	Single point concentration (time of sampling not reported)	Proportions of the hematological toxicity grade between neonates with intracellular concentrations above or below the observed median	NS
<b>NNRTI</b>							
Rotger et al. <sup>[90]</sup>	Yes PK Trial	EFV	EFV+ZDV+3TC EFV+ABC+3TC EFV+d4T+ddl +/- PI (doses not provided)	55	Intracellular AUC obtained by Bayesian estimation (number of samples per patient and sampling times not provided)	Presence of grade I to IV of (1) sleep disorder (2) mood disorder (3) fatigue	(1) NS (2) P = 0.02 (3) NS

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<sup>a</sup> Significant results always associated increased risk of toxicity with higher intracellular concentrations

**Abbreviations:** ABC = abacavir; AUC = area under the concentration time curve; bid = two times a day; tid = three times a day; ddl = didanosine; d4T = stavudine; EFV = efavirenz; IDV = indinavir; PI = protease inhibitors; PK = pharmacokinetic; NCA = non compartmental analysis; NS = non significant; ZDV = zidovudine; ZDV-P= total zidovudine phosphates; ZDV-TP = zidovudine triphosphate; 3TC = lamivudine; 3TC -TP= lamivudine triphosphate; W = week.



Figure 1

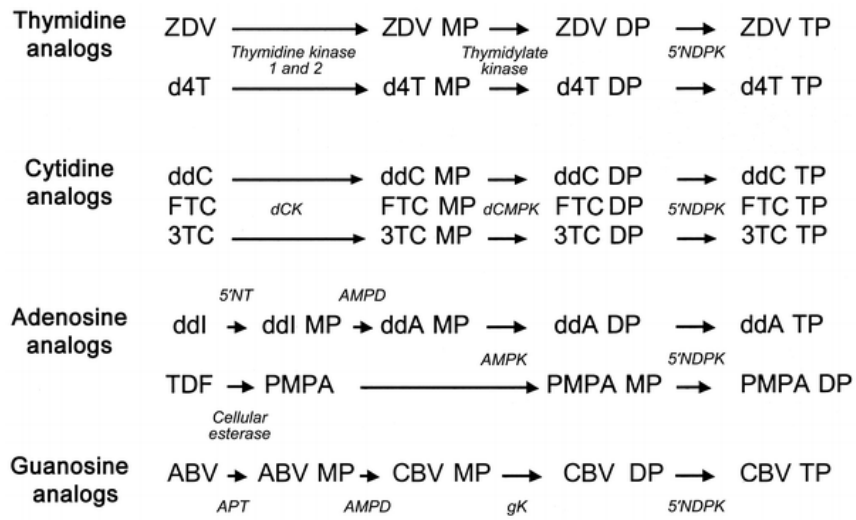


Figure 2

